

CIRCADIAN REGULATION AND NATURAL VARIATION OF LOW TEMPERATURE
SIGNALING IN *ARABIDOPSIS*

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ABSTRACT

CIRCADIAN REGULATION AND NATURAL VARIATION OF LOW TEMPERATURE SIGNALING IN *ARABIDOPSIS*

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Many plants increase in freezing tolerance in response to low non-freezing temperature through a process known as cold-acclimation. In *Arabidopsis*, cold acclimation is associated with the induction or repression of over a thousand genes. The *CBF* cold response pathway has a central role in these changes. Within minutes of transfer to low temperature, genes encoding three closely related transcription factors, *CBF1-3*, are induced and alter expression of more than one hundred target genes, which go on to impart freezing tolerance. An abiotic stress such as freezing can limit the productivity and relative fitness of a plant. Consequently, there is considerable interest in finding upstream regulators of the *CBF* pathway in hopes of expanding the geographical range and yield of important food and biofuel crops. Previous studies have shown that *CBF1-3* are subject to circadian regulation and that cold induction of *CBF1-3* is gated by the circadian clock. We identify specific clock components involved in the circadian oscillations, and thus, upstream regulation of *CBF* genes. We also demonstrate the involvement of these identified clock components in gating cold-regulated expression of *CBF1-3* and *CBF*-target-genes. Furthermore, we show that these clock components affect plant freezing tolerance.

Investigating natural variation of cold response pathways in ecotypes of *Arabidopsis* provides another means of distinguishing genes important for freezing

tolerance. In previous studies, two *Arabidopsis* populations collected from Sweden (SW) and Italy (IT) were tested for fitness (survival and seed set) in reciprocal transplant experiments. Reciprocal transplant experiments revealed that home accessions at both sites had a strong advantage in terms of seed-set and survival. SW and IT recombinant inbred lines (RILs) were used to define fitness QTL in both locations. Since there is substantial variation in temperature across latitudes, genes associated with freezing tolerance may potentially underlie identified fitness QTL. This study shows that there are differences in the cold-acclimated freezing tolerance of SW and IT ecotypes under laboratory conditions. Through RNA-seq experiments, a set of genes that may contribute to differences in freezing tolerance between SW and IT is defined. RILs are used to map expression QTL (eQTL) for a subset of these low-temperature associated genes. Some of the eQTL mapped for these low-temperature associated genes overlap with previously identified fitness QTL and this study offers hypotheses as to the genes underlying these eQTL.

This dissertation is dedicated to all of my family, but particularly to my parents, Jim and Gail, and my new parents John and Sara for their encouragement, to my undergraduate advisor J. Gary Tallman for his role in sparking my enthusiasm for scientific research, and especially to my husband Jackson Gehan for his support and the tent-cot.

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PREFACE

In Chapter Two, *cca1-1*, and the restored *cca1-1 CCA1p:CCA1-GFP* lines were procured by Eva Farre and generously donated by Jose Pruneda-Paz of the Kay Lab (San Diego, California). Other T-DNA mutant lines, used in Chapters Two and Four, were obtained from the Arabidopsis Biological Resource Center unless otherwise specified. In Chapter Three, the Schemske and Agren labs conducted reciprocal transplant experiments with the Swedish (SW) and Italian (IT) ecotypes and provided data for fitness QTL. Recombinant inbred lines (RILs) were generated and genotyped by members of the Schemske Lab. Avery Mendelsen and Nicolas Batora aided in sterilization, planting and transplanting of RILs used in eQTL analysis. SW and IT versions of *CBF2* were sequenced by Chin-Mei Lee. Alignment and assembly of SW and IT RNA-seq data to the Columbia-0 (COL-0) genome was performed by Pingsha Hu.

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KEY TO ABBREVIATIONS

ABA	ABSCISIC ACID
ACT	ACTIN
AP2	APETALA2
BOA	BROTHER OF LUX ARRHYTHMO
BP	BASE PAIR
CAMTA	CALMODULIN-BINDING TRANSCRIPTIONAL ACTIVATOR
CBF	C-REPEAT BINDING FACTOR
CBS	CCA1 BINDING SITE
CCA1	CIRCADIAN CLOCK ASSOCIATED 1
CHE	CCA1 HIKING EXPEDITION
CHIP	CHROMATIN IMMUNOPRECIPITATION
COR	COLD-RESPONSIVE GENE
COS	COLD-STANDARD GENE
CRT	C-REPEAT
CVI	CAPE VERDE ISLANDS
DAVID	DATABASE FOR ANNOTATION VISUALIZATION AND INTEGRATED DISCOVERY
DGDG	DI-GALACTOSYLDIACYLGLYCEROL
DRE	DROUGHT RESPONSE ELEMENT
DREB	DEHYDRATION RESPONSIVE ELEMENT BINDING FACTOR
DUF	DOMAIN OF UNKNOWN FUNCTION
EE	EVENING ELEMENT
ELF	EARLY FLOWERING
EQTL	EXPRESSION QUANTITATIVE TRAIT LOCI
FDR	FALSE DISCOVERY RATE
FTD	FREEZING TOLERANCE DIFFERENCE
GI	GIGANTEA
GO	GENE ONTOLOGY
ICE1	INDUCER OF CBF EXPRESSION1
ICER	INDUCER OF CBF EXPRESSION REGION
IT	ITALIAN ECOTYPE
LEA	LATE EMBRYOGENESIS ABUNDANT
LER	LANSBERG ERRECTA
LHCB	LIGH HARVESTING COMPLEX
LHY	LATE ELONGATED HYPOCOTYL
LOD	LOGARITHM OF ODDS
LUX	LUX ARRHYTHMO
MGDG	MONO-GALACTOSYLDIACYLGLYCEROL
NLS	NUCLEAR LOCALIZATION SIGNAL

P5CS	Δ -PYRROLINE-5-CARBOXYLATE SYNTHASE
PAG	PHOTOSYNTHESIS ASSOCIATED GENE
PBS	PUTATIVE BOA BINDING SITE
PR	PATHOGENESIS-RELATED
PRR	PSEUDO-RESPONSE REGULATOR
QTL	QUANTITATIVE TRAIT LOCI
RILS	RECOMBINANT INBRED LINES
RVE	REVEILLE
SFR2	SENSITIVE TO FREEZING 2
SUMO	SMALL UBIQUITIN-RELATED MODIFIER
SW	SWEDISH ECOTYPE
T1ME	TOC1 MORNING ELEMENT
TBS	TCP BINDING SITE
TF	TRANSCRIPTION FACTOR
TOC1	TIMING OF CAB 1
UBQ	UBIQUITIN
ZT	ZEITGEBER TIME

CHAPTER ONE
LITERATURE REVIEW

CHAPTER ONE

LITERATURE REVIEW

Cold-acclimation and freezing tolerance. The geographic range of plants is limited by a number of abiotic factors, including low temperature (1-3). Plants can vary greatly in their tolerance to temperature changes. In fact, in the deserts of Southern California plants thrive in an environment that goes from freezing to scorching in a single day, with temperature swings of approximately 50°C (4, 5). For plants growing in temperate and tropical climates the average difference between day and night temperatures is much narrower than these extreme desert conditions. However, temperate and tropical plants are still exposed to temperature fluctuations of approximately 10-15°C in a single day (5).

In addition to constraining the geographical locations that a plant can grow, low temperatures can also decrease plant productivity and cause crop loss (6). Most temperate plants, such as winter wheat, barley and *Arabidopsis*, can increase their tolerance to freezing temperature by prior exposure to low non-freezing temperatures, a process known as cold-acclimation (6-8). Many economically important crops such as commercial citrus trees (9), maize (10), tomato (11) and soybean (12) are chilling-sensitive and cannot cold-acclimate. In fact, in 2007, after 4 days of freezing temperatures, the California citrus industry suffered \$480 million dollars of damage (13). Chilling-sensitive crops cannot cold acclimate but can reduce damage by cold through prior exposure to low temperatures (8). Consequently, a better understanding of the mechanisms of cold-acclimation may also lead to enhancements in chilling tolerance (8,

14, 15).

Freezing tolerance is an agronomically important trait and considerable effort has gone into breeding plants with greater freezing tolerance (6, 16-19). Closely related species of plants can differ significantly in their ability to cold-acclimate (15). For example, the common cultivated potato, *Solanum tuberosum*, and the wild potato *Solanum commersonii*, are closely related evolutionarily (15). Both species are chilling tolerant and have similar basal freezing tolerance, but *Solanum commersonii* can cold-acclimate while *Solanum tuberosum* cannot (15). Even ecotypes within a species can vary greatly in their basal and cold-acclimated freezing tolerance (2, 3, 20-24).

Traditional breeding methods, thus far, have only achieved limited success in increasing plant freezing tolerance (19). In comparison to traditional breeding methods, transgenic plants have been much more successful in increasing freezing tolerance of important crop species (25-30). Therefore there is considerable interest in continuing to elucidate the molecular mechanisms underlying the process of cold-acclimation.

Molecular changes associated with plant cold-acclimation. At freezing temperatures ice forms on the outside of leaf surfaces and subsequently propagates to the intercellular space of plant tissue (31-33). This intercellular ice can cause physical damage to plant cells and tissue (31-33). However, the dehydration that accompanies freezing is the primary source of damage (31-33). The severe cellular dehydration that accompanies freezing, is the result of the chemical potential of the cell moving unfrozen water from the intracellular (cytoplasm) space to the intercellular space where the water

is frozen (31-33).

Under non-acclimated conditions (no prior exposure to low temperatures), dehydration as a result of freezing can also instigate two forms of membrane damage. First, is endocytotic vesiculation, which alone is not damaging (31-33). However, the surface area reduction is not reversible, and cell lysis occurs when cells regain their volume during thawing (31-33). Secondly, during severe dehydration hexagonal II phase regions can form between the plasma membrane and endomembranes, such as the chloroplast envelope (31-33). These hexagonal II phase regions can result in a loss of osmotic responsiveness (31-33). Therefore, the plant cell can never recover from freeze-induced dehydration.

Cold-acclimated (prior exposure to low, non-freezing temperatures) plant cells respond differently to cellular dehydration as a result of freezing (31-33). Rather than endocytotic vesiculation, exocytotic vesicles continuous with the plasma membrane are formed (31-33). Therefore cells are not disrupted when they regain their volume during thawing. Hexagonal II phase regions in the plasma membrane also do not form with cold-acclimation (31-33). Consequently, assessment of membrane damage after freeze/thaw is a means of quantitatively measuring plant freezing tolerance (electrolyte leakage assay) and distinguishing plants that can and cannot cold-acclimate.

It has long been observed that there are significant changes in membrane lipid composition during cold-acclimation (31, 32, 34, 35). These changes include an increase in fatty acid unsaturation and phospholipid content as well as a drastic decrease in chloroplast-specific mono-galactosyldiacylglycerol (MGDG; (35)). A recent

study by Moellering et al. 2010 found that the gene SENSITIVE TO FREEZING 2 (SFR2) encodes a galactolipid remodeling enzyme vital to this chloroplast-specific decrease in MGDG (35). MGDG is prone to the formation of hexagonal II phase lipid structures (35). As previously mentioned, these hexagonal II phase regions can result in a loss of osmotic responsiveness during freezing (31-33). SFR2 is thus hypothesized to increase plant freezing tolerance by decreasing the formation of hexagonal II phase lipids by converting MGDG to di-galactosyldiacylglycerol (DGDG) and oligogalactolipids, which are not prone to hexagonal II phase formation *in vitro* (35).

Along with changes in lipid composition, Guy et al. 1985, showed through *in vitro* translation of mRNA in non-acclimated and cold-acclimated samples, that there are many gene expression changes during the process of cold-acclimation (36). Since then, many studies have detailed the changes in gene expression that occur during the process of cold acclimation (37-45). These studies found that many of the cold-responsive genes encode hydrophilic proteins. These hydrophilic COLD-RESPONSIVE (COR) polypeptides have simple, repetitive, amino-acid composition (37-45).

COR6.6, also known as KIN2, is almost identical to KIN1, a small hydrophilic protein that is hypothesized to act as an antifreeze protein due to sequence similarity with arctic fish antifreeze proteins (37, 39, 46-48). COR6.6 is targeted to the cytoplasm and has been shown to have cryoprotective properties by reducing the amount of freeze-induced membrane fusions *in vitro* (46-48). *COR15A* is a cold-regulated gene specifically localized to the stroma of chloroplasts (38). Current data support the hypothesis that COR15A increases freezing tolerance by preventing the formation of

hexagonal II phase regions, which normally form between the plasma membrane and chloroplasts (41, 46). Fittingly, overexpression of *COR15A* leads to an increase in non-acclimated freezing tolerance of chloroplasts *in vitro* and an increase in the acclimated and non-acclimated freezing tolerance of protoplasts (41). *COR47* has sequence homology with LATE EMBRYOGENESIS ABUNDANT (LEA) proteins, which respond to ABSCISIC ACID (ABA), water stress, and drought (39). Like *COR47*, *COR78* is another hydrophilic protein that resembles LEA proteins, which responds to cold, ABA and drought (49, 50). Mutation or overexpression of *COR78* has little impact on plant freezing tolerance (51). Accordingly, the function of *COR78* and many of these *COR* proteins in freezing tolerance have yet to be entirely elucidated (51).

Transcriptional changes at low temperature are also accompanied by extensive changes in metabolic profile, which include the accumulation of certain amino acids and sugars, such as proline and sucrose (44, 52, 53). Free proline has been shown to have cryoprotective properties in plants (52) and the low-temperature induced increase in free proline has been linked to the increase in transcripts for Δ^1 -pyrroline-5-carboxylate synthase (*P5CS*), a key enzyme in the proline biosynthetic pathway (54). Fructose, glucose, and sucrose are sugars up-regulated by low-temperature and the synthesis of these sugars is associated with the raffinose pathway (44, 52, 53). These solutes accumulate in the cytoplasm, decreasing the chemical potential driving dehydration (31, 34, 53). The increase in these solutes with cold-acclimation also possibly reduces hexagonal II phase formation by increasing the distance between membranes (34). The up-regulation of the raffinose pathway by cold has been specifically linked to the gene

GALACTINOL SYNTHASE3 (GOLS3), a key enzyme in the raffinose biosynthetic pathway (55). *GOLS1* and *GOLS2* are regulated by other stresses (55). While these are two examples of genes driving metabolic changes during cold-acclimation, the genes responsible for many of the other metabolic changes remain unidentified (44).

PATHOGENESIS-RELATED (PR) proteins accumulate in the intercellular fluid (apoplasts) of plants at low-temperature and have been shown to have antifreeze activity (56-58). High concentrations of these PR antifreeze proteins in the apoplast can decrease the freezing temperature of the intercellular fluid, thus preventing ice formation (56). At lower concentrations, the growth of ice in the intercellular space is also limited by PR antifreeze proteins, which adhere onto ice and prevent large ice crystals from forming (56). This decrease in intercellular ice formation by PR proteins thus, decreases the amount of membrane damage by intercellular ice and suggests an interesting relationship between cold and biotic stresses (56).

CBF pathway of cold-acclimation. Promoter analysis of the *COR* genes identified the C-REPEAT/DEHYDRATION RESPONSE ELEMENT (CRT/DRE) as a novel and significantly overrepresented cis-element (40, 50). This element was found to be responsive to cold and drought but not ABA (40, 50). A transcriptional activator, C-REPEAT BINDING FACTOR1 (CBF1), was identified as binding to the CRT/DRE through a yeast-1-hybrid screen (59). Two closely related genes *CBF2* and *CBF3*, located in tandem to *CBF1* on chromosome 4, were subsequently identified and shown to bind to the CRT/DRE (42). *CBF1-3* all contain a NUCLEAR LOCALIZATION SIGNAL

(NLS), an APETALA2 (AP2) DNA binding domain, followed by an acidic activation domain (59).

Within minutes of exposure to cold, *CBF1-3* gene expression is rapidly induced in *Arabidopsis* (42). CBF transcription factors bind to the CRT/DRE element in the promoters of *COR* genes, which leads to activation of *COR* gene expression (42). Constitutive over-expression of *CBFs* in *Arabidopsis* results in expression of *COR* genes under warm temperatures and constitutive freezing tolerance without prior cold acclimation (45, 54, 60-62). Phenotypically, *CBF* over-expressing plants are dwarf compared to wild-type plants. *CBF1-3* also regulate many of the metabolic changes associated with cold acclimation (44, 62). In fact, both *P5CS* and *GOLS3* are CBF target genes (44, 54, 55).

Genomic analysis also revealed the existence of a closely related AP2 transcription factor on chromosome 5 named *CBF4* (63). Although closely related in terms of transcript sequence, the regulation of *CBF4* seems to have diverged significantly from *CBF1-3*, since *CBF4* appears to be specifically up-regulated by drought but not low-temperature (63). Overexpression of *CBF4* leads to the activation of genes containing CRT/DRE elements (63). Consequently, both drought and freezing tolerance are increased in *CBF4* overexpressing plants (63).

CBF pathway in important plant species. Although the CBF pathway was identified in the model plant, *Arabidopsis*, it appears to be conserved in many economically important plant species that can cold-acclimate, such as wheat, rye,

poplar and barley (64-66). Overexpression of the *CBF* genes in these plants also leads to an increase in freezing tolerance (64-66). The *CBF* genes are also present in plants that do not cold-acclimate such as rice, and tomato (67, 68). Interestingly, overexpression of *CBF* genes in rice and tomato does not lead to an increase in freezing tolerance, but the CBF regulons of these two species are significantly smaller than other species that do cold-acclimate (67, 68). *Solanum commersonii* and *Solanum tuberosum*, two evolutionarily close potato species, differ in their ability to cold acclimate and also have significant differences in the genes that comprise their CBF regulons, although the number of CBF regulated genes is similar (15). Chapter Three of this study will detail variations in the low-temperature response of two *Arabidopsis* ecotypes with differing freezing tolerance.

Natural variation of CBF genes in Arabidopsis. Previous studies on the natural variation of freezing tolerance in *Arabidopsis* have implicated the *CBFs* as a source of significant variation. Alonso-Blanco et al. 2005 mapped differences in freezing tolerance using recombinant inbred lines (RILs) from *Arabidopsis* accessions LANDSBERG ERECTA (LER) and CAPE VERDE ISLANDS (CVI) (20). Seven different quantitative trait loci (QTL) on chromosomes 1,4, and 5 were found, but the single QTL on chromosome 4 explained the most variance and also contained the *CBF* locus (20). CVI freezing tolerance was significantly reduced in comparison to LER, as was *CBF2* expression. Introduction of the Ler-*CBF2* transgene into CVI significantly reduced differences in freezing tolerance between LER and CVI (20).

CBF expression in the Versailles collection of *Arabidopsis* was also found to vary significantly (24). Although freezing tolerance did not simply correlate with *CBF* expression in the 8 accessions tested, the most freezing-sensitive Versailles accessions were also the accessions with the lowest cold-induced *CBF* expression levels (24). The most freezing tolerant Versailles accessions varied significantly in their *CBF* expression, suggesting the importance of CBF-independent pathways of freezing tolerance for some accessions (24). Freezing tolerance QTL mapped for two Versailles RIL populations were not completely overlapping (24). However, both populations had a common QTL on chromosome 4 that overlapped with the *CBF* locus and explained the most variance (24). Further analysis of *CBF* gene sequence in the 48 Versailles accessions showed a number of polymorphisms in both the promoter and coding sequences of *CBF1-3* (22).

A study by Zhen et al. 2008 further analyzed *CBF* sequence in 24 accession of *Arabidopsis* and found a large bias toward nonsynonymous changes in the activation domain compared to the DNA binding domain of *CBF* genes (23). Zhen et al. 2008 also showed that the *CBF* genes undergo relaxed purifying selection in *Arabidopsis* accession native to southern latitudes in comparison to accessions native to northern latitudes (23). The 10 southern latitude accessions tested had 1.5 to 4.6-fold more polymorphisms in the individual *CBF* genes in comparison to the 14 northern latitude accessions tested (23). The four accessions (CO-1, CVI-0, CVI-1, and ITA-0) with the lowest *CBF* expression originated from southern latitudes and had the lowest survival rates in freezing tolerance assays (23). CO-1, CVI-1, ITA-0 and two additional

southern accessions, Can-0 and L1-0, also had reduced expression of three CBF target genes (COR6.6, COR15A, COR78) in comparison to northern accessions (23).

CBF-independent pathways of cold-acclimation in Arabidopsis. Microarray analysis of *CBF* overexpressing plants revealed that only 12% of the cold-responsive transcriptome is part of the CBF regulon (69). Furthermore, 28% of cold-responsive genes are not affected by expression of the *CBF* genes (69). This indicates that there are CBF-independent pathways regulating expression of cold-induced genes.

Through an EMS mutant screen, ESKIMO1 (ESK1) was identified as a plant with constitutive freezing tolerance without prior cold acclimation (70). The *esk1* mutant has constitutively high levels of proline (70), similar to *CBF* overexpressing plants (62). However, ESK1-regulated genes do not overlap significantly with CBF regulated genes, suggesting that ESK1 is part of a CBF-independent pathway (70, 71). ESK1 encodes DOMAIN OF UNKNOWN FUNCTION 231 (DUF231) protein and it was recently shown that ESK1 is important for production of functional xylem and consequently water transport (71, 72).

Hos9 and *hos10* mutant plants were identified in a mutant screen using plants expressing the *COR78* promoter driving the expression of a luciferase reporter gene (73-75). The *hos9* and *hos10* mutants both have reduced freezing tolerance compared to wild-type plants in whole plant freeze tests (73, 74). Though the gene underlying the *hos10* mutation has yet to be found (75), the gene underlying the *hos9* mutation is a constitutively active homeodomain transcription factor (73). Microarray analysis of the

hos9 mutant reveals that genes regulated by HOS9 are significantly different from those regulated by the CBFs (73).

Upstream regulation of CBF pathway in Arabidopsis. CBF genes

significantly impact freezing tolerance in economically important species, and appear to be under substantial selective pressure (22-24). Consequently, there is considerable interest in finding upstream regulators of the *CBF* genes. In 2004, Novillo et al. isolated a *cbf2* mutant that had increased freezing tolerance and higher expression of *CBF1* and *CBF3* (76). This therefore suggests that *CBF2* negatively regulates *CBF1* and *CBF3*. However, this hypothesis was not substantiated by microarray studies (45, 61). Plants overexpressing *CBF2* did not have significantly reduced *CBF1* or *CBF3* expression (45, 61).

In 2003, Zarka et al. used 5' and 3' deletions of the *CBF2* promoter to identify a ~155 base pair (bp) fragment required for cold induction (77). Interestingly, the *Arabidopsis* accession, CVI, which was found to have reduced *CBF2* expression in comparison to LER, has a 1630 bp deletion in the *CBF2* promoter that intrudes on this important ~155 bp region (20). Zarka et al. 2003 also identified two short elements, INDUCTION OF CBF EXPRESSION REGION 1 (ICER1) and ICER2, within this ~155 bp region important for cold-regulated expression (77). Subsequent mutations performed within this ~155 bp region further identified sequences important for cold-induced *CBF* expression (78). One element found through this mutational analysis was identified as a binding site for the CALMODULIN BINDING TRANSCRIPTION

ACTIVATOR (CAMTA) family of calmodulin binding transcription factors (78). CAMTA3 was found to positively regulate *CBF1* and *CBF2*, but not *CBF3*, which does not contain a CAMTA binding site (78). Interestingly, exposure to cold temperature results in a rapid spike of cytosolic calcium (79). Although the connection between calcium and cold is still not well understood, these cold-induced increases in cytosolic calcium are also required for normal activation of CBF regulated genes (79). In plants, calcium can be perceived by calmodulin (78). Therefore, as a calmodulin binding transcription factor, CAMTA3, is a putative connection between calcium, cold and the *CBF* genes (80).

In 2003, INDUCER OF CBF EXPRESSION1 (ICE1) was identified through a mutant screen using plants expressing a luciferase reporter gene under the control of the *CBF3* promoter (81). *ICE1*, is a constitutively expressed, nuclear localized, MYC family transcription factor that positively regulates *CBF3*, but has little effect on *CBF1* or *CBF2* (81). Upon exposure to cold, *ICE1* transcripts are only slightly up-regulated (81), but ICE1 is post-transcriptionally regulated through sumoylation by SIZ1, a SMALL UBIQUITIN-RELATED MODIFIER (SUMO) E3 ligase (82). Sumoylation is a reversible post-transcriptional modification that is most notably known to affect subnuclear targeting, transcriptional regulation and ubiquitin mediated-protein degradation (82). Miura et al. 2007 demonstrated that cold-induced sumoylation of ICE1 leads to enhanced protein stability by reducing polyubiquitination, which normally leads to degradation (82).

MYB15 was identified through microarray experiments as a cold-regulated gene with higher expression in *ice1* mutants (83). This suggests that ICE1 normally represses expression of *MYB15*. Consistent with this idea, ICE1 sumoylation by SIZ1 also represses expression of *MYB15* (82). Subsequent yeast-2-hybrid and pull-down assays also suggested interaction of MYB15 with ICE1 (83). Overexpression of *MYB15* reduces expression of *CBFs* and MYB15 RNAi lines have been shown to increase *CBF* expression (83). *CBF1-3* promoters contain the known binding site for MYB15 and *in vitro* binding assays suggest that MYB15 negatively regulates *CBF* genes directly (83). But CBF target genes are not affected by overexpression or reduction of *MYB15* expression (83). Agarwal et al. 2006 explain this confounding result by suggesting that MYB15 negatively regulates *CBF* genes, but induces expression of other cold-regulated transcription factors, which up-regulate *COR* genes (83). This is a reasonable hypothesis, however, *in vivo* binding assays are needed to confirm the MYB15-CBF interaction.

ZAT12 is a transcription factor that was found to have a similar cold induction pattern to the *CBF* genes (45, 69). *ZAT12* overexpressing plants had significantly reduced *CBF* expression but only slightly reduced CBF target gene expression (45). ZAT12 is thus another potential negative regulator of the *CBF* genes. *ZAT12* overexpressing plants are dwarf as are *CBF* overexpressing plants, but are only slightly more freezing tolerant than wild-type plants (45). Microarray analysis of *ZAT12* overexpressing plants further supports the role of ZAT12 in cold signaling (45).

However, further evidence is needed to show that ZAT12 is directly involved in the negative regulation of *CBF* genes.

Upstream CBF regulation by the circadian clock. Circadian clocks give an internal estimate of time, allowing organisms to predict daily changes in their environment such as the onset of day or night, or changes between day and night temperature (84, 85). Not surprisingly, the ability to anticipate and respond to environmental changes provides an adaptive advantage for plants (84, 86, 87).

The *Arabidopsis* circadian clock is composed of several interlocking feedback loops and additional components of the *Arabidopsis* clock continue to be found each year (88, 89). Morning-expressed MYB transcription factors CIRCADIAN CLOCK ASSOCIATED 1 (*CCA1*) and LATE ELONGATED HYPOCOTYL (*LHY*) directly repress the expression of *TIMING OF CAB1* (*TOC1*) by binding to an EVENING ELEMENT (EE; AAAATATCT) in the *TOC1* promoter (90). EARLY FLOWERING 3 (*ELF3*), *ELF4*, *LUX* ARRHYTHMO (*LUX*), and *GIGANTEA* (*GI*) promote expression of *CCA1* and *LHY*, but there is no evidence that these transcription factors directly regulate *CCA1* or *LHY* (90-92). Recently, BROTHER OF *LUX* ARRHYTHMO (*BOA*) was identified as a clock component that positively regulates *CCA1* expression by binding to PUTATIVE *BOA* BINDING SITE 3 (*PBS3*) and *PBS4* elements in the *CCA1* promoter (89).

TOC1, up until very recently (93), was also widely hypothesized to promote expression of *CCA1* and *LHY* since *toc1* mutants have reduced expression of *CCA1* and *LHY* (90). However, this hypothesis was confounded for many years by the result

that *TOC1* overexpressing plants also have low expression of *CCA1* and *LHY* (94). In 2012, Gendron et al. showed that *TOC1* generally acts as a repressor, which can bind to a new cis-element *TOC1 MORNING ELEMENT* (T1ME) through its CCT domain (93). Although, Gendron et al. 2012 also demonstrated that *TOC1* represses expression of *CCA1* and *LHY*, they do not negate genetic data (90) that suggests that *TOC1* is somehow necessary for activation of *CCA1* and *LHY* (93). Although the role of *TOC1* in the circadian clock still seems unsettled there is evidence that suggests that the increase in *CCA1* and *LHY* could be partially explained by the repression of *CCA1* *HIKING EXPEDITION* (CHE) by *TOC1* (93, 95).

Pruneda-Paz et al. 2009 demonstrated that CHE represses *CCA1* through the TCP-BINDING SITE (TBS; GGTCCCAC) in the *CCA1* promoter (95). *CCA1* and *LHY* also repress *CHE* through the *CCA1*-BINDING SITE (CBS; AAAAATCT) in the *CHE* promoter (95). The *TOC1* protein binds directly to the N-terminal domain of CHE, which binds directly to TBS, and chromatin immunoprecipitation (ChIP) experiments have shown enrichment of *TOC1* at TBS elements in the *CCA1* promoter (95). *TOC1* is thus hypothesized to promote expression of *CCA1* by antagonizing the *CCA1* inhibitor, CHE (95). *PRR1* was found to be identical to *TOC1* (96) and *PRR7* and 9, were subsequently identified as components of the circadian clock that form a 'morning expressed' feedback loop with *CCA1* and *LHY* (97-102). *CCA1* and *LHY* positively regulate *PRR7*, and 9, which then directly repress *CCA1* and *LHY* expression (97-102).

In 2000, Harmer et al. demonstrated that components of key transcriptional pathways such as photosynthesis, phenylpropanoid biosynthesis, cell elongation, and

starch mobilization, are all under concerted regulation by the circadian clock (84). Harmer et al. 2000 also showed that *CBF3* expression is rhythmic under warm conditions (84). This result led Fowler et al. 2005 to examine the expression of *CBF* genes under circadian conditions (103). In 2005, Fowler et al. used northern blot analysis to demonstrate that the amplitude of *CBF1-3* cold-induction is dependent on the time of day that the plants are exposed to cold (103). This gating of cold-induced gene expression was observed under both diurnal and light-entrained circadian conditions (103). Peak *CBF* expression was at ZEITGEBER TIME 4 (4 hours after dawn; ZT4) and ZT28, and troughs of expression were at ZT16 and ZT40 (103).

Fowler et al. 2005 used *CCA1* overexpressing plants to show that gated cold-induced expression of *CBF* genes is dependent on the circadian clock (103), since overexpression of central clock component *CCA1* disrupts normal clock function (90, 104). However, the specific clock components regulating *CBF* expression are still unclear. A study by Nakamichi et al. 2009 showed that the *prr9/prr7/prr5* triple mutant disrupted gated cold-induced expression of *CBF* genes. The triple mutant had constitutively high *CBF* expression (105). Therefore, PRR9, PRR7 and PRR5, potentially act as negative regulators of *CBF* expression, though no evidence of in vivo interaction with the *CBF* genes was provided (105). In Chapter Two of this study an alternate mechanism is suggested for both the circadian regulation and gated cold-induction of *CBF* genes.

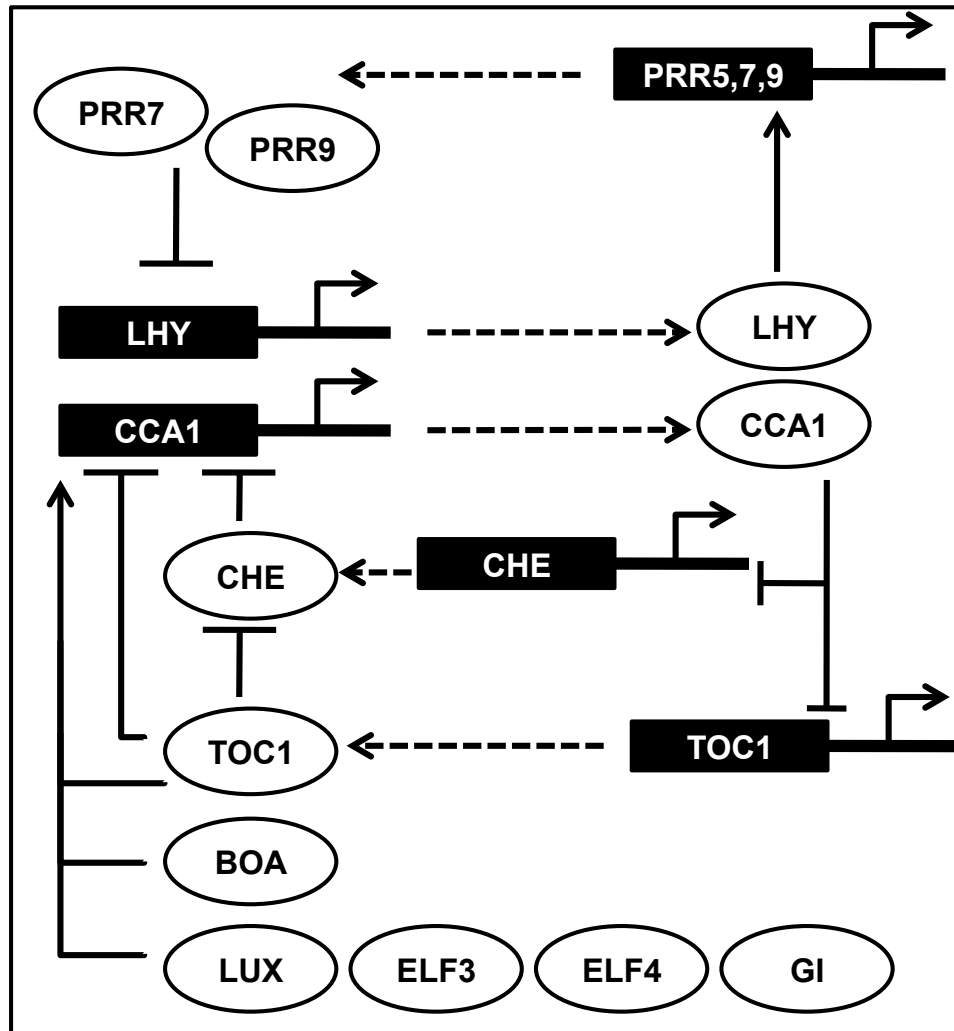


Figure 1.1. Regulation of central components of the *Arabidopsis* circadian clock based on previous literature. Ovals represent proteins, and rectangles represent a gene and promoter region. Solid lines ending in arrows (that are not attached to genes) represent positive regulation, while solid lines ending in a perpendicular line represent negative regulation. Dotted lines represent transcription and translation of a gene to a protein. For details on the regulation of central clock components please refer 'upstream CBF regulation by the circadian clock' section of Chapter One, which begins on page 15.

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CHAPTER TWO

CIRCADIAN CLOCK-ASSOCIATED 1 and LATE ELONGATED HYPOCOTYL regulate expression of the CBF cold response pathway and freezing tolerance in *Arabidopsis*

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CHAPTER TWO

CIRCADIAN CLOCK-ASSOCIATED 1 and LATE ELONGATED HYPOCOTYL regulate expression of the CBF cold response pathway and freezing tolerance in *Arabidopsis*

SUMMARY

The C-REPEAT BINDING FACTOR (CBF) cold response pathway has a prominent role in cold acclimation, the process whereby certain plants increase in freezing tolerance in response to low non-freezing temperatures. In *Arabidopsis*, the CBF pathway is characterized by rapid induction of the *C-REPEAT BINDING FACTOR 1* (*CBF1*), *CBF2* and *CBF3* genes, which encode transcriptional activators, followed by induction of the CBF-targeted genes known as the CBF regulon. Expression of the CBF regulon results in an increase in freezing tolerance. Previous studies established that *CBF1*, *CBF2* and *CBF3* are subject to circadian regulation and that their cold induction is gated by the circadian clock.

Here we present the results of genetic analysis and chromatin-immunoprecipitation experiments indicating that both of these forms of regulation involve direct positive action of two transcription factors that are core components of the clock, i.e., CIRCADIAN CLOCK ASSOCIATED 1 (CCA1) and LATE ELONGATED HYPOCOTYL (LHY). In plants carrying the *cca1-11/lhy-21* double mutation, cold induction of *CBF1*, *CBF2* and *CBF3* was greatly impaired and circadian regulation of *CBF1* and *CBF3* was essentially eliminated; circadian regulation of *CBF2* continued, though with significantly reduced amplitude. Circadian regulation and cold induction of the CBF regulon genes *COLD-REGULATED GENE15A* (*COR15A*), *COR47* and

COR78 were also greatly diminished in plants carrying the *cca1-11//hy-21* double mutation. Furthermore, the *cca1-11//hy-21* double mutation resulted in impaired freezing tolerance in both non-acclimated and cold-acclimated plants. These results indicate that CCA1/LHY-mediated output from the circadian clock contributes to plant cold tolerance through regulation of the CBF cold response pathway.

INTRODUCTION

A general feature of plants from temperate environments is that they increase in freezing tolerance in response to low non-freezing temperatures, a process called cold acclimation (1, 2). It is now well established that cold acclimation involves extensive changes in gene expression (3-6). The best understood cold regulatory pathway is the CBF pathway. This pathway, which is widely conserved in plants (7), is best characterized in *Arabidopsis* (8, 9). When *Arabidopsis* plants are transferred from warm to cold temperature, *C-REPEAT BINDING FACTOR 1* (*CBF1*), *CBF2* and *CBF3*—also known as *DROUGHT RESPONSE ELEMENT BINDING FACTOR1B* (*DREB1B*), *DREB1C* and *DREB1A*, respectively—are rapidly induced. These genes, which are physically linked in tandem array, encode transcription factors that are members of the AP2/ERF family of DNA binding proteins (10). The CBF proteins bind to the CRT/DRE regulatory element present in the promoters of about 100 cold-regulated (COR) genes, known as the CBF regulon, and induce their expression (4, 6, 11). Constitutive overexpression of *CBF1*, *CBF2* and *CBF3* at warm temperature results in constitutive expression of the CBF regulon and an increase in freezing tolerance (12-14). The mechanisms whereby expression of the CBF regulon promotes freezing tolerance are not completely understood, but involve the synthesis of low molecular weight cryoprotectants such as sucrose and raffinose and proteins that have cryoprotective properties (1, 2).

Given their importance in cold acclimation, efforts have been directed at understanding the mechanisms involved in cold-induction of *CBF1*, *CBF2* and *CBF3*. To date, two positive regulators have been identified: INDUCER OF CBF EXPRESSION

1 (*ICE1*), a MYC family transcription factor that positively regulates *CBF3* (15), and CALMODULIN BINDING TRANSCRIPTION ACTIVATOR 3 (*CAMTA3*), a CAMTA family transcription factor that positively regulates *CBF1* and *CBF2* (16). The *ICE1* and *CAMTA3* genes are transcribed at warm temperature, indicating that their activities involve posttranscriptional regulatory mechanisms that are responsive to low temperature (16-18).

Another factor that affects the expression of *CBF1*, *CBF2* and *CBF3* is the circadian clock (19-22). At warm temperature, the transcript levels for *CBF1*, *CBF2* and *CBF3* oscillate with a peak at about 8 h after dawn (ZEITGEBER TIME 8; ZT8), and a trough at about ZT20. Moreover, cold-induction of *CBF1*, *CBF2* and *CBF3* is “gated” by the clock (22); if plants are exposed to low temperature at ZT4, the increase in *CBF1*, *CBF2* and *CBF3* transcript levels is much greater than if plants are exposed to low temperature at ZT16. These results indicate that cold induction of *CBF1*, *CBF2* and *CBF3* involves the integration of low temperature and clock regulatory pathways.

The circadian clock of *Arabidopsis* consists of multiple interlocking regulatory feedback loops (23, 24). Key components of the core feedback loop are CIRCADIAN CLOCK ASSOCIATED 1 (*CCA1*) and LATE ELONGATED HYPOCOTYL (*LHY*), MYB transcription factors that have partially overlapping functions (25-28), and TIMING OF CAB 1 (*TOC1*), a PSEUDO RESPONSE REGULATOR (PRR) protein (29). Expression of *CCA1* and *LHY* peaks just after dawn, while the expression of *TOC1* peaks in the early evening. *CCA1* and *LHY* bind to the Evening Element (EE) (19) present in the promoter of *TOC1* and repress its transcription (30). *TOC1* is necessary for the

induction of both *CCA1* and *LHY* (28). TOC1 is known to inhibit the repression of *CCA1* by the TCP transcription factor CCA1 HIKING EXPEDITION 1 (CHE1) (31), but the means by which *LHY* expression is activated by TOC1 remains unknown. *CCA1* and *LHY* also regulate expression of PSEUDO RESPONSE REGULATOR 7 (PRR7) and PRR9 (32, 33), two components of the morning regulatory loop. *CCA1* and *LHY* bind to the promoters of these two genes to induce their expression and the PRR7 and PRR9 proteins then negatively regulate *CCA1* and *LHY* (32, 33).

It was recently reported that *Arabidopsis* plants carrying the *prp5/prp7/prp9* triple mutation constitutively express *CBF1*, *CBF2* and *CBF3* at high levels and display constitutively high levels of freezing tolerance (34). Thus, it was proposed that PRR5, PRR7 and PRR9 might act as direct negative regulators of *CBF1*, *CBF2* and *CBF3* (34). Here we present results indicating that the clock also provides positive regulation of the CBF cold response pathway and enhances freezing tolerance through action of the core clock components *CCA1* and *LHY*.

RESULTS

CCA1 and LHY have a direct role in circadian regulation of CBF1, CBF2 and CBF3. Consistent with previous reports (19-21), we found that *CBF1*, *CBF2* and *CBF3* are subject to circadian regulation (Fig. 2.1). Transcript levels for *CBF1*, *CBF2* and *CBF3* oscillated with a peak occurring at about ZT8 followed by a second peak about 24 h later. The oscillation patterns for the three *CBF* genes were similar, though we observed one consistent difference; whereas the ZT8 peak for *CBF1* was lower than the second peak, the ZT8 peaks for *CBF2* and *CBF3* were greater than the second peak. Thus, the transition from dark to light may have a specific effect on the regulation of *CBF1*.

Three lines of evidence led us to think that *CCA1* and *LHY* might drive circadian regulation of *CBF1*, *CBF2* and *CBF3*. First, the protein levels for *CCA1* and *LHY* peak in the early morning (ZT1-3) (26, 35, 36), just prior to when the transcript levels of *CBF1*, *CBF2*, and *CBF3* begin to increase. Second, the transcript levels for *PRR7* (32, 36) and *LIGHT HARVESTING COMPLEX B (LHCB)* (24, 35, 37), both of which are induced by *CCA1* and *LHY*, peak similarly to the *CBF* genes. Lastly, the promoter regions of *CBF1*, *CBF2* and *CBF3* have several EE (AAAATATCT) (19) and *CCA1*-Binding Sites (CBS; AATCT) (35) (Fig. 2.2), which mediate binding of *CCA1* and *LHY* (30, 35, 38) to target promoters. To determine whether *CCA1* and *LHY* were involved in circadian regulation of *CBF1*, *CBF2* and *CBF3*, we asked whether their expression was affected in plants carrying either the single *cca1-11* or *lhy-21* null mutations or the *cca1-11/lhy-21* double mutation. We found that the single mutations had differing effects on

the three *CBF* genes; whereas cycling of *CBF1* was severely disrupted, *CBF2* and *CBF3* transcript levels cycled with approximately the same amplitudes as in the wild-type plants, though the peaks were about 2 h earlier than in the wild-type plants (Fig. A2.1). Period shortening of output genes such as *LHCB* has been previously observed in *cca1* and *lhy* mutant plants (39, 40), which we also observed in our experiments (Fig. A2.2). The *cca1-11/lhy-21* double mutations also had differing effects on the circadian regulation of the three *CBF* genes (Fig. 2.1); whereas circadian regulation of *CBF1* and *CBF3* was essentially eliminated in the double mutant plants, *CBF2* transcript levels clearly continued to cycle, though the amplitude was diminished and the period was greatly shortened. Period shortening of output genes such as *LHCB* has also been previously observed in *cca1-11/lhy-21* double mutant plants (40, 41) and in our experiments (Fig. A2.3). From these results we concluded that circadian regulation of *CBF1* and *CBF3* is dependent on the action of either CCA1 or LHY and that circadian regulation of *CBF2* involves action of CCA1 and LHY, but can be driven to a considerable degree by other unknown factors.

CCA1 and LHY may impart circadian regulation of *CBF1*, *CBF2* and *CBF3* by binding to the EE and CBS motifs present in the *CBF* promoters and act as positive regulators stimulating transcription. To test this hypothesis, we conducted chromatin immunoprecipitation (ChIP) experiments to determine whether CCA1 binds directly to the promoter regions of *CBF1*, *CBF2* and *CBF3*. This was assessed by comparing ChIP results obtained with plants carrying the *cca1-1* mutation and *cca1-1* plants that had been restored with a construct encoding the CCA1 protein tagged with green

fluorescent protein (GFP) under the endogenous CCA1 promoter (31). Chromatin was isolated from plants harvested at ZT4, the point when *CBF* transcript levels begin to rise. In mock experiments where rabbit immunoglobulin was used for precipitation, no specific binding was detected for any of the sub-regions (A to O) of the *CBF1-3* locus tested or for *TOC1*, *ACTIN7* or *UBIQUITIN10* (Fig. A2.4). In contrast, test experiments indicated that specific binding of the CCA1-GFP protein occurred throughout most of the *CBF1-3* locus and as well as within the promoter region of *TOC1*, a positive control, but not in the promoters of *ACTIN7* or *UBIQUITIN10*, two negative controls (31) (Fig. 2.2). Significant CCA1-GFP binding occurred in the promoter regions of *CBF1* (C), *CBF2* (L), and *CBF3* (G and I). We also observed CCA1 associated with the coding region of *CBF1* (E). This could be due to the tandem connection of the CBF genes and consequently, close proximity to several CCA1 binding sites located in the adjacent *CBF3* promoter (Fig. 2.2). Thus, although it is possible that there is CCA1 binding within the *CBF1* transcript region, the actual binding site could be downstream in the *CBF3* promoter. In sum, the results of our genetic and ChIP experiments supported the model that circadian regulation of *CBF1*, *CBF2* and *CBF3* involves action of CCA1 and LHY binding to the promoters of these genes and up-regulating their transcription during the morning hours.

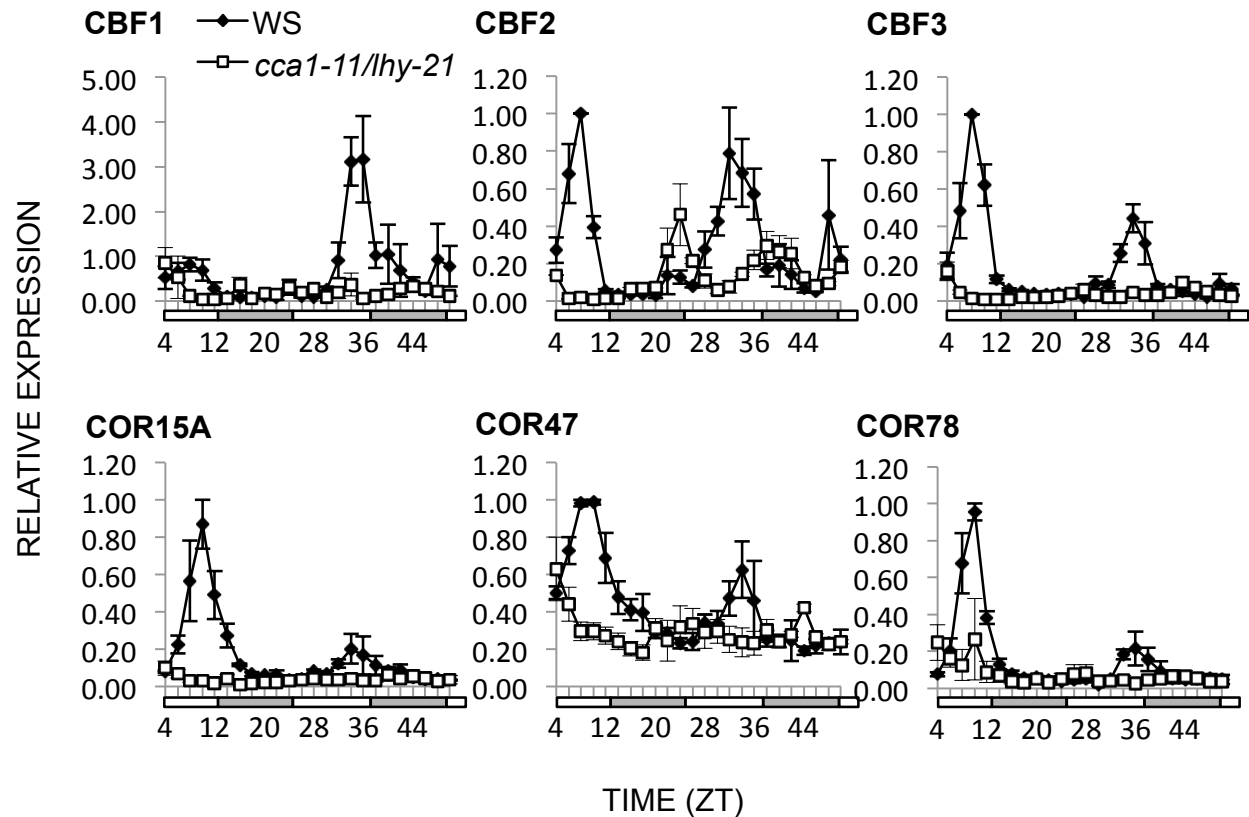


Figure 2.1. Effects of the *cca1-11/lhy-21* double mutation on circadian regulation of *CBF1*, *CBF2*, *CBF3* and CBF-targeted genes *COR15A*, *COR47* and *COR78*. Wild-type Ws-2 (WS) and *cca111/lhy-21* double mutant plants were grown at 22°C under a 12 h photoperiod to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were harvested every 2 h and the transcript levels for the indicated genes were determined by qRT-PCR. Gene expression was normalized to IPP2 for each sample. Gene expression is relative to one wild-type sample set to a value of 1 for each biological replicate. Values are averages from three independent biological experiments (n=3). Error bars indicate \pm SEM.

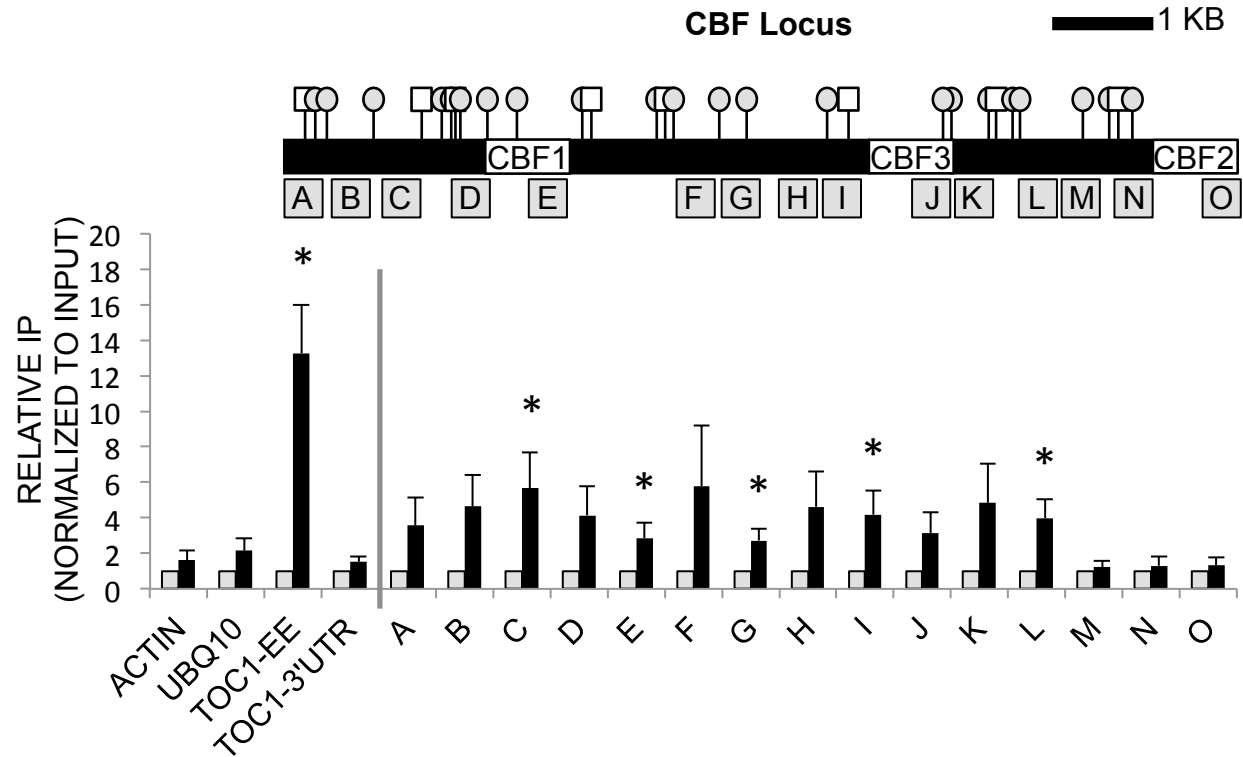


Figure 2.2. Binding of CCA1 at the *CBF1-3* locus. *cca1-1* and *cca1-1 CCA1p:CCA1-GFP* plants were grown at 22°C under a 12 h photoperiod to the four leaf stage. Tissue was fixed at ZT4 and ChIP was performed using anti-GFP antibody. Immunoprecipitated DNA was quantified by qRT-PCR using primers specific to regions within the *CBF1-3* locus (boxes A through O). The levels of immunoprecipitated DNA were normalized to the respective input DNA. Immunoprecipitation in *cca1-1 CCA1p:CCA1-GFP* plants (black bars) is relative to *cca1-1* plants (gray bars) set to a value of 1. Primer pairs directed to the 3'-untranslated region of *TOC1* (*TOC1-3'UTR*), *ACTIN7* (*ACTIN*) and *UBIQUITIN10* (*UBQ10*) were used as negative controls (31). Primers near the EE element in the *TOC1* promoter (*TOC1-EE*) were used as a positive control. Values represent the average of five independent biological experiments (n=5). Error bars indicate \pm SEM. *=p<0.05 using a paired, one-tailed t-test. In the *CBF* locus diagram, the transcribed regions are indicated with white boxes and the approximate positions of CBS (AATCT), and EE (AAAATATCT) motifs are indicated by gray circles, and white squares respectively.

Rhythmic expression of CBF regulon COR genes and freezing tolerance is impaired in plants carrying the cca1-11/lhy-21 double mutation. Harmer *et al.* (19) suggested that the circadian regulation of *CBF1*, *CBF2* and *CBF3* could potentially result in rhythmic expression of CBF regulon *COR* genes. To test this, we examined the transcript levels for three CBF-inducible genes—*COR15A*, *COR47* and *COR78*—in wild-type plants and in plants carrying the *cca1-11/lhy-21* double mutation. The results indicated that the transcript levels for all three *COR* genes oscillated with a period of about 24 h in wild-type plants, though the amplitude of the second peak was much less than that of the first for *COR15A* and *COR78* (Fig. 2.1). For all three genes, the first peak was at about ZT10, which was consistent with the transcript levels of *CBF1*, *CBF2* and *CBF3* peaking just before this at about ZT8 (19, 20) (Fig. 2.1). Moreover, the oscillation in *COR* transcript levels was largely reduced in the *cca1-11/lhy-21* double mutant plants (Fig. 2.1). These results were consistent with the model that circadian-regulated expression of *CBF1*, *CBF2* and *CBF3* imparts rhythmic expression of CBF-targeted *COR* genes at “basal” non-acclimating temperatures.

The decrease in expression of the CBF-targeted *COR* genes in the *cca1-11/lhy-21* double mutant plants could potentially result in a decrease in basal freezing tolerance. We tested this using the electrolyte leakage assay to compare the freezing tolerance of wild-type and *cca1-11/lhy-21* plants. The results indicated that the *cca1-11/lhy-21* double mutation reduced freezing tolerance by about 50%; whereas the EL₅₀ (temperature at which cell damage results in release of 50% of total electrolytes) of wild-

type plants was about -4°C, it was about -2°C in the *cca1-11/hy-21* mutant plants (Fig. 2.3). Thus, the circadian clock is required for maximum basal freezing tolerance.

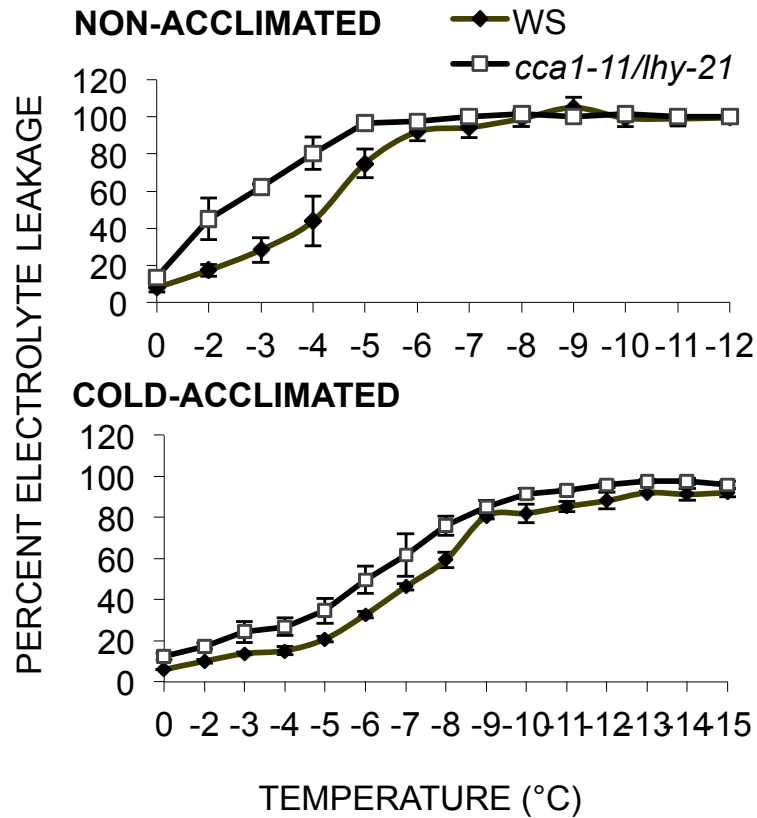


Figure 2.3. Effect of the *cca1-11/lhy-21* double mutation on plant freezing tolerance. *cca1-11/lhy-21* double mutant and wild-type Ws-2 (WS) plants were grown at 22°C under a 12 h photoperiod for approximately 18 days and either tested directly for freezing tolerance (non-acclimated plants; top graph) or were transferred at ZT4 to 4°C for 7 days under a 12 h photoperiod and then tested for freezing tolerance (cold-acclimated plants; bottom graph). Freezing tolerance was tested using the electrolyte leakage test. The results presented are average values from three independent experiments (n=3). Error bars indicate \pm SEM.

CCA1 and LHY regulate cold induction of CBF1, CBF2 and CBF3. Fowler *et al.* (22) reported that cold induction of *CBF1*, *CBF2*, and *CBF3* is gated by the circadian clock. Given our results indicating a role for CCA1 and LHY in the circadian regulation of *CBF1*, *CBF2* and *CBF3*, we asked whether these transcription factors also had a role in the gating phenomenon. As previously reported (22, 34), cold-induction of *CBF1*, *CBF2*, and *CBF3* was much greater in the subjective day than in the subjective evening (Fig. 2.4). This cold induction was little affected by the single *cca1-11* and *lhy-21* mutations (Fig. A2.1), but was greatly reduced in the *cca1-11/lhy-21* double mutants and the period of cycling was shortened (Fig. 2.4) as was the period of cycling for *LHCB* (Fig. A2.3). Thus, CCA1 and LHY have a major role in the induction of *CBF1*, *CBF2* and *CBF3* expression in response to low temperature.

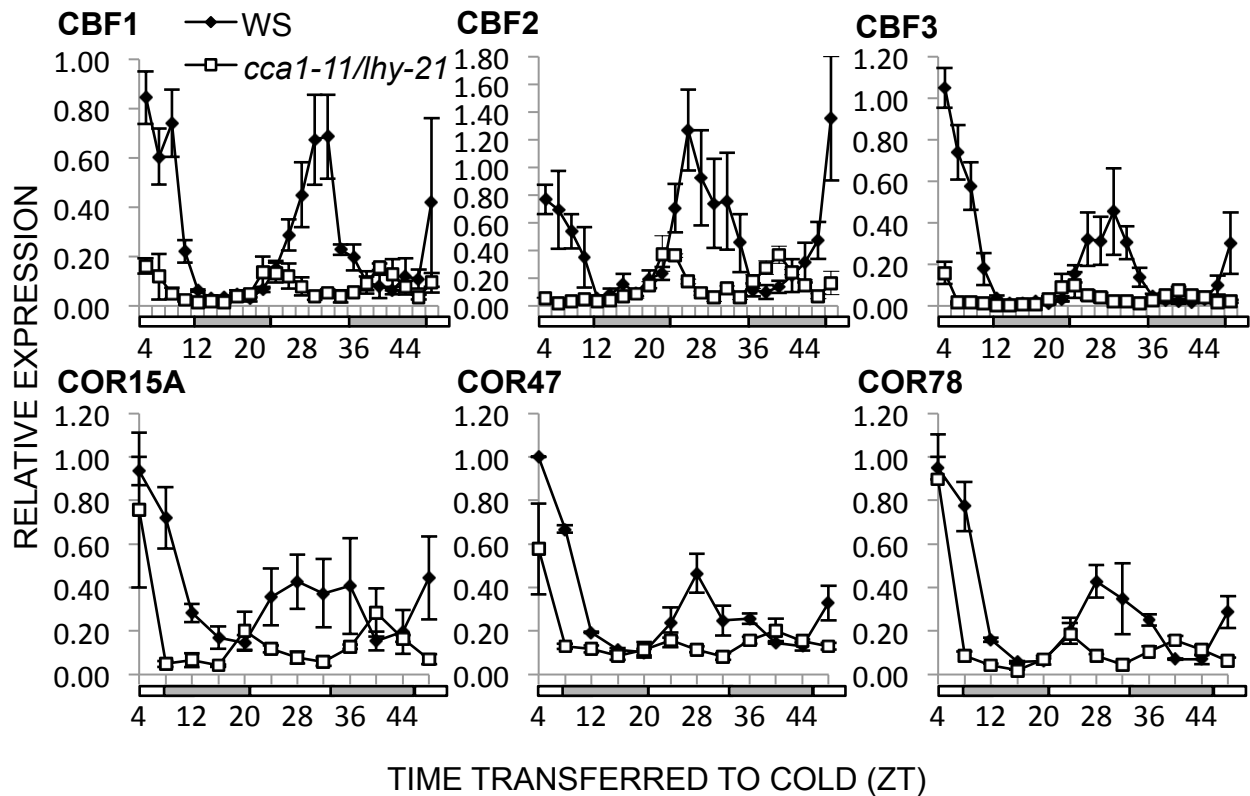


Figure 2.4. Effects of the *cca1-1/lhy-21* double mutation on cold induction of *CBF1*, *CBF2*, *CBF3* and *CBF*-targeted genes *COR15A*, *COR47* and *COR78*. Wild-type *Ws-2* (WS) and *cca1-1/lhy-21* double mutant plants were grown at 22°C under a 12 h photoperiod to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were transferred to cold temperature (4°C) for 2 h, every 2 h (*CBF* genes) or for 4 h, every 4 h (*COR* genes) at the start of constant light conditions. Transcript levels for the indicated genes were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were transferred to cold temperature. Gene expression was normalized to UBQ10 for each sample. Gene expression is relative to one wild-type sample set to a value of 1 for each biological replicate. Values are averages from three independent biological experiments (n=3). Error bars indicate \pm SEM. See Table A2.1 for primer pair sequences.

Cold induction of CBF-targeted COR genes and freezing tolerance is impaired in plants carrying the cca1-11/lhy-21 double mutation. The finding that cold induction of *CBF1*, *CBF2* and *CBF3* was impaired in plants carrying the *cca1-11/lhy-21* double mutation prompted us to determine whether the double mutation also impaired cold induction of CBF-targeted *COR* genes and the freezing tolerance of cold-acclimated plants. In wild-type plants, the degree to which *COR15A*, *COR47* and *COR78* were induced by low temperature cycled with peaks in the late day and troughs in the late subjective evening (Fig. 2.4), times that were consistent with the cycling of *CBF1*, *CBF2* and *CBF3* cold induction (Fig. 2.4). In plants carrying the *cca1-11/lhy-21* double mutation, the peak following the night to day transition was little affected, but the subsequent peaks were greatly diminished and the period of cycling was shortened (Fig. 2.4). In addition, the freezing tolerance of cold-acclimated plants carrying the *cca1-11/lhy-21* double mutation was about 1°C less than that of cold-acclimated wild-type plants (Fig. 2.3). These results indicate that CCA1 and LHY are required for *Arabidopsis* plants to attain maximum levels of *COR* gene induction and freezing tolerance in response to low temperature.

DISCUSSION

The CBF cold response pathway is highly conserved among plants and has a major role in plant freezing tolerance (1-3, 9). Accordingly, there is considerable interest in understanding the mechanisms that control expression of this stress response pathway. Here we establish that the CBF pathway is subject to positive regulation by the circadian clock components CCA1 and LHY. We show that these factors have roles in both circadian regulation and cold induction of the pathway and that they are required for plants to attain maximum freezing tolerance at both basal and cold acclimating temperatures.

At basal growth temperature, the transcript levels for *CBF1*, *CBF2*, and *CBF3* oscillate with peaks and troughs occurring at about ZT8 and ZT20, respectively (19-21) (Fig. 2.1). Our genetic and ChIP analyses indicate that this circadian regulation is due to direct action of CCA1 and LHY binding at the *CBF1-3* locus—presumably at the EE, CBS, and related motifs—and inducing transcription of the *CBF* genes. In the morning hours when CCA1 and LHY protein levels peak (25, 35, 36), the transcript levels of *CBF1*, *CBF2*, and *CBF3* peak; and in the evening hours, when CCA1 and LHY protein levels are low, the transcript levels for *CBF1*, *CBF2*, and *CBF3* are low (Fig. 2.5). The finding that circadian regulation of *CBF1* and *CBF3* is nearly eliminated in plants carrying the *cca1-11/hy-21* double mutation indicates that no other regulatory proteins are sufficient to impart positive circadian regulation of these genes. In contrast, circadian regulation of *CBF2* continues in plants carrying the *cca1-11/hy-21* double mutation, albeit with reduced amplitude and shortened periodicity. Thus, at least one

additional regulatory protein appears to drive positive circadian regulation of *CBF2*.

Prime candidates for this residual regulation are the four REVEILLE (RVE) proteins RVE1, RVE3, RVE4 and RVE8 (42). These MYB-like transcription factors fall into the CCA1 subfamily, bind to the EE motif, are circadian-regulated, and like CCA1 and LHY, have peak transcript levels at dawn (42).

Kidokoro *et al.* (21) reported that circadian regulation of *CBF1*, *CBF2*, and *CBF3* also involves negative regulation. These investigators found that PIF7 binds to a G-box element in the promoter of *CBF2* and that this element was required for down-regulation of the *CBF2* promoter during the subjective evening. In addition, it was found that PIF7 physically interacts with TOC1 (21). Thus, circadian-controlled down-regulation of the *CBF* genes appears to involve action of a PIF7-TOC1 protein complex binding to G-box elements in their promoters (Fig. 2.5).

In addition to establishing a role for CCA1 and LHY in circadian regulation of *CBF1*, *CBF2* and *CBF3*, our results indicate that CCA1 and LHY also act as positive regulators of *CBF* cold induction. This is evidenced by the finding that cold induction of *CBF1*, *CBF2* and *CBF3* is greatly impaired in plants carrying the *cca1-11/lhy-21* double mutation (Fig. 2.4). We propose that the gating of *CBF1*, *CBF2* and *CBF3* cold induction results, in part, from positive synergistic interaction between cold-signaling and clock output pathways, the former mediated by ICE1 and CAMTA3, and the latter by CCA1 and LHY (Fig. 2.5). If the temperature drops in the morning, CCA1 and LHY are present at the *CBF* locus and can act with ICE1 and CAMTA3 to induce high level expression of *CBF1*, *CBF2* and *CBF3*. In contrast, if the temperature drops in the

evening, *CCA1* and *LHY* are at low levels and consequently, there is little synergy between the low temperature and clock pathways and the induction of *CBF1*, *CBF2* and *CBF3* is low, approximating the peak levels obtained with circadian regulation (Fig. 2.5).

Cold induction of *CBF1*, *CBF2* and *CBF3* during the evening hours may also involve negative regulation. Such regulation would not appear to involve *PIF7*, as Kidokoro *et al.* (21) showed that the gating of *CBF1*, *CBF2*, and *CBF3* expression is not impaired in plants carrying the *pif7-2* mutation (21). However, Nakamichi *et al.* (34) found that circadian regulation of *CBF1*, *CBF2* and *CBF3*, and the gating of their cold induction, does not occur in plants carrying the *prr9-11/prr7-10/prr5-10* triple mutation. When plants were grown at basal temperature, the transcript levels for *CBF1*, *CBF2* and *CBF3* remained high throughout the day in the triple mutant plants (34). Similarly, the cold induction of *CBF1*, *CBF2* and *CBF3* in the triple mutant plants remained at about the peak levels observed in wild-type plants regardless of the time of day at which the mutant plants were exposed to low temperature (34). Nakamichi *et al.* (34) concluded that *PRR9*, *PRR7* and *PRR5* are negative regulators of *CBF1*, *CBF2* and *CBF3* and proposed two possible mechanisms. One was that *PRR9*, *PRR7* and *PRR5* directly repress expression of the *CBF* genes. Alternatively, they suggested that aberrant expression of the *CBF* genes might result from the “circadian disorder” caused by the *prr9-11/prr7-10/prr5-10* triple mutation. Our results provide no direct evidence in favor of, or against, the first model. However, the consistently high *CBF* expression may be partially explained by the constitutively elevated expression of *CCA1* and *LHY* in the triple mutant plants (34, 43).

A final point regards the role of the clock in freezing tolerance. Our results indicate that CCA1 and LHY are required for *Arabidopsis* to attain maximum levels of freezing tolerance at both non-acclimating and cold-acclimating temperatures (Fig. 2.3). Recently, Espinoza et al. (44) independently reached the same conclusion as they also found that the *cca1-11/hy-21* double mutation resulted in impaired freezing tolerance. Our results also indicate a mechanism whereby cold-signaling and clock regulatory pathways are integrated to condition freezing tolerance; the positive regulation of the CBF cold response pathway mediated through CCA1 and LHY binding at the *CBF1-3* locus and inducing expression of *CBF1*, *CBF2* and *CBF3* (Fig. 2.5). Taken together, our results suggest that the integration of cold signaling pathways with the circadian clock may have been an important evolutionary event that has contributed to plant adaptation to cold environments.

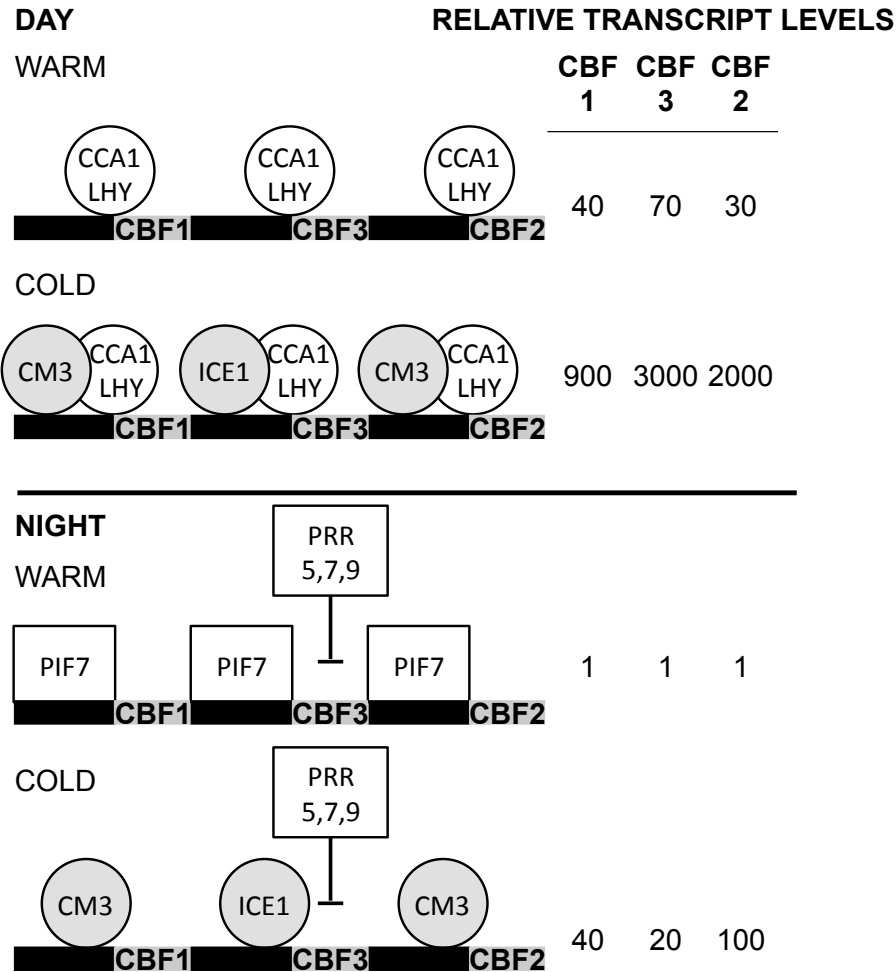


Figure 2.5. Model for circadian regulation and gated cold-induction of *CBF1*, *CBF2* and *CBF3*. During the day, CCA1 and LHY bind throughout the *CBF* locus and promote *CBF* transcription. In the evening, CCA1 and LHY are at low levels and have little effect on *CBF* expression. Oscillations in CCA1 and LHY binding at the *CBF* locus largely account for the circadian regulation of the *CBF* genes. Circadian regulation also involves repression in the evening hours mediated by PIF7 (21) and PRR5, PRR7 and PR9 (34). Transfer of plants to low temperature in the day or evening results in activation of ICE1 (15) and CAMTA3 (CM3) (16), and possibly other transcription factors, that stimulate transcription of the *CBF* genes. If the temperature drops during the day, the clock and cold-signaling pathways act synergistically to induce *CBF* expression to high levels. If plants are exposed to cold temperatures in the evening, there is no positive synergy between the two pathways and there is repression by PRR5, PRR7 and PR9 (34) leading to *CBF* induction at moderate levels. Relative transcript levels were calculated using peak and trough values obtained for the *CBF* genes in the experiments presented in Fig. 2.2.1 and Fig. 4 (the values obtained for plants grown at warm temperature and harvested in the evening were set to 1). See Discussion for details.

MATERIALS AND METHODS

Plant Material and Growth Conditions. *Arabidopsis thaliana* ecotype WS-2 and mutants in this background were grown as described previously (16). Homozygous T-DNA mutant lines were obtained from the *Arabidopsis* Biological Resource Center (45). Null mutations were checked by qRT-PCR. These lines were *cca1-11*(CS9378), *lhy-21* (CS9379) and *cca1-11/lhy-21*(CS9380). Restored *cca1-1* line, CCA1p:CCA1-GFP under the CCA1 endogenous promoter and *cca1-1* (31), used in ChIP experiments, were generously donated by the Kay Lab (University of California, San Diego).

All seeds were stratified for 3 to 5 days in the dark at 4°C. Except for freezing tolerance tests, plants were grown at 22°C under sterile conditions on Gamborg's B5 medium (Caisson Laboratories) without sucrose at $\sim 100 \mu\text{mol m}^{-2}\text{s}^{-1}$ in a 12 h photoperiod. For circadian experiments, plants were sampled at 22°C in $100 \mu\text{mol m}^{-2}\text{s}^{-1}$ constant light or at 4°C in $35 \mu\text{mol m}^{-2}\text{s}^{-1}$ constant light. For electrolyte leakage experiments plants were grown as described (16) at $\sim 100 \mu\text{mol m}^{-2}\text{s}^{-1}$ under a 12 h photoperiod. Cold temperature treatment for plants grown on soil was at 4°C in light at $35 \mu\text{mol m}^{-2}\text{s}^{-1}$ under a 12 h photoperiod.

RNA Analysis. RNA extraction was performed as described (16). For qRT-PCR (Applied Biosystems 7500 FAST Real-Time PCR System in FAST mode), cDNA was made as described in (16) except that total RNA of either 0.2 or 0.025 μg was used for a

40 μ L reverse-transcription reaction. In the 10 μ L PCR reactions, 2 μ L of diluted cDNA was used. UBQ10 or IPP2 were used as reference genes. All primer sets can be found in Table A2.1.

Chromatin Immunoprecipitation. ChIP experiments were carried out as described by Pruneda-Paz et al. (31) with a few modifications. CCA1p:CCA1-GFP and *cca1-1* lines were sampled at ZT4 instead of ZT3. DNA was also purified by PCR Clean-Up Kit (Qiagen) instead of phenol-chloroform extraction. Immunoprecipitated DNA was analyzed with Applied Biosystems FAST real-time PCR in FAST mode (using presets). For each biological replicate immunoprecipitated DNA was normalized to the input DNA as in (31) and each of these values were expressed relative to the *cca1-1* line set to a value of 1. A one-tailed paired t-test was performed to assess the statistical significance of enrichment in the CCA1p:CCA1-GFP line compared to *cca1-1* plants for each primer pair used across biological replicates. Primers pairs used in ChIP experiments can be found in Table A2.1.

Freezing Tolerance Tests. Electrolyte leakage assays were performed as described in (16). For cold acclimation, plants were transferred to 4°C at ZT4 for 7 days under a 12 h photoperiod. Assays for acclimated and non-acclimated plants started at ~ZT2 in all biological replicates.

ACKNOWLEDGMENTS

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APPENDIX

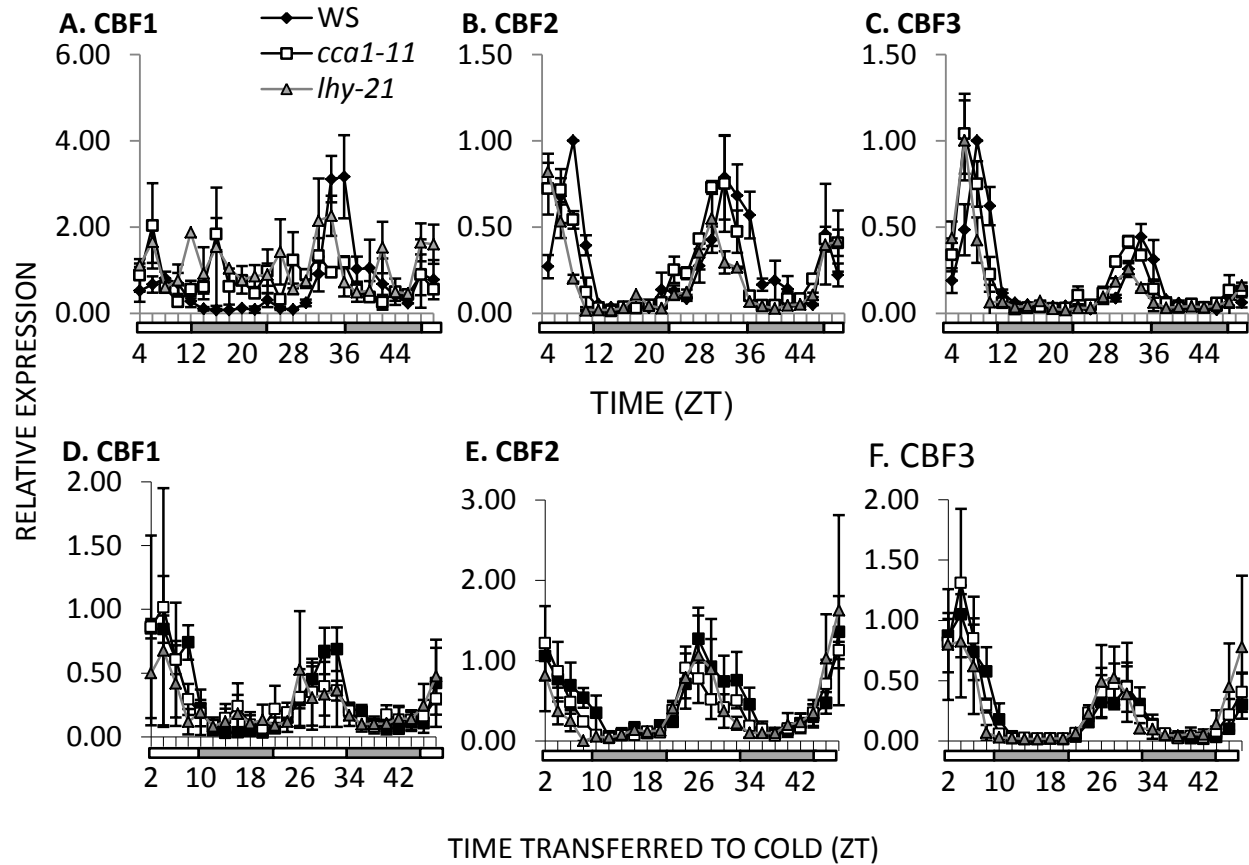


Figure A2.1. Circadian rhythms and gated expression of CBF genes in *cca1-11* and *lhy-21*. qRT-PCR analysis for *CBF1*, *CBF2* and *CBF3* in *cca1-11* (white boxes), *lhy-21* (grey triangles) and wild-type (WS) (black diamonds). Expression for each gene was relative to one wild-type sample (WS) set to a value of 1 in each biological replicate. Values are averages of three independent experiments (n=3) for WS, error bars indicate \pm SEM. Values are average of two independent experiments (n=2) for *cca1-11* and *lhy-21*, error bars indicate \pm range. (A-C) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were harvested every 2 h and the transcript levels for the indicated genes were determined by qRT-PCR. Gene expression was normalized to IPP2 for each sample. (D-F) Plants were grown with a 12 h-photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were dropped to cold temperature (4°C) for 2 h, every 2 h, for 48 h at the start of constant light conditions and the transcript levels for the indicated genes were determined by qRT-PCR. Gene expression was normalized to UBQ10 for each sample. See Table A2.1 for primer pair sequences.

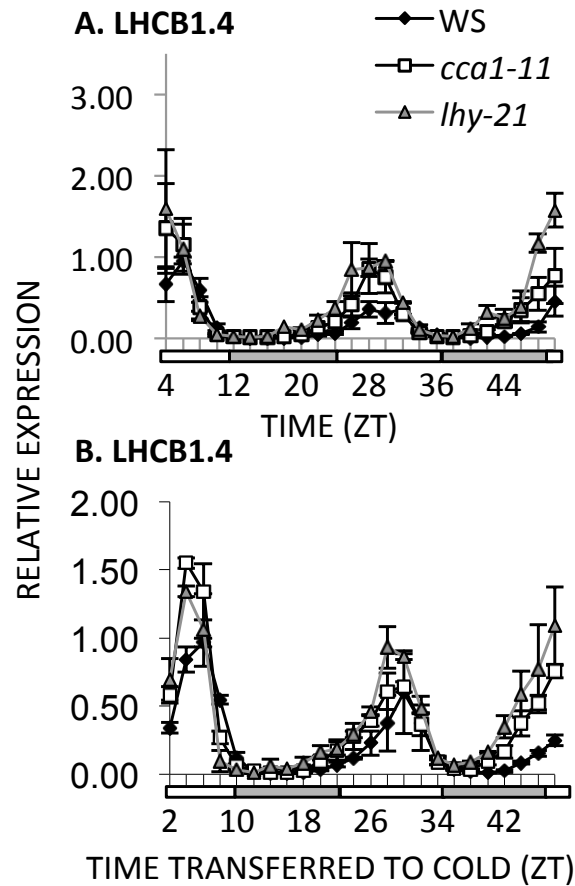


Figure A2.2. Robust circadian rhythms and gated expression of *LHCb1.4* in *cca1-11* and *lhy-21* plants. qRT-PCR analysis of *LHCb1.4* in *cca1-11* (white boxes), *lhy-21* (grey triangles) and wild-type (WS) (black diamonds). Expression for *LHCb1.4* was relative to one wild-type sample (WS) set to a value of 1 in each biological replicate. Values are averages from three independent experiments (n=3) for WS, error bars indicate \pm SEM. Values are averages from two independent experiments (n=2) for *cca1-11* and *lhy-21*, error bars indicate \pm range. (A) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were harvested every 2 h and the transcript levels for *LHCb1.4* were determined by qRT-PCR. Gene expression was normalized to IPP2 for each sample. (B) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were dropped to cold temperature (4°C) for 2 h, every 2 h, for 48 h at the start of constant light conditions and the transcript levels for *LHCb1.4* were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were dropped to cold temperature. Gene expression was normalized to UBQ10 for each sample. See Table A2.1 for primer pair sequences

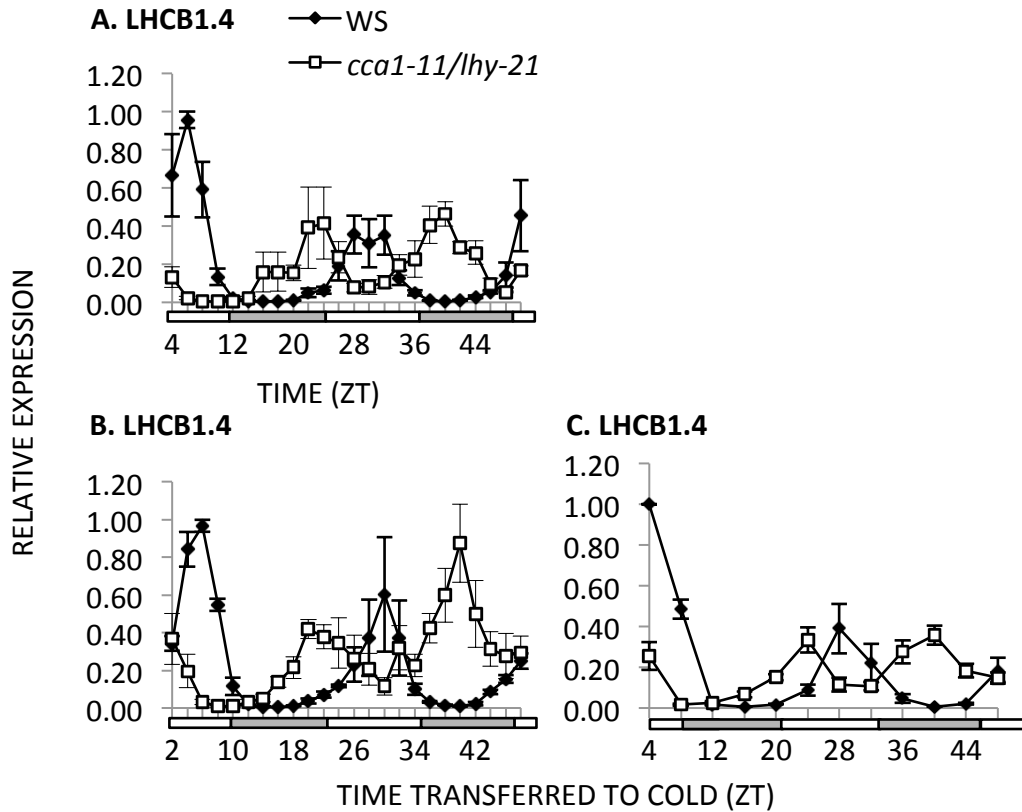


Figure A2.3. Robust circadian rhythms and gated expression of *LHCb1.4* in *cca1-11/lhy-21* plants. qRT-PCR analysis *LHCb1.4* in *cca1-11/lhy-21* (white boxes), and wild-type (WS) (black diamonds). Expression for *LHCb1.4* was relative to one wild-type sample (WS) set to a value of 1 in each biological replicate. Values are averages of three independent experiments (n=3). Error bars indicate \pm SEM. (A) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were harvested every 2 h and the transcript levels for *LHCb1.4* were determined by qRT-PCR. Gene expression was normalized to IPP2 for each sample. (B) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were dropped to cold temperature (4°C) for 2 h, every 2 h, for 48 h at the start of constant light conditions and the transcript levels for the indicated genes were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were dropped to cold temperature. Gene expression was normalized to UBQ10 for each sample. (C) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were dropped to cold temperature for 4 h, every 4 h, for 48 h at the start of constant light conditions and the transcript levels for *LHCb1.4* were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were dropped to cold temperature. Gene expression was normalized to UBQ10 for each sample. See Table A2.1 for primer pair sequences.

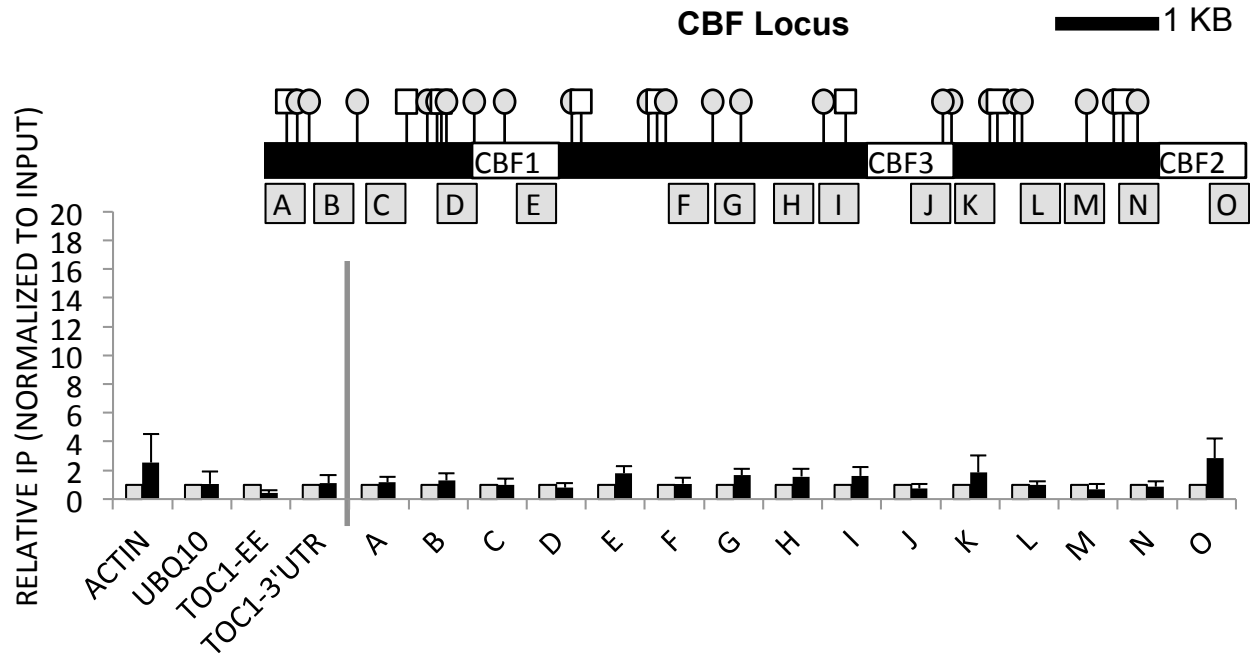


Figure A2.4. Mock experiments testing CCA1 binding at the *CBF1-3* locus. *cca1-1* and *cca1-1 CCA1p:CCA1-GFP* plants were grown at 22°C under a 12 h photoperiod to the four leaf stage. Tissue was fixed at ZT4 and ChIP was performed using rabbit-IgG (Jackson ImmunoResearch) antibody. Immunoprecipitated DNA was quantified by qRT-PCR using primers specific to regions within the *CBF1-3* locus (boxes A through O). The levels of immunoprecipitated DNA were normalized to the respective input DNA. Immunoprecipitation in *cca1-1 CCA1p:CCA1-GFP* plants (black bars) is relative to *cca1-1* plants (gray bars) set to a value of 1. Primer pairs directed to the 3'-untranslated region of *TOC1* (TOC1-3'UTR), *ACTIN7* (ACT) and *UBQUITIN10* (UBQ10) were used as negative controls (32). Primers near the EE element in the *TOC1* promoter (TOC1-EE) were used as a positive control. Values represent the average of five independent biological experiments (n=5). Error bars indicate \pm SEM. *= $p < 0.05$ using a paired, one-tailed t-test. See Table A2.1 for primer pair sequences. In the *CBF* locus diagram, the transcribed regions are indicated with white boxes and the approximate positions of CBS (AATCT), and EE (AAAATATCT) motifs are indicated by gray circles, and white squares.

Table A2.1. Primer pairs used for qRT-PCR and ChIP experiments. Primers used in this study were designed with Primer Express 3.0, using presets optimized for qRT-PCR. For locations of primer pairs on *CBF1-3* please see Fig. 2.2 or Fig. A2.4.

qRT-PCR Primers					
Name	Forward (FW)	FW	Reverse (RV)	RV	ATG
CBF1	GGAGACAATGTTTGGGATGC	+671	CGACTATCGAATATTAGTAACTCC	+793	AT4G25490
CBF2	CGACGGATGCTCATGGTCTT	+560	TCTTCATCCATATAAAACGCATCTTG	+628	AT4G25470
CBF3	TTCCGTCCGTACAGTGGAAT	+694	AACTCCATAACGATACGTCGTC	+741	AT4G25480
COR15A	GAAAAAACAGTGAAACCGCAGAT	+704	CCACATACGCCGCAGCTT	+750	AT2G42540
COR47	CGGTACCAGTGTCGGAGAGT	+748	ACAGCTGGTGAATCCTCTGC	+855	AT1G20440
COR78	GAAAGGAGGAGGAGGAATGG	+2251	AACCAGCCAGATGATTTTGG	+2351	AT5G52310
IPP2	ATTTGCCCATCGTCCTCTGT	+115	GAGAAAGCACGAAAATTCGGTAA	+155	AT3G02780
UBQ10	GGCCTTGATAATCCCTGATGAATAAG	+1833	AAAGAGATAACAGGAACGGAAACATAGT	+1866	AT4G05320
LHCB1.4	GCCTTCGCTACCAACTTCGTC	+829	AACCGGATACACAACTCGATC	+873	AT2G34430
ChIP Primers					
Name	Forward (FW)	FW	Reverse (RV)	RV	ATG
ACTIN	CGTTTCGCTTTCCTTAGTGTTA	+54	AGCGAACGGATCTAGAGACTC	+167	AT5G09810
UBQ10	TCCAGGACAAGGAGGTATTCCTCCG	+1616	CCACCAAAGTTTTACATGAAACGAA	+1796	AT4G05320
TOC1	TTTTATGGCCTGCACTTTTTATTG	-186	GGTGGGACTTGGGATATTTTAGG	-144	AT5G61380
TOC1 3'UTR	GCTACAGCCAAAAAACATCGA	+3212	GAGCCGCAAGAGCCAACAT	+3256	AT5G61380
CBF1-A	AGAACCACGACGATATAGAGAGTGAA	-1997	CGCCTGCCAAAATCATTCTAC	-1951	AT4G25490
CBF1-B	TGCTTTCAAGGCCGAATGAT	-1312	CGTTCTCATTCCACGTGTGATG	-1274	AT4G25490

Table A2.1. (cont'd)

CBF1-C	TTACCACTCTTTTTTCCCTCTTTG	-845	CTCGCTCTCACGTTATTGACATTT	-801	AT4G25490
CBF1-D	TCTTTACAAGGGTCAAAGGACACA	-186	GCGAAGCAATCCCACGAT	-142	AT4G25490
CBF1-E	GGAGACAATGTTTGGGATGC	+671	CGACTATCGAATATTAGTAACTCC	+793	AT4G25490
CBF3-F	AGTTCTATCGGACTAATTCTTGGCTTA	-1859	GATGATCAAGCGTAATTGCTTTGT	-1752	AT4G25480
CBF3-G	TGACTAAGGACGTGGTGGTTGA	-1235	AGCGCACTTCCTTCTCACTCA	-1178	AT4G25480
CBF3-H	TGTTACATTTGATCATTCACCCAAA	-604	CGTATATAAGCACGTAAGTCACCAAGT	-550	AT4G25480
CBF3-I	CGTGGCATTACCAGAGACACA	-124	GCGGAAGATATTTTAGAGGCAAAA	-83	AT4G25480
CBF3-J	TTCCGTCCGTACAGTGGAAT	+694	AACTCCATAACGATACGTCGTC	+741	AT4G25480
CBF2-K	CAAGAGAGCACTGTCCGTAGCTT	-1851	TGGTTACAAGAGGAGCCACGTA	-1811	AT4G25470
CBF2-L	TTTGCCGGAAACTCAACTCA	-1147	CCTTCTTTTTGGTCTGAAA	-1108	AT4G25470
CBF2-M	GAGAGATGCTGGAAATTGTGATCA	-943	AAATATGGTAAGTGGTTAGGCGAAA	-897	AT4G25470
CBF2-N	GGGATCGCTTAGCTGTTTCTTA	-142	TCGAACGCGGAGTTTCTGT	-100	AT4G25470
CBF2-O	CGACGGATGCTCATGGTCTT	+560	TCTTCATCCATATAAAACGCATCTTG	+628	AT4G25470

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CHAPTER THREE

Natural variation in the low temperature response of *Arabidopsis* ecotypes from Sweden and Italy

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SUMMARY

In previous studies, three accessions from two *Arabidopsis* ecotypes collected from Sweden (SW) and Italy (IT) were tested for fitness (survival and fruit-number) in reciprocal transplant experiments. There is substantial variation in temperature across latitudes and minimum winter soil temperature was found to correlated with fitness in these experiments. Consequently, genes associated with freezing tolerance may underlie identified fitness QTLs. This project assessed differences in freezing tolerance between SW and IT accessions using electrolyte leakage assays under laboratory conditions. The SW accession was able to maintain greater freezing tolerance, compared to IT, after 2 weeks of cold-acclimation.

RNA-seq experiments on non-acclimated, 1 week cold-acclimated, and 2 week cold-acclimated SW and IT accessions were analyzed to find genes that may contribute to differences in freezing tolerance. This transcriptomic data was also used to determine general differences between SW and IT and assess if SW and IT reach similar levels of freezing tolerance with similar transcriptional changes.

SW and IT recombinant inbred lines (RILs) were used in expression QTL (eQTL) analysis on genes potentially contributing to differences in freezing tolerance. CBF regulon genes were overrepresented in the list of genes that may potentially contribute to differences in freezing tolerance. Variation in CBF regulon genes, which were

selected for eQTL analysis, indeed mapped to the CBF locus. Interestingly, the location of the CBF locus also overlaps with a fitness QTL.

Based on gene expression levels, CBF2 regulatory sequences may have diverged between SW and IT. However, there is also a 13 base pair deletion in the coding region of IT-CBF2 that results in a predicted protein with a DNA binding domain but without an activation domain. This truncated protein could potentially act as a dominant negative version of CBF2 in IT, able to bind to downstream target genes but incapable of activating transcription. Consistent with this hypothesis, expression of up-regulated CBF target genes is generally lower in IT compared to SW. This would potentially be the first example of a naturally occurring dominant negative version of a CBF. Further work is necessary to confirm this hypothesis. Altogether, results from this study suggest that CBF2 is an excellent candidate gene for differences in freezing tolerance and possibly differences in fitness between SW and IT.

INTRODUCTION

Adaptation of plants and other organisms to their local environment has been frequently observed and is driven by random mutation and natural selection (1-5). With rapid change in global environmental conditions, understanding the molecular mechanisms underlying local adaptation are of particular interest (1-5). *Arabidopsis* is a model plant native to a wide geographical range (Europe to Central Asia) and thus wide range of climatic conditions (6, 7). Two naturally occurring populations of *Arabidopsis* from Rodasen, Sweden (SW) and Castelnuovo di Porto, Italy (IT) were recently collected (8). In reciprocal transplant experiments with three accessions from each SW and IT population, accessions originating from the experimental sites had a strong advantage in terms of fruit-number and survival throughout five years of experimentation (8). Barrett et al. 2011 defined an adaptive allele as one that functionally affects a phenotypic trait to increase fitness (5). Consequently, it can be very difficult to demonstrate that specific alleles are adaptive (1-5). Reciprocal transplant experiments with recombinant inbred lines (RILs) allow fitness to be measured then mapped to specific genetic regions (quantitative trait loci; QTL) and for segregating traits to be identified (5, 9). This method thus connects specific genetic regions to a phenotypic trait that impacts fitness, though these regions can still often span several hundred genes (5).

The Schemske lab sequenced SW and IT accessions and 141,437 single nucleotide polymorphisms (SNPs) were found (Schemske, unpublished). A population of RILs was created by crossing a randomly selected individual from the SW ecotype

(male parent) to a randomly selected individual from the IT ecotype (female parent), seeds from the F1 generation were used to establish a large number of lines that were selfed by single-seed descent for nine generations (Schemske and Agren, unpublished). SW and IT RILs used in this study were from the F10 generation. 384 of the 141,437 SNPs with an average spacing of ~1.1 cm across the *Arabidopsis* genome were selected to genotype the RIL population (Schemske, unpublished). 400 genotyped RILs were then grown at both the Swedish and Italian sites and QTL for fitness were mapped by measuring both fruit-number and survival over these 384 markers (Table 3.1; Schemske and Agren, unpublished).

Both minimum winter soil temperature and flowering-time were found to correlate with fitness of the parent lines during previous reciprocal transplant experiments (8). Other recent studies on local adaptation in various *Arabidopsis* ecotypes have also shown temperature variation to be particularly linked to identified 'local alleles' (1, 2). Sweden has significantly lower average air and soil temperatures in comparison to Italy, especially during winter months (8). Therefore, freezing tolerance is a potential trait underlying fitness QTL mapped in the RILs. Ideally, the SW and IT RILs would be quantitatively (or even semi-quantitatively) tested for differences in freezing tolerance (10) in order to map freezing tolerance QTL and determine if they overlap with fitness QTL. The Schemske lab is currently optimizing freezing tolerance protocols for mass screening of RILs. Keeping this in mind, a reductionist approach has been taken to determine if freezing tolerance related genes underlie previously identified fitness QTL (Table 3.1; Schemske and Agren, unpublished).

Results and analyses are broken into three sections. First, significant differences in SW and IT freezing tolerance are shown under controlled laboratory conditions. Second, RNA-seq experiments are used to define a set of genes possibly contributing to differences in freezing tolerance. This is the first look at the transcriptomes of these two *Arabidopsis* accessions. Therefore in this second section, basal differences between SW and IT transcriptomes are observed. Further analysis of RNA-seq data in this section also suggests that the cold-response of the SW and IT accessions is significantly different. Third, a subset of the genes that possibly contribute to differences in freezing tolerance are used to map QTL for differences in gene expression (expression QTL; eQTL; (11-14)). Some of the eQTL were found to overlap with previously defined fitness QTL (Table 3.1). Consequently, the genes underlying eQTL mapped for genes possibly contributing to differences in freezing tolerance might also underlie fitness QTL. This study therefore provides evidence in support of freezing tolerance as a trait underlying SW and IT fitness QTL (Table 3.1) and suggests candidate genes contributing to these QTL.

Table 3.1. Locations of fitness QTL. 400 RILs were grown at both Swedish and Italian field sites in 2009. Fruit-number and survival-until-fruit-set were fitness traits measured (Data courtesy of the Schemske Lab, unpublished). RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>) for each fitness trait at each location. To detect QTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LOD) values were calculated for each trait using 1000 permutations in RQTL. To estimate the location of each QTL a Bayes credible interval (99%) was calculated in RQTL. No QTL are reported for survival-until-fruit-set at the Italian field site because no QTL were detected above the calculated significance threshold of 3.42 ($\alpha=0.01$) or even a less stringent LOD threshold of 2.54 ($\alpha=0.05$). For locations of markers based on the Col-0 genome please see Table A3.1. 'MB' indicates the megabases contained within the QTL. '# GENES' indicates the number of genes contained within the QTL based on the Col-0 genome. %VAR indicates the estimated proportion of phenotypic variance explained by the QTL.

FRUIT-NUMBER ITALY 2009 (LOD THRESHOLD 3.17)							
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	LOD	%VAR
1	57.35	58.00	62.48	2.00	584	18.35	11.66
2	22.00	26.84	46.00	6.59	1803	8.04	4.81
4	44.31	57.69	60.00	6.91	2065	14.37	8.91
5	0.00	1.40	4.59	1.16	365	9.18	5.52
5	68.69	75.80	77.42	3.19	988	18.14	11.52
FRUIT-NUMBER SWEDEN 2009 (LOD THRESHOLD 3.08)							
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	LOD	%VAR
2	50.00	55.27	58.59	4.32	1427	12.26	11.68
SURVIVAL SWEDEN 2009 (LOD THRESHOLD 2.99)							
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	LOD	%VAR
1	76.9	78.6	83	2.31	695	6.26	7.27
5	64	71.3	78.2	4.89	1505	4.63	4.8

RESULTS AND DISCUSSION: FREEZING TOLERANCE OF SW AND IT

Summary: Freezing tolerance of SW and IT. Minimum winter soil temperature was correlated with the relative fitness of the parent lines during previous reciprocal transplant experiments (8). If freezing tolerance is a trait underlying fitness QTL mapped for the RILS, then SW and IT accessions should differ in freezing tolerance. SW and IT have similar freezing tolerance without cold-acclimation and with 1 week of cold-acclimation (Fig 3.1). But SW was significantly more freezing tolerant than IT with both 2 and 3 weeks of cold-acclimation (Fig. 3.2). Consequently, genes contributing to differences in freezing tolerance between SW and IT may also be candidates for genes underlying fitness QTL.

Freezing tolerance of SW and IT. If freezing tolerance is a trait underlying fitness QTL found using the SW and IT RILs, then a significant difference in the freezing tolerance of the SW and IT accessions might be expected. Electrolyte leakage assays were used to assess the freezing tolerance of both SW and IT accessions (16). The EL₅₀ (temperature at which cell damage results in release of 50% of total electrolytes) of SW and IT accessions was the same under non-acclimated conditions (EL₅₀=-2.5; Fig. 3.1). Hannah et al. 2006 found that nine accessions of *Arabidopsis* generally had similar non-acclimated freezing tolerance despite differing levels of freezing tolerance at 2 weeks of cold-acclimation (6). Accordingly, differences in the cold-acclimated freezing tolerance of the SW and IT accessions were determined.

After 1 week of cold-acclimation at 4°C, both SW and IT can increase their freezing tolerance to a similar degree (EL₅₀=-5.5; Fig. 3.1). However after 2 weeks of

cold-acclimation at 4°C, IT is unable to maintain freezing tolerance similar to 1 week of cold-acclimation (Fig 3.1) and decreases in freezing tolerance ($EL_{50}=-4.5$; Fig. 3.2). On the other hand, the SW accession increases in freezing tolerance after 2 weeks of cold-acclimation ($EL_{50}=-6.5$; Fig. 3.2) in comparison to 1 week of cold-acclimation (Fig 3.1). After 3 weeks of cold-acclimation at 4°C there is still a substantial difference in freezing tolerance between SW ($EL_{50}=-5.5$) and IT ($EL_{50}=-4.5$) though the SW line has decreased its freezing tolerance in comparison to 2 weeks of cold-acclimation (Fig 3.2).

The study by Hannah et al. 2006 also found that the cold-acclimated freezing tolerance of nine accessions of *Arabidopsis* correlated considerably with the accession's average home temperature in the coldest month of the growing season (6). A separate study using 71 accessions of *Arabidopsis* found a similar correlation with the latitude of origin (17). Data from this study corroborates both of these previous studies. SW, the accession with the colder home winter temperature and higher latitude (8), has greater freezing tolerance than IT after 2 weeks of cold-acclimation (Fig. 3.2).

The SW accession was significantly more freezing tolerant than the IT accession at both 2 ($p \leq 0.026$ at EL_{50} calculated by Student's t test) and 3 ($p \leq 0.015$ at EL_{50} calculated by Student's t test) weeks of cold-acclimation (Fig 3.2). Therefore, freezing tolerance is a potential trait contributing to local adaptation (1-5). Consequently, genes contributing to differences in freezing tolerance between SW and IT may also be candidates for genes underlying fitness QTL.

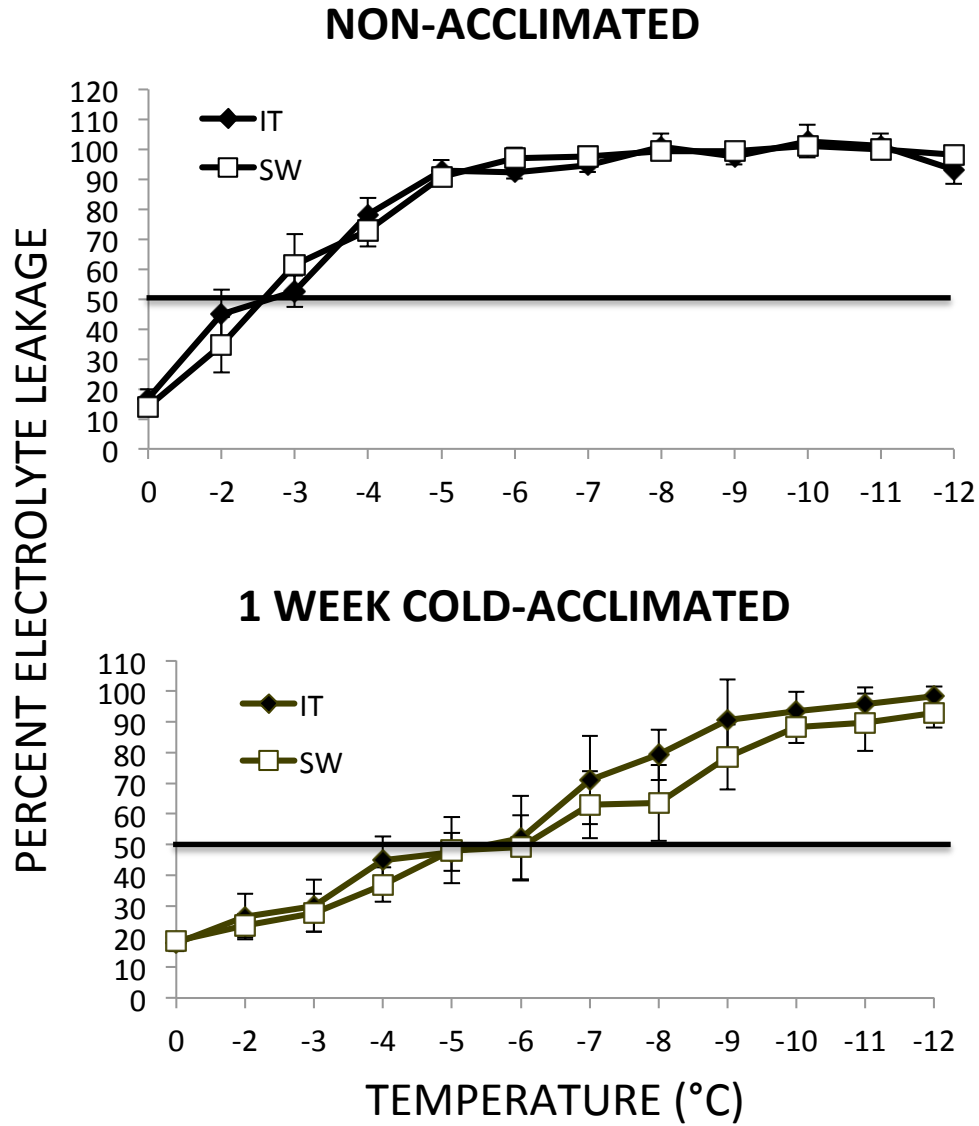


Figure 3.1. Freezing tolerance of SW and IT accessions without cold-acclimation or with 1 week of cold-acclimation. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days and either tested directly for freezing tolerance (non-acclimated plants; top graph) or were transferred at ZT4 to 4°C for 7 days under a 12 h photoperiod and then tested for freezing tolerance (cold-acclimated plants; bottom graph). Freezing tolerance was tested using the electrolyte leakage test. For non-acclimated samples the results presented are average values from six independent experiments (n=6). For 1 week cold-acclimated samples the results presented are average values from four independent experiments (n=4). Error bars indicate \pm SEM.

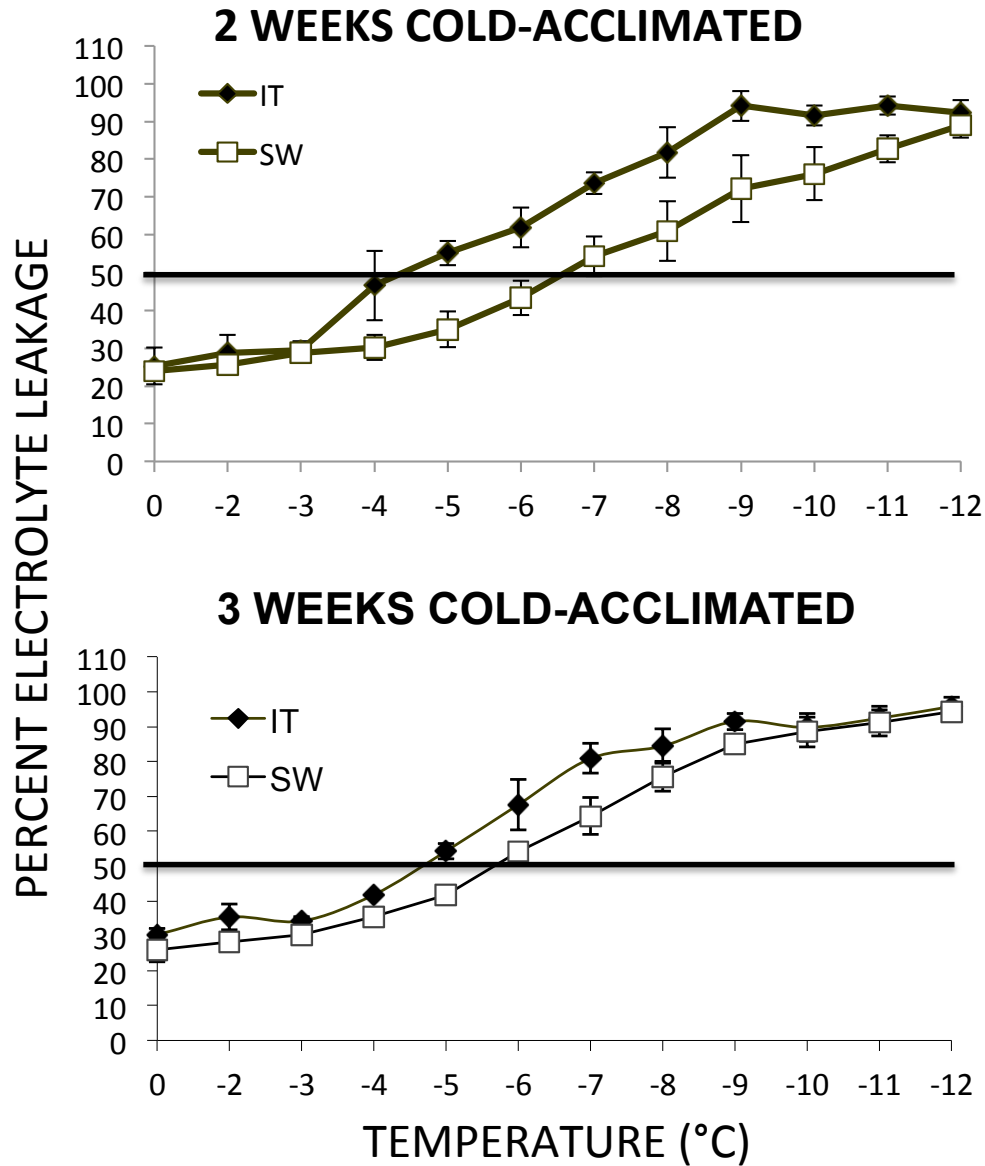


Figure 3.2. Freezing tolerance of SW and IT accessions with 2 or 3 weeks of cold-acclimation. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days then transferred at ZT4 to 4°C for 2 weeks (2 week cold-acclimated plants; top graph) or 3 weeks (3 week cold-acclimated plants; bottom graph) under a 12 h photoperiod and then tested for freezing tolerance. Freezing tolerance was tested using the electrolyte leakage test. For 2 week cold –acclimated samples the results presented are average values from five independent experiments (n=5). For 3 week cold-acclimated samples the results presented are average values from four independent experiments (n=4). Error bars indicate \pm SEM.

RESULTS AND DISCUSSION: SW AND IT RNA-SEQ EXPERIMENTS

This second section of experiments analyzes RNA-seq data from non-acclimated, 1 week cold-acclimated and 2 week cold-acclimated SW and IT samples. The ultimate goal of these experiments is to determine the genes that potentially contribute to differences in freezing tolerance seen after 2 weeks of cold-acclimation in SW and IT. However, the transcriptomes of these accessions have never been examined and transcriptional changes prior to 2 weeks of cold-acclimation may contribute to differences in freezing tolerance. Furthermore, the freezing tolerance of SW and IT are the same at 1 week of cold-acclimation, which allows the opportunity to determine if SW and IT attain similar levels of freezing tolerance with similar transcriptional changes. Therefore analysis of RNA-seq data is partitioned into four subsections. First, basal differences between SW and IT transcriptomes are examined. Second, sets of cold-regulated genes for SW and IT are defined. Third, SW and IT cold-regulated transcripts at 1 week of cold-acclimation are compared. Fourth, two sets of genes that may contribute to differences in freezing tolerance are defined.

ANALYSIS OF SW AND IT BASAL TRANSCRIPTOMES

Summary: Analysis of SW and IT basal (non-acclimated) transcriptomes. In the first section of results it is demonstrated that SW has significantly ($p \leq 0.026$) higher freezing tolerance in comparison to IT after 2 weeks of cold-acclimation (Fig 3.2). Although SW and IT have similar levels of freezing tolerance under non-acclimated conditions, basal (non-acclimated) transcriptome differences were first evaluated to examine general variance and to determine if genes previously associated with cold-

acclimation are enriched in differentially expressed genes. There are a total of 1259 genes differentially expressed between SW and IT under non-acclimated conditions. Analysis of these differentially expressed genes does not overtly suggest that there will be greater freezing tolerance in SW after 2 weeks of cold-acclimation but does indicate that there might be differences in known pathways of cold-acclimation between SW and IT.

Analysis of SW and IT basal transcriptomes. SW has significantly ($p \leq 0.026$) higher freezing tolerance in comparison to IT after 2 weeks of cold-acclimation (Fig 3.2). RNA-seq experiments were conducted on rosette tissue collected before the start of electrolyte leakage assays (Fig 3.1 and 3.2). For RNA-seq experiments average correlation between biological replicates was $R^2 = 0.90$, suggesting that RNA-seq data in this study is repeatable (Table A3.9). Altogether, SW and IT transcriptomes under non-acclimated, 1 week cold-acclimated, and 2 week cold-acclimated conditions, represent a total of 25,678 genes (76.4%) from the *Arabidopsis* Col-0 genome. On average, there were 23,264 genes (69.2%) represented in each sample (Table A3.9). Pairwise comparisons between non-acclimated and cold-acclimated samples or between SW and IT samples were calculated using a two-tailed Student's t test in the cuffdiff program within Cufflinks (18). Genes were considered differentially expressed between non-acclimated and cold-acclimated samples or between SW and IT samples if the Benjamini-Hochberg (false discovery rate; FDR) corrected p-value was ≤ 0.050 . While analyzing differentially expressed genes a ≥ 3 fragments per kilobase of exon per million

fragments mapped (FPKM) threshold (i.e. one sample in the pairwise comparison had to be ≥ 3 FPKM) was used (19, 20).

Genes differentially expressed between SW and IT without treatment (non-acclimated conditions) were first evaluated to examine general variance and to determine if genes previously associated with cold-acclimation are enriched in differentially expressed genes. Altogether, 1259 genes were differentially expressed between SW and IT under non-acclimated conditions that met the ≥ 3 FPKM criteria (19, 20) in either SW or IT (Fig. 3.3). Of these differentially expressed genes, 887 genes have higher expression in IT and 372 are higher in SW (Fig. 3.3).

Although these basal differences in gene expression do not manifest into differences in non-acclimated freezing tolerance, it is of interest to determine if the 1259 genes differentially expressed between SW and IT have previously been correlated with increases in freezing tolerance. In 2006, Hannah et al. analyzed the freezing tolerance and transcriptomes of nine accessions of *Arabidopsis* after two weeks of cold acclimation to define a set of genes (henceforth referred to as Hannah genes) positively and negatively correlated with increases in freezing tolerance associated with cold-acclimation (6). The Hannah gene set is significantly enriched in genes from the CBF regulon (6). The CBFs are a fundamental component of the most well described pathway of cold-acclimation. Overexpression of *CBF* genes leads to constitutive activation of CBF target genes and a plant with much higher freezing tolerance (21-25). The Hannah gene set is also enriched in genes from the ZAT12 regulon, and overexpression of ZAT12 leads to a slight increase in plant freezing tolerance (25).

There significant overlap (40 genes; $p=1.5E-08$; calculated using a hypergeometric distribution in R) between the 887 genes with higher expression in IT and Hannah genes positively correlated with freezing tolerance (Fig. 3.3). A significant number of these (36 of the 40) overlapping genes have greater than 2-fold higher expression in IT compared to SW (Fig 3.4). Since the CBF regulon is enriched in the Hannah gene set, it was determined if the genes with higher basal expression in IT are also enriched in CBF regulated genes (25). Indeed, the genes with higher basal expression in IT are enriched (26 genes; $p=0.00056$) in genes up-regulated by CBFs (25). The CBF pathway of cold-acclimation appears to be differentially regulated between SW and IT. Since there is no difference in freezing tolerance between SW and IT under non-acclimated conditions (Fig 3.1) significant overlap between genes with higher in expression in SW, Hannah genes positively correlated with freezing tolerance, and CBF regulon genes, might also be expected. This was not the case. There is not significant overlap between Hannah genes positively or negatively correlated with freezing tolerance and the 372 genes with higher expression in SW (Fig 3.3). These 372 genes also are not enriched in CBF regulon genes. Consequently, it was of interest to determine what types of genes are enriched.

Gene ontology (GO) enrichment analysis showed that no cold related GO categories were specifically enriched in the 372 genes with higher expression in SW (Table 3.2). 'Response to abiotic stimulus' and 'response to water deprivation' were enriched GO categories in the 887 genes with higher expression in IT (Table 3.3). Defense response related GO categories were significantly enriched in both sets of

genes (Table 3.2 and 3.3). Cold-related GO categories were not specifically enriched in SW or IT, but previous research by Hon et al. 1995 showed that defense response related genes, PR2, PR3 and PR5 encode proteins with antifreeze properties (26). Interestingly, PR3 expression is significantly higher (17.5 fold; $p \leq 0.05$) in SW compared to IT at non-acclimated conditions.

In summary, SW has higher basal expression of 372 genes, which includes a pathogenesis related gene with antifreeze properties. IT has higher basal expression of 887 genes, which are significantly enriched in genes previously positively correlated with cold-acclimation. Given this enrichment of genes positively correlated with freezing tolerance, is it interesting that IT is not more freezing tolerant than SW under non-acclimated conditions. Ultimately, comparison of SW and IT basal transcriptomes does not reveal any obvious suggestion for the higher level of freezing tolerance in SW seen later at 2 weeks of cold-acclimation, but does suggest that genes associated with cold-acclimation may be differentially regulated between SW and IT.

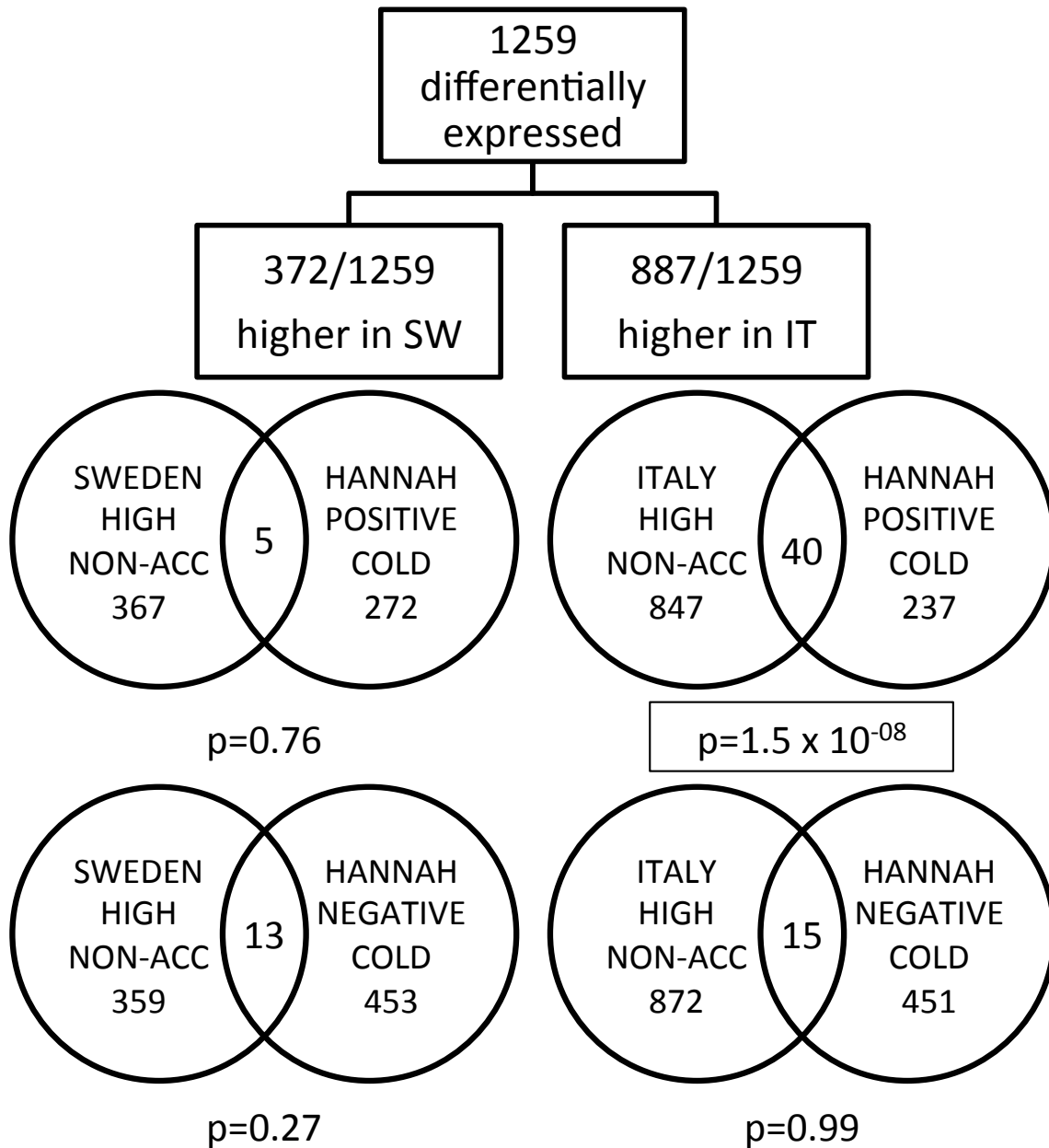


Figure 3.3 1259 genes differentially expressed between SW and IT under non-acclimated conditions. (Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression. P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes; 277 Hannah genes positively correlated with freezing tolerance and 466 Hannah genes negatively correlated with freezing tolerance; (6)).

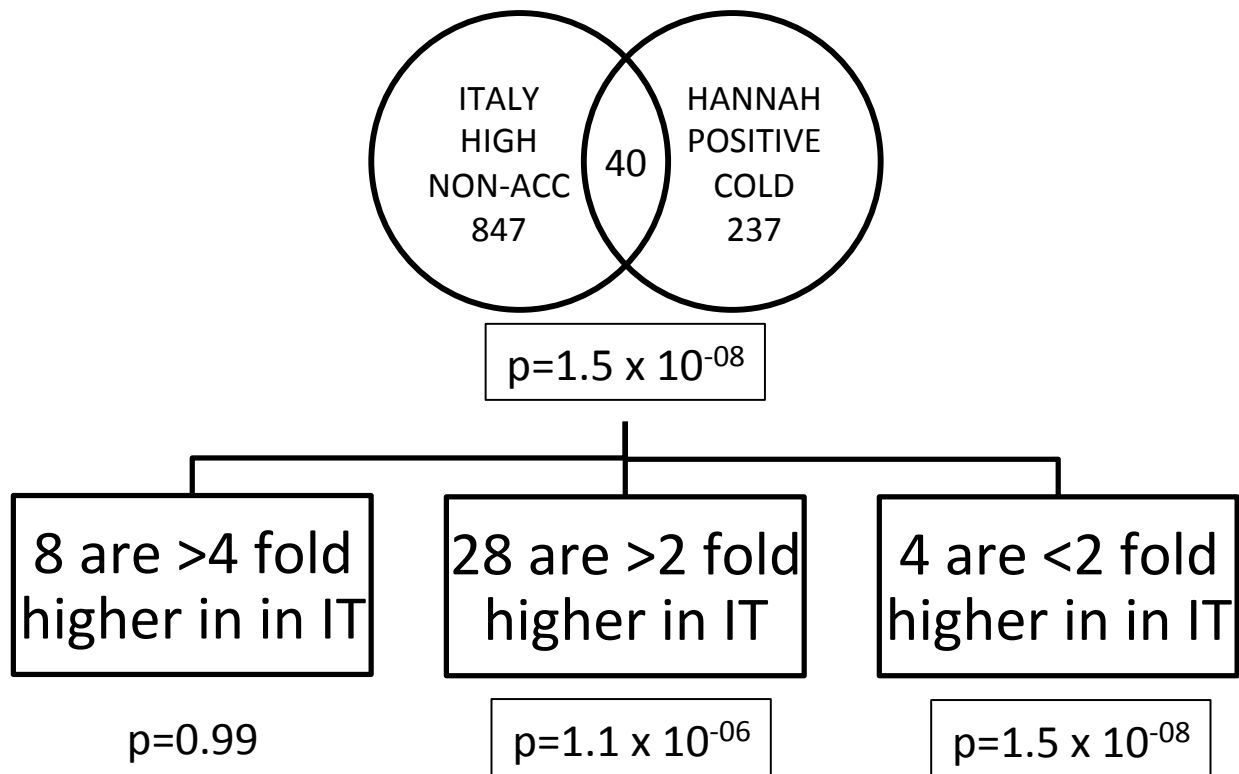


Figure 3.4. Fold change of 40 Hannah genes positively correlated with freezing tolerance that have higher expression in IT. P-values represent the significance of overlap between 'IT High' and Hannah genes if 4- fold, 2-fold, or no fold change cut-offs were applied (from left to right).

Table 3.2 GO categories significantly enriched in 372 genes with higher expression in SW under non-acclimated conditions. GO enrichment was determined using the database for annotation visualization and integrated discovery (DAVID; david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in the 372 genes with higher expression in SW. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	HIGH IN SW (372 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Apoptosis	15	172	3.45E-05
BP	Programmed Cell Death	16	223	4.76E-05
BP	Defense Response	36	1030	5.30E-05
BP	Cell Death	16	254	1.87E-04
MF	Amine Biosynthetic Process	13	204	1.79E-03
BP	Carbohydrate Binding	14	215	2.62E-03
BP	Nitrogen Biosynthetic Process	19	506	1.16E-02
CC	Oxidation Reduction	32	1186	1.23E-02
BP	Biogenic Amine Biosynthetic Process	6	40	1.25E-02
CC	External Encapsulating Structure	22	620	1.26E-02
BP	Cell Wall	22	611	2.05E-02
MF	Immune Response	13	293	3.00E-02
BP	Amino Acid Biosynthetic Process	10	184	3.86E-02
MF	Amino Acid Derivative Biosynthetic Process	10	194	4.31E-02
BP	Innate Immune Response	12	275	4.59E-02
MF	Sugar Binding	9	124	4.74E-02
BP	Biogenic Amine Metabolic Process	6	59	4.85E-02

Table 3.3. GO categories significantly enriched in 887 genes with higher expression in IT under non-acclimated conditions. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in the 887 genes with higher expression in IT. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	HIGH IN IT (887 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Secondary Metabolic Process	37	423	1.24E-04
BP	Defense Response	65	1030	1.62E-04
BP	Apoptosis	21	172	2.49E-04
BP	Phenylpropanoid Metabolic Process	20	166	4.47E-04
BP	Programmed Cell Death	23	223	6.89E-04
BP	Flavonoid Biosynthetic Process	11	53	1.03E-03
BP	Cell Death	24	254	1.25E-03
BP	Flavonoid Metabolic Process	11	57	1.53E-03
BP	Phenylpropanoid Biosynthetic Process	16	128	1.80E-03
BP	Immune Response	25	293	2.99E-03
BP	Amino Acid Derivative Metabolic Process	24	280	3.77E-03
BP	Response To Water Deprivation	18	176	4.74E-03
BP	Response To Abiotic Stimulus	64	1197	6.22E-03
BP	Oxidation Reduction	63	1186	7.34E-03
BP	Response To Water	18	185	7.55E-03
BP	Amino Acid Derivative Biosynthetic Process	18	194	1.17E-02
BP	Innate Immune Response	22	275	1.43E-02
BP	Response To Salicylic Acid Stimulus	15	150	1.97E-02
BP	Aromatic Compound Biosynthetic Process	18	215	3.23E-02
BP	Response To Organic Substance	59	1176	4.06E-02

SW AND IT COLD-INDUCED GENES

Summary: SW and IT cold-induced genes. In this second subsection of RNA-seq analysis cold-regulated genes for SW and IT are defined. SW and IT cold-regulated genes are also compared in greater detail in the third and fourth RNA-seq experiment subsections. However general analysis reveals that SW cold-regulated genes are more enriched in genes previously associated with increases in freezing tolerance than IT cold-regulated genes. The quantity of cold-regulated genes in SW and IT also suggests that there could be differences their capacity to cold-acclimate.

SW and IT cold-induced genes. To find genes contributing to differences in freezing tolerance seen after 2 weeks of cold-acclimation (Fig 3.2) sets of cold-responsive genes were first defined for both SW and IT. RNA-seq data generated from SW and IT rosette tissue under non-acclimated, 1 week cold-acclimated and 2 week cold acclimated conditions, was collected before electrolyte leakage experiments.

Fold change cut-offs were not used while defining sets of cold-responsive genes. A fold change threshold is an arbitrary value and not necessarily biologically significant especially since transcription factors can have compounding downstream effects (28). Furthermore, if fold change cut-offs were applied when defining cold-induced genes important similarities between SW and IT cold-responsive genes might later have been missed. For example, a gene up-regulated 2-fold by cold in SW and 1.9-fold in IT would have been removed from the list of IT cold-responsive genes if a 2-fold threshold had been applied. Instead, genes were considered differentially expressed in response to cold if the FDR corrected p-value comparing non-acclimated to cold samples was

≤ 0.050 (as determined by cuffdiff program within Cufflinks). Although a fold-change threshold was not used, a ≥ 3 FPKM cut-off for differential expression was used while defining cold-responsive genes (19, 20). This means that the sample with higher expression (FPKM) was greater or equal to ≥ 3 FPKM, otherwise interesting genes may have been removed. For example, a gene down-regulated from 15,000FPKM to 1FPKM, after 1 week of cold-acclimation, would have been filtered out if both non-acclimated and cold samples were required to have ≥ 3 FPKM. While a threshold of ≥ 3 FPKM (19, 20) for differentially expressed genes is still an arbitrary cut-off, it allows small but significant (≤ 0.050) fold-change differences to be included in analysis.

Based on the methods above, there are a total of 2307 genes up-regulated by cold in SW and 2310 genes up-regulated by cold in IT (Fig. 3.5 and 3.6). There are also a total of 3252 genes down-regulated by cold in SW and 2658 genes down-regulated by cold in IT (Fig. 3.5 and 3.6). Both SW and IT increase their freezing tolerance after 1 and 2 weeks of cold-acclimation in comparison to basal freezing tolerance (Fig. 3.1 and 3.2). The defined cold-regulated genes are presumed to instigate these increases in freezing tolerance associated with cold-acclimation. A previous study by Hannah et al. 2006 defined a set of genes positively and negatively correlated with increases in freezing tolerance associated with cold acclimation (this set of genes is detailed in earlier analysis of basal transcriptomes; (6)). Therefore it was determined if SW and IT cold-regulated genes are enriched in genes previously associated with cold-acclimation. Fittingly, genes up-regulated by cold in SW and IT are enriched for Hannah genes positively correlated but not negatively correlated with increases in freezing tolerance

(Fig 3.7 and 3.8). Interestingly, the number of Hannah genes positively correlated with freezing tolerance, which overlap with SW genes up-regulated by cold, is more significant than with the genes up-regulated by cold in IT (Fig. 3.7 and 3.8). In SW, genes down-regulated by cold are significantly enriched in Hannah genes negatively correlated with freezing tolerance (Fig. 3.6). However, this is not also the case for the 2658 IT genes down-regulated by cold (Fig. 3.8). This suggests that there may be significant differences in the genes that comprise SW and IT cold-regulated transcriptomes, which will be further analyzed in the following subsections.

SW and IT have similar freezing tolerance at 1 week of cold-acclimation and differing freezing tolerance at 2 weeks of cold-acclimation. Therefore it was also of interest to determine if there is significant overlap between cold-regulated genes at 1 and 2 weeks of cold-acclimation, or if discrete sets of genes are cold-regulated with 1 and 2 weeks of cold-acclimation (Fig 3.5 and 3.6). There is significant overlap in genes up-regulated by cold at 1 week and 2 weeks of cold-acclimation in both SW (1013 genes; Fig 3.5) and IT (1007; Fig. 3.6). There is also significant overlap in genes down-regulated by cold at 1 and 2 weeks of cold acclimation in both SW (1275 genes; Fig 3.5) and IT (1376 genes; Fig 3.6).

Hannah et al. 2006 found that the cold-acclimation capacity of nine accessions of *Arabidopsis* was positively correlated with the number of genes that significantly change in expression in response to cold (6). Consequently, it was of interest to determine if this correlation was also corroborated with sets of SW and IT cold-regulated genes. At 2 weeks of cold-acclimation there are more cold-regulated genes in SW (1978 genes

up-regulated by cold; 3205 genes down-regulated by cold; Fig 3.5) than in IT (1552 genes up-regulated by cold; 1796 genes down-regulated by cold; Fig 3.6). Based on this result Hannah et al. 2006 would hypothesize that SW has a greater capacity to cold acclimate than IT. This hypothesis is substantiated in electrolyte leakage assays performed in this study. SW has greater freezing tolerance than IT with 2 weeks of cold-acclimation (Fig. 3.2).

The number of gene expression changes from 1 to 2 weeks of cold-acclimation are also consistent with the 'gene number to cold-acclimation correlation' hypothesis by Hannah et al. 2006. Electrolyte leakage assays show that SW increases in freezing tolerance from 1 to 2 weeks of cold-acclimation (Fig 3.1 and 3.2), and the number of gene expression changes in SW also increases from 2664 cold-regulated genes at 1 week of cold-acclimation to 5183 cold-regulated genes at 2 weeks of cold-acclimation (Fig 3.5). Furthermore, IT decreases in freezing tolerance from 1 to 2 weeks of cold-acclimation and the number of gene expression changes also decreases from 4011 cold-regulated genes at 1 week of cold-acclimation to 3348 cold-regulated genes at 2 weeks of cold-acclimation.

Unfortunately, the 'gene number to cold-acclimation correlation' hypothesis by Hannah et al. 2006 is not infallible and is inconsistent with gene expression changes at 1 week of cold-acclimation. At 1 week of cold-acclimation there are 2664 cold-regulated genes in SW (1342 genes up-regulated by cold; 1322 genes down-regulated by cold; Fig 3.5), and 4011 cold-regulated genes in IT (1746 genes up-regulated by cold; 2265 genes down-regulated by cold; Fig 3.6). According to Hannah et al. 2006, this would

suggest that IT has a greater capacity to cold-acclimate than SW. But electrolyte leakage assays determined that SW and IT have similar freezing tolerance with 1 week of cold-acclimation (Fig 3.1).

Altogether, SW cold-regulated genes appear to be more enriched in genes previously associated with increases in freezing tolerance than IT cold-regulated genes. The quantity of cold-regulated genes in SW and IT, also suggests that there could be differences their capacity to cold-acclimate. Therefore in the next subsections of this study SW and IT cold-regulated genes will be compared in more detail. Since both SW and IT have similar freezing tolerance after 1 week of cold-acclimation, a more detailed comparison of transcriptomes at this time-point may reveal if SW and IT achieve this level of freezing tolerance by similar or divergent mechanisms.

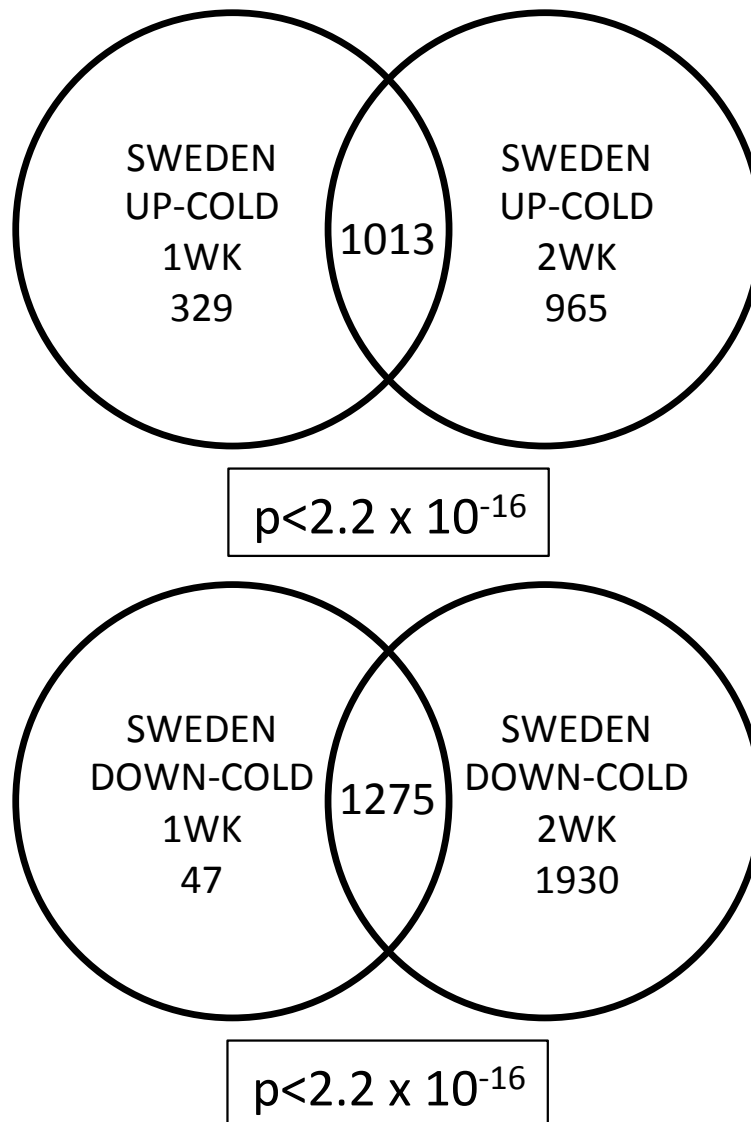


Figure 3.5 Cold-responsive genes in SW. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 or 2 weeks under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values represent overlap between differentially expressed cold-responsive genes at 1 and 2 weeks of cold-acclimation (calculated using a hypergeometric distribution in R).

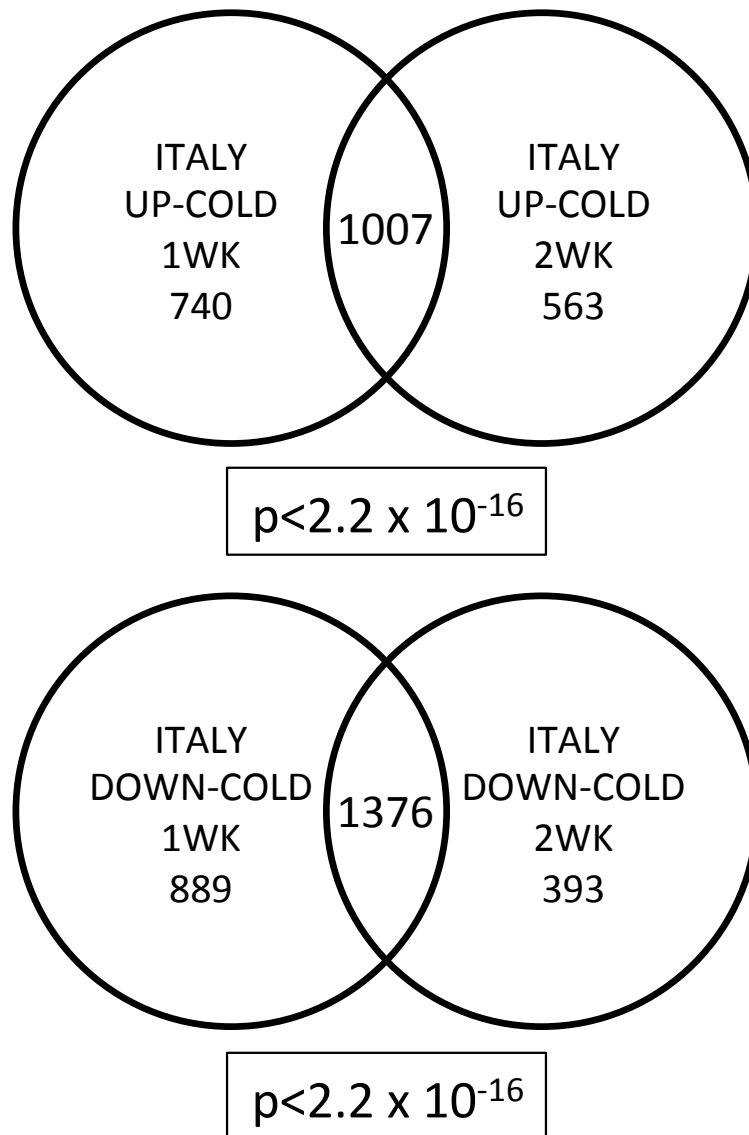


Figure 3.6. Cold-responsive genes in IT. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 or 2 weeks under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values represent overlap between differentially expressed cold-responsive genes at 1 and 2 weeks of cold-acclimation (calculated using a hypergeometric distribution in R).

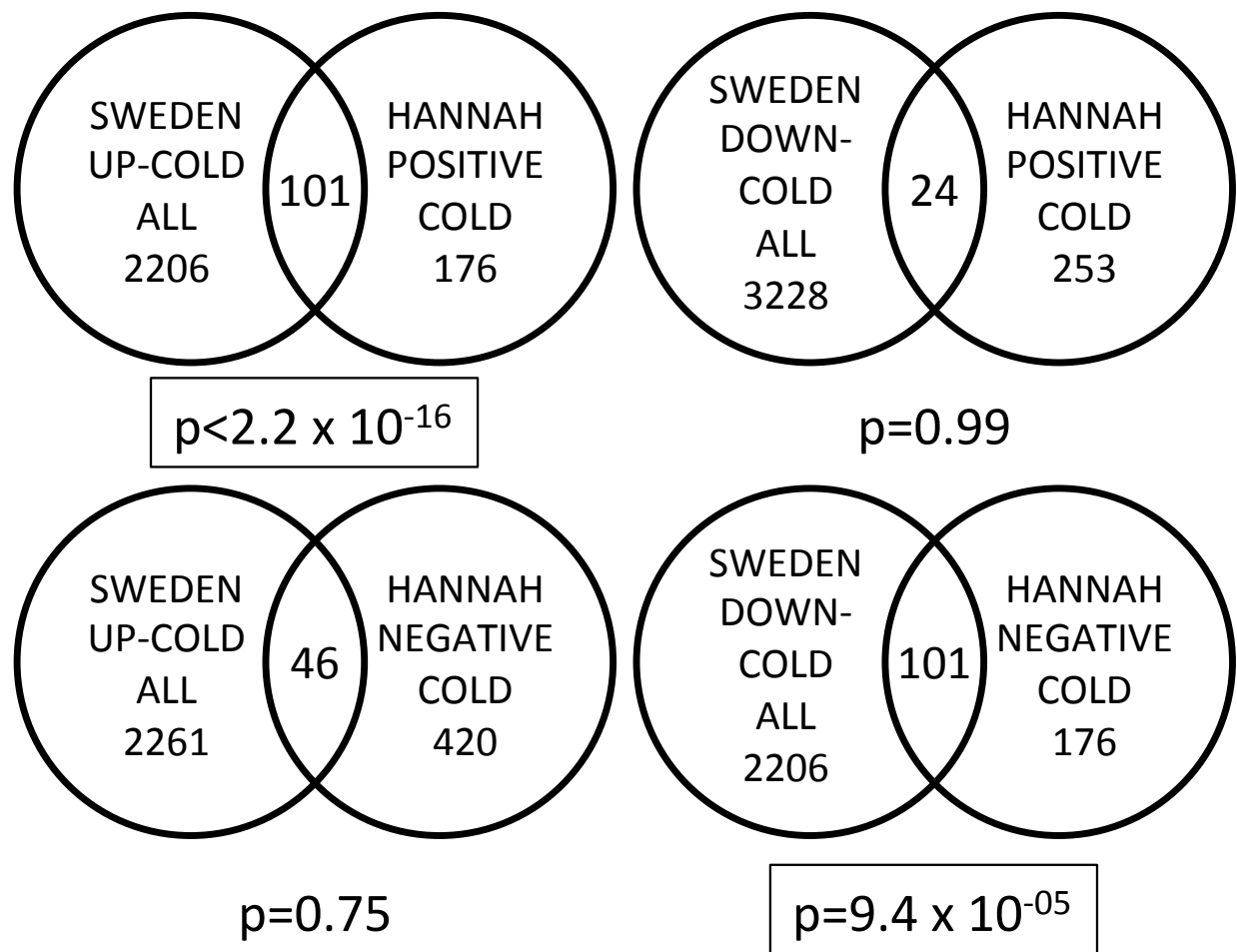


Figure 3.7. Overlap between the SW cold-responsive genes set and the Hannah genes set (6). Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 Hannah genes positively correlated with freezing tolerance and 466 Hannah genes negatively correlated with freezing tolerance; (6)).

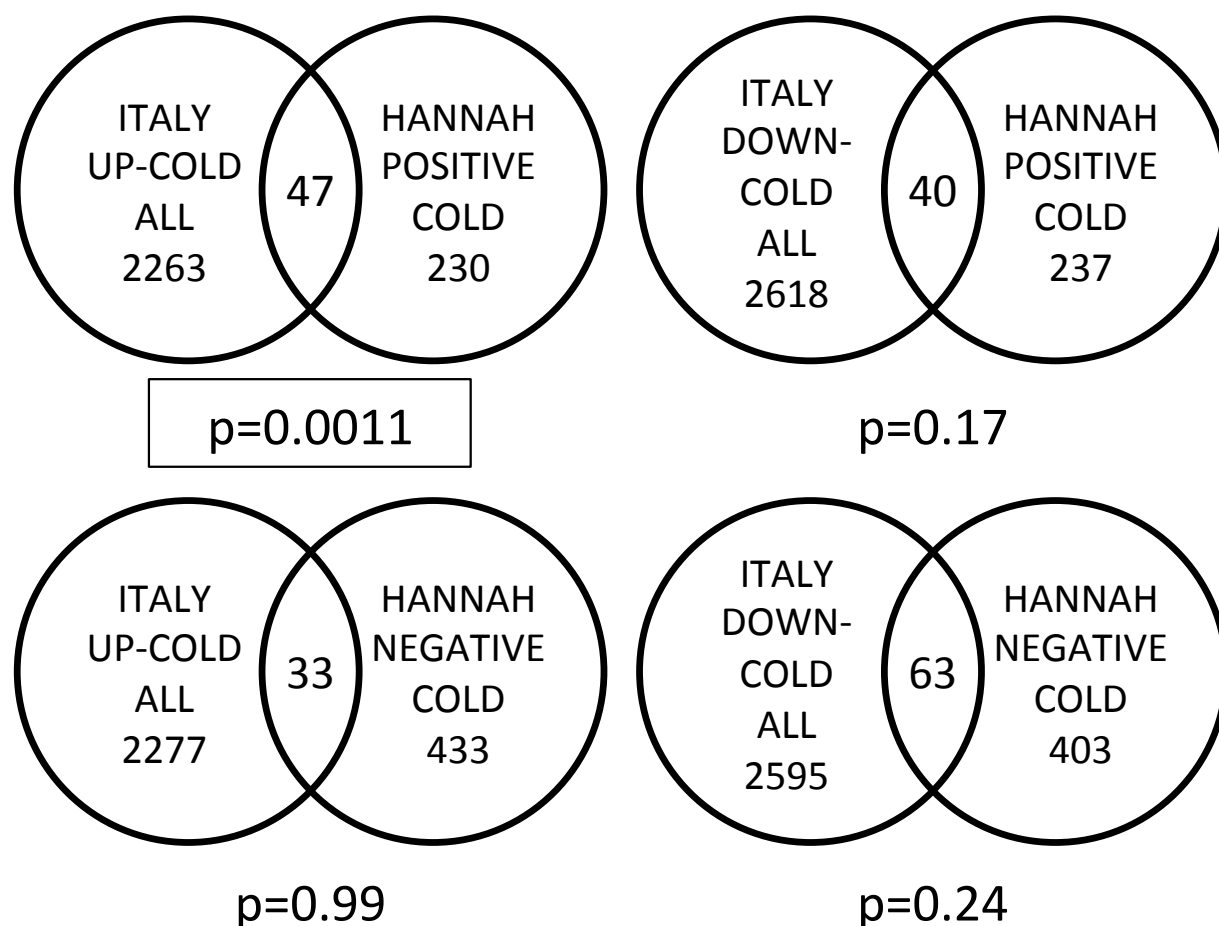


Figure 3.8. Overlap between the IT cold-responsive genes set and the Hannah genes set (6). Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 Hannah genes positively correlated with freezing tolerance and 466 Hannah genes negatively correlated with freezing tolerance; (6)).

SW AND IT TRANSCRIPTOMES AT 1 WEEK OF COLD-ACCLIMATION.

Summary: SW and IT transcriptomes at 1 week of cold-acclimation. SW and IT have similar freezing tolerance after 1 week of cold-acclimation. Therefore, the SW and IT cold-response at 1 week of cold-acclimation was examined to determine if this level of freezing tolerance is attained by similar transcriptional changes. Although there are a significant number of genes that respond similarly to cold in SW and IT, the majority of cold-responsive genes are differentially regulated, either in terms of induction pattern (up-regulated by cold / down-regulated by cold / not regulated by cold) or in terms of expression level (FPKM). Furthermore, these differentially expressed cold-regulated genes are enriched in genes that have previously been correlated with cold-acclimation. Analysis of RNA-seq data also suggests that there may be differences in the regulation of the CBF pathway of cold-acclimation between SW and IT. Overall, data suggests that SW and IT may reach similar levels of freezing tolerance through significantly different transcriptional changes at 1 week of cold-acclimation.

SW and IT transcriptomes at 1 week of cold-acclimation: Analysis of non-overlapping cold-regulated genes. SW and IT have similar freezing tolerance after 1 week of cold-acclimation. Therefore, the SW and IT cold-response at 1 week of cold-acclimation was examined to determine if this level of freezing tolerance is attained by similar transcriptional mechanisms. First, for a gene to ‘respond similarly’ to cold it must have a similar induction pattern (increase in response to cold or decrease in response to cold). Even if a gene ultimately has a similar level of expression at 1 week of cold-

acclimation despite dissimilar induction patterns (i.e. the gene is constitutively on in one ecotype) the change in induction would indicate altered regulation of that gene.

There are a significant number of genes commonly up-regulated (734 genes) and down-regulated (946 genes) by cold between SW and IT with 1 week of cold-acclimation (Fig. 3.9). But there are also 608 genes only up-regulated by cold in SW and 1012 genes only up-regulated by cold in IT at 1 week of cold-acclimation (Fig. 3.9). There are also 376 genes only down-regulated by cold in SW and 1319 genes only down-regulated by cold in IT at 1 week of cold-acclimation (Fig. 3.9). To determine if SW and IT use similar transcriptional mechanisms to attain similar levels of freezing tolerance, the cold-genes, which are uniquely expressed in SW or IT, were first examined. The genes with overlapping induction patterns in SW and IT were next examined to determine the genes that have significantly different levels of expression. Lastly, genes that do appear to respond similarly to cold in SW and IT, in terms of induction pattern and expression level, were further analyzed.

It is possible that cold-responsive genes that are differentially regulated between SW and IT do not play a role in increases in freezing tolerance associated with cold-acclimation. To appraise if non-overlapping cold-responsive genes have been previously associated with cold-acclimation enrichment of Hannah genes (6), which have been previously positively and negatively correlated with increases in freezing tolerance associated with cold-acclimation, was tested (Fig. 3.10). The 608 genes uniquely up-regulated by cold in SW at 1 week of cold-acclimation are enriched (55 genes overlapping) in Hannah genes that are positively correlated with freezing

tolerance (Fig. 3.10). Of the 55 overlapping Hannah genes, 20 are greater than 4-fold induced by cold in SW, 33 are greater than 2-fold induced and only 2 are less than 2-fold induced (Fig. 3.11). This suggests that genes uniquely up-regulated by cold in SW may contribute to cold-acclimation in some way. The 608 genes only up-regulated in SW are also enriched (56 genes; $p < 2.2E-16$) in genes up-regulated by the CBFs (CBF regulon; (25)). The CBFs are central components of the most well described pathway of cold-acclimation. Since these CBF regulon genes are only expressed in SW it suggests that there might be some difference in the CBF pathway of cold-acclimation between SW and IT. 'Response to cold' and other related abiotic stress GO categories are also enriched in the 608 genes only up-regulated in SW (Table 3.4). Interestingly, the 1012 genes that are exclusively up-regulated by cold in IT at 1 week of cold-acclimation are not enriched in Hannah genes (Fig. 3.10) or in CBF regulon genes. 'Response to cold' or other related abiotic stress categories are also markedly absent from the GO terms enriched in the 1012 genes only up-regulated by cold in IT (Table 3.5).

The 376 genes that are uniquely down-regulated by cold in SW are enriched (40 genes) in Hannah genes that negatively correlate with freezing tolerance (Fig 3.12). Of the 40 overlapping Hannah genes, 5 are greater than 4-fold reduced by cold, 30 of these genes are greater than 2-fold reduced and 5 are less than 2-fold reduced (Fig 3.13). For these 376 genes there are no biological process GO categories significantly enriched above the threshold (FDR p -value ≤ 0.050) but molecular function GO categories enriched include 'water transport' (Table 3.6). The 1319 genes only down-regulated by cold in IT are not significantly enriched (5 genes) in Hannah genes

negatively correlated with freezing tolerance (Fig. 3.12). Remarkably, the 1319 genes only down-regulated in IT are enriched (32 genes) in Hannah genes that are positively correlated with increases in freezing tolerance (Fig. 3.12). Of the 32 overlapping Hannah genes, 6 are greater than 4-fold reduced by cold, 20 are greater than 2-fold reduced 6 are less than 2-fold reduced (Fig. 3.14). Furthermore, these 1319 gene that are only down-regulated by cold in IT are enriched (23 genes; $p=0.021$) in genes up-regulated by the CBFs (25). This indicates that CBF regulon genes may be uniquely repressed in IT and again suggests that the CBF pathway of cold-acclimation may be regulated differently between SW and IT. For the 1319 genes only down-regulated in IT, 'defense response' related GO categories such as 'innate immune response' and 'response to biotic stimulus' are significantly enriched (Table 3.7). This is interesting considering that 'defense response' was also enriched in genes with higher expression in IT under basal conditions (Table 3.3).

Altogether, analysis of RNA-seq data at 1 week of cold-acclimation indicates that genes uniquely cold-regulated in SW are enriched in genes previously associated with cold acclimation. This enrichment of genes correlated with cold-acclimation in cold-regulated genes uniquely expressed SW is curious considering that there is no significant difference in freezing tolerance between SW and IT at 1 week of cold acclimation (Fig. 3.1). Therefore cold-regulated genes uniquely expressed in IT at 1 week of cold-acclimation may potentially be part of pathways of cold-acclimation currently not well described allowing IT to achieve a similar level of freezing tolerance to SW at 1 week of cold-acclimation (Fig 3.1). Differences in gene expression at 1 week of

cold-acclimation may also take time to manifest into differences in freezing tolerance (Fig 3.2). Consequently genes differentially regulated between SW and IT at 1 week of cold-acclimation, which remain differentially regulated at 2 weeks of cold-acclimation, are included in the list of genes that potentially contribute to differences in freezing tolerance. Genes that potentially contribute to differences in freezing tolerance will be described in more detail in the next subsection of RNA-seq analysis but first genes with similar cold induction patterns but significantly different levels of expression will be examined.

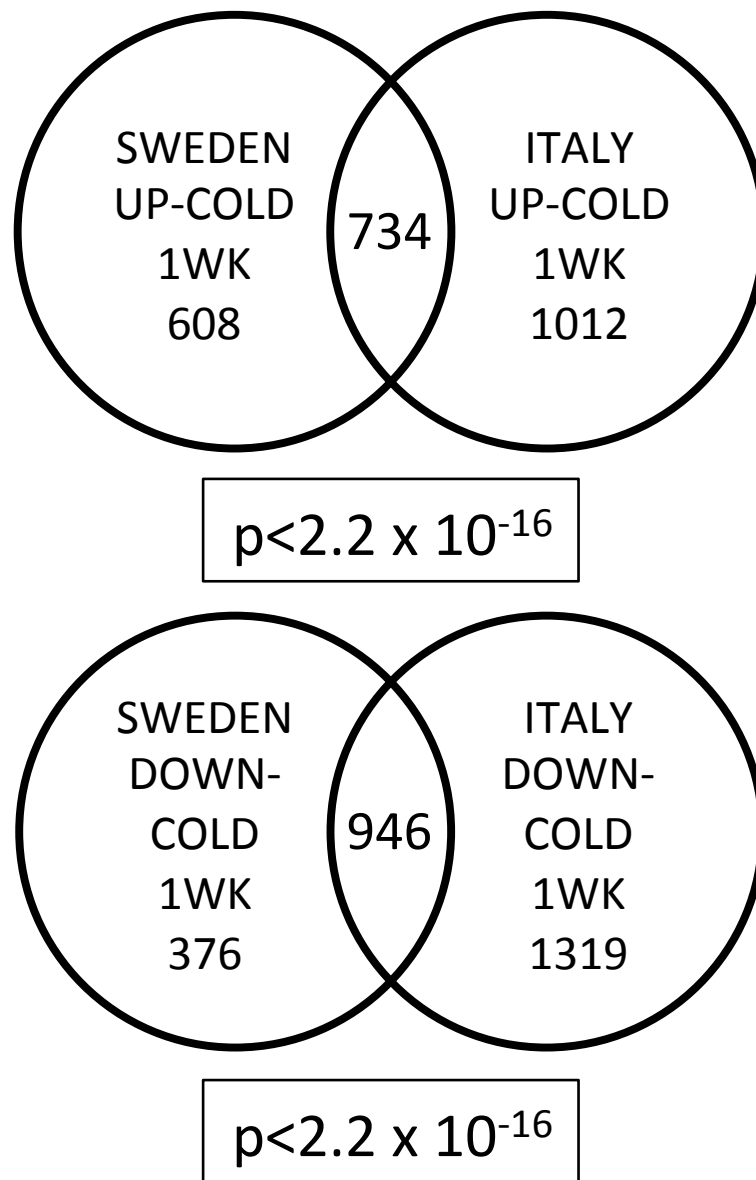


Figure 3.9. Overlap between SW and IT response at 1 week of cold-acclimation. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values represent overlap between SW and IT cold-responsive genes (calculated using a hypergeometric distribution in R).

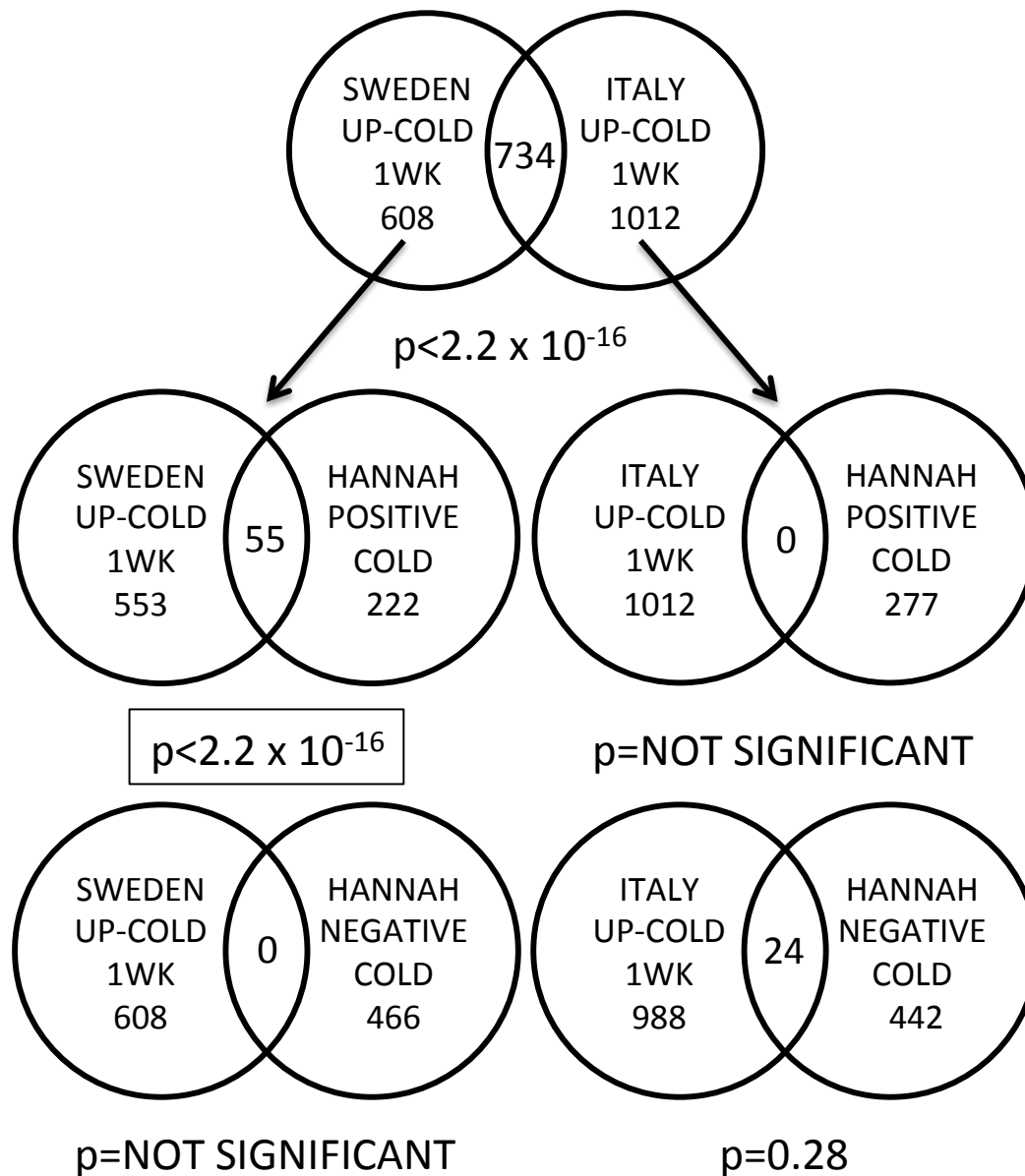


Figure 3.10. Overlap between genes uniquely up-regulated in SW or IT at 1 week of cold-acclimation and the Hannah genes set. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 genes positively correlated with freezing tolerance and 466 genes negatively correlated with freezing tolerance; (6)).

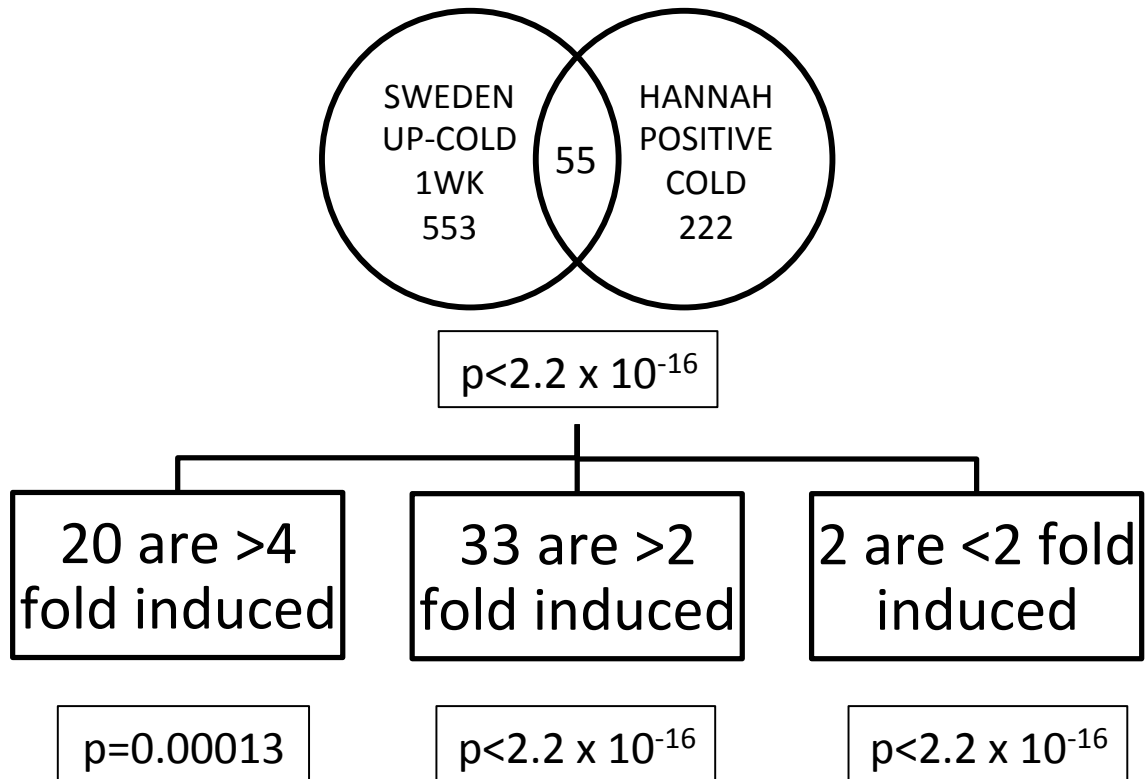


Figure 3.11. Fold change of 55 Hannah genes, which overlap with 608 genes only up-regulated in SW at 1 week of cold-acclimation. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-offs were applied (from left to right).

Table 3.4. GO categories significantly enriched in 608 genes only up-regulated in SW at 1 week of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). ‘GO TERM CATEGORY’ denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. ‘# HITS’ denotes the number of genes with that GO category in the 608 genes only up-regulated in SW at 1 week of cold-acclimation. ‘# GENES IN GO’ denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	SW ONLY UP-COLD (608 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Response To Organic Substance	66	1176	1.70E-07
BP	Response To Endogenous Stimulus	59	975	2.24E-07
BP	Response To Ethylene Stimulus	22	256	2.96E-04
BP	Secondary Metabolic Process	29	423	3.36E-04
BP	Phenylpropanoid Biosynthetic Process	15	128	4.21E-04
BP	Ethylene Mediated Signaling Pathway	17	170	4.63E-04
BP	Response To Abiotic Stimulus	56	1197	5.27E-04
BP	Response To Jasmonic Acid Stimulus	16	156	6.27E-04
BP	Response To Absciscic Acid Stimulus	22	285	6.41E-04
BP	Response To Water	17	185	9.88E-04
BP	Phenylpropanoid Metabolic Process	16	166	9.97E-04
BP	Response To Cold	19	233	1.16E-03
BP	Response To Temperature Stimulus	24	354	1.33E-03
BP	Response To Water Deprivation	16	176	1.59E-03
MF	UDP-Glucosyltransferase Activity	13	119	3.36E-03
MF	Glutathione Transferase Activity	9	49	3.57E-03
BP	Response To Oxidative Stress	20	287	4.32E-03
BP	Two-Component Signal Transduction	17	218	4.42E-03
BP	Amino Acid Derivative Biosynthetic Process	16	194	4.55E-03
BP	Toxin Metabolic Process	8	46	4.92E-03
MF	Quercetin 3-O-Glucosyltransferase Activity	7	23	4.94E-03
MF	Transcription Regulator Activity	72	1913	5.96E-03
BP	Hormone-Mediated Signaling	25	432	7.29E-03
BP	Cellular Response To Hormone Stimulus	25	432	7.29E-03
BP	Carbohydrate Transport	10	83	7.33E-03
MF	Glucosyltransferase Activity	13	137	8.06E-03
MF	Transcription Factor Activity	64	1678	8.12E-03
BP	Response To Wounding	13	147	9.93E-03
BP	Response To Chitin	12	127	1.01E-02
MF	Solute: Cation Symporter Activity	12	126	1.15E-02
BP	Response To Carbohydrate Stimulus	15	199	1.31E-02

Table 3.4 (cont'd)

BP	Response To Inorganic Substance	28	540	1.32E-02
BP	Regulation Of Transcription	73	2024	2.14E-02
BP	Aromatic Compound Biosynthetic Process	15	215	2.58E-02
CC	Vacuole	30	643	2.80E-02
MF	Sugar: Hydrogen Symporter Activity	10	99	2.81E-02
BP	Negative Regulation Of ABA Signaling	5	19	2.83E-02
MF	Symporter Activity	12	146	3.14E-02
CC	Plasma Membrane	76	2228	3.85E-02
BP	Flavonoid Biosynthetic Process	7	53	4.65E-02

Table 3.5. GO categories significantly enriched in 1012 genes only up-regulated in IT at 1 week of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in the 1012 genes only up-regulated in IT at 1 week of cold-acclimation. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	IT ONLY UP-COLD (1012 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
MF	Structural Constituent Of Ribosome	147	394	1.30E-97
CC	Non-Membrane-Bounded Organelle	232	1144	6.95E-96
CC	Cytosolic Part	126	271	1.09E-95
CC	Ribosomal Subunit	128	294	1.94E-93
CC	Nucleolus	103	332	3.13E-57
CC	Cytosol	145	708	1.01E-56
CC	Membrane-Enclosed Lumen	134	667	4.44E-51
BP	Ribosome Biogenesis	84	242	1.21E-50
CC	Nuclear Lumen	113	472	1.54E-50
CC	Intracellular Organelle Lumen	132	659	3.72E-50
BP	Translation	173	1231	3.19E-46
BP	RNA Processing	72	436	4.78E-21
MF	RNA Binding	128	1351	5.49E-16
CC	Mitochondrion	114	1349	1.58E-10
MF	Purine NTP-Dependent Helicase Activity	26	113	8.97E-10
MF	ATP-Dependent Helicase Activity	26	113	8.97E-10
MF	Translation Factor Activity	27	149	7.50E-08
CC	Plastid	205	3259	1.76E-07
CC	Chloroplast	197	3192	1.67E-06
CC	Cell Wall	52	611	1.06E-04
MF	ATPase Activity	44	467	1.22E-04
CC	External Encapsulating Structure	52	620	1.51E-04
BP	Embryonic Development, Seed Dormancy	37	372	3.10E-04
BP	RNA Modification	20	133	3.47E-04
MF	Macromolecule Transmembrane Transporter	12	51	3.83E-04
BP	Chromatin Assembly	14	71	6.54E-04
CC	Plasma Membrane	135	2228	7.25E-04
BP	Nucleosome Assembly	13	69	1.94E-03
BP	Seed Development	38	432	2.29E-03
BP	Mitochondrion Organization	9	32	2.48E-03
MF	RNA Methyltransferase Activity	8	26	3.05E-03

Table 3.5 (cont'd)

CC	Small Nucleolar Ribonucleoprotein Complex	14	100	3.83E-03
BP	Cellular Macromolecular Complex Assembly	23	208	3.93E-03
BP	Nucleotide Biosynthetic Process	20	170	4.66E-03
MF	Pseudouridine Synthase Activity	7	20	4.81E-03
BP	Fruit Development	38	452	4.87E-03
BP	Nitrogen Compound Biosynthetic Process	41	506	4.87E-03
BP	Pseudouridine Synthesis	7	21	6.82E-03
BP	Response To Cadmium Ion	29	327	1.01E-02
MF	DNA-Directed RNA Polymerase Activity	11	63	1.02E-02
CC	Vacuole	46	643	1.22E-02
MF	Exonuclease Activity	8	34	1.52E-02
BP	Regulation Of Macromolecule Biosynthetic Process	11	69	1.68E-02
MF	RNA Polymerase Activity	11	69	1.91E-02
CC	CUL4 RING Ubiquitin Ligase Complex	14	122	1.98E-02
MF	Ribonuclease Activity	12	84	2.26E-02
BP	Purine Nucleotide Metabolic Process	16	140	2.28E-02
MF	GTPase Activity	13	97	2.36E-02
BP	Protein Folding	23	251	2.52E-02
BP	Translational Initiation	11	74	2.60E-02
MF	Nuclease Activity	19	188	2.80E-02
BP	Protein Localization In Organelle	11	76	3.11E-02
BP	Negative Regulation Of DNA Replication	4	6	3.19E-02
MF	3'-5'-Exoribonuclease Activity	7	30	3.29E-02
MF	Intramolecular Transferase Activity	9	53	3.70E-02
BP	Response To Metal Ion	30	384	4.14E-02
CC	Nucleosome	9	63	4.20E-02
BP	Pyrimidine Nucleotide Metabolic Process	6	22	4.23E-02
MF	GTP Binding	23	263	4.29E-02
BP	Regulation Of DNA Replication	5	14	4.61E-02
MF	Exoribonuclease Activity	7	33	4.84E-02

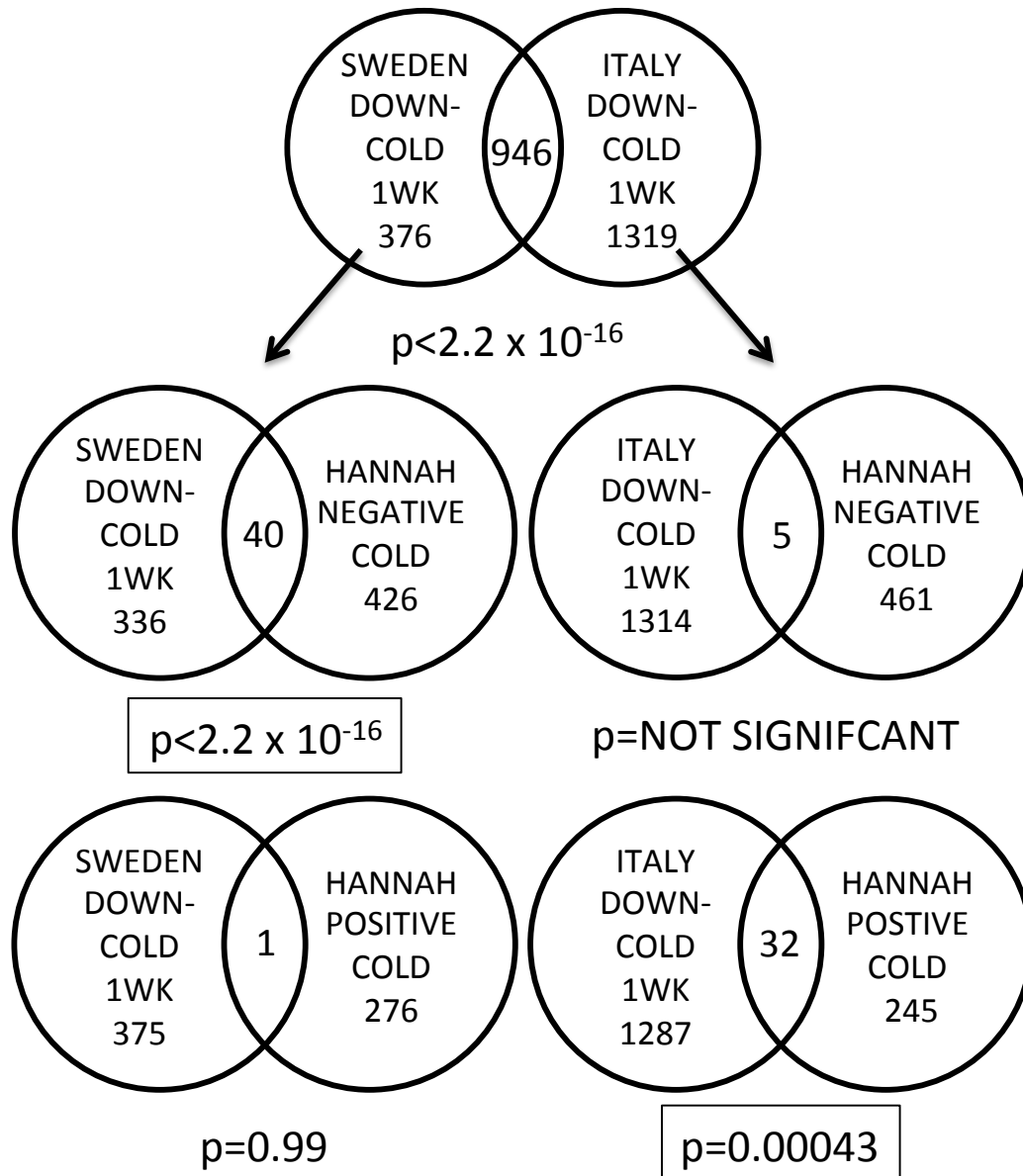


Figure 3.12. Overlap between genes uniquely down-regulated in SW or IT at 1 week of cold-acclimation and the Hannah genes set. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 genes positively correlated with freezing tolerance and 466 genes negatively correlated with freezing tolerance; (6)).

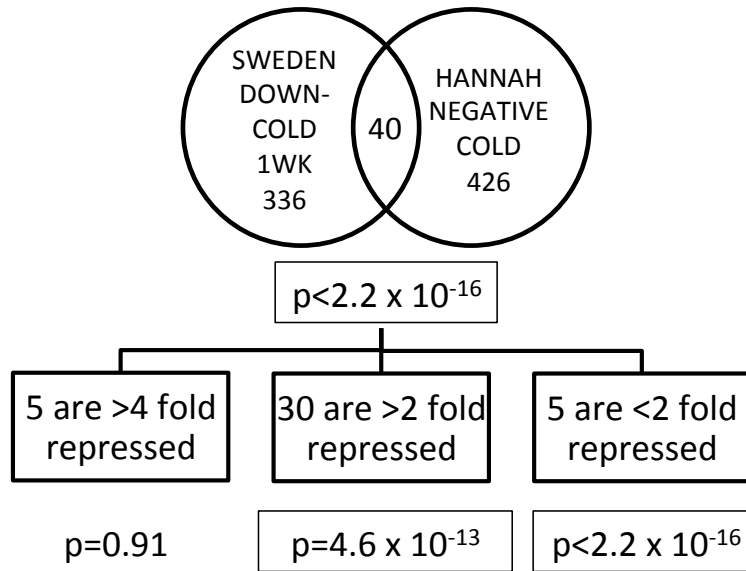


Figure 3.13. Fold change of 40 Hannah genes that overlap with 376 genes only down-regulated in SW at 1 week of cold-acclimation. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-offs were applied (from left to right).

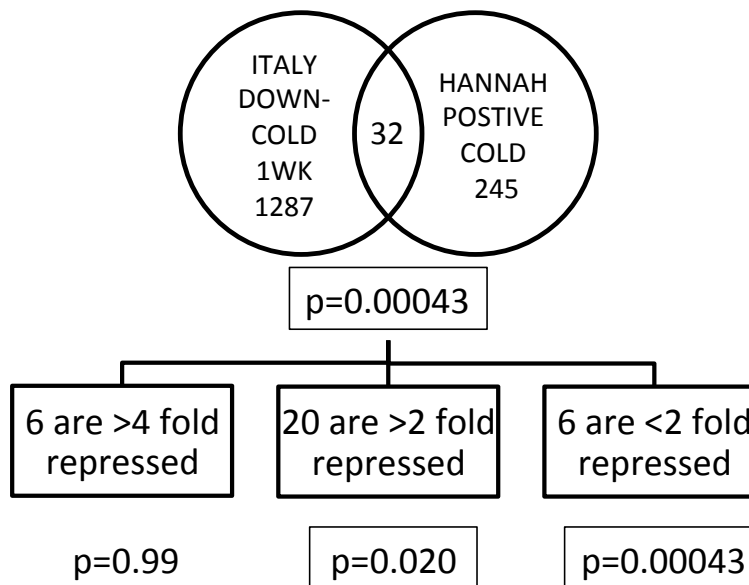


Figure 3.14. Fold change of 32 Hannah genes that overlap with 1319 genes only down-regulated in IT at 1 week of cold-acclimation. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-offs were applied (from left to right).

Table 3.6. GO categories significantly enriched in 376 genes only down-regulated in SW at 1 week of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in the 376 genes only down-regulated in SW at 1 week of cold-acclimation. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	SW ONLY DOWN-COLD (376 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
CC	Anchored To Plasma Membrane	9	66	2.08E-03
CC	Anchored To Membrane	18	320	2.44E-03
CC	External Encapsulating Structure	26	620	2.70E-03
CC	Cell Wall	25	611	4.29E-03
MF	Channel Activity	11	126	4.76E-03
MF	Passive Transmembrane Transporter Activity	11	126	4.76E-03
CC	Apoplast	18	382	8.16E-03
CC	Intrinsic To Plasma Membrane	9	103	8.33E-03
MF	Substrate Specific Channel Activity	11	125	8.87E-03
CC	Intrinsic To Membrane	66	2658	2.21E-02
MF	Water Transporter Activity	6	37	2.89E-02
MF	Water Channel Activity	6	37	2.89E-02
CC	Plasma Membrane	56	2228	4.16E-02

Table 3.7. GO categories significantly enriched in 1319 genes only down-regulated in IT at 1 week of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in 1319 genes only down-regulated in IT at 1 week of cold-acclimation. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	IT ONLY DOWN-COLD (1319 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Immune Response	50	293	1.06E-10
BP	Cell Death	38	254	2.26E-06
BP	Response To Organic Substance	102	1176	1.37E-05
BP	Apoptosis	28	172	2.85E-05
BP	Response To Chitin	21	127	9.75E-04
BP	Defense Response	84	1030	1.82E-03
BP	Response To Carbohydrate Stimulus	26	199	3.29E-03
CC	Intrinsic To Membrane	169	2658	1.79E-02
BP	Response To Wounding	20	147	1.89E-02
CC	CCAAT-Binding Factor Complex	6	13	2.50E-02
BP	Plant-Type Hypersensitive Response	10	45	3.97E-02
BP	Amine Transport	12	67	4.65E-02
BP	Protein Ubiquitination	17	125	4.83E-02

SW and IT transcriptomes at 1 week of cold-acclimation: Analysis of cold-induced genes with overlapping induction patterns but different levels of expression. Though there are 1620 genes exclusively up-regulated by cold in either SW or IT there are also a significant number of genes commonly up-regulated by cold (734 genes) between SW and IT (Fig. 3.9). There are also a significant number of genes commonly down-regulated by cold (946 genes) in addition to the 1695 genes exclusively down-regulated in either SW or IT (Fig. 3.9). The 734 commonly up-regulated and 946 commonly down-regulated genes have similar cold-induction patterns and thus met the first requirement of 'responding similarly' (Fig. 3.9). The second measure for 'responding similarly' was that the gene must have a similar level of expression between SW and IT. Genes must have similar levels of expression in order to 'respond similarly' because statistically significant variances would again indicate that the gene has altered regulation between SW and IT. Statistically significant differences in expression level (FPKM) were measured by a FDR corrected p-value of ≤ 0.050 .

A significant number of genes (105 genes out of 734) commonly up-regulated by cold in SW and IT have different induction levels (Fig. 3.15). There are 77 differentially expressed up-regulated genes with higher expression in SW and 28 with higher expression in IT (Fig 3.15). Again, these differentially expressed genes may not be associated with cold-acclimation. Therefore it was determined if these differentially expressed genes were enriched in Hannah genes that have been previously positively and negatively correlated with increases in freezing tolerance associated with cold-acclimation (6). The 77 genes with higher expression in SW are significantly enriched

(13 genes) in Hannah genes positively correlated with increases in freezing tolerance (Fig. 3.16). Of the 13 overlapping Hannah genes, 7 have greater than 4-fold higher expression in SW compared to IT, 5 are greater than 2-fold higher and 1 is less than 2-fold higher (Fig. 3.17). The 77 genes up-regulated by cold with higher expression in SW are enriched in cold-related GO terms such as 'response to cold', 'response to abiotic stimulus' and 'cold-acclimation' (Table 3.8). The 77 genes with higher expression in SW are also significantly enriched (14 genes; $p=1.3E-13$) in genes up-regulated by the CBFs (25). The 28 genes up-regulated by cold with higher expression in IT do not include any genes regulated by the CBFs. This suggests that SW and IT may differ in the CBF pathway of cold-acclimation, which is similar what was observed in genes that were uniquely cold regulated in either SW or IT. Furthermore, there is also no overlap between the 28 up-regulated genes with higher expression in IT and Hannah genes positively correlated with freezing tolerance (Fig. 3.16). These 28 genes are also not enriched in GO terms related to stress (Table 3.9).

A significant number of genes (105 out of 946 genes) commonly down-regulated by cold in SW and IT have significant differences in expression level (Fig. 3.18). There are 36 differentially expressed genes with higher expression in SW (more repressed in IT) and 69 with higher expression in IT (more repressed in SW; Fig 3.18). The 36 genes that are more repressed by cold in IT (higher expression in SW) do not significantly overlap with Hannah genes negatively correlated with increases in freezing tolerance or with CBF regulon genes (Fig. 3.19). These 36 genes do not have any statistically overrepresented GO terms. There is significant overlap (6 genes) between the 63

genes more repressed by cold in SW (higher in IT) and Hannah genes negatively correlated with freezing tolerance (Fig. 3.19). Of the 6 overlapping Hannah genes, 2 are greater than 4-fold repressed in SW compared to IT and 4 are greater than 2-fold repressed (Fig. 3.20). These 69 genes are not enriched in GO terms specifically related to cold but are enriched in GO terms such as 'response to endogenous stimulus' (Table 3.10). The 69 genes down-regulated by cold that are more repressed in SW (higher in IT) are also significantly enriched (4 genes; $p=0.0072$) in genes down-regulated by CBFs, again suggesting that SW and IT have differences in the CBF pathway of cold-acclimation.

Analysis of both non-overlapping cold genes and genes with common induction patterns but significantly different levels of expression suggests that the differentially expressed genes in SW are more enriched in genes previously correlated (Hannah genes) and associated (CBF regulon) with cold-acclimation. There is no difference in freezing tolerance between SW and IT at 1 week of cold-acclimation (Fig. 3.1). This suggests that IT may reach a similar level of freezing tolerance to SW through pathways of cold-acclimation not yet well defined. However, there are a significant number of genes with similar induction patterns and no significant difference in expression level (629 genes commonly up-regulated, and 841 genes commonly down-regulated). Next, these cold-regulated genes that respond similarly in SW and IT will be further analyzed to determine if they have been previously associated with cold-acclimation.

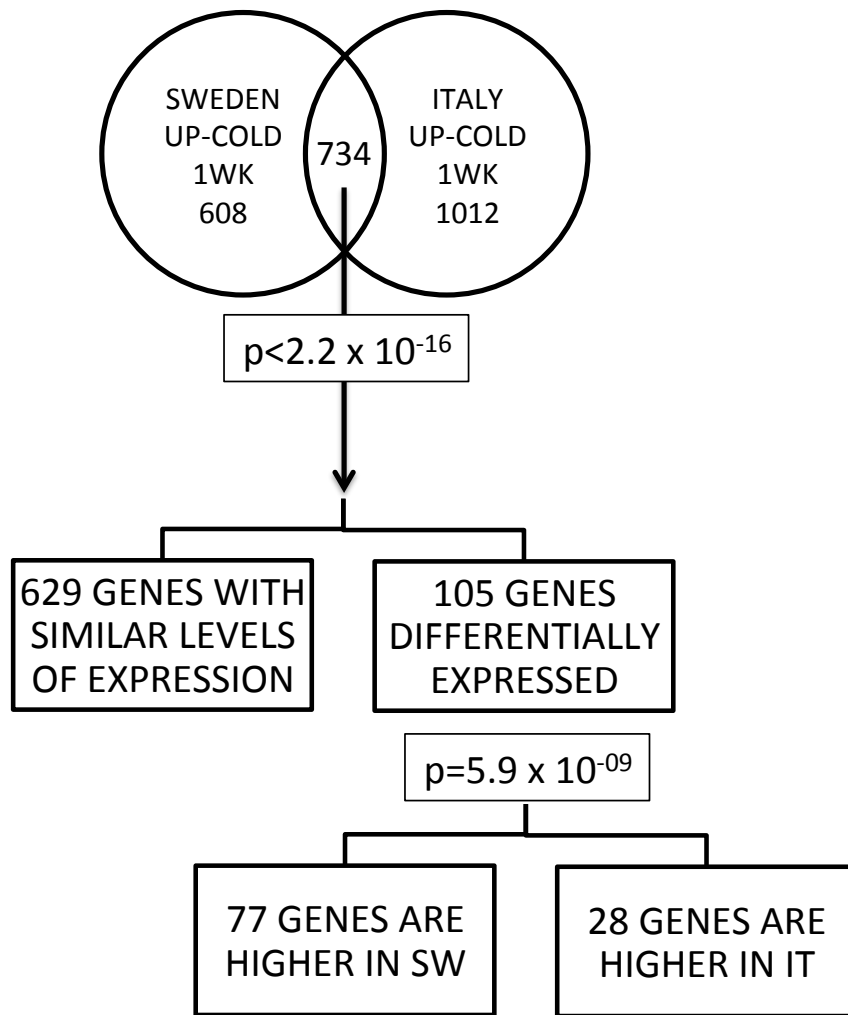


Figure 3.15. 105 genes commonly up-regulated by cold but significantly different in expression level. 77 genes are higher in expression in SW and 28 genes are higher in IT. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . Genes were considered differentially expressed between SW and IT if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values represent overlap between SW and IT cold-responsive genes (calculated using a hypergeometric distribution in R).

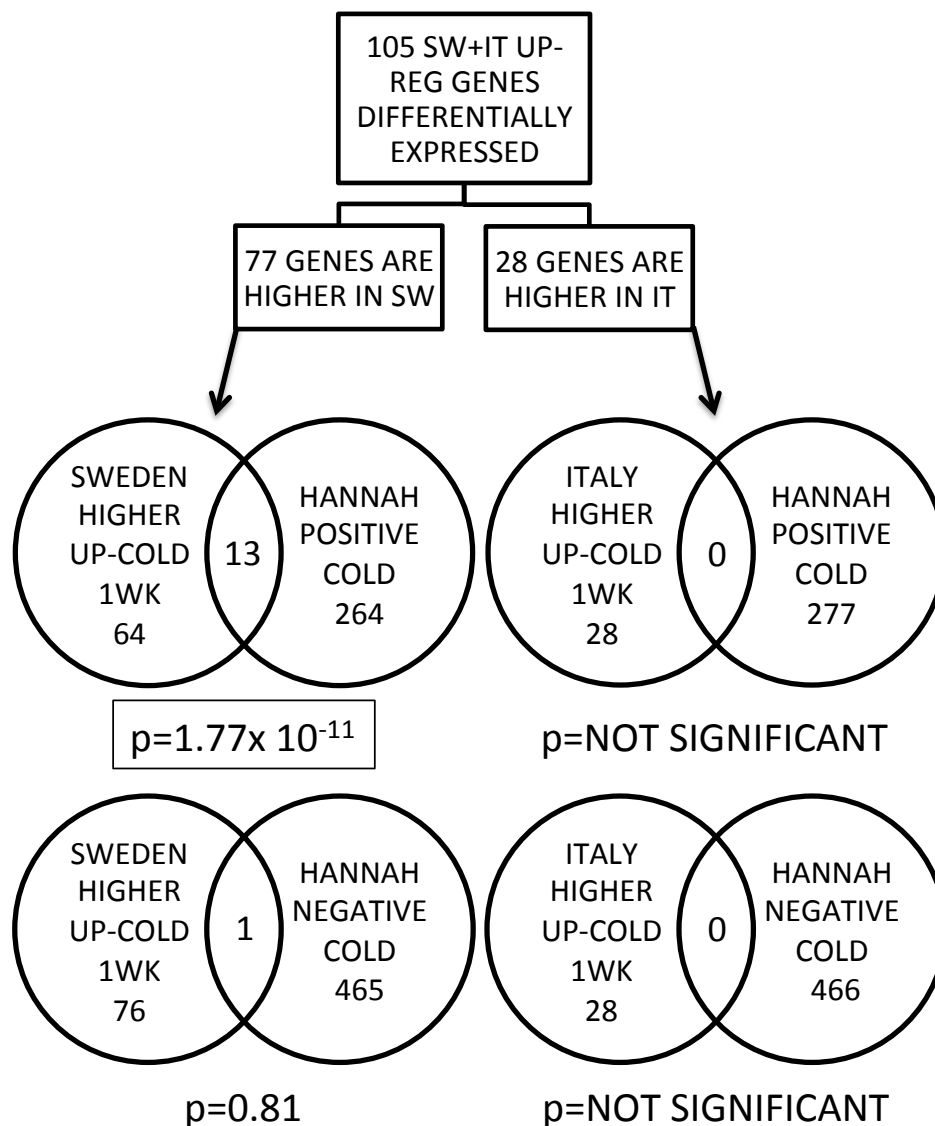


Figure 3.16. 105 genes commonly up-regulated by cold but significantly different in expression level and their overlap with the Hannah genes set. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . Genes were considered differentially expressed between SW and IT if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 genes positively correlated with freezing tolerance and 466 genes negatively correlated with freezing tolerance; (6)).

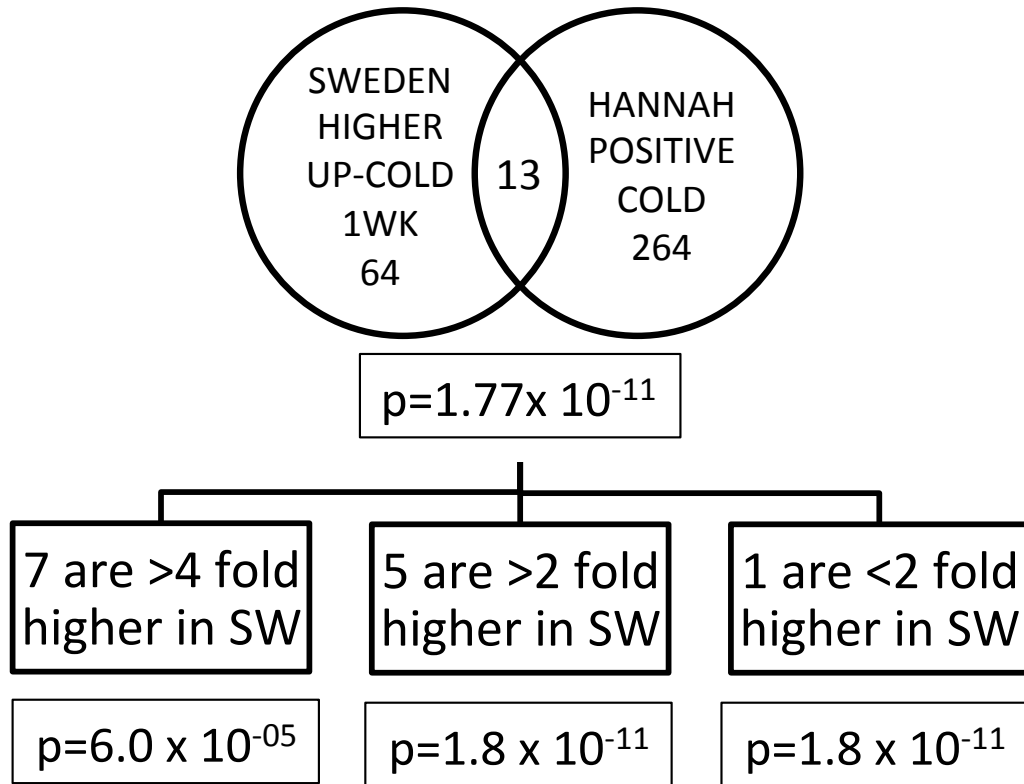


Figure 3.17. Fold change of 13 Hannah genes that overlap with 77 genes with up-regulated induction pattern, but higher expression level in SW. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-offs were applied (from left to right).

Table 3.8. GO categories significantly enriched in 77 genes with up-regulated induction pattern and higher expression level in SW at 1 week of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in 77 genes with up-regulated induction pattern and higher expression level in SW at 1 week of cold-acclimation. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	SW HIGHER UP-COLD (77 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Response To Temperature Stimulus	11	354	1.92E-04
BP	Response To Cold	9	233	3.22E-04
BP	Response To Abiotic Stimulus	17	1197	3.93E-04
BP	Response To Water	8	185	5.03E-04
BP	Response To Absciscic Acid Stimulus	9	285	5.66E-04
BP	Response To Water Deprivation	7	176	2.17E-03
BP	Oxidation Reduction	14	1186	1.46E-02
BP	Secondary Metabolic Process	8	423	3.21E-02
BP	Aromatic Compound Biosynthetic Process	6	215	3.50E-02
BP	Heat Acclimation	3	16	3.81E-02
BP	Response To Desiccation	3	18	4.38E-02
BP	Cold Acclimation	3	20	4.95E-02

Table 3.9. GO categories significantly enriched in 28 genes with up-regulated induction pattern and higher expression level in IT at 1 week of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in 28 genes with up-regulated induction pattern and higher expression level in IT at 1 week of cold-acclimation. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	IT HIGHER UP-COLD (28 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Ribonucleoprotein Complex Biogenesis	5	249	1.02E-02

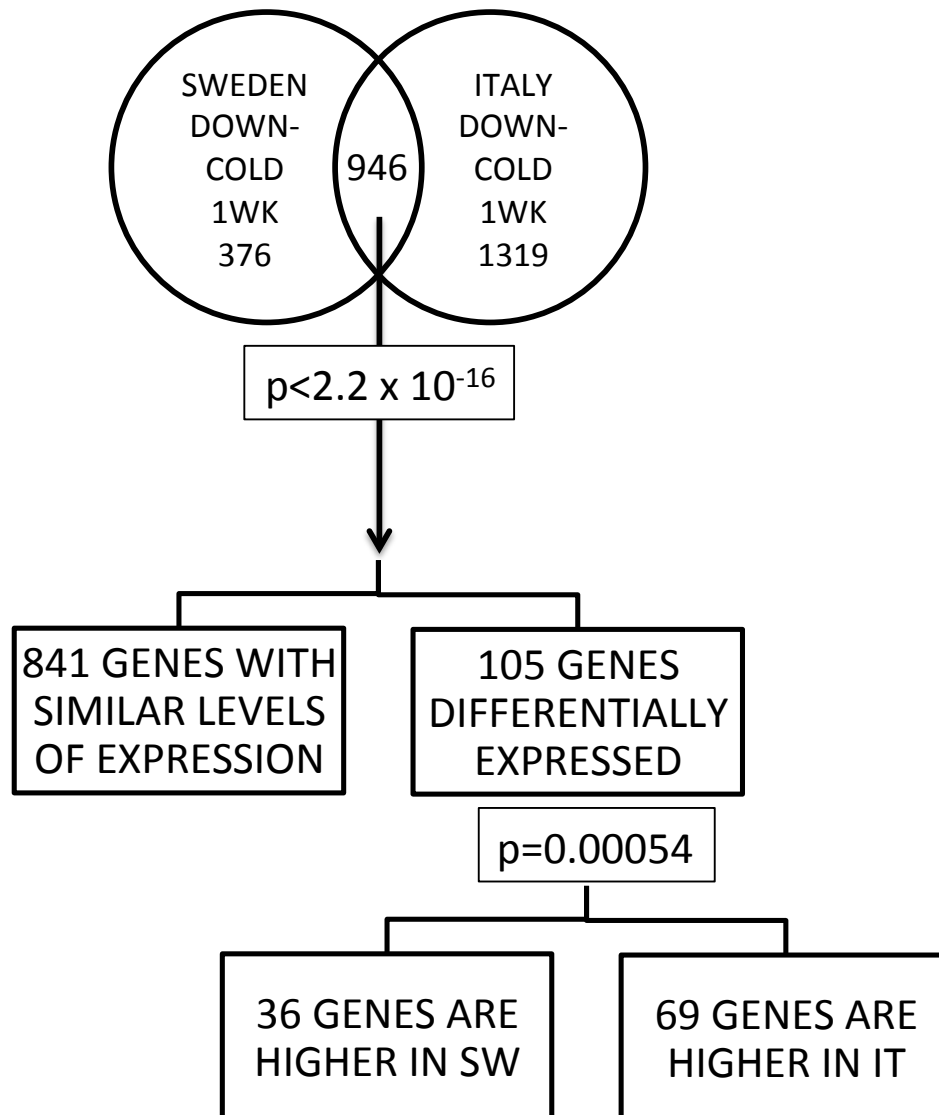


Figure 3.18. 105 genes commonly down-regulated by cold but significantly different in expression level. 36 genes more repressed in IT (higher in expression in SW) and 69 genes more repressed in SW (higher expression in IT). Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . Genes were considered differentially expressed between SW and IT if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values represent overlap between SW and IT cold-responsive genes (calculated using a hypergeometric distribution in R).

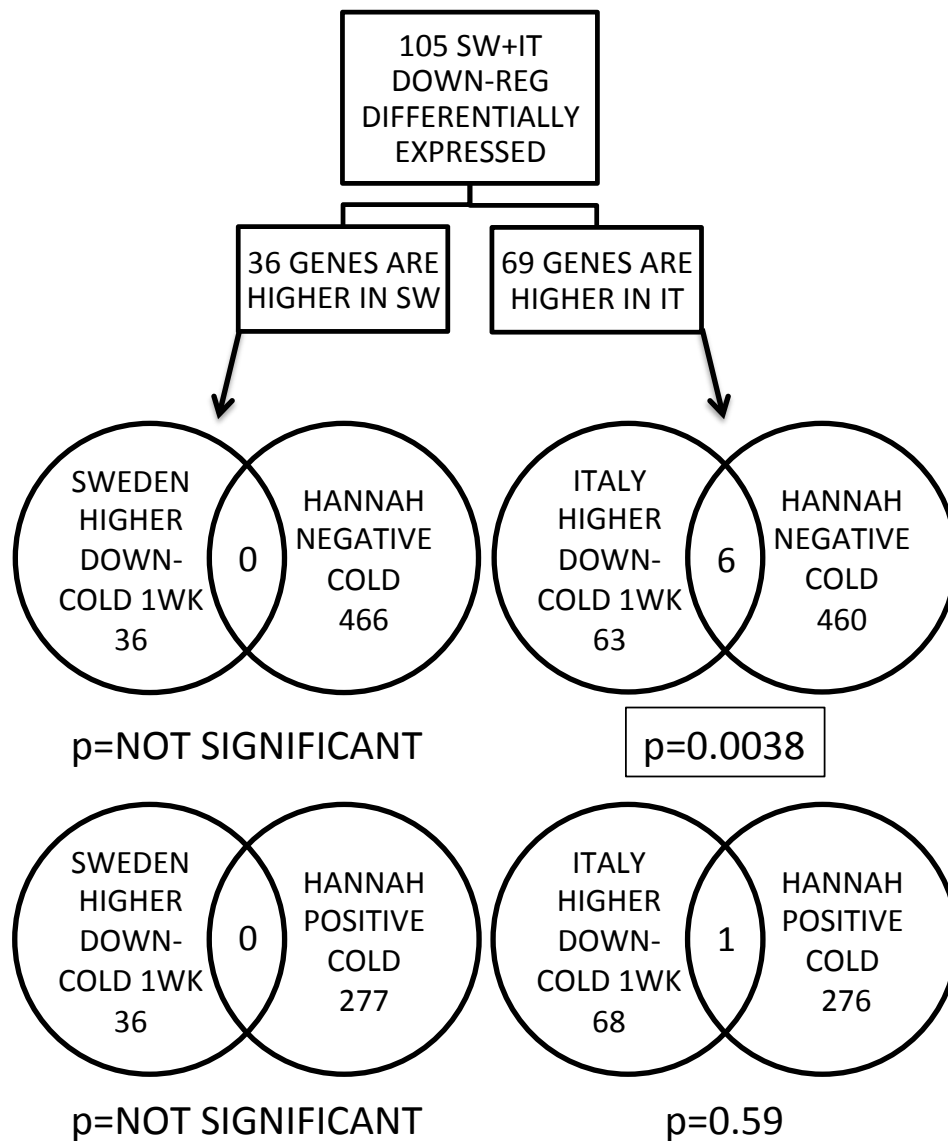


Figure 3.19. 105 genes commonly down-regulated by cold but significantly different in expression level and their overlap with the Hannah genes set. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . Genes were considered differentially expressed between SW and IT if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 genes positively correlated with freezing tolerance and 466 genes negatively correlated with freezing tolerance; (6)).

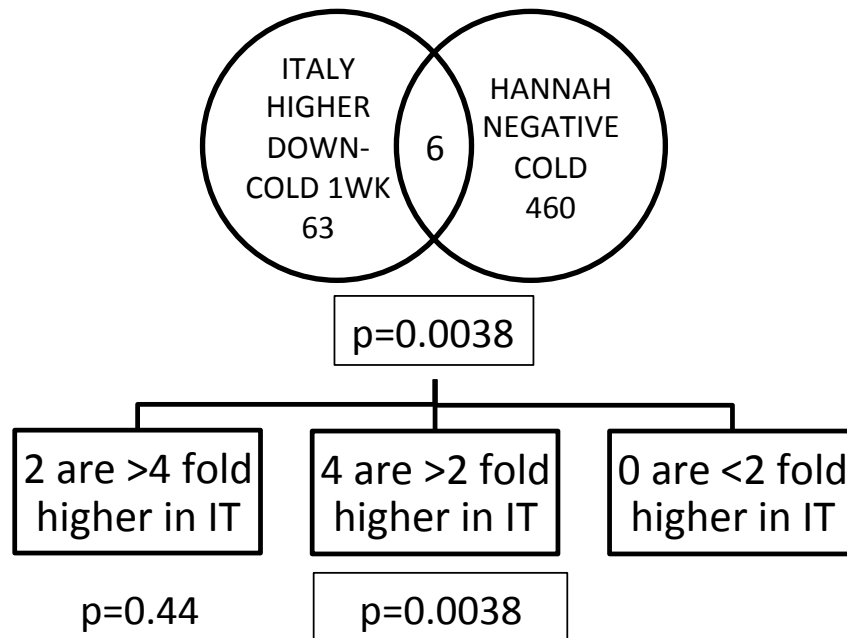


Figure 3.20. Fold change of 6 Hannah genes that overlap with 69 genes with down-regulated induction pattern, but higher expression level in IT. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-offs were applied (from left to right).

Table 3.10. GO categories significantly enriched in 69 genes with down-regulated induction pattern and higher expression level in IT at 1 week of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). ‘GO TERM CATEGORY’ denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. ‘# HITS’ denotes the number of genes with that GO category in 69 genes with down-regulated induction pattern and higher expression level in IT at 1 week of cold-acclimation. ‘# GENES IN GO’ denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	IT HIGHER UP-COLD (69 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Response To Organic Substance	19	1176	1.81E-06
BP	Response To Endogenous Stimulus	13	975	5.03E-03
BP	Response To Hormone Stimulus	12	909	8.78E-03

SW and IT transcriptomes at 1 week of cold-acclimation: Analysis of genes that respond similarly to cold in SW and IT. Analysis of cold-responsive genes, which are differentially regulated between SW and IT, suggests that SW and IT may differ in their transcriptional mechanisms of cold-acclimation. However, there are a significant number of genes that are regulated similarly between SW and IT. There are 629 genes up-regulated by cold in both SW and IT with similar expression level. However, these genes may not be associated with increasing freezing tolerance. Therefore it was subsequently determined if genes commonly up-regulated by cold are enriched in genes previously associated with cold-acclimation. Hannah genes have been positively and negatively correlated with increases in freezing tolerance associated with cold-acclimation (6). The 629 genes commonly up-regulated by cold in SW and IT are not significantly enriched in Hannah genes positively or negatively correlated with increases in freezing tolerance (Fig. 3.21). However, these 629 up-regulated genes are enriched (15 genes; $p=0.0045$) in genes up-regulated by the CBFs. Abiotic stress GO terms, such as 'response to cold', 'response to temperature stimulus', 'cold-acclimation', 'response to salt stress' and 'response to osmotic stress' are significantly enriched in these 629 genes (Table 3.11). Similar GO terms are also enriched in the 277 Hannah genes positively correlated with increases in freezing tolerance (Table A3.2).

The 841 genes commonly down-regulated by cold with similar expression levels in SW and IT are not significantly enriched in Hannah genes (Fig. 3.22) or in CBF regulon genes (11 genes; $p=0.33$). Noteworthy overrepresented GO terms for these 841 genes include biotic stress related categories such as 'immune response', 'defense

response to bacterium' and 'apoptosis' (Table 3.12). Photosynthesis and light related GO categories are also enriched (Table 3.12). Defense response and photosynthesis related GO terms are also overrepresented in the 466 Hannah genes negatively correlated with increases in freezing tolerance (Table A3.2).

In summary, 1725 out of 2354 genes up-regulated by 1 week of cold are exclusively expressed in either SW or IT (Fig. 3.9) or have significantly different levels of expression between SW and IT (Fig. 3.15). 1800 out of 2641 genes down-regulated by 1 week of cold are exclusively expressed in either SW or IT (Fig 3.9) or have significantly different levels of expression between SW and IT (Fig 3.18). Therefore, the majority of cold-responsive genes are not regulated similarly between SW and IT. Although all of these differentially regulated genes may not play a role in cold-acclimation, comparisons of differentially-regulated genes to the Hannah gene set (6) and to the CBF regulon (25) suggests enrichment of genes correlated with cold-acclimation. There are a significant number of genes regulated similarly by cold in SW and IT. Though genes commonly up-regulated by cold are enriched in CBF regulon genes, common cold-regulated genes between SW and IT are not as enriched in genes previously associated with cold-acclimation in comparison to differentially regulated genes (Fig 3.10, 3.12, 3.16, 3.19, 3.21, 3.22). Overall, analysis of RNA-seq data suggests that SW and IT reach similar levels of freezing tolerance at 1 week of cold-acclimation (Fig. 3.2) through different transcriptional changes. Furthermore, IT appears to be consistently less enriched in Hannah genes and CBF regulated genes than SW, suggesting that IT possibly reaches a similar level of freezing tolerance

through CBF-independent pathways, which are much less well described. These differences in cold-response will be taken into account while defining sets of candidate genes that may contribute to differences in freezing tolerance (FTD genes) in the next section of this study.

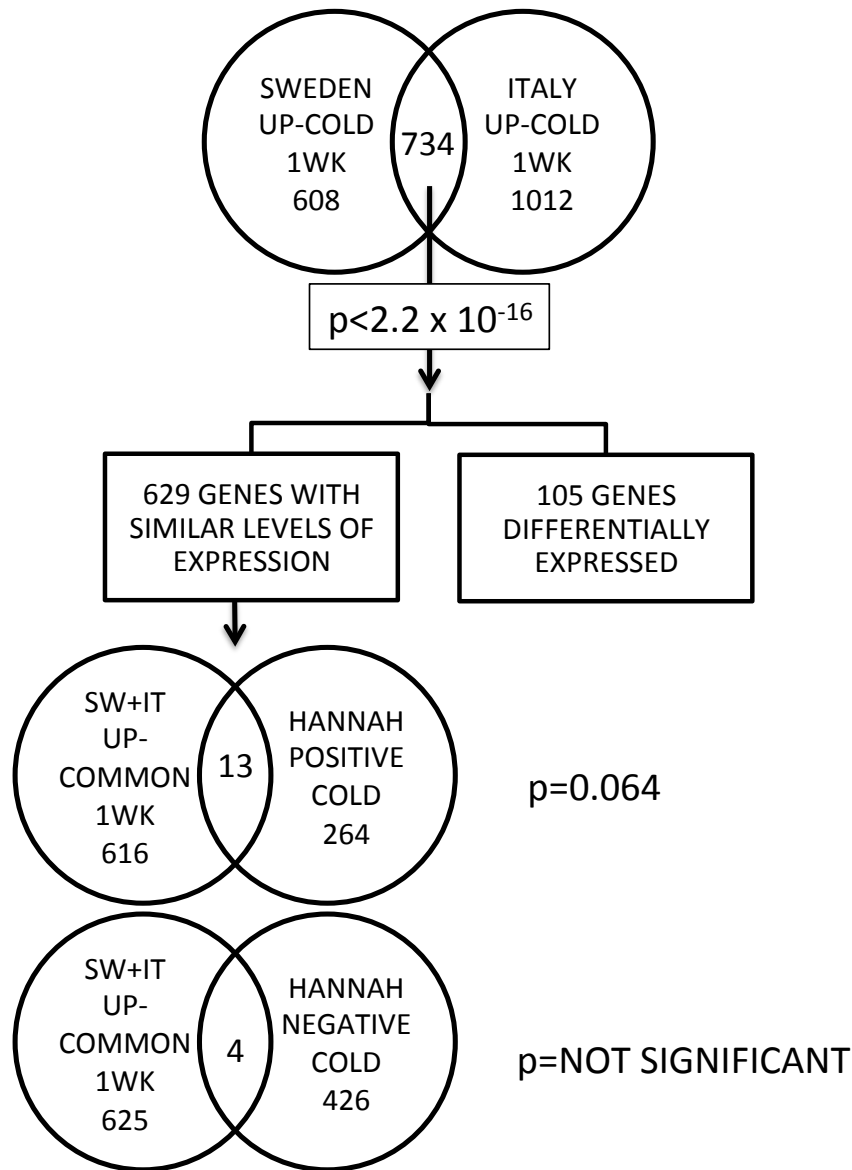


Figure 3.21. 629 genes with up-regulated induction patterns and similar expression levels and their overlap with the Hannah genes set. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . Genes were not differentially expressed between SW and IT if the FDR corrected p-value was ≥ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 genes positively correlated with freezing tolerance and 466 genes negatively correlated with freezing tolerance; (6)).

Table 3.11 GO categories significantly enriched in 629 genes with up-regulated induction patterns and similar expression levels in SW and IT. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). ‘GO TERM CATEGORY’ denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. ‘# HITS’ denotes the number of genes with that GO category in 629 genes with up-regulated induction patterns and similar expression levels in SW and IT. ‘# GENES IN GO’ denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	SW+IT COMMON UP-COLD (629 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
CC	Cytosolic Ribosome	62	317	2.73E-31
CC	Ribosome	73	470	9.99E-31
MF	Structural Constituent Of Ribosome	65	394	9.01E-29
CC	Ribonucleoprotein Complex	81	671	7.15E-27
MF	Structural Molecule Activity	71	538	7.25E-26
CC	Non-Membrane-Bounded Organelle	103	1144	1.55E-24
CC	Cytosol	75	708	1.99E-21
CC	Membrane-Enclosed Lumen	63	667	1.78E-15
CC	Nucleolus	44	332	1.89E-15
CC	Nuclear Lumen	51	472	1.92E-14
CC	Organelle Lumen	60	659	8.27E-14
BP	Ribosome Biogenesis	31	242	2.45E-09
BP	Circadian Rhythm	12	39	1.11E-06
BP	Translation	69	1231	1.77E-06
BP	Response To Abiotic Stimulus	65	1197	1.17E-05
BP	Response To Osmotic Stress	31	399	3.87E-05
BP	Response To Temperature Stimulus	29	354	3.89E-05
BP	Response To Cold	23	233	4.19E-05
CC	Plastid	128	3259	1.05E-03
CC	External Encapsulating Structure	36	620	1.27E-03
CC	Chloroplast	125	3192	1.37E-03
BP	Response To Salt Stress	26	369	1.96E-03
CC	Plant-Type Cell Wall	20	270	3.06E-03
BP	Response To Cadmium Ion	23	327	6.06E-03
CC	Plasma Membrane	89	2228	8.74E-03
BP	Cold Acclimation	6	20	1.03E-02
CC	Mitochondrion	58	1349	1.69E-02
BP	Response To Metal Ion	24	384	1.96E-02
BP	Response To Inorganic Substance	30	540	2.02E-02
BP	Protein Folding	18	251	2.57E-02

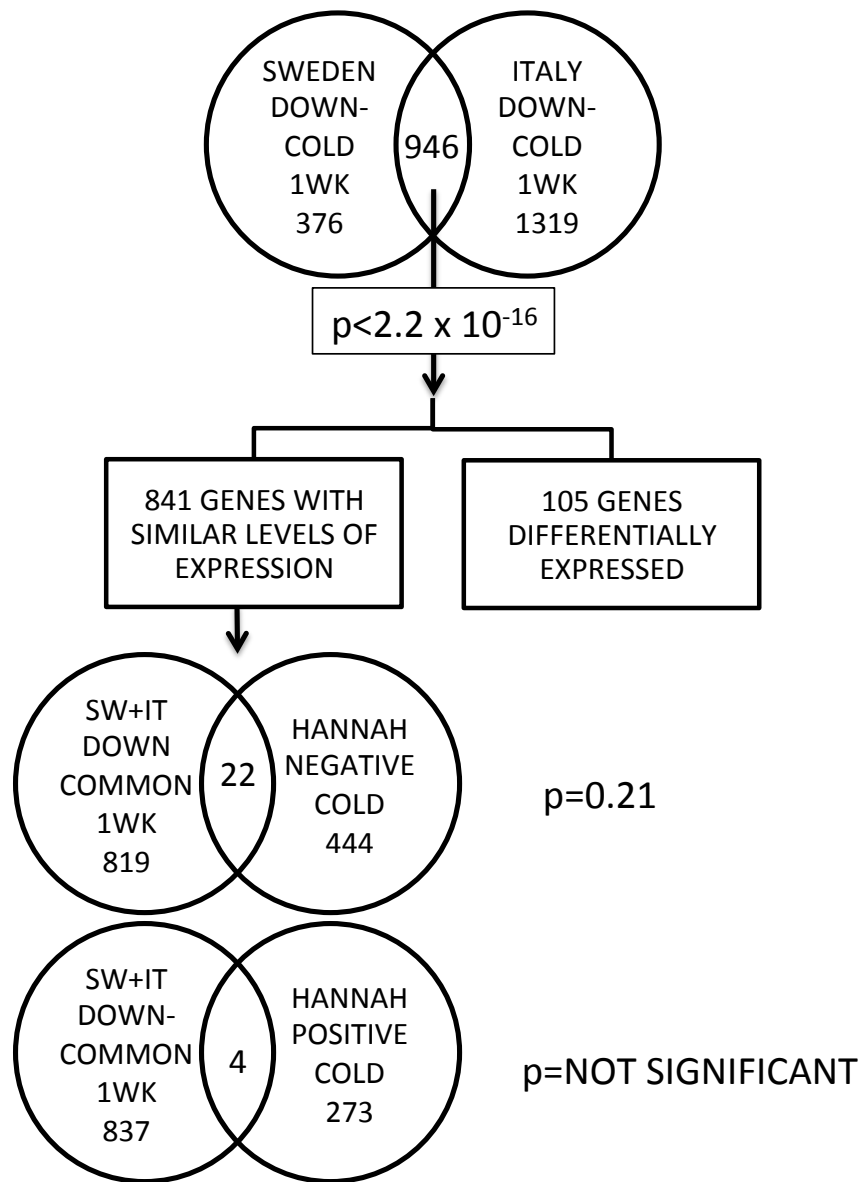


Figure 3.22. 841 genes with down-regulated induction patterns and similar expression levels and their overlap with the Hannah genes set. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . Genes were not differentially expressed between SW and IT if the FDR corrected p-value was ≥ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 genes positively correlated with freezing tolerance and 466 genes negatively correlated with freezing tolerance; (6)).

Table 3.12. GO categories significantly enriched in 841 genes with down-regulated induction patterns and similar expression levels in SW and IT. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in 841 genes with down-regulated induction patterns and similar expression levels in SW and IT. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	SW+IT COMMON DOWN-COLD 841 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Response To Abiotic Stimulus	75	1197	5.31E-04
CC	Organelle Subcompartment	30	349	5.70E-04
CC	Thylakoid Membrane	27	305	6.39E-04
BP	Response To Light Stimulus	39	466	6.43E-04
BP	Programmed Cell Death	25	223	7.54E-04
BP	Response To Radiation	39	482	8.59E-04
BP	Response To Bacterium	25	244	1.21E-03
CC	Photosynthetic Membrane	27	330	1.37E-03
BP	Cell Death	27	254	1.45E-03
CC	Chloroplast	145	3192	1.71E-03
BP	Response To Blue Light	11	55	2.34E-03
BP	Response To Far Red Light	10	44	2.52E-03
BP	Protein Amino Acid Phosphorylation	65	1065	2.68E-03
BP	Immune Response	27	293	2.76E-03
CC	Plastid	146	3259	2.87E-03
MF	Protein Serine/Threonine Kinase Activity	58	975	9.41E-03
BP	Response To Red Light	10	56	1.05E-02
MF	Protein Kinase Activity	66	1110	1.10E-02
BP	Apoptosis	18	172	1.11E-02
BP	Photosynthesis	18	175	1.18E-02
MF	Nucleoside Binding	123	2536	1.20E-02
MF	Adenyl Ribonucleotide Binding	115	2368	1.29E-02
MF	Adenyl Nucleotide Binding	123	2528	1.54E-02
BP	Phosphorylation	66	1178	1.57E-02
BP	Response To Organic Substance	66	1176	1.60E-02
CC	Vacuole	38	643	2.02E-02
MF	ATP Binding	112	2340	2.38E-02
BP	Phosphate Metabolic Process	69	1273	2.68E-02
CC	Cell Wall	35	611	4.49E-02
CC	Interchromatin Granule	3	3	4.79E-02

GENES CONTRIBUTING TO DIFFERENCES IN FREEZING TOLERANCE

Summary: Genes contributing to differences in freezing tolerance. In the next section of RNA-seq analysis two sets of genes that may possibly contribute to differences in freezing tolerance between SW and IT at 2 weeks of cold-acclimation are defined (Fig. 3.2). Category 1 FREEZING TOLERANCE DIFFERENCE (FTD) candidate genes are cold-regulated genes, which are differentially expressed between SW and IT at only 2 weeks of cold-acclimation, the point at which freezing tolerance differs. Analysis of SW and IT transcriptomes with 1 week of cold- suggests that SW and IT may potentially differ in their transcriptional mechanisms of cold-acclimation. Therefore, Category 2 FTD candidate genes are differentially expressed between SW and IT at both 1 week and 2 weeks of cold-acclimation. Analysis of Category 1 and Category 2 FTD genes suggests that the CBF pathway of cold-acclimation may be differentially regulated between SW and IT.

Genes contributing to differences in freezing tolerance: Category 1

FREEZING TOLERANCE DIFFERENCE (FTD) candidate genes. SW and IT have no difference in freezing tolerance with 1 week of cold-acclimation or under non-acclimated conditions (Fig. 3.1). But after 2 weeks of cold-acclimation there is a significant difference in freezing tolerance between SW and IT (Fig 3.2; $p \leq 0.026$). To determine if genes associated with differences in freezing tolerance possibly underlie fitness QTL, candidate genes contributing to differences in freezing tolerance were first defined. It is possible that differentially expressed cold genes between SW and IT are involved in differences between SW and IT not related to freezing tolerance. However, even if

these differentially expressed genes were not related to freezing tolerance, these genes would still represent differences between SW and IT that could potentially contribute to differences in fitness between SW and IT. Examination of SW and IT transcriptomes at 1 week of cold suggests that SW and IT may differ in their mechanisms of cold-acclimation, since the majority of cold-regulated genes are differentially regulated between SW and IT. Although it is possible that genes expressed similarly between SW and IT are responsible for cold-acclimation (Fig. 3.21-3.22), it seems likely that at least a portion of differentially regulated cold genes between SW and IT contribute cold-acclimation in some way since differentially regulated cold genes at 1 week of cold-acclimation are significantly enriched in genes regulated by the CBFs, master regulators of the most well defined pathway of cold-acclimation (21, 24, 29). Consequently, differences in gene expression at 1 week of cold-acclimation may contribute to the differences in freezing tolerance seen later at 2 weeks of cold-acclimation. These genes were defined as Category 2 FTD genes and will be described later. However, genes that are differentially expressed only at 2 weeks of cold-acclimation also seem like good candidates for genes contributing to differences in freezing tolerance. Therefore, Category 1 FTD candidate genes are cold-regulated genes (in either SW or IT) that have no difference in expression level between SW and IT at 1 week of cold-acclimation and non-acclimated conditions, but different levels of expression at 2 weeks of cold-acclimation. Again, differential expression was determined by a FDR corrected p-value ≤ 0.050 (30) and a ≥ 3 FPKM threshold for either sample in the pairwise comparison (19, 20).

There are 2130 Category 1 FTD candidate genes (Fig. 3.23; Table A3.3 and A3.4). Of these 2130 Category 1 FTD genes, 474 have higher expression in SW at 2 weeks of cold-acclimation (Fig. 3.23, Table A3.3). Again, all these differentially expressed genes may not necessarily contribute to cold-acclimation. Consequently, enrichment of genes that have been previously associated with cold-acclimation was tested. The CBFs are central components of the most well described pathway of cold-acclimation. The 474 Category 1 FTD candidate genes with higher expression in SW are significantly enriched (33 genes; $p=3.2E-17$; Table A3.3) in genes up-regulated by the CBFs. This suggests that the CBF pathway of cold-acclimation differs between SW and IT, which is similar to what was seen at 1 week of cold-acclimation. These 474 genes are also significantly enriched in Hannah genes positively and negatively correlated with freezing tolerance (Fig 3.23). There are 15 Hannah genes positively correlated with freezing tolerance that overlap with the Category 1 genes with higher expression in SW. This overlap is statistically significant however the majority of these genes have less than 2-fold higher expression in SW compared to IT (Fig 3.24). There are 20 Hannah genes negatively correlated with freezing tolerance that overlap with the Category 1 genes with higher expression in SW. This overlap is again statistically significant but all of these genes are less than 2-fold higher in SW compared to IT (Fig. 3.25). Interesting GO categories overrepresented in these 474 genes include 'response to temperature', 'response to cold' and 'photosynthesis' (Table 3.13).

Out of 2130 Category 1 FTD genes, 1656 Category 1 FTD candidate genes have higher expression in IT at 2 weeks of cold-acclimation (Fig 2.23; Table A.3.4). These

1656 genes are not significantly enriched in Hannah genes positively or negatively correlated with freezing tolerance (Fig. 3.23) and are also not enriched in genes up-regulated (6 genes; $p=0.99$) or down-regulated (21 genes; $p=0.29$) by the CBFs. However, this genes set is significantly enriched in a number of notable GO terms, most of which related to defense response and biotic stress (Table 3.14). Temperature-related GO terms are not enriched in these 1656 Category 1 genes, but other abiotic stress categories such as 'response to salt stress' and 'response to osmotic stress' are enriched. Altogether, Category 1 FTD candidate genes with higher expression in SW appear to be more enriched in genes that have been previously associated with cold-acclimation than IT. This difference is consistent with analysis of SW and IT transcriptomes at 1 week of cold-acclimation and again suggests that IT may cold-acclimate using pathways not yet well defined.

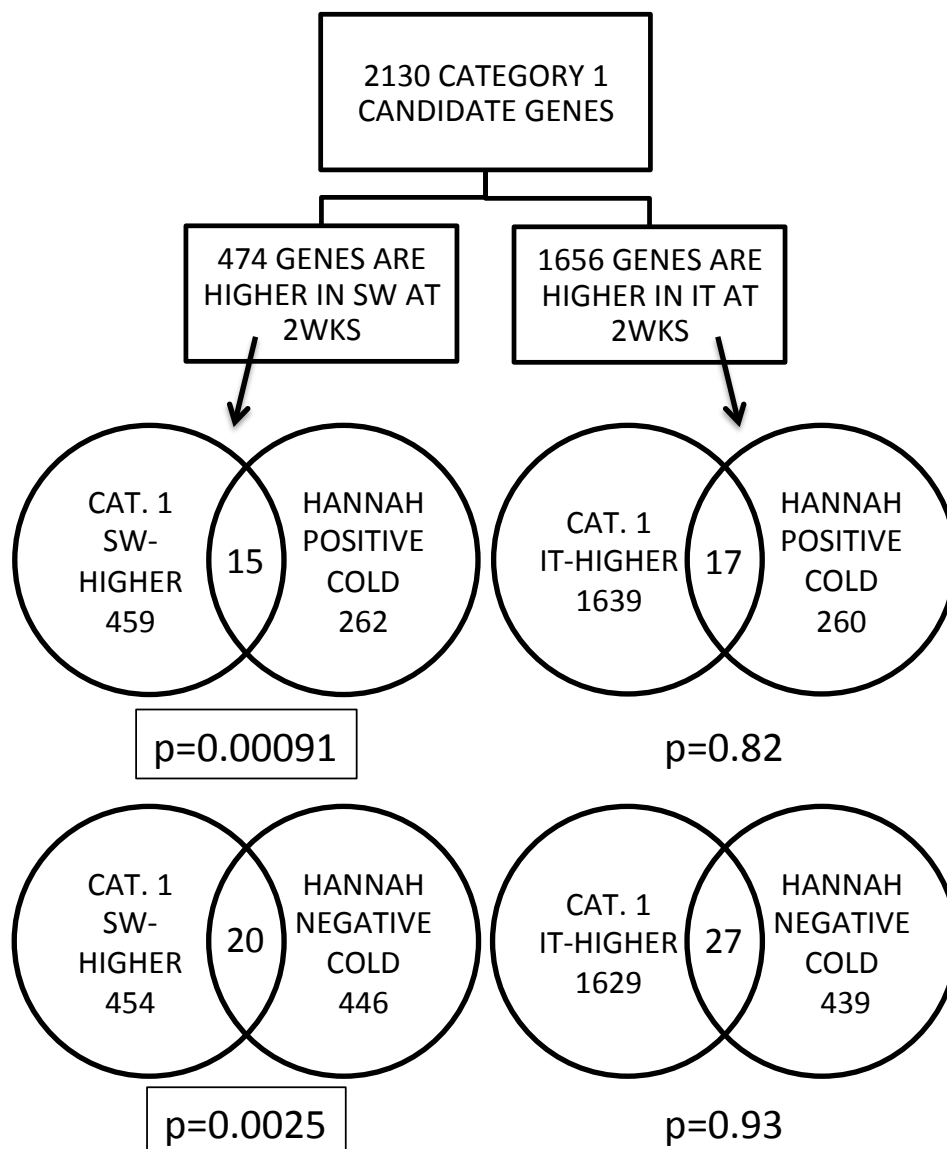


Figure 3.23. 2130 Category 1 FTD candidate genes and overlap with the Hannah gene set. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 Hannah genes positively correlated with freezing tolerance and 466 Hannah genes negatively correlated with freezing tolerance; (6)).

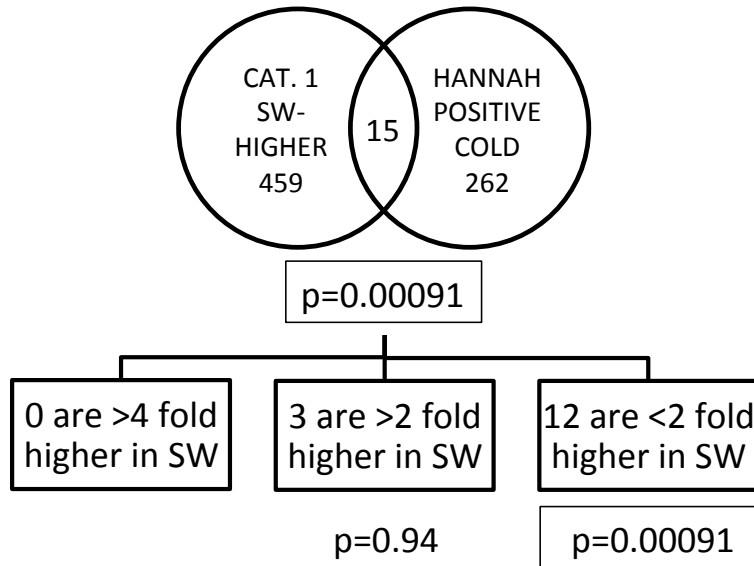


Figure 3.24. Fold change of 15 Hannah genes that overlap with 474 Category 1 FTD genes with higher expression in SW. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-offs were applied (from left to right).

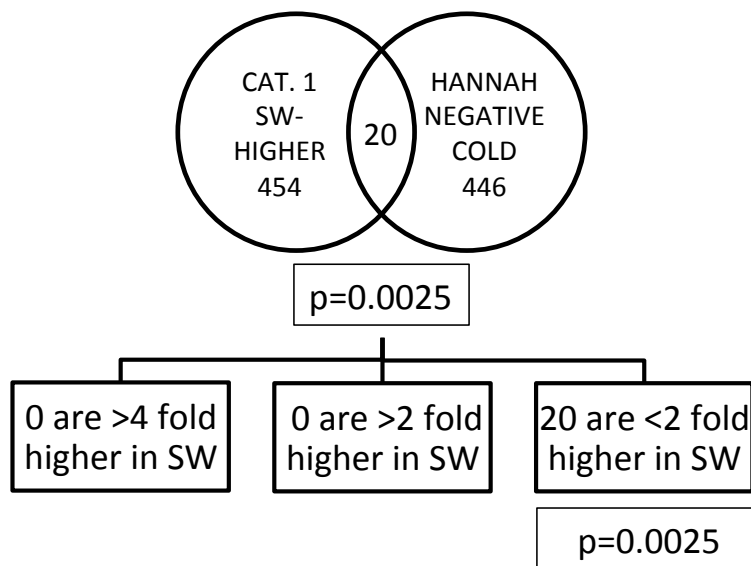


Figure 3.25. Fold change of 20 Hannah genes that overlap with 474 Category 1 FTD genes with higher expression in SW. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-offs were applied (from left to right).

Table 3.13. GO categories significantly enriched in 474 Category 1 genes with higher expression in SW at 2 weeks of cold-acclimation. ‘GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). GO TERM CATEGORY’ denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. ‘# HITS’ denotes the number of genes with that GO category in 474 Category 1 genes with higher expression in SW at 2 weeks of cold-acclimation. ‘# GENES IN GO’ denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	SW HIGH CATEGORY 1 (474 GENES)	# HITS	# GENES IN GO	FDR P- VALUE
CC	Chloroplast	155	3192	3.54E-24
CC	Plastid	156	3259	5.20E-24
CC	Organelle Subcompartment	32	349	6.21E-10
CC	Thylakoid Membrane	29	305	2.44E-09
CC	Photosynthetic Membrane	29	330	1.24E-08
CC	Organelle Membrane	45	849	9.29E-07
BP	Starch Metabolic Process	10	34	3.53E-06
BP	Photosynthesis	19	175	4.41E-06
CC	Plastid Envelope	28	461	3.35E-05
CC	Chloroplast Envelope	26	440	1.23E-04
CC	Photosystem	9	72	1.43E-03
BP	Response To Abiotic Stimulus	45	1197	2.26E-03
BP	Carbohydrate Biosynthetic Process	15	189	2.75E-03
BP	Cellular Glucan Metabolic Process	12	124	2.89E-03
BP	Response To Temperature Stimulus	20	354	3.83E-03
BP	Response To Cold	16	233	4.10E-03
BP	Oligosaccharide Metabolic Process	8	51	4.22E-03
BP	Starch Catabolic Process	5	13	5.73E-03
BP	Carbohydrate Biosynthetic Process	16	254	6.92E-03
BP	Cellular Polysaccharide Catabolic Process	5	15	8.16E-03
BP	Glucan Metabolic Process	12	151	8.73E-03
BP	Cellular Polysaccharide Metabolic Process	12	154	8.90E-03
BP	Photosynthesis, Light Reaction	9	85	9.97E-03
BP	Starch Biosynthetic Process	5	17	1.12E-02
BP	Generation Of Precursor Metabolites And Energy	19	404	3.06E-02
CC	Photosystem II	6	52	3.53E-02
CC	Plastid Outer Membrane	5	34	3.86E-02
CC	Chloroplast Photosystem II	4	18	3.94E-02
CC	Oxygen Evolving Complex	4	18	3.94E-02
BP	Glycoside Metabolic Process	8	84	4.23E-02

Table 3.14. GO categories significantly enriched in 1656 Category 1 genes with higher expression in IT at 2 weeks of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in 1656 Category 1 genes with higher expression in IT at 2 weeks of cold-acclimation. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	IT HIGH CATEGORY 1 GENES (1656 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
CC	Plasma Membrane	277	2228	9.35E-25
BP	Response To Chitin	43	127	1.38E-14
MF	Nucleoside Binding	288	2536	2.55E-13
BP	Response To Carbohydrate Stimulus	50	199	6.49E-12
BP	Immune Response	63	293	6.57E-12
BP	Protein Amino Acid Phosphorylation	146	1065	6.95E-12
BP	Response To Organic Substance	157	1176	7.34E-12
MF	Protein Kinase Activity	150	1110	2.27E-11
MF	Purine Nucleotide Binding	300	2785	2.77E-11
MF	Adenyl Ribonucleotide Binding	262	2368	6.58E-11
BP	Innate Immune Response	58	275	6.76E-11
BP	Response To Wounding	40	147	1.11E-10
CC	Endoplasmic Reticulum	74	446	1.12E-10
MF	ATP Binding	257	2340	2.42E-10
BP	Phosphate Metabolic Process	159	1273	6.15E-10
BP	Response To Bacterium	52	244	6.63E-10
CC	Intrinsic To Membrane	262	2658	9.77E-10
BP	Phosphorylation	148	1178	2.08E-09
BP	Cell Death	52	254	2.15E-09
BP	Defense Response To Bacterium	42	184	6.69E-09
BP	Ribonucleotide Binding	274	2621	8.30E-09
BP	Nucleotide Binding	332	3333	1.08E-08
BP	Programmed Cell Death	45	223	8.10E-08
BP	Defense Response	127	1030	1.94E-07
BP	Callose Deposition During Defense Response	12	18	4.77E-07
BP	Plant-Type Hypersensitive Response	17	45	3.31E-06
BP	Polysaccharide Localization	12	21	3.45E-06
CC	External Encapsulating Structure	78	620	6.92E-06
CC	Integral To Membrane	207	2193	7.12E-06
CC	Cell Wall	76	611	1.06E-05
BP	Amino Acid Derivative Metabolic Process	44	280	1.14E-04

Table 3.14 (cont'd)

CC	Endoplasmic Reticulum Part	20	92	2.89E-04
CC	Vacuole	72	643	6.41E-04
BP	Phospholipid Transport	8	14	1.16E-03
BP	Indole Derivative Metabolic Process	13	42	1.42E-03
BP	Aromatic Compound Biosynthetic Process	34	215	1.50E-03
BP	Response To Inorganic Substance	66	540	1.52E-03
BP	Secondary Metabolic Process	55	423	1.52E-03
BP	Apoptosis	29	172	1.88E-03
MF	Calcium Ion Binding	54	405	1.95E-03
BP	Response To Abiotic Stimulus	123	1197	2.22E-03
BP	Response To Endogenous Stimulus	104	975	2.24E-03
BP	Defense Response To Fungus	10	26	2.28E-03
BP	Cell Wall Modification	24	135	3.87E-03
BP	Response To Jasmonic Acid Stimulus	26	156	5.30E-03
MF	Phospholipid-Translocating ATPase Activity	7	12	6.64E-03
MF	Aminophospholipid Transporter Activity	7	12	6.64E-03
BP	Response To Salt Stress	47	369	7.50E-03
BP	Indole Derivative Biosynthetic Process	11	37	7.62E-03
BP	Sulfur Metabolic Process	27	171	8.46E-03
BP	Response To Endoplasmic Reticulum Stress	7	14	8.55E-03
MF	FAD Binding	27	164	9.39E-03
BP	Response To Osmotic Stress	49	399	1.16E-02
BP	Response To Oxidative Stress	38	287	1.41E-02
BP	Positive Regulation Of Response To Stimulus	13	55	1.45E-02
BP	Regulation Of Innate Immune Response	10	35	2.02E-02
BP	Response To Light Stimulus	54	466	2.06E-02
BP	Salicylic Acid Mediated Signaling Pathway	9	29	2.27E-02
MF	Calmodulin Binding	27	175	2.46E-02
BP	S-Glycoside Metabolic Process	11	44	2.71E-02
BP	Glucosinolate Metabolic Process	11	44	2.71E-02
BP	Positive Regulation Of Defense Response	9	31	3.50E-02
MF	Protein Tyrosine Kinase Activity	40	312	3.85E-02
BP	Response To Radiation	54	482	4.00E-02
BP	Response To Ozone	8	25	4.01E-02
MF	Acid-Amino Acid Ligase Activity	38	294	4.27E-02
BP	Amino Acid Derivative Biosynthetic Process	27	194	4.35E-02
MF	Ligase Activity	43	349	4.62E-02
MF	Cofactor Binding	59	525	4.63E-02
BP	Intracellular Signaling Cascade	80	790	4.81E-02
BP	Response To Hormone Stimulus	90	909	4.87E-02

Genes contributing to differences in freezing tolerance: Category 2 FTD

candidate genes. Though there is no difference in freezing tolerance between SW and IT with 1 week of cold-acclimation, analysis of SW and IT transcriptomes suggests that they reach this similar level of freezing tolerance by different mechanisms (Fig 3.9). It is possible that transcriptional differences at 1 week of cold-acclimation do not result in differences in freezing tolerance until 2 weeks of cold-acclimation. It is also possible that differences in cold-regulated gene expression at 1 week and 2 weeks of cold-acclimation couple to result in differences in freezing tolerance at 2 weeks of cold-acclimation. Based on this, Category 2 FTD candidate genes are cold-induced genes (in either SW or IT) with significantly different levels of expression at both 1 and 2 weeks of cold-acclimation.

There are 798 Category 2 FTD candidate genes (Fig 3.26). Out of 798 Category 2 genes, 294 have higher expression in SW compared to IT at 2 weeks of cold-acclimation (Fig 3.26; Table A3.5). It was also determined if these genes have been previously associated with cold-acclimation. These 294 genes are significantly enriched (39 genes; $p < 2.2 \times 10^{-16}$) in genes up-regulated the CBFs (Table A3.5). These 294 genes are also significantly enriched in Hannah genes positively correlated with freezing tolerance (Fig 3.26). There are 32 Hannah genes positively correlated with freezing tolerance that overlap with the Category 2 genes with higher expression in SW. This overlap is statistically significant, and 8 genes have greater than 4-fold higher expression in SW compared to IT, 15 are greater than 2-fold higher and 9 are less than 2-fold higher (Fig 3.27). 'Response to cold' is the GO category most overrepresented in

these 294 genes, and other abiotic stress GO terms such as ‘response to water deprivation’ are also enriched (Table 3.15).

There are 504 out of 798 Category 2 FTD candidate genes with higher expression in IT compared to SW at 2 weeks of cold-acclimation (Fig. 3.26; Table A3.6). The 504 genes with higher expression in IT are not enriched in genes up-regulated (2 genes; $p=0.97$) or down-regulated (3 genes; $p=0.92$) by the CBFs (Table A3.6). However, these 504 genes are significantly enriched in Hannah genes both positively and negatively correlated with increases in freezing tolerance. There are 12 Hannah genes positively correlated with freezing tolerance that overlap with the Category 2 genes with higher expression in IT than SW. This overlap is statistically significant, but less significant than the overlap between Category 2 genes with higher expression in SW and Hannah genes positively correlated with freezing tolerance (Fig 3.27). Of the 12 genes high in IT that overlap with the Hannah gene set, 2 are greater than 4-fold higher in IT compared to SW, 7 are greater than 2 fold higher, and 3 are less than 2-fold higher (Fig. 3.28). There are 25 Hannah genes negatively correlated with freezing tolerance that overlap with the Category 2 genes with higher expression in IT compared to SW. These 25 genes negatively correlated with freezing tolerance thus, have lower expression in SW. Of these 25 genes, 4 have greater than 4-fold higher expression in IT, 10 are greater than 2-fold higher, and 11 are less than 2-fold higher (Fig 3.29). The 504 Category 2 FTD candidate genes are not enriched in temperature-related GO terms. But ‘defense response’ and ‘biotic interaction’ related GO terms are highly enriched (Table 3.16).

In Summary, Category 2 FTD candidate genes with higher expression in SW are more enriched in genes that have been previously associated with cold-acclimation than IT. This is consistent with analysis of Category 1 FTD genes and cold-regulated transcriptomes at 1 week of cold-acclimation. This again suggests that IT may cold-acclimate using pathways not yet well defined. Since IT decreases in freezing tolerance from 1 to 2 weeks of cold-acclimation it also suggests that IT pathways of cold-acclimation are not as efficient as pathways in SW.

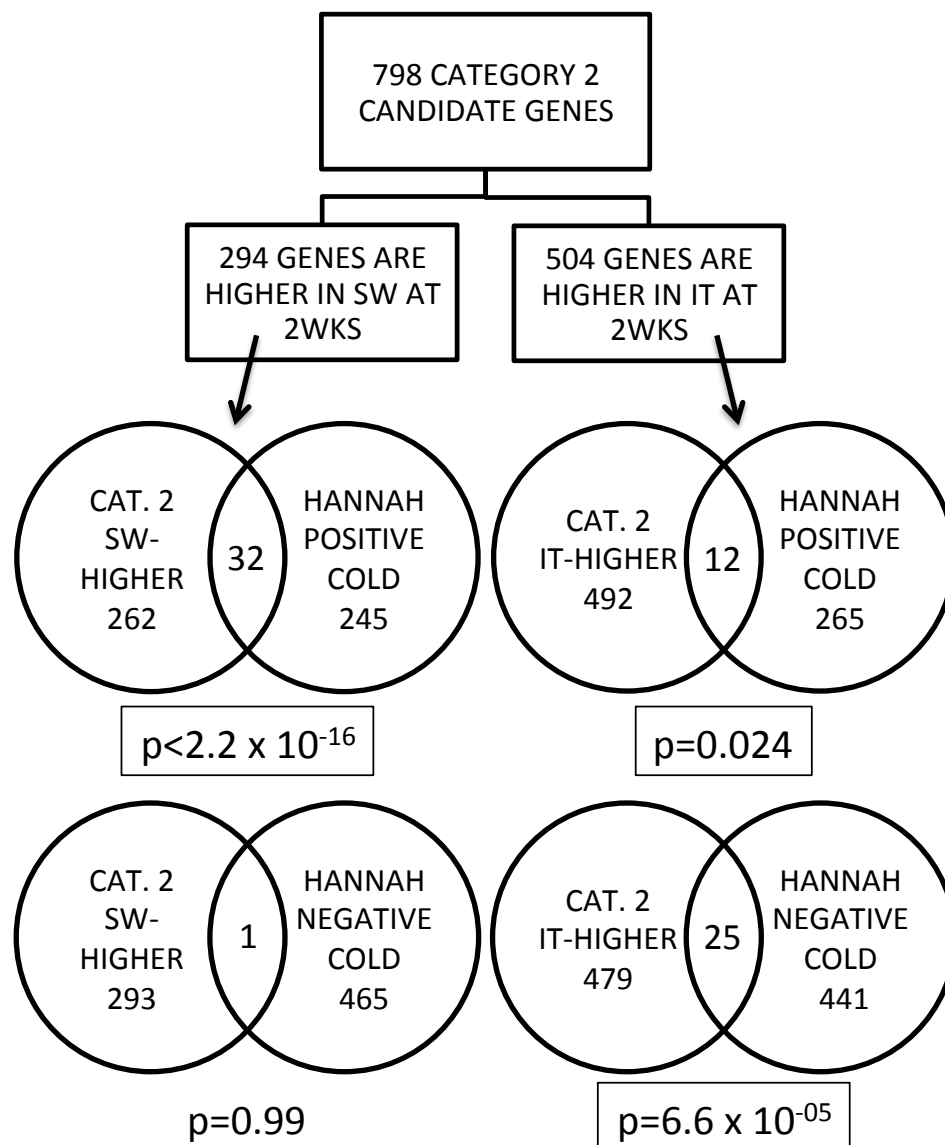


Figure 3.26. 798 Category 2 FTD candidate genes and their overlap with the Hannah gene set. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 week under a 12 h photoperiod then sampled. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Genes were considered differentially expressed from non-acclimated conditions if the FDR corrected p-value was ≤ 0.05 . ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). Genes were considered differentially expressed between SW and IT if the FDR corrected p-value was ≤ 0.05 . P-values (calculated using a hypergeometric distribution in R) represent overlap between differentially expressed genes and Hannah genes (277 genes positively correlated with freezing tolerance and 466 genes negatively correlated with freezing tolerance; (6)).

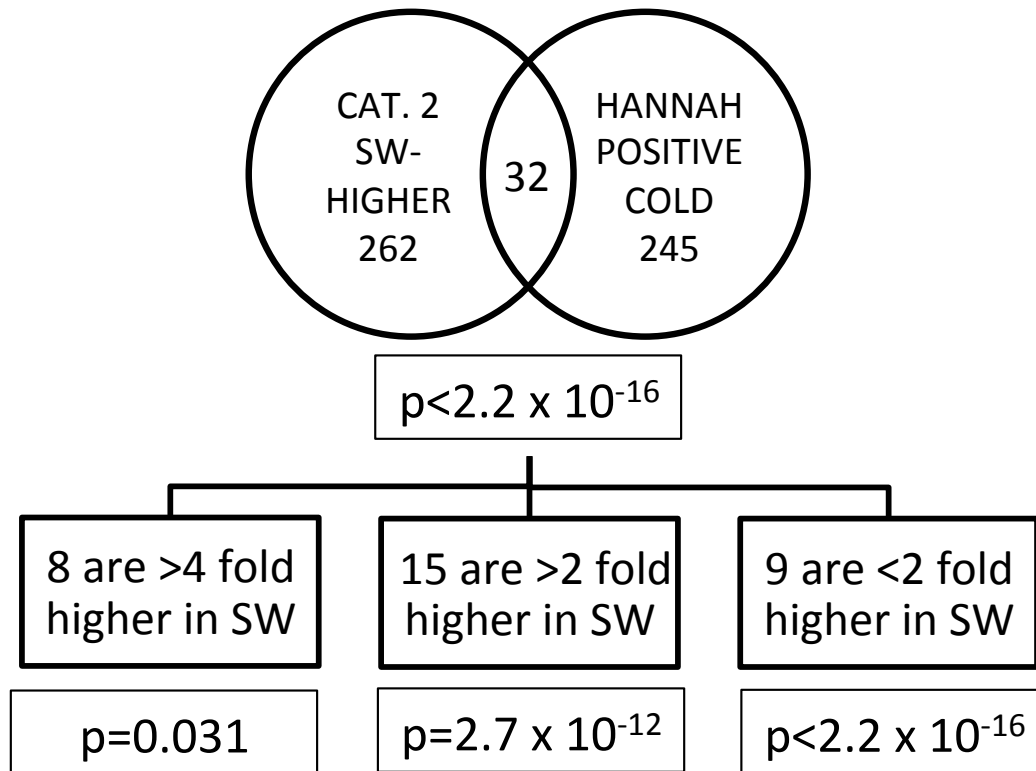


Figure 3.27. Fold change of 32 Hannah genes that overlap with 294 Category 2 FTD genes with higher expression in SW. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-off were applied (from left to right).

Table 3.15. GO categories significantly enriched in 294 Category 2 genes with higher expression in SW at 2 weeks of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in 294 Category 2 genes with higher expression in SW at 2 weeks of cold-acclimation. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	SW HIGH CATEGORY 2 (294 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Response To Cold	22	233	1.81E-09
BP	Response To Temperature Stimulus	25	354	1.04E-08
BP	Response To Water	17	185	4.77E-07
BP	Response To Water Deprivation	16	176	1.41E-06
BP	Response To Abiotic Stimulus	40	1197	8.06E-06
BP	Cold Acclimation	6	20	4.48E-04
BP	Response To Absciscic Acid Stimulus	13	285	2.96E-02
BP	Programmed Cell Death	11	223	4.33E-02
BP	Response To Organic Substance	30	1176	4.46E-02

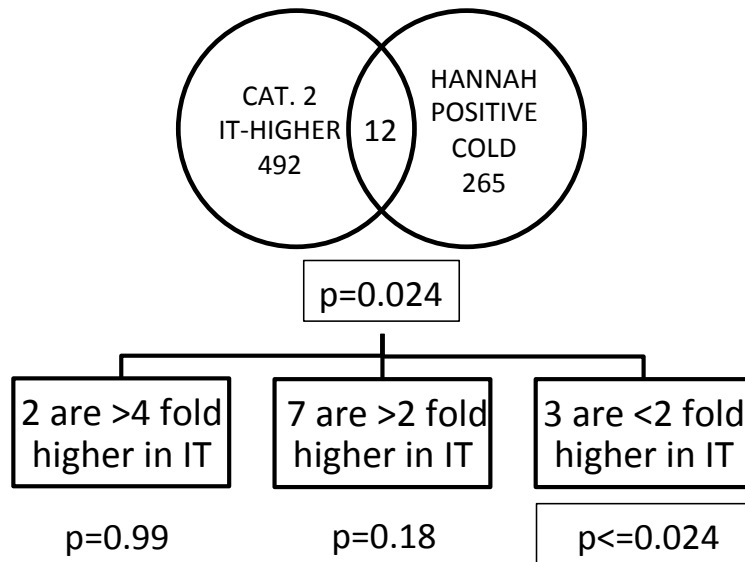


Figure 3.28. Fold change of 12 Hannah genes that overlap with 504 Category 2 FTD genes with higher expression in IT. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-off were applied (from left to right).

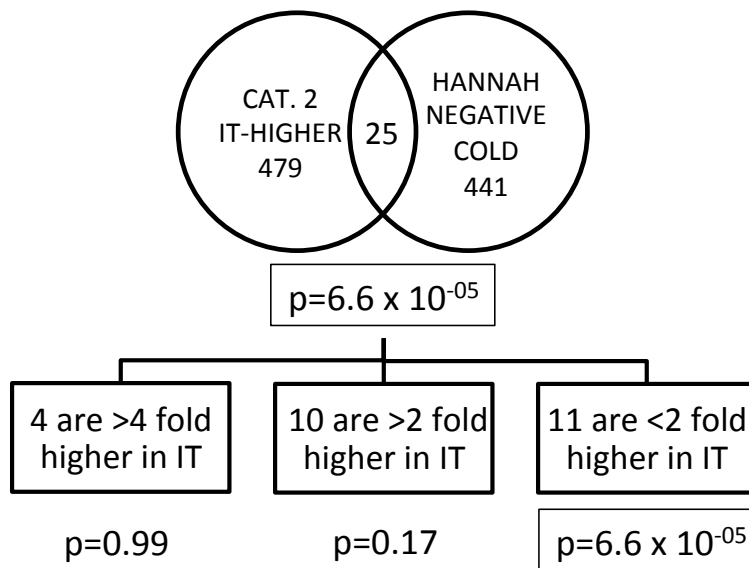


Figure 3.29. Fold change of 25 Hannah genes that overlap with 504 Category 2 FTD genes with higher expression in IT. P-values represent the significance of overlap between gene only up-regulated in SW and Hannah genes if 4-fold, 2-fold, or no fold change cut-off were applied (from left to right).

Table 3.16. GO categories significantly enriched in 504 Category 2 genes with higher expression in IT at 2 weeks of cold-acclimation. GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in 504 Category 2 genes with higher expression in IT at 2 weeks of cold-acclimation. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	IT HIGH CATEGORY 2 (504 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Innate Immune Response	26	275	1.55E-06
BP	Defense Response	56	1030	1.74E-06
BP	Response To Bacterium	21	244	1.51E-04
BP	Response To Organic Substance	54	1176	1.77E-04
BP	Response To Endogenous Stimulus	44	975	2.46E-03
BP	Secondary Metabolic Process	25	423	3.26E-03
BP	Response To Salicylic Acid Stimulus	14	150	3.59E-03
CC	Vacuole	31	643	4.25E-03
BP	Response To Hormone Stimulus	39	909	1.60E-02
BP	Oxidation Reduction	47	1186	1.70E-02
CC	Extracellular Region	44	1225	2.13E-02
CC	Apoplast	20	382	2.82E-02
BP	Toxin Metabolic Process	7	46	3.46E-02
CC	Intrinsic To Plasma Membrane	9	103	3.75E-02
BP	Cell Death	16	254	3.82E-02
BP	Systemic Acquired Resistance	6	32	3.96E-02
CC	External Encapsulating Structure	26	620	4.25E-02
BP	Response To Jasmonic Acid Stimulus	12	156	4.29E-02
CC	Anchored To Plasma Membrane	7	66	4.76E-02
CC	Cell Wall	25	611	4.91E-02

Summary of SW and IT transcriptomes. Through analysis of basal, as well as 1 and 2 week cold-acclimated transcriptomes two sets of FTD candidate genes have been defined. Furthermore, several interesting trends regarding differences between SW and IT seem to have emerged through this investigation. Analysis of RNA-seq data suggests that SW and IT cold-acclimate through significantly different transcriptional changes. Comparisons of SW and IT cold-regulated transcriptomes to CBF regulon genes (25) and Hannah genes (6), which are enriched in CBF regulon genes, seem to indicate that SW is more enriched in genes that have been previously associated with cold-acclimation. This suggests that SW may use previously defined pathways of cold-acclimation, such as the CBF pathway, while IT may use pathways of cold-acclimation not yet well defined. However, differences between SW and IT were determined without fold-change thresholds. Therefore, the fold-change of expression data was visually examined using heatmaps to ensure that these trends are consistent.

Visual analysis of cold-regulated genes suggests that most genes have similar rather than opposite cold-induction patterns in SW and IT, however most cold-regulated genes differ between SW and IT in terms of expression level (Fig 3.9, 3.23, 3.26). This seems consistent with previous analysis using FDR corrected p-value thresholds. Under non-acclimated conditions, genes up-regulated by the CBFs (25) appear to have higher expression in IT, which is consistent with previous analysis of basal transcriptomes using FDR corrected p-value thresholds. The difference in basal expression of CBF regulon genes again suggests that the CBF pathway of cold-acclimation is regulated differently between SW and IT. The CBFs are cold-responsive

genes (29). With the addition of cold, genes up-regulated by the CBFs generally have higher expression in SW than IT, and genes down-regulated by the CBFs are more repressed in SW (Fig 3.31). There also appears to be more CBF regulon genes responsive to cold in SW in comparison to IT (Fig. 3.3.1). Altogether, analysis of RNA-seq data using fold-change values still indicates that the CBF pathway of cold-acclimation may be differentially regulated between SW and IT. Fold-change analysis of Hannah genes (6), shows that SW generally has higher expression of genes previously positively correlated with freezing tolerance, and has lower expression of genes previously negatively correlated with freezing tolerance in comparison to IT (Fig. 3.32). Altogether, analysis of RNA-seq data by fold-change still suggests that SW is more enriched in genes that have been previously associated with cold-acclimation than IT (Fig. 3.31 and 3.32).

GO enrichment analysis also revealed several trends in SW and IT transcriptomes. Under all conditions, IT seems to be enriched in genes related to biotic stress. Therefore, biotic stress related genes were pulled from TAIR (www.arabidopsis.org), to determine if this is generally the case. Differentially expressed biotic genes between SW and IT were defined by a FDR corrected p-value ≤ 0.050 (384 genes differentially expressed out of 925). A heatmap was generated for these biotic stress genes to visually compare fold-change differences between SW and IT expression. IT does appear to have higher expression of biotic stress related genes under both warm and cold conditions in comparison to SW (Fig 3.33), but this difference is most pronounced at 2 weeks of cold-acclimation. Also, more biotic stress related

genes appear to be up-regulated by cold in IT at 2 weeks of cold-acclimation in comparison to SW (Fig. 3.33). Examination of IT transcripts suggests that IT is less enriched in pathways of cold-acclimation that have been previously defined. Consequently, some of these biotic stress related genes may function in cold-acclimation, especially since pathogens related proteins have previously been shown to have antifreeze properties (26). Furthermore, resistance to biotic stress may be an interesting trait segregating between the SW and IT ecotypes and should be tested in the SW and IT RILs in the future.

Photosynthesis was another interesting reoccurring GO term for SW and IT. It is well known that many transcriptional as well as metabolic changes occur during cold-acclimation (23, 31-34). Photosynthesis provides the energy for these necessary changes to occur (35). Increased photosynthetic capacity is consequently correlated with cold-acclimation (24, 36-39). Therefore, differentially expressed photosynthesis associated genes (list of genes provided by Hu and Thomashow, unpublished) between SW and IT were determined by a FDR corrected p-value ≤ 0.050 (52 genes differentially expressed out of 244). A heatmap was generated for these photosynthesis genes to visually compare the fold-change differences between SW and IT expression (Fig 3.34). Although most photosynthesis related genes appear to be down-regulated by cold in both SW and IT, SW appears to have higher expression of photosynthesis related genes under both warm and cold conditions in comparison to IT (Fig 3.34). This may indicate that there are differences in the photosynthetic capacity of SW and IT. Differences in photosynthetic capacity would correlate with differences in freezing

tolerance between SW and IT at 2 weeks of cold-acclimation. Nevertheless, differences in photosynthetic capacity would need to be measured experimentally. Variations in photosynthetic capacity would also be an interesting trait to screen and map in the SW and IT RILs.

In the next section of this study, expression of Category 1 and Category 2 FTD genes are examined in the SW and IT RILs, which were constructed by the Schemske Lab. This eQTL analysis allows expression differences to be mapped to specific genetic locations, and genes potentially important for differences in freezing tolerance as well as fitness QTL can also be determined (Table 3.1). FTD candidate genes that are also CBF regulon genes will be targeted for this study since CBF regulon genes appear to be enriched in differentially expressed genes. Furthermore, FTD candidate genes annotated as 'photosynthesis related' and 'biotic stress related' will be included in eQTL analysis since those GO terms appear to be overrepresented.

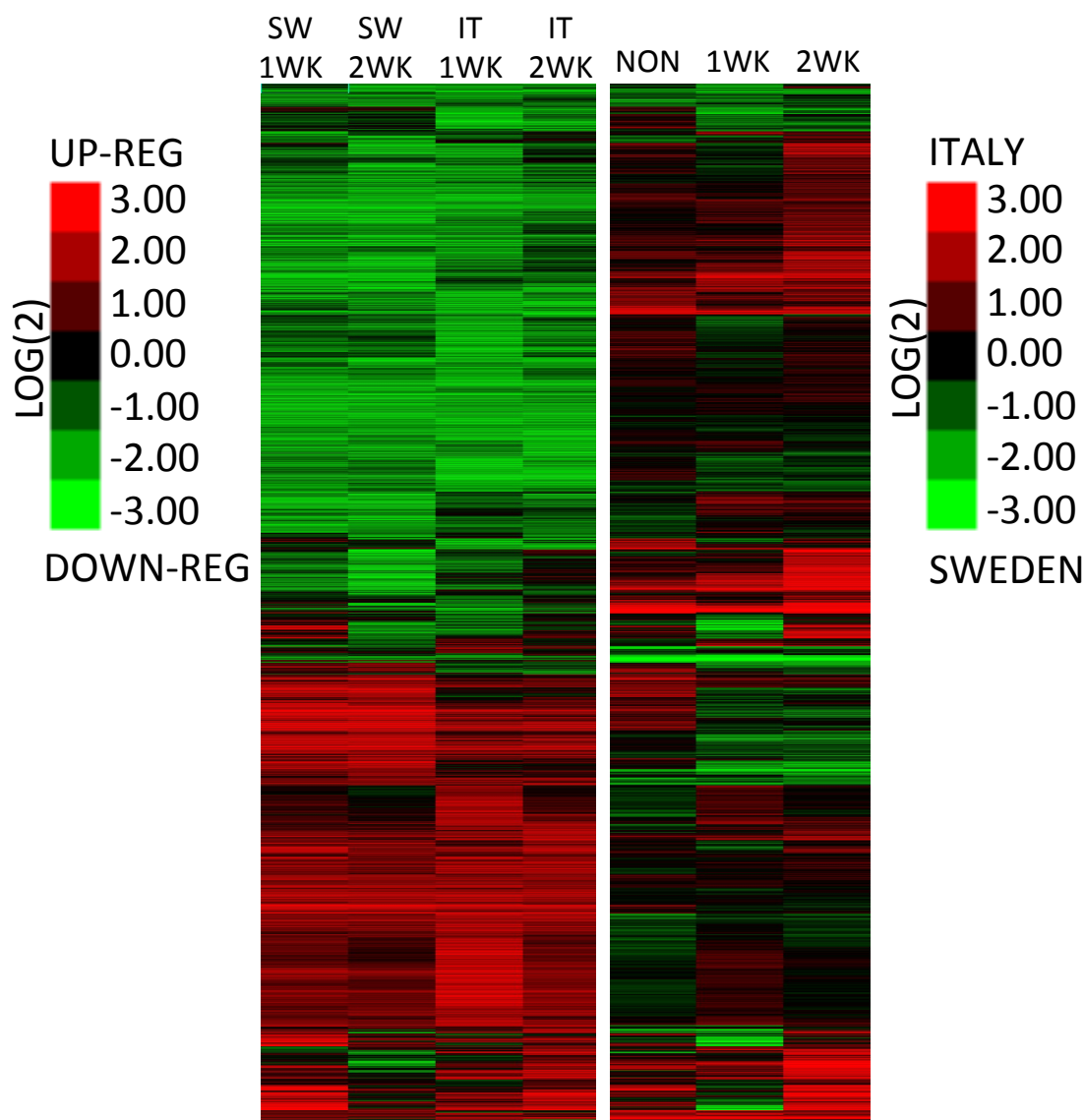


Figure 3.30. Fold change of cold regulated genes in SW and IT (7863 genes). Gene expression was measured by RNA-seq. The results are averages from three independent experiments ($n=3$). ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). Genes were considered differentially regulated between non-acclimated and cold conditions if the FDR corrected p-value was ≤ 0.050 . Left panel values are the logarithm (base=2) of the fold-change between non-acclimated and cold conditions; negative values denote genes down-regulated by cold (green), positive values denote genes up-regulated by cold (red). Right panel values are the logarithm (base=2) of the fold-change between SW and IT; negative values denote higher fold expression in SW (green) and positive values denote higher fold expression IT (red). Hierarchical clustering of genes was performed with the program, Cluster (40), and heatmap was generated using MapleTree (40). For interpretation of the references to color in this and all other figures, the reader is referred to the electronic version of this dissertation.

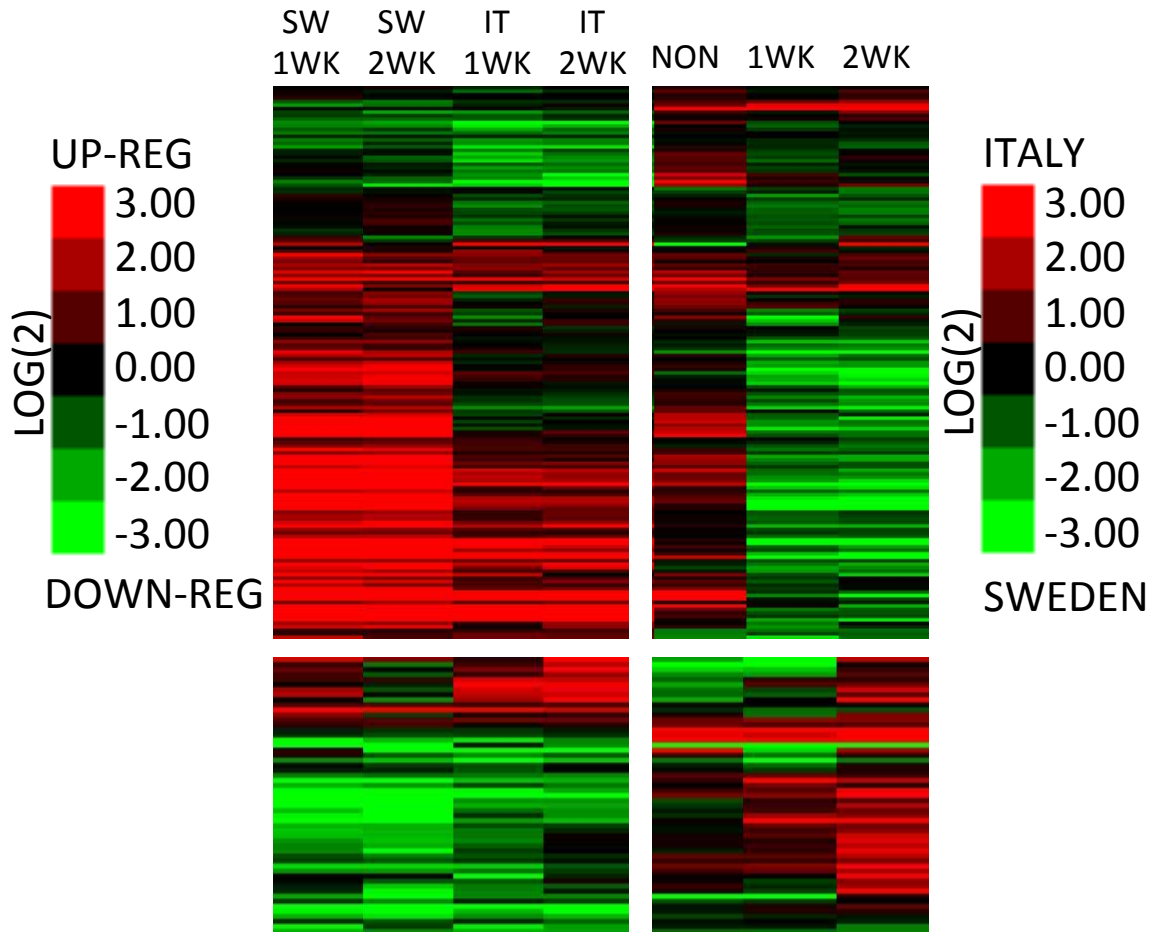


Figure 3.31. Fold change of CBF regulon genes in SW and IT. Top panels: 159 genes up-regulated by CBFs; Bottom panels: 55 genes down-regulated by CBFs (25). Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). Left panels are the logarithm (base=2) of the fold-change between non-acclimated and cold conditions; negative values denote genes down-regulated by cold (green), positive values denote genes up-regulated by cold (red). Right panels are the logarithm (base=2) of the fold-change between SW and IT; negative values denote higher fold expression in SW (green) and positive values denote higher fold expression IT (red). Hierarchical clustering of genes was performed with the program, Cluster (40), and heatmap was generated using MapleTree (40).

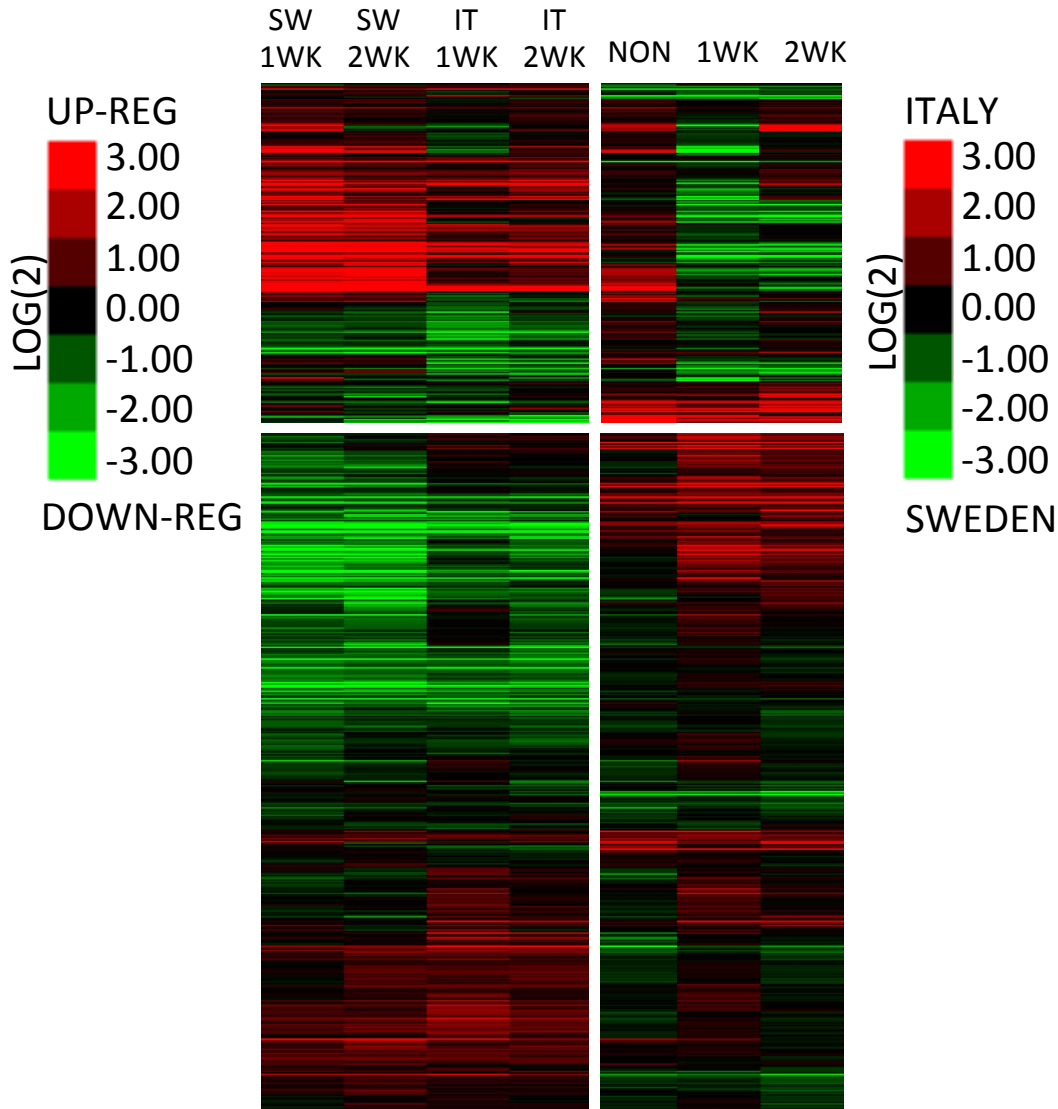


Figure 3.32. Fold change of Hannah genes in SW and IT. Top panels: 253 Hannah genes positively correlated with freezing tolerance; Bottom panels: 461 Hannah genes negatively correlated with freezing tolerance; (6). Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments ($n=3$). ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). Left panels are the logarithm (base=2) of the fold-change between non-acclimated and cold conditions; negative values denote genes down-regulated by cold (green), positive values denote genes up-regulated by cold (red). Right panels are the logarithm (base=2) of the fold-change between SW and IT; negative values denote higher fold expression in SW (green) and positive values denote higher fold expression IT (red). Hierarchical clustering of genes was performed with the program, Cluster (40), and heatmap was generated using MapleTree (40).

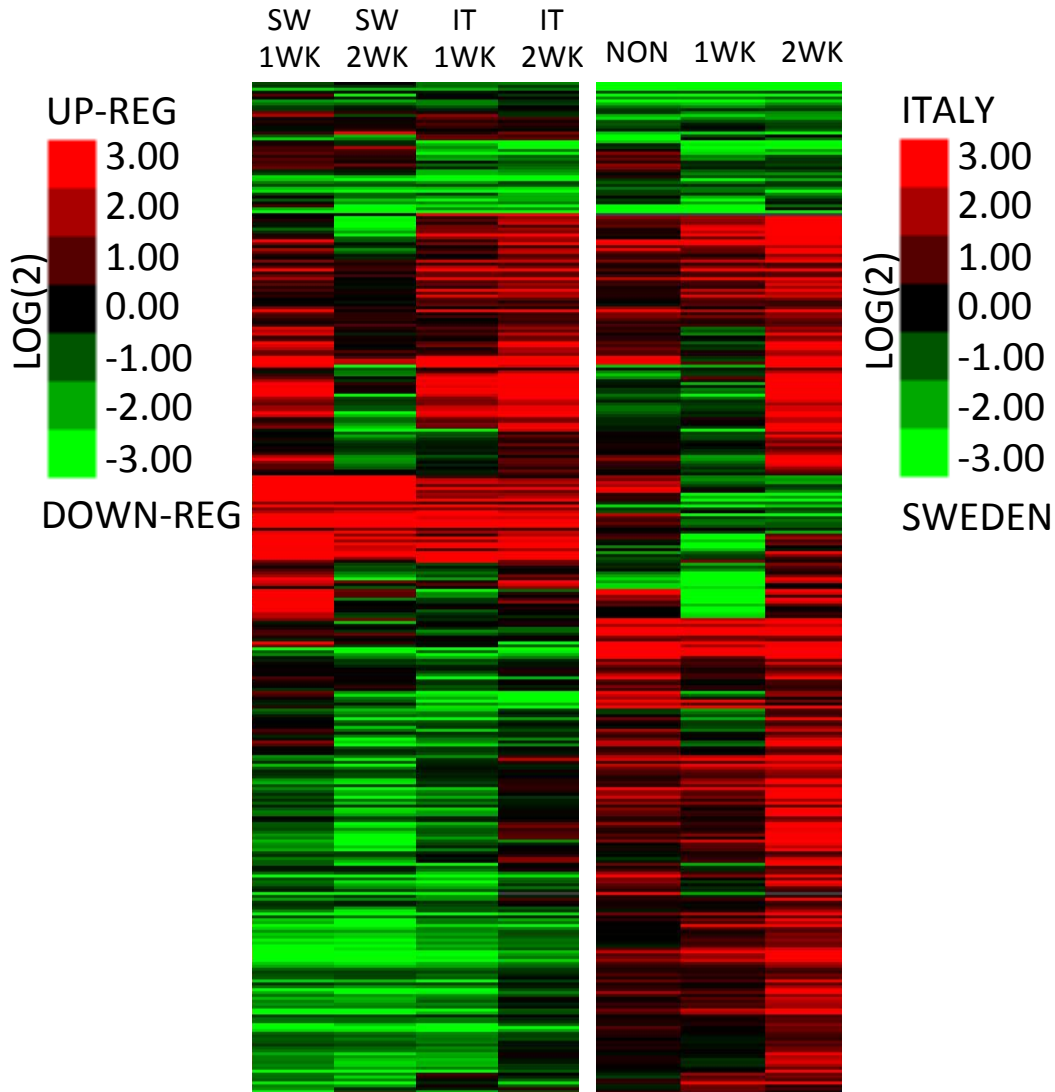


Figure 3.33. Fold change of differentially expressed biotic stress genes in SW and IT (348 genes). Gene expression was measured by RNA-seq. The results are averages from three independent experiments ($n=3$). ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). Genes were included if they were differentially expressed (FDR corrected p -value < 0.05) between SW and IT in any sample (non-acclimated, 1 week of cold-acclimation, or 2 weeks of cold-acclimation). Left panels are the logarithm (base=2) of the fold-change between non-acclimated and cold conditions; negative values denote genes down-regulated by cold (green), positive values denote genes up-regulated by cold (red). Right panels are the logarithm (base=2) of the fold-change between SW and IT; negative values denote higher fold expression in SW (green) and positive values denote higher fold expression IT (red). Hierarchical clustering of genes was performed with the program, Cluster (40), and heatmap was generated using MapTree (40).

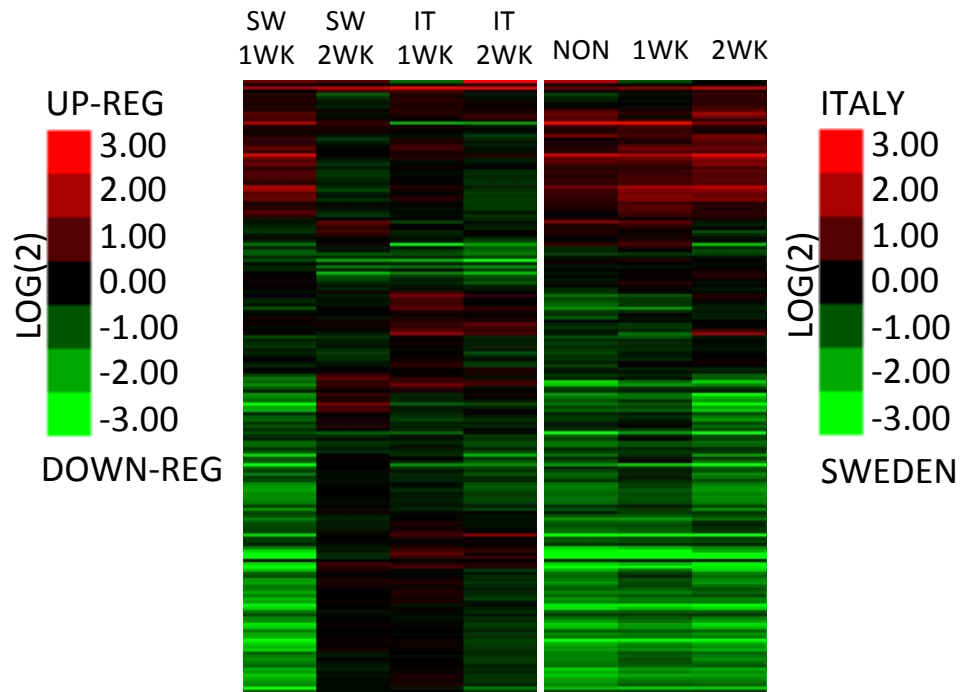


Figure 3.34. Fold change of differentially expressed photosynthesis associated genes in SW and IT (53 genes). Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). ≥ 3 FPKM was used as the cut-off for differential expression (19, 20). Genes were included in the heatmap if they were differentially expressed (FDR corrected p-value<0.05) between SW and IT in any sample (non-acclimated, 1 week of cold-acclimation, or 2 weeks of cold-acclimation). Left panels are the logarithm (base=2) of the fold-change between non-acclimated and cold conditions; negative values denote genes down-regulated by cold (green), positive values denote genes up-regulated by cold (red). Right panels are the logarithm (base=2) of the fold-change between SW and IT; negative values denote higher fold expression in SW (green) and positive values denote higher fold expression IT (red). Hierarchical clustering of genes was performed with the program, Cluster (40), and heatmap was generated using MapleTree (40).

RESULTS AND DISCUSSION: SW AND IT EQTL MAPPING

Summary: SW and IT eQTL mapping: In the second section of this study Category 1 and Category 2 FTD genes, which possibly contribute to differences in freezing tolerance between SW and IT accessions at 2 weeks of cold-acclimation, were determined. FTD genes are enriched in CBF regulon genes and QTL mapped for differences in expression (eQTL) for several CBF-regulated FTD candidate genes mapped to the CBF locus on chromosome 4. This eQTL on chromosome 4 also overlaps with a fitness QTL mapped in the SW and IT RILs (Table 3.1). Consequently, the CBFs are candidates for regulating differences in FTD gene expression and fitness between SW and IT. Analysis of IT-CBF2 coding sequence reveals a deletion in the activation domain. Based on previous studies conducted in the Thomashow lab, it is hypothesized that IT-CBF2 functions as a dominant negative version of CBF2. RNA-seq data is generally consistent with this hypothesis.

SW and IT eQTL mapping: The ultimate goal of this project is to determine the genes underlying the fitness QTL mapped using the SW and IT RILs (Table 3.1). In general, QTL peaks can span a large genomic region encompassing numerous genes (12, 13, 41). The fitness QTL found for the SW and IT RILs are certainly no exception (Table 3.1). For example, the fruit-set fitness QTL on chromosome 4 spans a region containing 2065 genes (Table 3.1). Many of these genes might contain polymorphisms, which may or may not contribute to fitness (12, 13). Moreover, 24% of genes in the *Arabidopsis* (Col-0) genome are included within SW and IT fitness QTL (Table 3.1).

Consequently, determining the specific genes responsible for a QTL can be extremely challenging (5, 12, 13, 41).

Minimum winter temperature was strongly correlated with fitness of the SW and IT parent lines in reciprocal transplant experiments (8), and freezing tolerance was found to be significantly different ($p \leq 0.026$ at 2 weeks of cold-acclimation) between SW and IT in laboratory experiments (Fig. 3.2). From analysis of RNA-seq data, genes that may contribute to differences in freezing tolerance between SW and IT have been defined (Table A3.3-A3.6). Similar to traditional QTL, an eQTL marks a position in the genome where a polymorphism is associated with differences in the relative abundance of a specific gene (12). Although correlative, eQTL mapped for FTD candidate genes, which also overlap with specific fitness QTL, can be determined. This will help to prioritize the tedious construction of near-isogenic lines (9) or complementation lines (41), which are needed to confirm if a gene affects freezing tolerance as well as plant fitness.

An eQTL is normally classified as being cis- or trans- to the gene used for mapping. Trans-eQTL are not physically linked to the gene of interest, and thus, are indicative of a polymorphism that alters the regulation of the gene of interest (12). Likely candidates for trans-eQTL are transcription factor polymorphisms. A cis-eQTL is located in or near the gene of interest. This can result when there is a deletion of an entire gene or a polymorphism in the promoter or coding region that affects regulation or function of that gene (12).

10 FTD candidate genes (Table 3.17; Fig. 3.35 and 3.36) were selected for eQTL analysis with the SW and IT RIL population generated by the Schemske Lab (Schemske and Agren, unpublished). From analysis of SW and IT transcriptomes in the second section of this study, CBF regulon genes are consistently differentially regulated between SW and IT. Therefore, 1-Category 1 and 3-Category 2 FTD genes that are also regulated by the CBFs, were selected for eQTL analysis (Table 3.17). Even if eQTL mapped for these CBF regulated genes do not overlap with fitness QTL, it will determine if differences in CBF regulated genes map cis- to the gene itself, or trans- to novel loci or the CBF locus. COR314, SZF1, and DI21 were chosen for eQTL analysis because they are FTD candidate genes not included in the CBF regulon, and thus represent genes that are potentially regulated by CBF-independent (42) pathways of cold-acclimation (Table 3.17). ZAT12, a Category 1 FTD gene was selected for eQTL analysis because ZAT12 regulon genes are enriched in the Hannah gene set (6), which was found to have differing enrichment in SW and IT in the second section of this study. Biotic stress was a GO category overrepresented in Category 2 FTD genes (Table 3.16) and a number of biotic stress genes appear to be differentially regulated by cold between SW and IT (Fig 3.33). Therefore, PR5, a Category 2 FTD gene that encodes a protein, which has been previously been shown to have antifreeze properties (26), was selected for eQTL analysis (Table 3.17). Photosynthesis was a GO category overrepresented in Category 1 FTD candidate genes (Table 3.13), and a number of photosynthesis related genes are differentially regulated between SW and IT (Fig. 3.34). Therefore ZAT10, a Category 1 FTD gene and a photosynthesis-related transcription

factor, was selected for eQTL analysis (Table 3.17). Altogether, 4 of these FTD genes have higher expression in IT and 6 have higher expression in SW at 2 weeks of cold-acclimation (Fig 3.35 and 3.36; Table A3.3-3.6).

SW and IT accessions, sampled at the same time as RNA-seq experiments, were also analyzed by qRT-PCR (Fig. 3.35 and 3.36). This ensured that expression differences measured by qRT-PCR were similar to RNA-seq experiments (Fig. 3.35 and 3.36). 544 genotyped RILs were grown similarly to plants used in electrolyte leakage experiments and sampled at 2 weeks of cold-acclimation, which is the acclimation period with the greatest difference in freezing tolerance (Fig. 3.2). For the RILs, expression of the 10 FTD genes was measured using qRT-PCR and differences were mapped using RQTL (15). RQTL was also used to calculate genome wide logarithm of odds (LOD) threshold values as well as the percent of phenotypic variance explained by each QTL (Table 3.18 and 3.19). Distinct eQTL above LOD thresholds were found for all genes except ZAT10 (Table 3.18 and 3.19).

Quantitative traits like differences in gene expression are defined by small and intermediate effect QTL (43). The average percent of variance explained by each eQTL in this study (Table 3.18 and 3.19) is lower (5.32%) than previous genome wide eQTL studies in *Arabidopsis*, which see percent variance explained by eQTL averaging from 10-30% (13, 44). However, the 'Beavis effect' indicates that percent variance of a QTL is greatly overestimated if only 100 lines are tested, and are only slightly overestimated with 500 lines tested (43). This study uses 544 RILs, more than double the number of RILs used in eQTL studies by West et al. 2007 (211 lines used) and Keurentjes et al.

2007 (160 lines used) (13, 44). Consequently, the lower percent variance explained by eQTL in this study may be due to the Beavis effect (43). When 160 SW and IT RILs are randomly selected for COR15A eQTL analysis, the same QTL on chromosome 4 is found (Table 3.18). However, the LOD score for the COR15A eQTL decreases from 10.09 to 6.19 with 160 samples, while the percent variance explained by the eQTL increases from 8.39% to 16.33% with 160 samples suggesting that the lower percent variance explained by eQTL with 544 RILs may be due to the Beavis effect (Table 3.18). The eQTL, which were found to overlap with fitness QTL (Table 3.1), will be analyzed next and candidates for the genes underlying them will be provided. Genes with eQTL that do not overlap with fitness QTL will be discussed in more detail in Chapter Four of this study.

Expression differences in CBF regulon gene, COR78, map to two eQTL (Fig 3.37; Table 3.1 and 3.18). The first eQTL maps to a region on chromosome 5, which overlaps with a previously identified fitness QTL (Fig 3.37; Table 3.1 and 3.18). This may be a cis-eQTL since the COR78 gene is located (between markers 60.60 and 61.28) within the eQTL peak on chromosome 5. COR78 is a Hannah gene positively correlated with increases in freezing tolerance and is regularly used as a marker gene for both cold and salt stress (6, 16, 45, 46). Although the coding sequence of COR78 is conserved between SW, IT and Col-0, it is known that the response of COR78 to cold and salt stress is driven through its promoter (47). In fact, the COR78 promoter is often employed as a stress-inducible promoter for other genes (48-52). Therefore, polymorphisms in this region are likely to influence expression. According to the effect

plot, the SW genotype at this eQTL is associated with higher expression of COR78 (Fig. 3.35). Currently, there is little evidence to indicate that COR78 alone can affect freezing tolerance (53). However, these results advocate sequencing of the COR78 promoter, construction of near-isogenic lines (9) for the region containing COR78 and suggest that complementation lines (41) should be made.

There is an alternate hypothesis for the COR78 eQTL on chromosome 5. COR78 is known to be regulated by cold-induced transcription factors CBF1-3 (24), which are located on chromosome 4. But CBF4, a closely related transcription factor, is located within the peak of the chromosome 5 eQTL (Fig 3.37; Table 3.19), and moreover between the same SW and IT RIL markers as COR78 (markers 60.60 and 61.28). CBF4 has also been shown to regulate COR78, and overexpression of CBF4 results in constitutive freezing and drought tolerance (54). CBF4 expression is not known to be cold-regulated but it is regulated by drought (54). The RNA-seq data is consistent with this result and CBF4 is not significantly ($p>0.050$) induced by cold (Fig. 3.43; Table 3.20). There are also no differences in CBF4 expression level between SW and IT (Fig. 3.43; Table 3.20). However, CBF4 may be regulated by cold post-transcriptionally and this regulation may differ between SW and IT. Therefore, CBF4 coding and promoter regions in SW and IT should be sequenced and complementation lines (41) for CBF4 should be made.

COR78, also has a trans-eQTL on chromosome 4, which explains more of the expression variance than the trans/cis-eQTL on chromosome 5 (Fig 3.37; Table 3.18). Interestingly, this trans-eQTL on chromosome 4 overlaps with a previously defined

fitness QTL (Table 3.1 and 3.20). 5 other FTD genes used for eQTL analysis, *COR15A*, *COR47*, *COR314*, *EARLY RESPONSIVE TO DEHYDRATION10* (*ERD10*) and *ZAT12*, also have an eQTL that overlaps with this fitness QTL on chromosome 4 (Fig. 3.37-3.42; Table 3.1, 3.18 and 3.19). *CBF1-3* are located in tandem array on chromosome 4 between SW and IT RIL markers 50.20 and 50.60. The CBF locus is thus contained within the chromosome 4 eQTL of all 6 FTD genes (Fig. 3.37-3.42; Table 3.1, 3.18 and 3.19). Expression of *COR15A*, *COR47*, *COR78*, and *ERD10* was mapped knowing that they were previously found to be regulated by the CBF genes (Table 3.17; (25)). Based on the CBF regulon defined by Vogel et al. 2005, expression differences mapping to the CBF locus for *COR314* and *ZAT12* were not expected (25). However, in a separate analysis of *CBF* overexpression lines by Maruyama et al. 2004, *COR314* was found to be regulated by CBF genes (55). Furthermore, though *ZAT12* is not part of the CBF regulon, it does have a regulatory relationship with the transcription factors (25).

The 6 effect plots for the chromosome 4 eQTL also all indicate that the SW genotype is associated with higher expression (Fig. 3.37-3.42). This is interesting considering that *ZAT12* has an opposite induction pattern compared to the other genes used for mapping (Fig. 3.35 and 3.36). However, the details of the CBF-*ZAT12* relationship have yet to be elucidated. *ZAT12* is a cold-regulated transcription factor known to induce and repress cold-responsive genes (25, 42). CBF and *ZAT12* regulons overlap and both regulons are overrepresented in the Hannah gene set (6, 25). Overexpression of *ZAT12* down-regulates *CBF* expression and these plants are slightly more freezing tolerant than wild-type plants, but not nearly as freezing tolerant as *CBF*

overexpressing plants (25). *CBF* and *ZAT12* overexpressing plants both have a dwarf phenotype. However plants overexpressing *both ZAT12* and *CBF2* are similar in size to wild-type plants (Doherty, unpublished). Consequently, it may be possible that the CBFs regulate *ZAT12* in some way, but further work is necessary to elucidate the relationship of the CBFs to *ZAT12*.

Previous natural variation studies on freezing tolerance have also implicated the CBFs as a source of variation. Alonso-Blanco et al. 2005 used RILs constructed from *Arabidopsis* accessions LER and CVI to map differences in freezing tolerance (10). Seven different QTL on chromosomes 1,4, and 5 were found, but the single QTL on chromosome 4, which contained the CBF locus, explained the most variance (10). CVI freezing tolerance was significantly reduced in comparison to LER, as was *CBF2* expression. The CVI-*CBF2* promoter was found to have a 1630 bp deletion (starting at -160), reducing the promoter region to 378 bp (10). Introduction of the Ler-*CBF2* transgene into CVI significantly reduced differences in freezing tolerance between LER and CVI (10).

CBF expression in the Versailles core collection of *Arabidopsis* (48 accessions tested) was also found to vary significantly (56). However, freezing tolerance did not correlate simply with *CBF* expression level in the 8 accessions tested (56). The most freezing-sensitive Versailles accessions were also the accessions with the lowest maximal cold-induced *CBF* expression levels. But the most freezing tolerant Versailles accessions varied significantly in their *CBF* expression, suggesting the importance of CBF-independent pathways of freezing tolerance for some accessions (56). It should

be pointed out, however, that in this Versailles study, *CBF* expression measured after hours in cold was correlated with freezing tolerance assays conducted after 2 weeks of cold-acclimation (56). Therefore in the Versailles study, *CBF* expression and freezing tolerance assays were conducted at very different time-points. In future studies, *CBF* expression should be measured at several time-points, but especially at time-points just as to 2 week cold-acclimated freezing tolerance assays are conducted to determine if *CBF* expression and CBF regulon gene expression correlates with differences in freezing tolerance between accessions.

Freezing tolerance QTL differed between two Versailles RIL populations (56). However, in both populations a common QTL on chromosome 4 explained the most variance in freezing tolerance and overlapped with the CBF locus (56). Further analysis of CBF gene sequence in the 48 Versailles accessions showed a number of polymorphisms in both the promoter and coding sequences of CBF genes (57). The most notable of these accessions were BI-1 with a 10 amino acid deletion at the end of the activation domain in CBF1, Condara with a premature stop codon in the activation domain of CBF3, and Gre-0 with a glycine to arginine change in the activation domain of CBF2 (57). Unfortunately, these 3 particular accessions were not chosen for freezing tolerance assays or tested for CBF target gene expression (57).

A recent study by Zhen et al. 2008 further showed that CBF genes undergo relaxed selection in *Arabidopsis* accessions native to southern latitudes(58). Analysis of CBF sequence in 24 accession of *Arabidopsis* (9 accessions overlap with 48 Versailles accessions previously mentioned (56, 57)) found a large bias toward nonsynonymous

changes in the activation domain compared to DNA binding domain of CBF genes (58). Southern latitude accession had 1.3 to 4.6-fold more polymorphisms in the individual CBF genes and also had reduced expression of CBF target genes (58).

CBF1 and CBF3 coding sequences were found to be conserved between SW and IT. Transcripts for CBF1 and CBF3 are also not significantly ($p > 0.050$) differentially regulated between SW and IT at 2 weeks of cold-acclimation in RNA-seq experiments (Fig. 3.43; Table 3.20). This suggests that any polymorphisms in CBF1 and CBF3 promoters have little effect on gene expression. On the other hand, CBF2 transcripts are differentially regulated between SW and IT at 2 weeks of cold-acclimation (Fig. 3.43; Table 3.20). CBF2 expression is 1.5-fold higher in SW compared to IT at 2 weeks of cold-acclimation (Table 3.20). CBF1, 2 and 3 are known to regulate over a hundred target genes (25, 59, 60). Therefore, polymorphisms in the CBF2 promoter could contribute to differences in *CBF2* expression and have compounding effects on target gene regulation. The coding sequence of CBF2 is conserved between SW and Col-0 (Fig. 3.44). However, there is a 13 basepair (bp) deletion (+384 to +397) in the activation domain (61) of IT-CBF2, though surrounding coding sequences are conserved between SW, IT, and Col-0 (Fig 3.44). This leads to a predicted protein (Fig. 3.42) with a nuclear localization signal (NLS) and DNA binding domain, but missing activation domain (61). This predicted protein would be similar to the Versailles accession, Condara, which has a premature stop codon in the activation domain of CBF3 (57). Freezing tolerance and CBF target gene expression was not tested in

Condara (56, 57). However, Condara freezing tolerance and CBF target gene expression may be predicted to be diminished in comparison to SW, similar to IT.

A similar truncated version of CBF2 (*CBF2ΔC*), predicted to have a missing activation domain, was previously identified through an EMS screen of *CBF2* overexpressing plants (Gilmour and Thomashow, unpublished). At 2 weeks of cold-acclimation, the combined expression level of IT-CBF1 (3.02 FPKM) and IT-CBF3 (35.85 FPKM) is similar to IT-CBF2 (33.23 FPKM). Interestingly, *CBF2ΔC* was shown to function as a dominant negative protein in plants with equal expression of *CBF2* and *CBF2ΔC* (Doherty, unpublished). *CBF2ΔC* overexpressing plants are also significantly impaired in cold-acclimation (Doherty, unpublished) and IT is less freezing tolerant than SW at 2 weeks of cold-acclimation. Transcriptional analysis of *CBF2ΔC* overexpressing plants revealed that 40% of cold-induced genes were affected (Doherty, unpublished). CBF regulon genes are overrepresented in genes differentially expressed between SW and IT; Category 1 FTD genes include 61 CBF regulon genes ($p=4.3E-13$; Table A3.3-3.4) and Category 2 FTD genes include 51 gene CBF regulon genes ($p<2.2E-16$; Table A3.5-A3.6). In general, cold-regulated gene expression is consistent with IT-CBF2 acting as a dominant negative version of CBF2; genes up-regulated by CBFs have lower expression in IT than SW and gene down-regulated by CBFs have higher expression in IT than SW (Fig 3.31). Furthermore, there are fewer CBF regulon genes up-regulated by cold in IT in comparison to SW (Fig 3.31). Based on these results, I postulate that IT-CBF2 may function as a natural version of *CBF2ΔC*, in addition to effects of possible promoter polymorphisms. More experiments are necessary to test

this hypothesis, which would include determining CBF1-3 protein levels in IT and SW, as well as transforming the IT-CBF2 and SW-CBF2 into SW and IT, respectively.

It is interesting that differences in freezing tolerance are not seen at both 1 and 2 weeks of cold-acclimation since IT-CBF2 would presumably act as a dominant negative protein at both times. Category 2 FTD candidate genes are enriched in CBF regulon genes and were defined as cold-regulated genes that are differentially expressed between SW and IT at both 1 and 2 weeks of cold acclimation. Therefore, IT-CBF2 does appear to be acting as a dominant negative version of CBF2 at both 1 and 2 weeks of cold-acclimation. However, there are also known to be CBF-independent pathways of cold acclimation (42), which are of varying importance to *Arabidopsis* accessions (56). Therefore, there may be CBF-independent pathways of cold-acclimation, which compensate for the dominant negative IT-CBF2 at 1 week of cold but not 2 weeks. The analysis of IT transcriptomes at 1 and 2 weeks of cold-acclimation did reveal numerous genes, which are exclusively cold-regulated at either 1 or 2 weeks of cold-acclimation (Fig. 3.6).

It is also of note that at 2 weeks of cold-acclimation SW freezing tolerance increases in addition to IT freezing tolerance decreasing. *SW-CBF2* expression is significantly higher in comparison to *IT-CBF2* at only 2 weeks of cold-acclimation. Therefore, in addition to IT-CBF2 possibly acting as a dominant negative protein, reducing freezing tolerance through interruption of the CBF pathway of cold-acclimation, polymorphisms in the *SW-CBF2* promoter could be contributing to higher *CBF2* expression and subsequent increase in freezing tolerance at 2 weeks of cold-

acclimation. Overall, CBF2 appears to be an excellent candidate gene to explain differences in cold-regulated gene expression and differences in freezing tolerance and fitness between SW and IT. Sequencing the CBF2 promoter in SW and IT, near-isogenic lines (9) for the region containing CBF2, and complementation lines (41) should be made a top priority in order to test how great a role CBF2 plays in differential expression of FTD candidate genes, freezing tolerance, and fitness.

Table 3.17. 10 FTD candidate genes for eQTL analysis. Genes are ordered by their AGI numbers. Genes were considered significantly different between SW and IT under non-acclimated conditions if they had a Benjamini-Hochberg corrected p-value (FDR P-VALUE) of <0.05 and a minimum of ≥ 3 FPKM in either SW or IT. The differential expression of each gene is also represented by the logarithm (base=2) of the fold change (LOG2.FC). This table also denotes if a gene was previously described as positively or negatively correlating with freezing tolerance (HANNAH;(6)), a transcription factor (TF), a gene previously described as up- or down-regulated by 1 week of cold (COS, (25)), identified as an up- or down-regulated component of the CBF regulon (CBF REGULON; (25)) or identified as a photosynthesis associated genes (PAG; Hu and Thomashow, unpublished). Shaded genes have higher expression in SW and non-shaded genes have higher expression in IT.

CATEGORY 1 FTD CANDIDATES FOR eQTL ANALYSIS

AGI	NAME	SHORT DESCRIPTION	SW.IT.2WK LOG2FC	SW.IT.2WK FDR P-VALUE	HANNAH?	TF?	COS?	CBF REG?	PAG?
AT1G27730	ZAT10	Salt Tolerance Zinc Finger	2.11	0.00E+00	NO	YES	UP	NO	YES
AT1G29390	COR314	Cold Regulated 314	-0.97	1.74E-08	NO	NO	NO	NO	NO
AT2G42540	COR15A	Cold-Regulated 15a	-1.88	2.90E-08	NO	NO	UP	UP	NO
AT3G55980	SZF1	Salt-Inducible Zinc Finger 1	1.63	0.00E+00	NO	YES	DOWN	NO	NO
AT5G59820	ZAT12	C2H2-Type Zinc Finger Family	1.75	1.85E-14	NO	YES	NO	NO	NO

CATEGORY 2 FTD CANDIDATES FOR eQTL ANALYSIS

AGI	NAME	SHORT DESCRIPTION	SW.IT.2WK LOG2FC	SW.IT.2WK. FDR P-VALUE	HANNAH?	TF?	COS?	CBF REG?	PAGs
AT1G20440	COR47	Cold-Regulated 47	-2.51	0.00E+00	NO	NO	UP	UP	NO
AT1G20450	ERD10	Dehydrin Family Protein	-1.96	0.00E+00	NO	NO	UP	UP	NO
AT1G75040	PR5	Pathogenesis-Related 5	4.50	0.00E+00	NO	NO	UP	NO	NO
AT4G15910	DI21	Drought-Induced 21	-2.58	0.00E+00	NO	NO	NO	NO	NO
AT5G52310	COR78	Cold-Regulated 78	-2.83	0.00E+00	POSITIVE	NO	UP	UP	NO

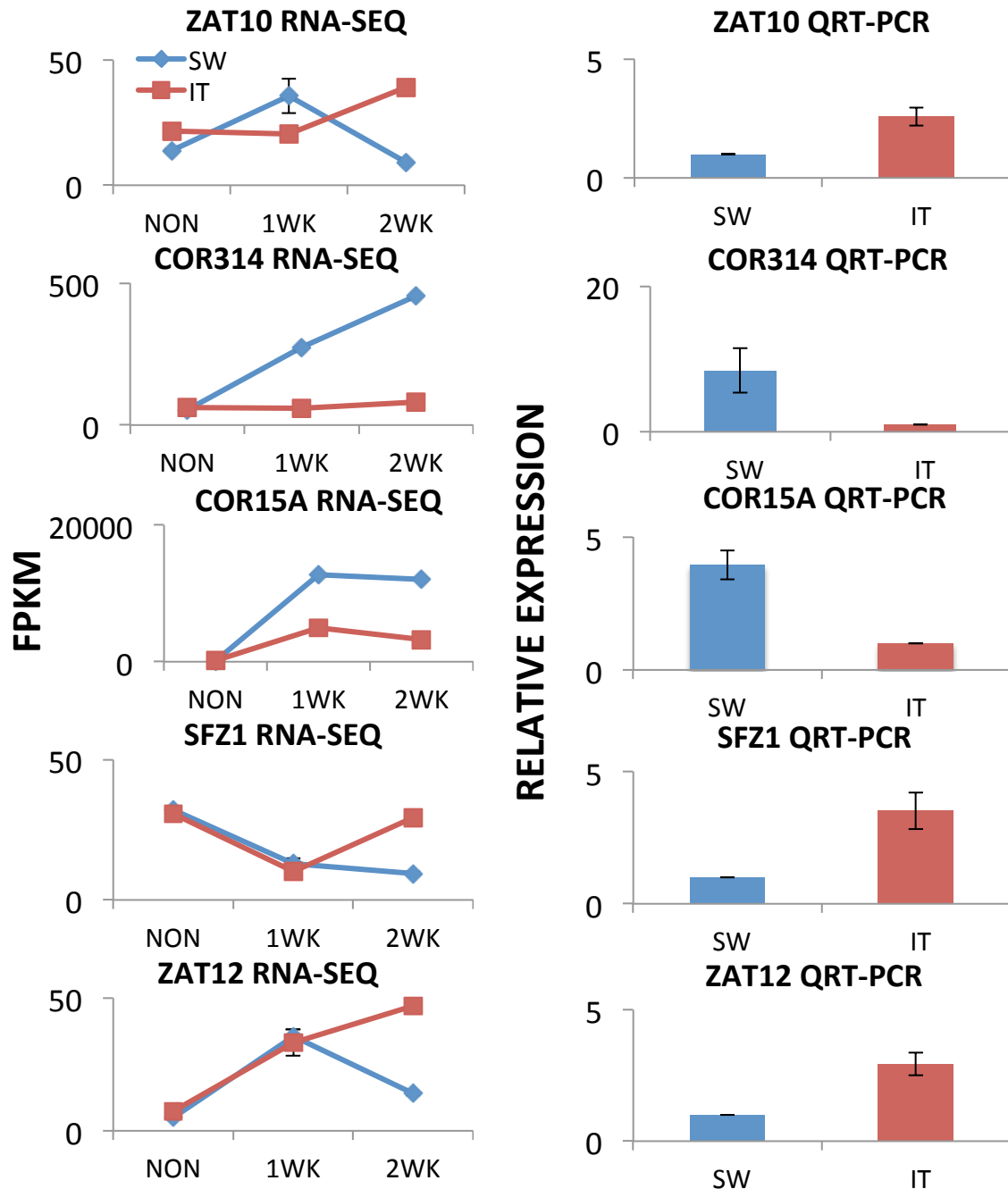


Figure 3.35. Category 1 FTD candidate genes for eQTL analysis. IT samples are shown in red, and SW in blue. The results presented are average values from three independent experiments (n=3). Error bars indicate \pm SEM. For RNA-seq experiments, plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 or 2 weeks under a 12 h photoperiod then sampled. For qRT-PCR experiments, plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was transferred to 4°C for 2 weeks under a 12 h photoperiod then sampled. IPP2 was used as a housekeeping gene. Primer sequences can be found in Table A3.6.

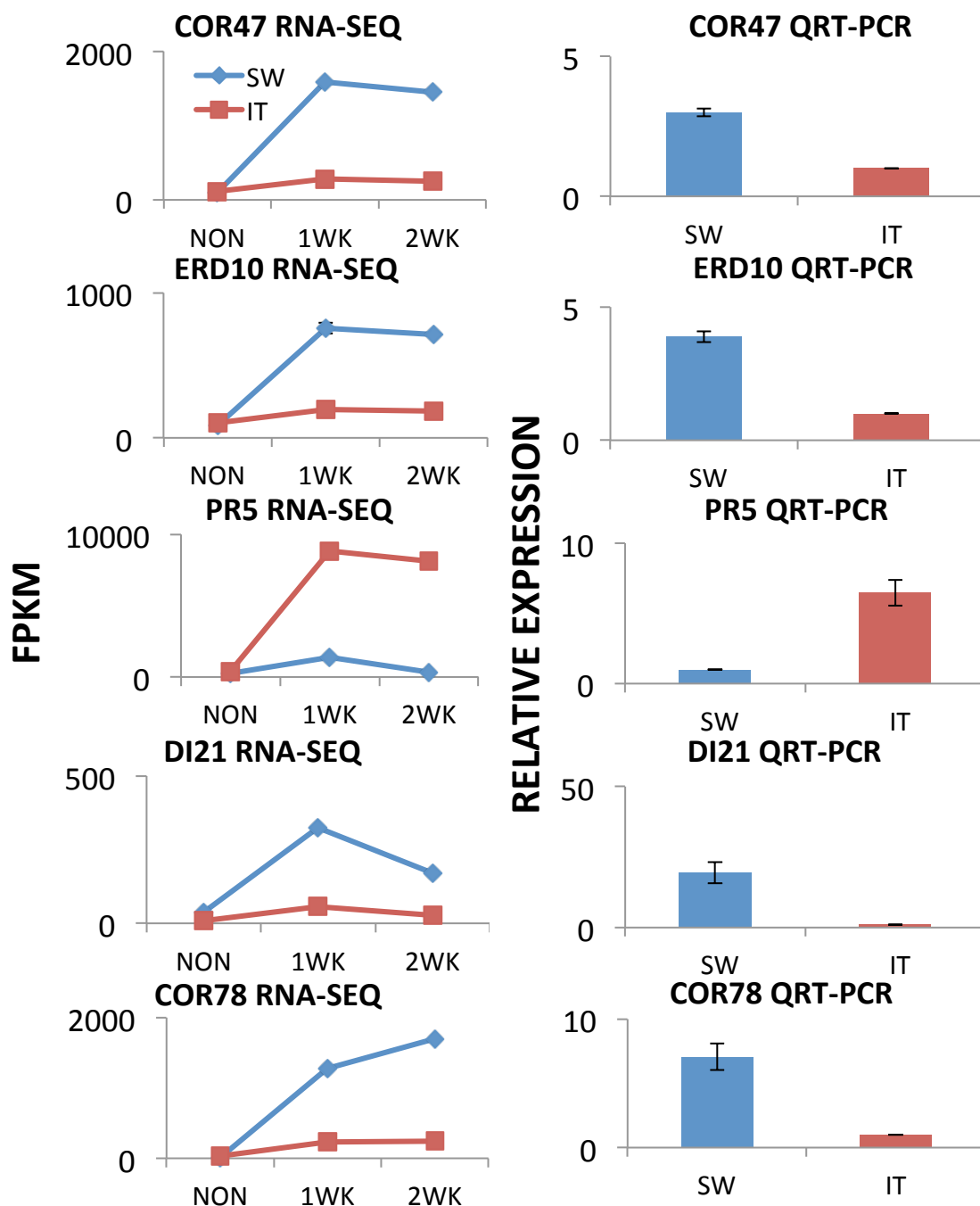


Figure 3.36. Category 2 FTD candidate genes for eQTL analysis. IT samples are shown in red, and SW in blue. The results presented are average values from three independent experiments (n=3). Error bars indicate \pm SEM. For RNA-seq experiments, plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 or 2 weeks under a 12 h photoperiod then sampled. For qRT-PCR experiments, plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was transferred to 4°C for 2 weeks under a 12 h photoperiod then sampled. IPP2 was used as a housekeeping gene. Primer sequences can be found in Table A3.6.

Table 3.18. Locations of eQTL for Category 1 FTD candidate genes. 544 RILs were used for eQTL mapping. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>). To detect eQTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LOD) values were calculated for each trait using 1000 permutations in RQTL. To estimate the location of each eQTL a Bayes credible interval (99%) was calculated in RQTL. For locations of markers based on the Col-0 genome please see Table A3.1. QTL are categorized as cis- or trans- (CIS/TRANS?). Shaded QTL overlap with previously identified fitness QTL (Schemske and Agren, unpublished). 'MB' indicates the number of megabases contained within the eQTL. '# GENES' indicates the number of genes contained within the eQTL based on the Col-0 genome. %VAR indicates the estimated proportion of phenotypic variance explained by the QTL.

AT1G27730-ZAT10 (LOD THRESHOLD=3.04)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
NO QTL								
AT1G29390-COR314 (LOD THRESHOLD=3.33)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
4	50.00	55.52	58.00	4.94	1575	TRANS	6.35	5.35
AT2G42540-COR15A (LOD THRESHOLD=3.13)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
4	50.000	50.004	55.520	3.790	1153	TRANS	10.09	8.390
AT3G55980-STZ1 (LOD THRESHOLD=2.81)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
4	32.00	33.42	36.00	1.73	477	TRANS	5.46	3.85
5	20.00	22.00	36.74	3.50	805	TRANS	3.87	2.40
AT5G59820-ZAT12 (LOD THRESHOLD=3.15)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
3	9.64	11.18	22.00	3.20	951	TRANS	5.25	4.10
4	50.00	50.60	56.60	4.66	1410	TRANS	4.46	4.37
5	10.00	48.00	56.60	15.66	4071	TRANS	6.54	4.91

Table 3.19. Locations of eQTL for Category 2 FTD candidate genes. 544 RILs were used for eQTL mapping. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>). To detect eQTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LOD) values were calculated for each trait using 1000 permutations in RQTL. To estimate the location of each eQTL a Bayes credible interval (99%) was calculated in RQTL. For locations of markers based on the Col-0 genome please see Table A3.1. QTL are categorized as cis- or trans- (CIS/TRANS?). Shaded QTL overlap with previously identified fitness QTL (Schemske and Agren, unpublished). ‘MB’ indicates the number of megabases contained within the eQTL. ‘# GENES’ indicates the number of genes contained within the eQTL based on the Col-0 genome. %VAR indicates the estimated proportion of phenotypic variance explained by the QTL.

AT1G20440-COR47 (LOD THRESHOLD=2.83)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
1	10	14.05	18.12	3.19	967	CIS	7.59	6.89
4	46.00	50.20	58.00	5.77	1977	TRANS	6.76	6.35

AT1G20450-ERD10 (LOD THRESHOLD=3.25)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
4	50.00	55.52	56.30	4.36	1322	TRANS	12.79	10.50

AT1G75040-PR5 (LOD THRESHOLD=3.05)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
2	40.25	46.95	48.04	1.73	450	TRANS	6.07	3.87
3	9.64	11.18	12.64	1.16	356	TRANS	7.63	4.25
5	42.00	54.27	56.60	4.68	1697	TRANS	5.87	4.63

AT4G15910-DI21 (LOD THRESHOLD=2.85)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
4	34.00	34.49	35.77	0.64	179	CIS	28.96	11.92

AT5G52310-COR78 (LOD THRESHOLD=3.00)								
CHR	LEFT (CM)	PEAK (CM)	RIGHT (CM)	MB	# GENES	CIS/ TRAN?	LOD	%VAR
4	48.00	55.52	60.44	6.35	1910	TRANS	6.39	5.18
5	54.83	64.62	78.17	7.83	2342	CIS	4.20	3.50

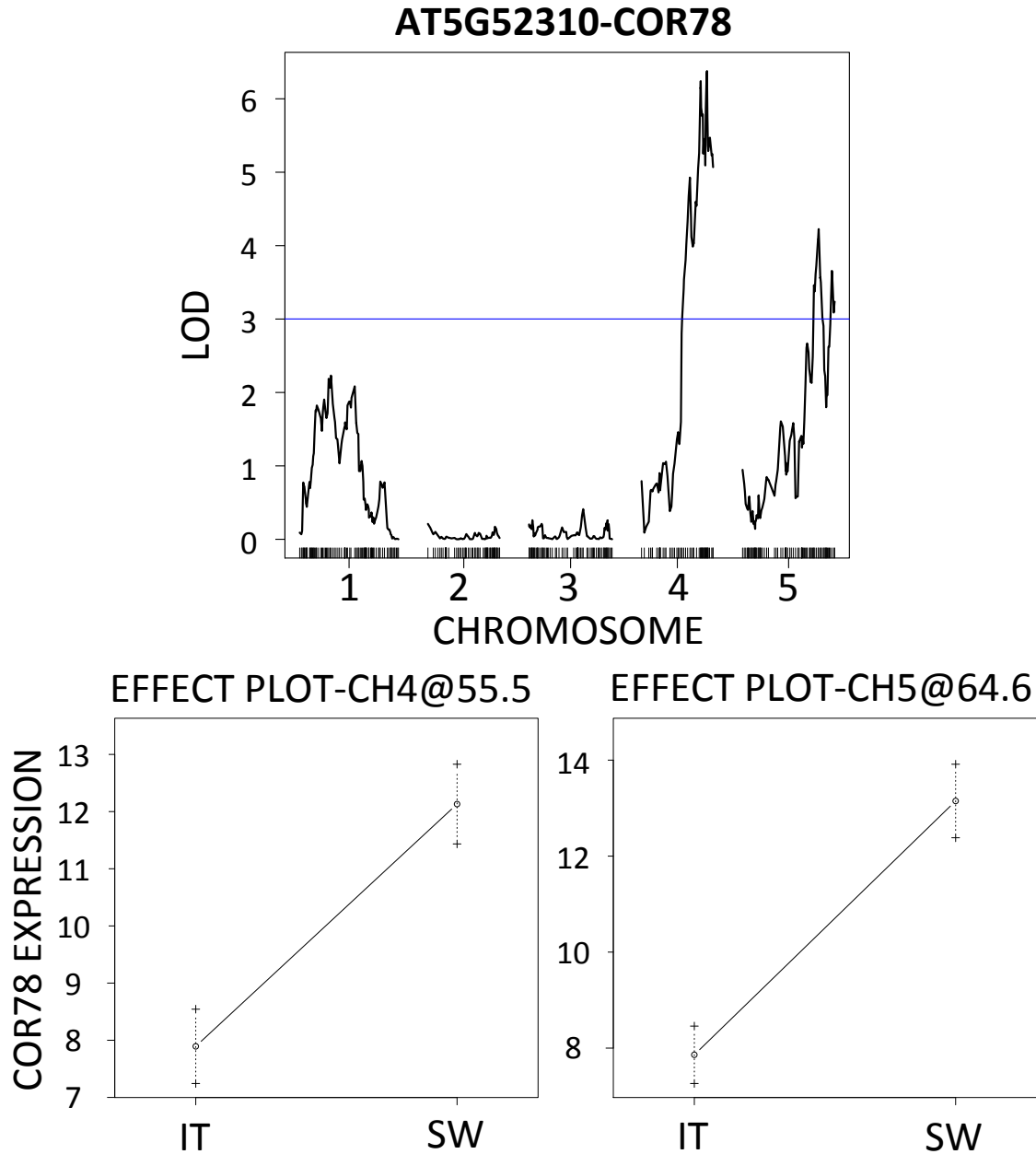


Figure 3.37. eQTL for Category 2 FTD gene COR78. 544 RILs were used for eQTL mapping by qRT-PCR. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>) for each fitness trait at each location. To detect QTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LODs) values were calculated for each trait using 1000 permutations in RQTL (shown in blue). Effects plots represent expression averages for the genotype groups (IT or SW) at eQTL on chromosome 4 and 5. Effect plots were constructed using 'effectplot' function in R/QTL(15).

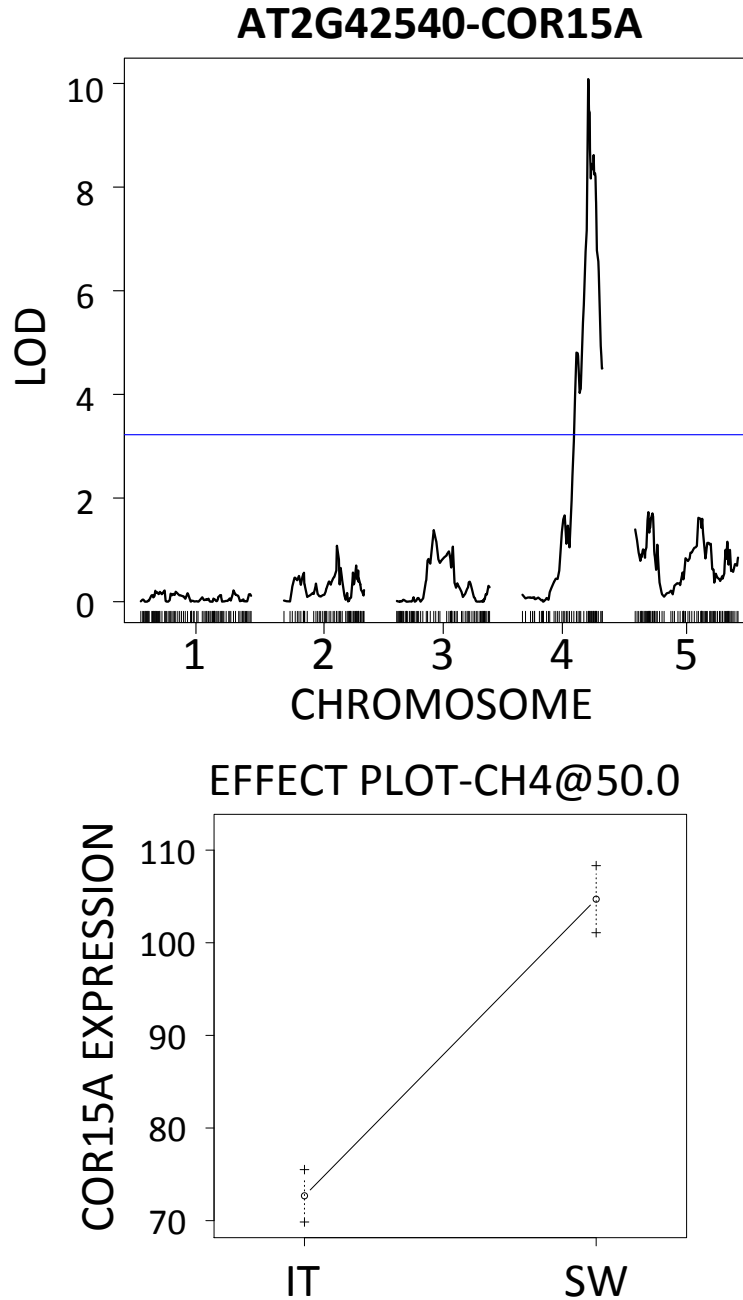


Figure 3.38. eQTL for Category 1 FTD gene COR15A. 544 RILs were used for eQTL mapping by qRT-PCR. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>) for each fitness trait at each location. To detect QTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LODs) values were calculated for each trait using 1000 permutations in RQTL (shown in blue). Effect plot represents expression averages for the genotype groups (IT or SW) at eQTL on chromosome 4. Effect plots were constructed using 'effectplot' function in R/QTL(15).

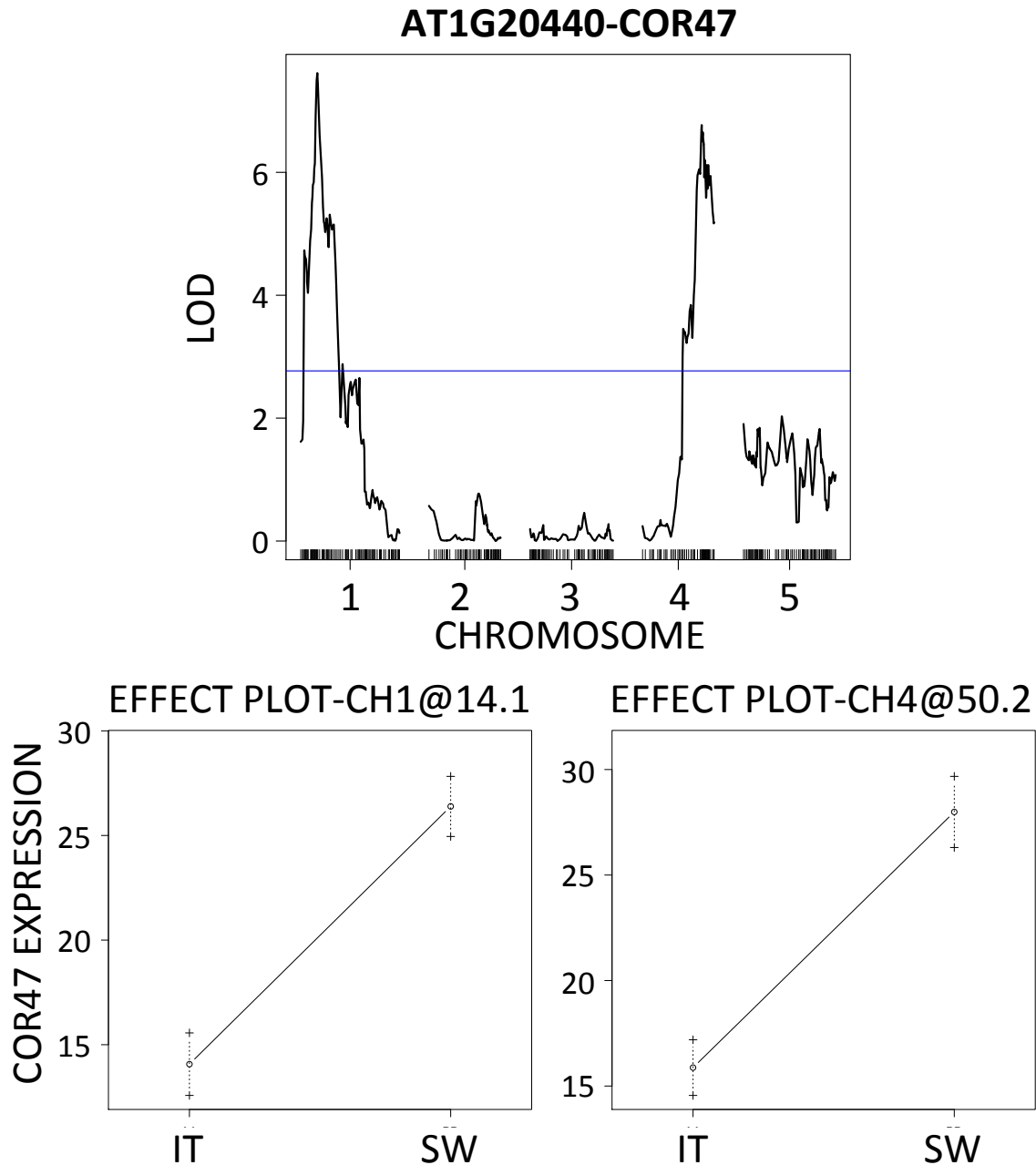


Figure 3.39. eQTL for Category 2 FTD gene COR47. 544 RILs were used for eQTL mapping by qRT-PCR. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>) for each fitness trait at each location. To detect QTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LODs) values were calculated for each trait using 1000 permutations in RQTL (shown in blue). Effect plots represent expression averages for the genotype groups (IT or SW) at eQTL on chromosome 1 and 4. Effect plots were constructed using 'effectplot' function in R/QTL(15).

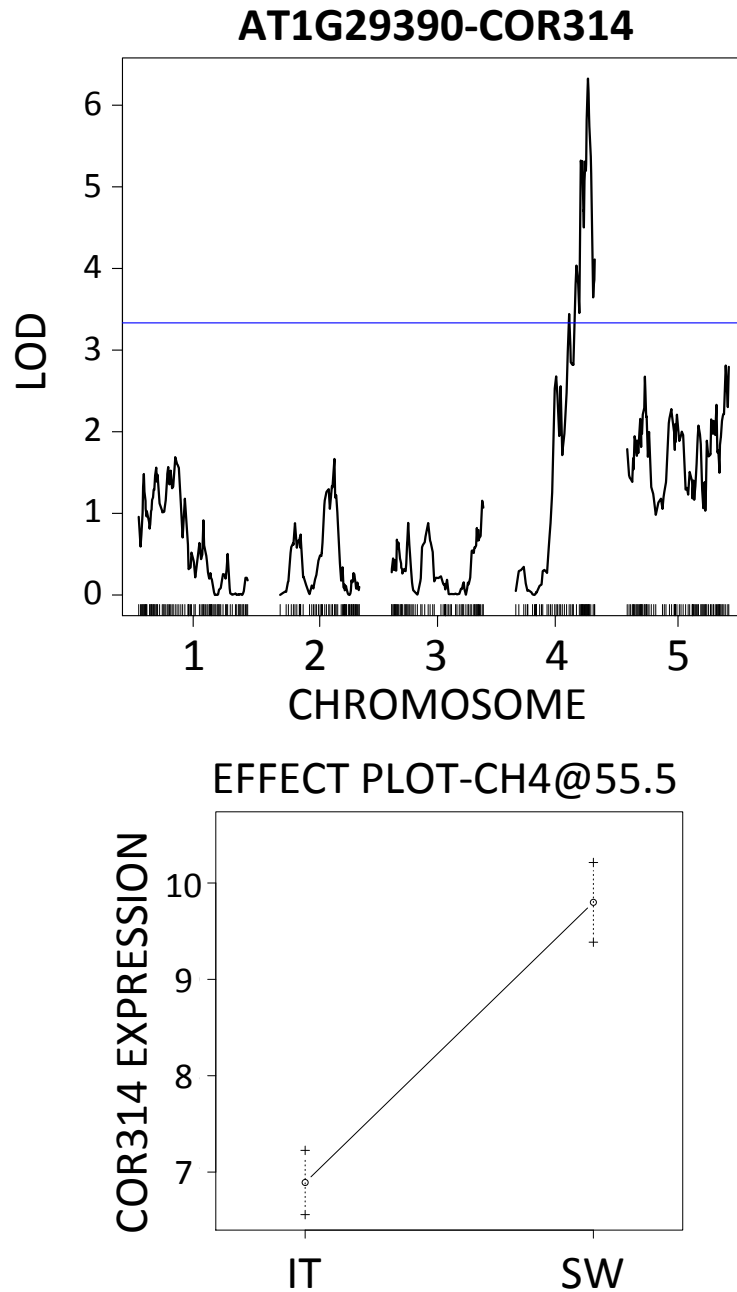


Figure 3.40. eQTL for Category 1 FTD gene COR314. 544 RILs were used for eQTL mapping by qRT-PCR. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>) for each fitness trait at each location. To detect QTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LODs) values were calculated for each trait using 1000 permutations in RQTL (shown in blue). Effect plot represents expression averages for the genotype groups (IT or SW) at eQTL on chromosome 4. Effect plots were constructed using 'effectplot' function in R/QTL(15).

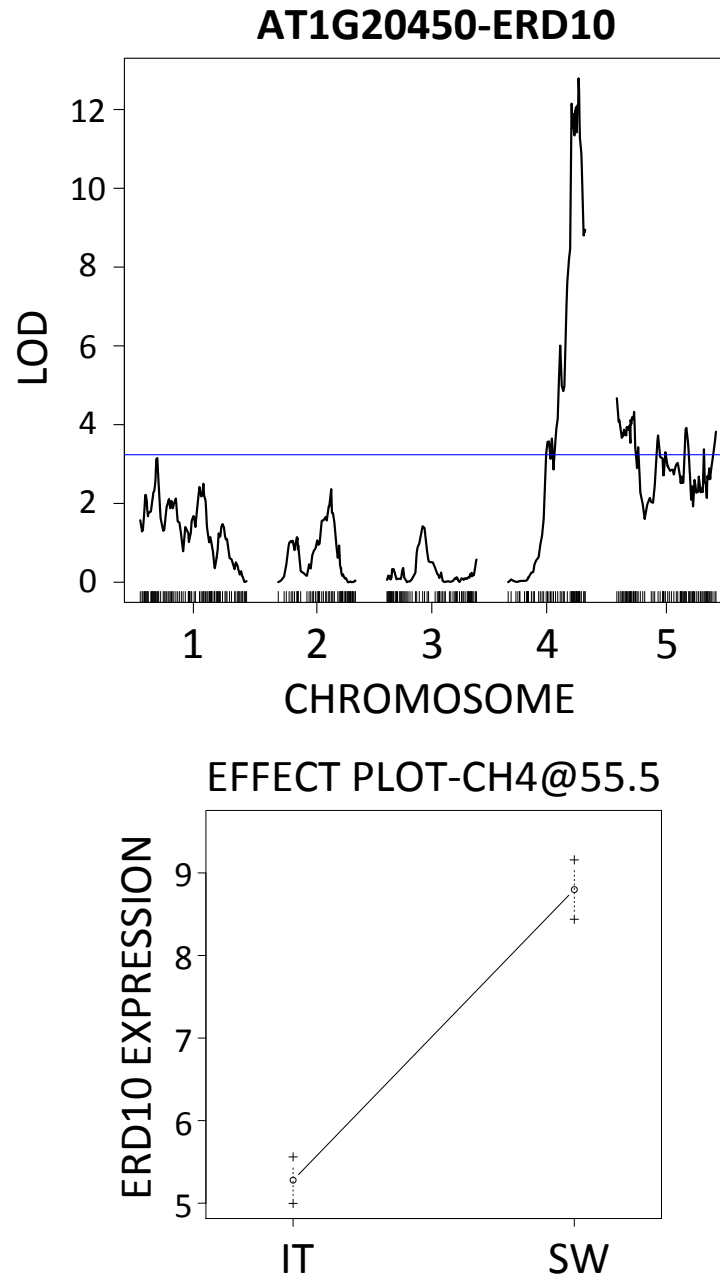


Figure 3.41. eQTL for Category 2 FTD gene ERD10. 544 RILs were used for eQTL mapping by qRT-PCR. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>) for each fitness trait at each location. To detect QTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LODs) values were calculated for each trait using 1000 permutations in RQTL (shown in blue). Effect plot represents expression averages for the genotype groups (IT or SW) at eQTL on chromosome 4. Effect plots were constructed using 'effectplot' function in R/QTL(15).

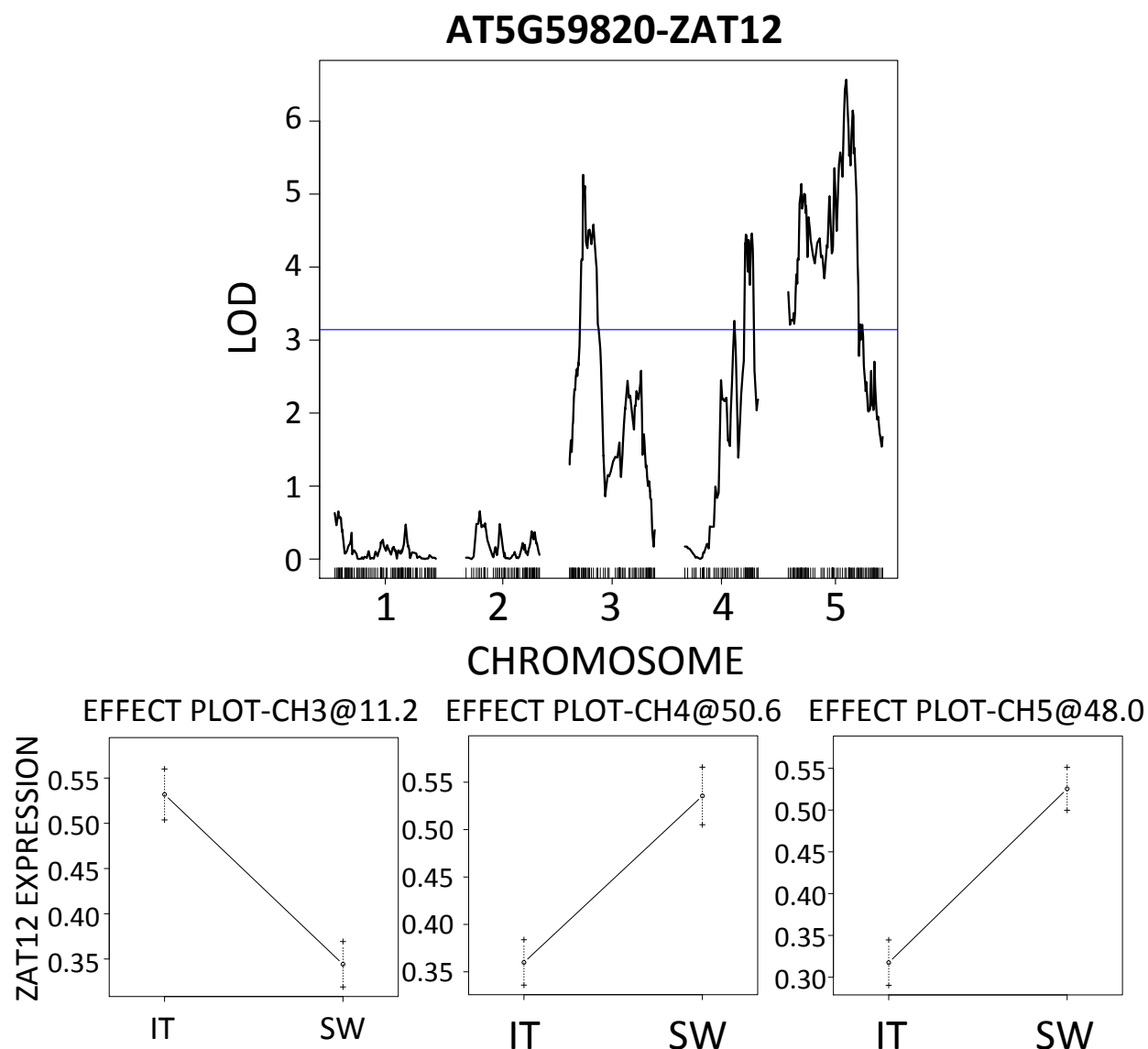


Figure 3.42. eQTL for Category 1 FTD gene ZAT12. 544 RILs were used for eQTL mapping by qRT-PCR. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>) for each fitness trait at each location. To detect QTL, estimated genome-wide significance thresholds ($\alpha=0.01$) for the logarithm of odds (LODs) values were calculated for each trait using 1000 permutations in RQTL (shown in blue). Effect plots represent expression averages for the genotype groups (IT or SW) at eQTL on chromosomes 1, 4 and 5. Effect plots were constructed using 'effectplot' function in R/QTL(15).

Table 3.20. Expression of CBF genes. Gene expression was measured by RNA-seq. The results presented are average values from three independent experiments (n=3). Comparisons of SW and IT under non-acclimated conditions, 1 week of cold-acclimation, and 2 weeks of cold-acclimation and the corrected p-value (FDR P-VALUE) for that comparison. The comparison of each gene is also represented by the logarithm (base=2) of the fold change (LOG2FC). This table also denotes if a gene was previously described as positively or negatively correlating with freezing tolerance (HANNAH;(6)), a transcription factor (TF), a gene previously described as up- or down-regulated by 1 week of cold (COS, (24)), identified as an up- or down-regulated component of the CBF regulon (CBF REGULON; (24)). Shaded genes have significantly different expression between SW and IT at 2 weeks of cold-acclimation.

AGI	NAME	SW.IT. NON	NON.FDR. P-VALUE	SW.IT. 1WK	1WK.FDR. P-VALUE	SW.IT. 2WK	2WK.FDR. P-VALUE	HANNAH?	TF?	COS?	CBF REG?
AT4G25490	CBF1	1.66	5.32E-01	-0.33	8.52E-01	-0.73	2.03E-01	POSITIVE	YES	NO	NO
AT4G25470	CBF2	0.55	7.41E-01	-0.40	5.24E-01	-0.58	8.06E-03	NO	YES	UP	UP
AT4G25480	CBF3	1.76	2.01E-02	-0.44	4.65E-01	-0.40	6.84E-02	NO	YES	UP	NO
AT5G51990	CBF4	0.00	1.00E+00	-1.96	3.85E-01	0.89	6.76E-01	NO	YES	NO	NO

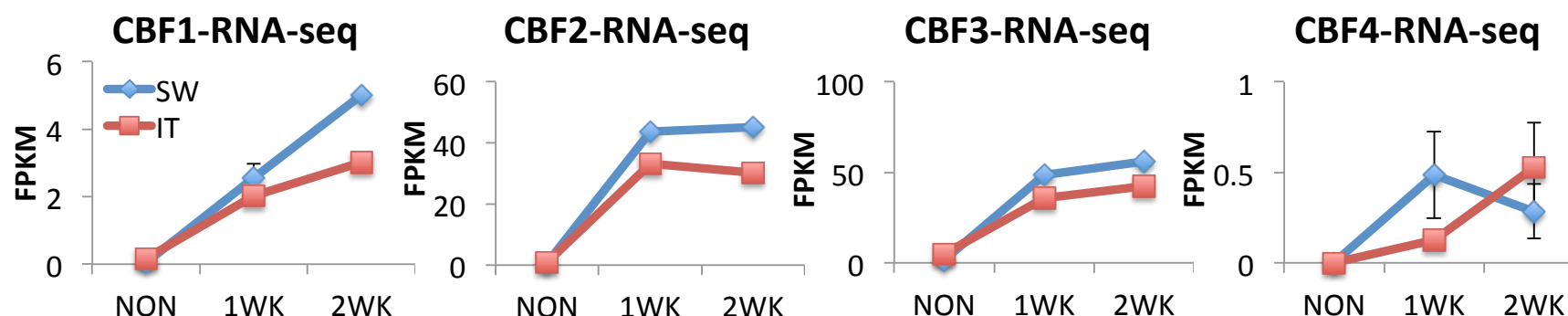


Figure 3.43. CBF gene expression. IT samples are shown in red, and SW in blue. Gene expression was measured by RNA-seq (FPKM). The results presented are average values from three independent experiments (n=3). Error bars indicate \pm SEM. Plants were grown at 22°C under a 12 h photoperiod for approximately 18 days before rosette tissue was sampled directly or transferred to 4°C for 1 or 2 weeks under a 12 h photoperiod then sampled.

MAJORITY	MNSFSAFSEMFGSDYESPVSSGGDYSPLATSCPCKPAGRKKFRETRHPIYRGVRQRNSGKWVCELREPNNKTRIWLGT	10	20	30	40	50	60	70	80
SW	MNSCSAFSEMFGSDYESPVSSGGDYSPLATSCPCKPAGRKKFRETRHPIYRGVRQRNSGKWVCELREPNNKTRIWLGT								
IT	MNSFSAFSEMFGSDYESPVSSGGDYSPLATSCPCKPAGRKKFRETRHPIYRGVRQRNSGKWVCELREPNNKTRIWLGT								
COL-0	MNSFSAFSEMFGSDYESPVSSGGDYSPLATSCPCKPAGRKKFRETRHPIYRGVRQRNSGKWVCELREPNNKTRIWLGT								
MAJORITY	QTAEMAARAHDVAAIALRGRSACLNFAADSAWRLRIPESTCAKEIQKAAAEALNFQDEMCHMTTDAHGLDMEETLVEAIY	90	100	110	120	130	140	150	160
SW	QTAEMAARAHDVAAIALRGRSACLNFAADSAWRLRIPESTCAKEIQKAAAEALNFQDEMCHMTTDAHGLDMEETLVEAIY								
IT	QTAEMAARAHDVAAIALRGRSACLNFAADSAWRLRIPESTCAKEIQKPR.								
COL-0	QTAEMAARAHDVAAIALRGRSACLNFAADSAWRLRIPESTCAKEIQKAAAEALNFQDEMCHMTTDAHGLDMEETLVEAIY								
MAJORITY	TPEQSQDAFYMDDEEAMLGMSLLDNMAEGMLLPSPSVQWNYNFDVEGDDDVSLWSY-	170	180	190	200	210			
SW	TPEQSQDAFYMDDEEAMLGMSLLDNMAEGMLLPSPSVQWNYNFDVEGDDDVSLWSY.								
COL-0	TPEQSQDAFYMDDEEAMLGMSLLDNMAEGMLLPSPSVQWNYNFDVEGDDDVSLWSY.								

Figure 3.44. Protein prediction for CBF2 in SW, IT and COL-0.

CONCLUSIONS

SW and IT ecotypes are adapted to their respective environments and both minimum winter temperature and flowering time were correlated with relative fitness of the SW and IT accessions in reciprocal transplant experiments (8). Temperature has previously been shown to be a strong selective pressure and freezing tolerance correlates with both latitude and minimum average temperature (6, 8, 17, 62). This study shows that SW and IT have significant differences in freezing tolerance at 2 weeks of cold acclimation, but not at 1 week of cold-acclimation or under non-acclimated conditions. This difference in freezing tolerance at 2 weeks of cold acclimation is due to both an increase in SW freezing tolerance as well as a reduction in IT freezing tolerance.

eQTL mapping revealed CBF2 as an excellent candidate gene to explain differences in cold-regulated gene expression and consequently freezing tolerance. Analysis of SW and IT CBF2 coding sequence exposed a 13 bp deletion in IT-CBF2 resulting in a predicted premature stop in the activation domain. Based on transcriptome and freezing tolerance analysis of a similar truncated version of CBF2, IT-CBF2 is predicted to function as a dominant negative version of CBF2. Transcriptional analyses are generally consistent with IT-CBF2 acting as a dominant negative version of CBF2, though additional experiments are needed to confirm. This is by no means the first example of CBF genes significantly contributing to differences in freezing tolerance between different accessions of *Arabidopsis* (10, 56-58). A similar premature stop in the activation domain of CBF3 was previously identified in the Condara accession, but

the effect of this truncation was not tested on CBF target genes or freezing tolerance (57). Therefore, this could possibly be the first natural example of a dominant negative version of CBF2.

The higher level of freezing tolerance in SW at 2 weeks of cold-acclimation could also be explained through *CBF2* expression, which is significantly higher than in IT at only this time-point. Therefore, the CBF2 promoter should be sequenced in SW and IT in the near future. SW photosynthetic capacity may also be greater than that of IT based on analysis of differentially regulated photosynthesis associate genes. However, photosynthetic capacity would need to be measured experimentally. Increased photosynthetic capacity is associated with higher levels of freezing tolerance, so differences in photosynthesis may also be a segregating trait between SW and IT.

Transcriptional analysis suggests that SW and IT reach similar levels of freezing tolerance at 1 week of cold-acclimation through significantly divergent transcriptional mechanisms. Both transcriptomic analysis and CBF2 sequence analysis suggest that IT may use CBF-independent mechanisms to cold-acclimate. These CBF-independent mechanisms might include genes annotated as biotic stress genes, since cold-regulated genes only found in IT were enriched in biotic stress GO terms. Biotic stress genes were also more highly expressed in IT compared to SW, regardless of cold-regulation. The mechanisms by which IT cold-acclimates do not seem to be as efficient as SW since there is a decrease in freezing tolerance at 2 weeks of cold-acclimation. I speculate that lower photosynthetic capacity in IT could be associated with the reduced number of cold-regulated genes from 1 to 2 weeks of cold-acclimation in IT, which may

synergistically combine with the dominant negative effect of IT-CBF2 to result in reduced freezing tolerance at 2 weeks of cold-acclimation.

Altogether, progress has been made towards discerning genetic differences between SW and IT that potentially contribute to differences in freezing tolerance and fitness. Other potentially segregating traits such as pathogen resistance, and photosynthetic capacity, that may contribute to fitness have also been identified. Genomic sequence of SW and IT parent lines should soon be available. This will make the RNA-seq data generated in this study more powerful, since locations of sequence differences across the SW and IT genomes will allow FTD candidate genes to be further prioritized for eQTL screening. In the near future, the Schemske lab plans to map differences in freezing tolerance in the SW and IT RILs. Based on results from this study overlap between freezing tolerance QTL and fitness QTL is expected (Table 3.1). A freezing tolerance QTL overlapping with the CBF locus is also expected. It will be interesting to see how much overlap there is between fitness and freezing tolerance QTL.

MATERIALS AND METHODS

Plant Material and Growth Conditions. Swedish (SW) and Italian (IT) accessions used in this study were collected from their native habitats by members of Schemske and Agren labs (Schemske and Agren, unpublished). Recombinant inbred lines (RILs) used in this study were constructed by the Schemske lab by crossing a randomly selected individual from the SW ecotype (male parent) to a randomly selected individual from the IT ecotype (female parent), seeds from the F1 generation were used to establish a large number of lines that were selfed by single-seed descent for nine generations (Schemske and Agren, unpublished). SW and IT RILs used in this study were from the F10 generation.

All seeds were stratified for 5 days in the dark at 4°C. For electrolyte leakage experiments, plants were grown as described by Dong et al. 2011(16), with slight modification. Plant were germinated on plates at 22°C under sterile conditions on Gamborg's B5 medium (Caisson Laboratories) with 1% sucrose at $\sim 100 \mu\text{mol m}^{-2} \text{s}^{-1}$ in a 12 h photoperiod for 7 days before transfer to soil for an additional 11 days, plants were then assayed for freezing tolerance directly (non-acclimated conditions) or transferred cold-acclimated conditions for 1, 2 or 3 weeks at $\sim 35 \mu\text{mol m}^{-2} \text{s}^{-1}$ in a 12 h photoperiod. For eQTL experiments RILs were grown as described by Dong et al. 2011(16), with slight modification. RILs were germinated on plates at 22°C under sterile conditions on Gamborg's B5 medium (Caisson Laboratories) with 1% sucrose at ~ 100

$\mu\text{mol m}^{-2}\text{s}^{-1}$ in a 12 h photoperiod for 7 days then transfer to soil for an additional 11 days before 2 week cold treatment at $\sim 35 \mu\text{mol m}^{-2}\text{s}^{-1}$ in a 12 h photoperiod.

Freezing Tolerance Tests. Electrolyte leakage assays were performed as described in Dong et al. 2011(16). For cold acclimation, plants were transferred to 4°C at ZT4 for 1 week, 2 weeks, or 3 weeks at $\sim 35 \mu\text{mol m}^{-2}\text{s}^{-1}$ under a 12 h photoperiod. Electrolyte leakage assays for acclimated and non-acclimated plants were started at \sim ZT4 in for all biological replicates. Tissue for RNA-seq experiments described below was collected before the start of each electrolyte leakage assay.

RNA-seq Experiments. Randomly selected rosette tissue (three technical replicates for each biological replication) from each electrolyte leakage experiment (non-acclimated, 1 week acclimated, 2 week acclimated, 3 week acclimated) was collected for RNA-seq analysis. RNA from 3 out of the 6 biological replication sets (SW, IT, at non-acclimated, 1 week acclimated, and 2 week acclimated) was chosen at random for further analysis. RNA was isolated as described in Dong et al. 2011 (16) from 2 technical replicates from selected biological replications; one technical replication was used for qRT-PCR analysis experiments and the other was submitted for RNA-seq analysis at MSU's Research Technology Support Facility (RTSF).

For qRT-PCR analysis, cDNA was made as described in Dong et al. 2011 (16). Expression of known cold-induced genes (CBF1, CBF2, CBF3, COR15A, COR47,

COR78) was tested using qRT-PCR (Applied Biosystems 7500 FAST Real-Time PCR System in FAST mode) as a means of ensuring that submitted RNA-seq samples were responding to cold. IPP2 (AT3G02780) was used as a reference gene.

For RNA-seq samples RNA quality was determined by an Agilent 2100 Bioanalyzer using manufacturer protocols ((Agilent RNA 6000 Nano Kit, Cat. 5067-1511, Agilent Technologies, Santa Clara, CA, USA). Sequencing was performed on an Illumina Genome Analyzer II (GA II). Like-treated samples (i.e. 3 biological replicates of SW and IT non-acclimated samples) were multiplexed in two lanes resulting in single-end reads ~75 bp in length with an average of 45,257,092 reads passing the Illumina purity filters for each sample (please see Table A3.8 for details). Sample preparation, including mRNA purification, cDNA preparation, end repair of cDNA, adaptor ligation, and cDNA amplification, was performed by MSU RTSF and conducted according to the manufacturer protocols (mRNA-Seq Sample Preparation Kit, Cat. RS-930-1001, Illumina Inc., San Diego, CA, USA).

RNA-seq Analysis. SW and IT RNA-seq data was aligned to TAIR10 sequence using Tophat (18). For Tophat, minimum intron length was set to 7 bp and maximum intron length was set to 12000 bp, all other parameters were set to defaults (18). Cufflinks was used to estimate transcript abundance based on unique reads in order to address the challenges associated with ‘multi-mapping reads’ (63), rRNA was also masked to improve robustness of transcript abundance estimates, all other parameters were set to default values (18). When comparing treatments for differential expression,

the cuffdiff program within Cufflinks was used for pairwise comparisons with default settings (18). Cuffdiff assigns p-values based a two-tailed Student's t-test; this value is then adjusted using a Benjamini-Hochberg correction for 'multiple-testing'(18).

While analyzing differentially expressed genes in RNA-seq data, a cut-off of ≥ 3 FPKM was used for differentially expressed genes (19, 20). This means that, when treatments were compared, one value in the comparison had to be ≥ 3 FPKM. When testing for significant overlap between two sets of genes, a hypergeometric probability was calculated using the 'stats' package in R (64). Gene ontology (GO) enrichment was determined using the database for annotation visualization and integrated discovery tool (DAVID; david.abcc.ncifcrf.gov; (27)). The default DAVID gene ontology settings 'GOTERM_BP_FAT', 'GOTERM_MF_FAT', and 'GOTERM_CC_FAT' were used for analysis (27). GO category was considered significantly enriched if its Benjamini-Hochberg corrected (FDR corrected) hypergeometric p-value was < 0.050 .

eQTL Experiments. RIL seed stratification, germination, and movement to low temperature conditions was staggered to allow all RILs to be grown then cold acclimated in the same set of growth chambers and to ensure that a manageable number of RILs, at the same age, could be samples within an hour. Plant tissue was collected and quickly frozen in liquid nitrogen between ZT4-ZT5 after 2 weeks of cold acclimation. RNA extraction and cDNA synthesis for RILs was performed as described by Dong et al. 2011(16). These samples were tested for expression of 10 Freezing Tolerance Difference (FTD) candidate genes (see Table 3.17) by qRT-PCR (Applied

Biosystems 7500 FAST Real-Time PCR System in FAST mode) with IPP2

(AT3G02780) as a housekeeping gene. All primer sets can be found in Table A3.7.

eQTL Mapping and Analysis. SW and IT parent lines were sequenced by the Schemske lab using the Illumina GAI platform. Paired end reads were mapped to the COL-0 reference sequence (TAIRv9) and approximately 141,437 single nucleotide polymorphisms (SNPs) were found (Schemske, unpublished). 384 of these SNPs with an average spacing of ~1.1 cm across the *Arabidopsis* genome were selected to genotype the RIL population (Schemske, unpublished). Expression data for the 10 FTD candidate genes generated during the eQTL experiments described above was used to map eQTL over these 384 markers. RQTL (15) was used to perform interval mapping using the multiple imputation method as described by the reference manual (<http://www.rqtl.org/>). To detect QTL, estimated genome-wide significance thresholds ($\alpha=0.01$ and $\alpha=0.05$) for the logarithm of odds (LOD) values were calculated for each FTD gene of interest using 1000 permutations in RQTL. To estimate the location of each QTL both a Bayes credible interval (99%) and LOD support interval were calculated in RQTL. The SeqViewer tool on TAIR (www.arabidopsis.org) was used to identify all genes within the QTL interval and this list was compared to SW and IT RNA-seq data described above to find candidate transcription factors.

CBF sequencing. Genomic DNA was extracted from SW, IT and COL-0 using a QIAGEN DNeasy Plant Mini Kit using manufacturer protocols (Cat. 69104, Qiagen,

Valencia, CA). CBF1-3, COR78, COR15A and COR47 coding sequences were amplified by PCR using TaKaRa LA Taq Hot-Start DNA Polymerase (Cat. RR042A, Mountain View, CA). Primers used to amplify CBF1, 2 and 3 coding sequence in SW, IT and Col-0, can be found in Table A3.8. PCR products were cloned into PCR2.1-TOPO, transformed into *E. coli* competent cells, and screened using manufacturer protocols (Cat. K4500-02, Invitrogen, Carlsbad, CA). Selected clones were sequenced by MSU RTSF. Sequence alignment was performed using the Lasergene Suite of tools (www.dnastar.com).

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I am especially grateful to Pingsha Hu who performed alignment and assembly of the RNA-seq data generated in this study. Chin-Mei Lee helped with the cloning and sequencing of SW and IT cold genes. Thanks to the Schemske lab for providing the *Arabidopsis* ecotypes from Sweden and Italy as well as the recombinant inbred lines. I am also grateful to Nicholas Bartoa and Avery Mendelson for their help with plant growth and care.

APPENDIX

Table A3.1. SW and IT markers used for mapping both fitness QTL and eQTL. Marker locations in centimorgans (CM), and marker locations in megabases (MB) based on the Col-0 genome.

CHROMOSOME	MARKER (CM)	MARKER LOCATIONS (MB)
1	0	0.371484
1	1.422	0.658928
1	2.075	0.95079
1	3.013	1.247666
1	3.384	1.545309
1	3.943	1.831853
1	4.408	2.112359
1	5.25	2.402241
1	5.343	2.689578
1	6.112	2.97831
1	6.206	3.270056
1	8.36	4.139064
1	8.842	4.434414
1	9.507	4.716592
1	9.916	5.008647
1	10.312	5.305697
1	11.03	5.588245
1	11.501	5.876257
1	11.97	6.486531
1	12.636	6.750734
1	13.483	7.043318
1	14.053	7.332032
1	14.525	7.621705
1	16.069	7.906548
1	18.117	8.199225
1	18.877	8.511642
1	19.638	8.846085
1	20.792	9.081675
1	21.941	9.389545
1	22.704	9.64586
1	23.652	9.948571
1	24.623	10.232305
1	25.681	10.517662
1	26.545	10.817536
1	28.103	11.101365
1	29.502	11.402119
1	30.882	11.697338
1	32.413	11.966262
1	33.796	12.344267
1	35.512	12.843656
1	37.885	13.156629
1	38.268	13.451072
1	38.363	13.753592
1	38.459	14.004113
1	38.459	15.522314
1	38.554	15.889605
1	39.911	16.136207
1	40.59	16.445556

Table A3.1 (cont'd)

1	42.502	16.923537
1	43.493	17.203021
1	46.721	17.775164
1	47.977	18.059668
1	48.931	18.352147
1	49.788	18.65907
1	50.352	18.926903
1	51.496	19.240146
1	52.543	19.529287
1	53.291	19.800653
1	54.325	20.108938
1	54.603	20.38011
1	55.197	20.668556
1	55.601	20.964571
1	55.904	21.251788
1	56.275	21.551115
1	57.347	22.152521
1	58.632	22.412767
1	58.823	22.723886
1	60.236	22.988965
1	60.984	23.292064
1	61.076	23.580047
1	61.824	23.857523
1	62.477	24.155803
1	63.146	24.435268
1	64.88	24.765127
1	66.846	25.032697
1	67.637	25.324254
1	68.419	25.609182
1	70.447	25.889912
1	71.997	26.181454
1	74.435	26.760155
1	75.399	27.057077
1	76.971	27.352821
1	77.163	27.626189
1	77.86	27.917735
1	78.551	28.220678
1	79.56	28.509198
1	80.38	28.806377
1	80.886	29.077284
1	82.142	29.384089
1	83.088	29.659444
1	83.845	30.245282
2	0	0.365473
2	4.606	0.661681
2	6.33	0.954162
2	8.55	1.258538
2	10.17	1.532339
2	11.4	1.871886
2	12.822	2.135186
2	14.736	2.433194
2	15.107	4.621443

Table A3.1 (cont'd)

2	15.2	5.049931
2	15.48	5.295348
2	15.761	5.587569
2	17.687	5.95692
2	22.613	6.764214
2	24.312	7.047117
2	25.601	7.340228
2	26.841	7.626054
2	27.893	7.920385
2	29.449	8.197294
2	30.714	8.502011
2	31.706	8.779839
2	31.992	9.065286
2	32.589	9.36707
2	34.308	9.647248
2	35.592	9.940989
2	37.365	10.225771
2	38.247	10.536215
2	39.837	10.837886
2	40.254	11.106932
2	41.761	11.38934
2	42.345	11.679435
2	43.114	11.966391
2	44.28	12.267422
2	46.948	12.547427
2	48.042	12.838094
2	48.734	13.135793
2	49.409	13.420085
2	49.516	13.716369
2	49.704	13.995766
2	50.366	14.319012
2	50.646	14.575482
2	51.02	14.868193
2	52.453	15.16817
2	53.036	15.447154
2	53.415	15.747054
2	54.399	16.06694
2	54.885	16.319037
2	55.269	16.60741
2	56.022	16.898396
2	56.402	17.18517
2	56.779	17.476046
2	56.874	17.779
2	58.025	18.063832
2	58.313	18.358707
2	58.594	18.639532
2	59.444	18.932927
2	60.487	19.233555
2	60.86	19.529865
3	0	0.370926
3	0.094	0.656579
3	1.032	0.961179

Table A3.1 (cont'd)

3	1.688	1.270326
3	2.155	1.525377
3	2.533	1.825965
3	2.82	2.112696
3	3.487	2.398036
3	3.96	2.692472
3	4.523	2.975352
3	5.092	3.269092
3	6.349	3.565378
3	7.208	4.141096
3	7.313	4.427835
3	8.176	4.718296
3	9.637	5.295658
3	10.681	5.619526
3	11.184	5.875889
3	11.853	6.175656
3	12.641	6.457498
3	13.533	6.74919
3	14.728	7.051377
3	15.618	7.329844
3	16.51	7.628106
3	18.033	7.914025
3	19.705	8.200995
3	22.343	8.497891
3	23.319	8.776721
3	25.381	9.13927
3	28.021	9.654017
3	29.512	9.940341
3	31.527	10.234401
3	32.697	10.587664
3	37.858	11.114247
3	39.55	11.399488
3	40.641	11.747164
3	41.121	12.041893
3	41.309	12.56772
3	41.402	15.210147
3	42.553	15.514979
3	43.813	15.759118
3	44.974	16.058567
3	45.949	16.36229
3	49.119	16.648133
3	50.183	16.899341
3	51.952	17.225206
3	53.323	17.486181
3	54.095	17.781386
3	54.672	18.077874
3	55.367	18.350962
3	57.064	18.651512
3	58.618	18.955604
3	59.188	19.220884
3	60.383	19.514107
3	61.53	19.799867

Table A3.1 (cont'd)

3	63.124	20.092191
3	63.604	20.401811
3	64.18	20.665136
3	64.928	20.960795
3	65.487	21.25848
3	65.858	21.558962
3	66.606	21.840004
3	66.791	22.14465
3	67.54	22.441322
3	68.386	22.70487
3	69.618	22.986945
3	70.466	23.275556
4	0	0.384888
4	2.329	0.680041
4	6.129	0.982791
4	7.648	1.238231
4	8.975	1.535171
4	9.255	2.717267
4	12.891	2.396469
4	14.399	2.133635
4	15.265	1.828832
4	15.674	3.900172
4	15.767	5.080132
4	18.335	5.587712
4	19.776	5.924099
4	20.73	6.171429
4	24.028	6.498644
4	25.332	6.754691
4	26.733	7.042854
4	27.984	7.346579
4	29.948	7.616384
4	31.047	7.918274
4	32.149	8.211444
4	33.418	8.498174
4	34.491	8.815569
4	35.766	9.140013
4	37.38	9.650672
4	39.077	9.942702
4	41.05	10.236673
4	42.292	10.522219
4	43.507	10.831675
4	44.312	11.15739
4	46.649	11.714851
4	48.883	12.292822
4	50.004	12.5471
4	50.195	12.843072
4	50.574	13.151997
4	51.142	13.421012
4	51.331	13.720933
4	51.993	14.008832
4	52.369	14.292115
4	52.847	14.584632

Table A3.1 (cont'd)

4	53.543	14.869919
4	53.935	15.162178
4	54.131	15.450267
4	54.523	15.735751
4	55.222	16.026122
4	55.523	16.337059
4	55.916	16.606958
4	56.299	16.90214
4	56.593	17.202249
4	57.696	17.486207
4	59.526	17.796648
4	60.435	18.063371
4	60.721	18.347312
5	0	0.367158
5	1.428	0.655985
5	2.63	0.949897
5	3.939	1.245151
5	4.594	1.528949
5	4.913	1.839919
5	5.695	2.111365
5	6.678	2.408921
5	7.459	2.685965
5	7.652	2.985496
5	8.525	3.270887
5	9.384	3.57043
5	10.048	3.852246
5	10.434	4.13647
5	10.717	4.426649
5	10.811	4.726262
5	11.39	5.013068
5	11.972	5.336868
5	13.126	5.656316
5	13.602	5.889105
5	14.471	6.473301
5	14.668	6.752582
5	14.963	7.071294
5	15.919	7.334179
5	16.811	7.618492
5	18.508	7.914975
5	20.419	8.232952
5	21.967	8.569949
5	27.139	9.0801
5	28.533	9.376062
5	29.833	9.6691
5	32.507	9.969215
5	34.144	10.233797
5	36.082	10.639041
5	36.645	10.929784
5	36.738	11.415254
5	37.016	13.106228
5	38.26	13.707451
5	40.107	14.015641

Table A3.1 (cont'd)

5	41.4	14.545953
5	43.043	14.876427
5	44.949	15.185072
5	46.891	15.464485
5	48.063	15.768401
5	49.947	16.043932
5	50.244	16.326685
5	50.244	16.611838
5	50.244	16.897426
5	50.45	17.196046
5	50.962	17.503065
5	51.576	17.787766
5	52.441	18.072654
5	53.407	18.382907
5	54.267	18.640297
5	54.836	18.926019
5	56.611	19.227434
5	57.476	19.550188
5	57.664	19.797944
5	58.578	20.09524
5	59.597	20.390151
5	60.101	20.67101
5	60.596	20.961565
5	61.278	21.249178
5	62.603	21.548963
5	64.099	21.864028
5	64.62	22.116026
5	66.055	22.406706
5	66.456	22.711593
5	67.732	23.006666
5	68.693	23.277096
5	69.359	23.580206
5	70.123	23.87063
5	70.596	24.172305
5	70.983	24.444383
5	71.373	24.725353
5	72.063	25.017566
5	72.931	25.350635
5	73.61	25.599724
5	74.474	25.889576
5	75.802	26.180944
5	77.422	26.468215
5	78.174	26.757861

Table A3.2. GO categories of Hannah genes positively (277 genes) and negatively (466 genes) correlated with freezing tolerance (6). GO enrichment was determined using DAVID (david.abcc.ncifcrf.gov; (27)). 'GO TERM CATEGORY' denotes if the GO term was categorized as a biological process (BP), cellular component (CC), or molecular function (MF) in the DAVID database. '# HITS' denotes the number of genes with that GO category in Hannah genes positively (277 genes) and negatively (466 genes) correlated with freezing tolerance. '# GENES IN GO' denotes the total number of genes in that GO category included in the DAVID database. GO category was considered significantly enriched if the FDR corrected hypergeometric p-value was ≤ 0.050 .

GO TERM CATEGORY	HANNAH POSITIVE (277GENES)	# HITS	# GENES IN GO	FDR P-VALUE
BP	Response To Water Deprivation	16	176	2.68E-07
BP	Response To Abiotic Stimulus	34	1197	1.22E-04
BP	Response To Cold	11	233	3.66E-02
BP	Response To Oxidative Stress	12	287	3.82E-02
BP	Carbohydrate Biosynthetic Process	11	254	4.13E-02
BP	Response To Osmotic Stress	14	399	4.43E-02

GO TERM CATEGORY	HANNAH NEGATIVE (466 GENES)	# HITS	# GENES IN GO	FDR P-VALUE
CC	Chloroplast	225	3192	3.34E-51
CC	Thylakoid	80	491	1.27E-37
CC	Photosynthetic Membrane	56	330	7.22E-27
MF	rRNA Binding	21	76	4.55E-15
CC	Organelle Membrane	65	849	8.10E-13
MF	Structural Constituent Of Ribosome	32	394	6.55E-09
MF	Structural Molecule Activity	37	538	1.04E-08
CC	Ribosome	38	470	5.16E-08
BP	Photosynthesis	18	175	7.19E-05
BP	Chlorophyll Biosynthetic Process	8	34	1.69E-03
BP	Pigment Metabolic Process	11	101	4.09E-03
BP	Nitrogen Compound Biosynthetic Process	26	506	4.52E-03
BP	Tetrapyrrole Biosynthetic Process	8	50	5.27E-03
CC	Anchored To Plasma Membrane	8	66	1.11E-02
CC	NAD(P)H Dehydrogenase Complex	4	10	1.14E-02
BP	Porphyrin Metabolic Process	8	61	1.71E-02
BP	Response To Cold	15	233	1.81E-02
BP	Heterocycle Biosynthetic Process	11	132	2.06E-02
BP	Ribosome Biogenesis	15	242	2.27E-02
CC	Non-Membrane-Bounded Organelle	46	1144	4.37E-02

Table A3.3. 474 Category 1 FTD Candidate Genes with higher expression in SW at 2 weeks of cold-acclimation. Genes are ordered by their AGI numbers. FPKM values are the average of three biological replicates (n=3). Standard error (SE) values are also given for each FPKM value. Genes were considered significantly different between SW and IT under non-acclimated conditions if they had a Benjamini-Hochberg corrected p-value (FDR P-VALUE) of <0.05 and a minimum of ≥ 3 FPKM in IT. The differential expression of each gene is also represented by the logarithm (base=2) of the fold change (LOG2.FC). This table also denotes if a gene was previously described as positively or negatively correlating with freezing tolerance (HANNAH;(6)), a transcription factor (TF), a gene previously described as up- or down-regulated by 1 week of cold (COS, (25)), identified as an up- or down-regulated component of the CBF regulon (CBF REGULON; (25)).

AGI	SW.IT.2WK					FDR P-VALUE	HANNAH?	TF?	COS?	CBF REG?
	SW.2WK FPKM	SW.2WK SE	IT.2WK FPKM	IT.2WK SE	LOG2FC					
AT1G01210	25.47	0.77	14.97	0.99	-0.77	1.60E-02	NO	NO	NO	NO
AT1G01640	10.27	0.35	6.01	0.37	-0.77	4.09E-02	NO	NO	NO	NO
AT1G02300	30.44	0.44	20.14	0.38	-0.60	6.08E-03	NO	NO	NO	NO
AT1G02460	17.43	0.27	10.21	0.22	-0.77	3.71E-04	NO	NO	UP	NO
AT1G03310	36.19	1.34	25.42	0.39	-0.51	1.53E-03	NO	NO	NO	NO
AT1G04240	61.42	1.64	35.21	0.91	-0.80	2.00E-05	NO	YES	NO	NO
AT1G04350	20.72	0.41	14.58	0.65	-0.51	3.30E-02	NO	NO	DOWN	NO
AT1G04570	132.07	1.41	40.21	0.74	-1.72	0.00E+00	NO	NO	NO	NO
AT1G04620	75.99	0.87	52.13	0.61	-0.54	3.74E-03	NO	NO	NO	NO
AT1G06360	67.20	1.07	44.29	2.60	-0.60	1.52E-03	NO	NO	NO	NO
AT1G06700	59.72	0.38	40.10	1.11	-0.57	4.17E-04	NO	NO	NO	NO
AT1G06980	16.78	0.31	6.63	0.48	-1.34	6.39E-05	NO	NO	NO	NO
AT1G07200	12.97	0.13	9.36	0.30	-0.47	1.02E-02	NO	NO	NO	NO
AT1G07280	131.04	0.92	96.96	0.44	-0.43	5.00E-03	NO	NO	NO	NO
AT1G08500	10.71	0.34	6.35	0.26	-0.75	4.21E-02	NO	NO	NO	NO
AT1G08570	53.28	0.33	39.07	0.74	-0.45	1.10E-02	NO	NO	NO	UP
AT1G09750	80.53	1.54	53.50	1.12	-0.59	1.43E-03	NO	NO	NO	NO
AT1G10522	64.52	1.31	47.36	0.80	-0.45	2.86E-02	NO	NO	NO	NO

Table A3.3 (cont'd)

AT1G10760	109.27	1.78	75.43	0.21	-0.53	1.53E-02	NO	NO	NO	NO
AT1G11700	23.34	0.66	15.93	0.52	-0.55	4.48E-02	NO	NO	NO	NO
AT1G12090	1320.94	3.95	788.36	2.37	-0.74	6.44E-04	NO	NO	NO	NO
AT1G12250	70.97	0.66	52.09	0.62	-0.45	1.99E-02	NO	NO	NO	NO
AT1G12710	47.22	1.05	27.27	0.94	-0.79	2.17E-05	NO	NO	NO	NO
AT1G13560	56.05	1.41	44.22	0.22	-0.34	3.15E-02	NO	NO	NO	NO
AT1G14150	156.49	3.14	87.89	0.69	-0.83	7.20E-10	NEGATIVE	NO	NO	NO
AT1G14200	20.79	0.48	13.80	0.25	-0.59	4.86E-02	NO	NO	NO	NO
AT1G15405	155.32	3.67	75.96	1.76	-1.03	2.04E-04	NO	NO	NO	NO
AT1G15470	25.73	0.62	18.39	0.19	-0.48	3.06E-02	NO	NO	NO	NO
AT1G16410	152.60	2.38	114.96	3.13	-0.41	6.93E-03	NO	NO	NO	NO
AT1G18360	31.45	0.78	17.57	0.35	-0.84	1.06E-05	NO	NO	NO	NO
AT1G19570	236.89	2.40	180.50	2.14	-0.39	1.65E-02	NO	NO	NO	NO
AT1G19960	407.82	4.55	204.76	3.28	-0.99	3.13E-08	NO	NO	NO	NO
AT1G20010	306.54	1.71	203.60	3.80	-0.59	3.35E-03	NO	NO	NO	NO
AT1G20610	9.37	0.37	6.25	0.12	-0.58	3.38E-02	NO	NO	NO	NO
AT1G21500	93.57	1.25	66.71	1.82	-0.49	2.30E-02	NEGATIVE	NO	DOWN	NO
AT1G21790	21.97	0.81	13.68	0.45	-0.68	2.96E-03	NO	NO	UP	UP
AT1G22540	5.53	0.05	2.57	0.22	-1.10	6.12E-04	NO	NO	NO	NO
AT1G22700	210.12	1.87	155.66	1.31	-0.43	1.49E-02	NEGATIVE	NO	NO	NO
AT1G22740	21.91	0.48	15.01	0.47	-0.55	4.26E-02	NO	NO	NO	NO
AT1G22940	37.65	0.29	22.38	0.42	-0.75	3.87E-05	NO	NO	NO	NO
AT1G23080	30.07	0.51	23.66	1.11	-0.35	4.99E-02	NO	NO	NO	NO
AT1G25422	3.05	0.18	1.05	0.17	-1.54	2.17E-02	NO	NO	NO	NO
AT1G26230	25.78	0.28	16.38	0.28	-0.65	3.47E-04	NO	NO	NO	NO
AT1G26761	42.88	0.49	29.43	0.43	-0.54	7.04E-03	NO	NO	NO	NO
AT1G27050	18.95	0.27	11.42	0.75	-0.73	1.56E-02	NO	YES	NO	NO
AT1G27385	39.61	1.11	28.36	0.48	-0.48	3.05E-02	NO	NO	NO	NO
AT1G27910	10.36	0.36	7.23	0.19	-0.52	2.46E-02	NO	NO	NO	UP

Table A3.3 (cont'd)

AT1G28070	23.20	1.28	10.83	0.12	-1.10	1.48E-04	NO	NO	NO	NO
AT1G28410	16.64	0.43	10.44	0.42	-0.67	5.00E-03	NO	NO	NO	NO
AT1G29390	119.79	1.15	61.03	0.97	-0.97	1.74E-08	NO	NO	NO	NO
AT1G30250	32.50	3.31	16.63	1.77	-0.97	2.57E-03	NO	NO	NO	NO
AT1G30500	6.98	0.10	2.87	0.13	-1.28	5.45E-03	POSITIVE	YES	NO	NO
AT1G30510	16.04	0.32	11.62	0.57	-0.47	3.60E-02	NO	NO	NO	NO
AT1G30910	21.79	0.33	14.17	0.25	-0.62	1.12E-02	NO	NO	NO	NO
AT1G31230	37.24	0.75	23.59	0.43	-0.66	2.71E-04	NO	NO	NO	NO
AT1G31460	6.20	0.22	3.68	0.23	-0.75	1.98E-02	NO	NO	NO	NO
AT1G31850	37.44	0.79	21.90	0.66	-0.77	1.75E-07	NO	NO	NO	NO
AT1G32070	67.46	0.29	41.51	0.68	-0.70	3.57E-06	NO	NO	NO	NO
AT1G32090	10.05	0.13	5.76	0.30	-0.80	2.72E-04	NO	NO	NO	NO
AT1G32860	47.13	0.47	20.18	0.94	-1.22	7.28E-12	POSITIVE	NO	UP	UP
AT1G33230	47.77	0.56	25.24	0.51	-0.92	2.64E-07	POSITIVE	NO	UP	UP
AT1G33265	96.99	2.21	60.14	0.97	-0.69	1.94E-04	NO	NO	NO	NO
AT1G35420	106.21	0.47	65.65	0.21	-0.69	7.39E-05	NO	NO	NO	NO
AT1G35720	1410.82	2.15	881.87	13.58	-0.68	1.10E-02	NO	NO	NO	NO
AT1G44800	32.53	1.45	24.01	0.58	-0.44	4.12E-02	NO	NO	NO	NO
AT1G45191	6.22	0.41	3.89	0.17	-0.68	3.94E-02	NO	NO	NO	NO
AT1G46480	9.94	0.05	5.86	0.26	-0.76	3.02E-02	NO	YES	NO	NO
AT1G47580	26.14	0.50	16.71	0.87	-0.65	1.52E-02	NO	NO	NO	NO
AT1G47710	97.26	1.00	62.81	1.00	-0.63	6.18E-04	POSITIVE	NO	UP	UP
AT1G48600	27.78	1.09	13.49	2.54	-1.04	3.27E-11	NO	NO	DOWN	NO
AT1G49160	10.37	0.47	5.44	0.16	-0.93	2.19E-05	NO	NO	NO	NO
AT1G49560	32.95	0.48	14.60	0.47	-1.17	9.27E-10	NO	YES	NO	NO
AT1G50840	16.86	0.55	12.58	0.17	-0.42	4.15E-02	NO	NO	NO	NO
AT1G51400	461.22	4.33	326.84	8.80	-0.50	8.19E-03	NO	NO	DOWN	NO
AT1G51610	78.50	0.56	52.66	0.20	-0.58	2.07E-03	NO	NO	UP	NO
AT1G51650	424.94	1.52	305.12	5.02	-0.48	1.31E-02	NO	NO	NO	NO

Table A3.3 (cont'd)

AT1G52220	644.30	1.76	383.59	3.87	-0.75	2.38E-05	NO	NO	NO	NO
AT1G53035	118.69	0.58	81.77	0.85	-0.54	4.63E-03	NO	NO	UP	NO
AT1G53160	28.70	0.45	11.38	0.44	-1.33	3.57E-08	NO	YES	NO	NO
AT1G53520	30.41	0.64	17.49	1.51	-0.80	3.78E-04	NEGATIVE	NO	NO	NO
AT1G53590	25.97	0.66	19.04	0.24	-0.45	2.64E-02	NO	NO	NO	NO
AT1G54000	6.48	0.16	2.46	0.35	-1.40	7.26E-05	NO	NO	NO	NO
AT1G54410	5206.40	93.83	3059.92	56.05	-0.77	1.53E-02	NO	NO	NO	UP
AT1G55690	30.71	0.68	18.98	0.47	-0.69	1.03E-07	NO	NO	NO	UP
AT1G56300	139.89	1.20	57.61	0.35	-1.28	1.91E-14	NO	NO	NO	NO
AT1G58235	19.26	0.37	12.75	0.23	-0.60	3.05E-02	NO	NO	NO	NO
AT1G60590	10.21	0.50	6.22	0.31	-0.71	4.05E-03	NO	NO	NO	NO
AT1G60680	5.12	0.26	2.45	0.14	-1.06	1.63E-02	NO	NO	NO	NO
AT1G61520	3383.70	29.25	2573.50	20.66	-0.39	4.52E-02	NO	NO	NO	NO
AT1G61740	29.12	0.79	18.64	0.19	-0.64	1.12E-03	NO	NO	NO	NO
AT1G62290	4.52	0.25	1.02	0.40	-2.15	1.62E-07	NO	NO	NO	NO
AT1G62960	21.40	0.28	15.66	0.53	-0.45	4.27E-02	NO	NO	NO	NO
AT1G63240	13.97	0.17	6.57	0.19	-1.09	7.02E-07	NO	NO	NO	NO
AT1G64770	162.85	1.36	105.16	0.89	-0.63	5.19E-06	NEGATIVE	NO	NO	NO
AT1G65590	8.75	0.17	4.97	0.51	-0.82	2.04E-03	NO	NO	NO	NO
AT1G65960	428.15	4.52	315.59	1.53	-0.44	1.43E-03	NO	NO	NO	UP
AT1G66430	84.72	0.21	48.52	1.26	-0.80	5.16E-06	NO	NO	NO	NO
AT1G67080	26.70	0.67	18.08	1.23	-0.56	1.87E-02	NO	NO	DOWN	NO
AT1G67105	10.03	0.25	4.38	0.75	-1.20	6.29E-03	NO	NO	NO	NO
AT1G69523	18.79	0.25	12.89	0.25	-0.54	2.88E-02	NO	NO	NO	NO
AT1G69830	74.14	1.14	49.51	0.25	-0.58	2.02E-03	NO	NO	NO	NO
AT1G69910	7.96	0.14	4.99	0.11	-0.67	1.21E-02	NO	NO	NO	NO
AT1G70560	9.96	0.47	4.21	0.28	-1.24	3.36E-05	NO	NO	NO	NO
AT1G70640	130.29	1.29	29.31	1.21	-2.15	0.00E+00	NO	NO	NO	NO
AT1G71000	5.59	0.75	0.45	0.04	-3.64	6.49E-04	NO	NO	NO	NO

Table A3.3 (cont'd)

AT1G71710	36.99	0.91	23.05	0.25	-0.68	1.14E-04	NO	NO	NO	UP
AT1G71720	61.67	1.19	42.54	0.72	-0.54	6.14E-03	NO	NO	NO	NO
AT1G73655	305.45	3.02	187.61	1.76	-0.70	6.71E-05	NEGATIVE	NO	NO	NO
AT1G74070	33.66	0.36	20.20	0.58	-0.74	2.03E-04	NEGATIVE	NO	DOWN	NO
AT1G74520	88.76	0.82	46.38	1.35	-0.94	3.37E-07	NO	NO	NO	UP
AT1G74560	103.91	1.15	63.58	0.98	-0.71	1.62E-05	NO	NO	NO	NO
AT1G74680	17.75	0.34	12.35	0.22	-0.52	2.03E-02	NO	NO	NO	NO
AT1G74840	79.97	0.42	48.23	0.36	-0.73	4.45E-05	NO	YES	NO	NO
AT1G75750	829.59	17.38	397.91	13.34	-1.06	0.00E+00	NO	NO	NO	NO
AT1G75780	8.56	0.28	5.13	0.16	-0.74	1.31E-02	NO	NO	NO	NO
AT1G76020	34.81	0.39	18.91	0.21	-0.88	1.38E-04	NO	NO	NO	NO
AT1G76790	39.90	0.40	22.05	0.50	-0.86	9.83E-06	NO	NO	NO	NO
AT1G77490	80.16	0.27	43.63	0.63	-0.88	3.79E-07	NO	NO	NO	NO
AT1G77540	80.90	0.53	55.05	1.53	-0.56	1.86E-02	NO	NO	NO	NO
AT1G77580	18.10	0.58	9.44	0.32	-0.94	5.71E-08	NO	NO	NO	NO
AT1G78610	21.05	0.41	13.62	0.52	-0.63	9.33E-04	NO	NO	NO	NO
AT1G78820	36.71	0.99	23.28	0.47	-0.66	6.50E-04	NO	NO	NO	NO
AT1G78995	92.65	1.12	63.84	0.17	-0.54	5.56E-03	NO	NO	NO	NO
AT1G79790	44.79	0.57	30.37	0.34	-0.56	5.87E-03	NO	NO	NO	NO
AT1G80940	28.45	0.61	15.51	0.42	-0.88	2.64E-06	NO	NO	NO	NO
AT2G01275	3.28	0.06	1.00	0.02	-1.71	3.80E-03	NO	NO	NO	NO
AT2G02080	48.63	7.69	33.63	6.37	-0.53	2.69E-03	NO	YES	NO	NO
AT2G02100	1972.26	17.76	1127.29	16.33	-0.81	6.08E-05	NO	NO	UP	NO
AT2G02510	218.11	1.02	144.88	2.60	-0.59	2.54E-03	NO	NO	NO	NO
AT2G02740	60.75	1.14	45.57	0.56	-0.41	4.83E-02	NO	YES	NO	NO
AT2G12400	6.92	0.26	2.77	0.06	-1.32	7.71E-06	NO	NO	NO	NO
AT2G13550	12.11	0.16	6.50	0.29	-0.90	1.66E-03	NO	NO	NO	NO
AT2G14045	36.12	0.61	25.84	0.58	-0.48	3.52E-02	NO	NO	NO	NO
AT2G14170	42.86	0.45	33.09	0.31	-0.37	4.08E-02	NO	NO	NO	NO

Table A3.3 (cont'd)

AT2G16600	3296.96	32.69	2148.89	37.50	-0.62	3.34E-02	NO	NO	NO	NO
AT2G18328	279.10	1.40	156.19	0.16	-0.84	1.94E-06	NO	NO	NO	NO
AT2G18890	18.94	0.81	12.95	0.52	-0.55	1.88E-02	NO	NO	NO	NO
AT2G19170	8.96	0.40	6.41	0.32	-0.48	4.55E-02	NO	NO	NO	NO
AT2G19860	38.60	0.69	25.07	0.73	-0.62	8.34E-04	NO	NO	NO	NO
AT2G20270	116.11	2.13	90.45	1.59	-0.36	3.40E-02	NO	NO	NO	NO
AT2G21130	94.42	0.81	69.47	1.15	-0.44	3.53E-02	NO	NO	UP	NO
AT2G21250	186.46	1.25	124.95	2.36	-0.58	1.36E-03	NO	NO	NO	NO
AT2G21320	143.22	1.58	107.75	0.65	-0.41	3.95E-02	NO	NO	UP	NO
AT2G21340	26.91	0.52	14.72	0.19	-0.87	2.93E-06	NO	NO	NO	NO
AT2G21370	38.41	0.32	24.58	0.05	-0.64	5.96E-05	NO	NO	NO	NO
AT2G21380	19.13	0.26	11.92	0.29	-0.68	1.98E-04	NO	NO	NO	NO
AT2G21590	21.42	0.90	7.47	0.30	-1.52	0.00E+00	NO	NO	NO	NO
AT2G22080	166.15	1.73	118.32	1.75	-0.49	9.38E-03	NO	NO	UP	UP
AT2G22122	38.51	0.36	20.34	0.34	-0.92	1.15E-03	NO	NO	NO	NO
AT2G22190	50.97	1.12	24.29	0.55	-1.07	7.57E-09	NO	NO	UP	NO
AT2G22240	28.57	0.59	19.55	1.04	-0.55	2.72E-03	NO	NO	NO	NO
AT2G22260	3.76	0.09	0.97	0.17	-1.96	1.19E-03	NO	NO	NO	NO
AT2G22540	84.61	0.49	61.71	0.44	-0.46	1.74E-02	NO	YES	NO	NO
AT2G23340	20.71	0.63	11.81	0.61	-0.81	3.15E-03	NO	YES	NO	UP
AT2G23670	257.19	2.55	192.22	6.07	-0.42	3.45E-02	NEGATIVE	NO	NO	NO
AT2G23910	13.22	0.17	8.49	0.20	-0.64	2.32E-02	NO	NO	NO	NO
AT2G24120	35.58	1.32	26.22	0.27	-0.44	2.04E-02	NO	NO	NO	NO
AT2G24150	25.03	0.54	17.12	0.86	-0.55	1.59E-02	NO	NO	NO	NO
AT2G24270	308.66	1.27	221.62	3.10	-0.48	3.31E-04	NO	NO	NO	NO
AT2G27530	746.39	6.61	599.41	9.34	-0.32	2.99E-02	NO	NO	NO	NO
AT2G28840	243.86	5.26	150.48	0.45	-0.70	7.13E-08	NO	NO	NO	NO
AT2G29170	43.52	4.05	22.93	2.63	-0.92	4.64E-02	NO	NO	NO	NO
AT2G29320	43.57	0.77	25.32	0.77	-0.78	6.28E-05	NO	NO	NO	NO

Table A3.3 (cont'd)

AT2G29630	895.67	12.20	601.79	3.34	-0.57	2.82E-02	NO	NO	NO	NO
AT2G30170	37.95	0.32	25.86	0.53	-0.55	2.65E-03	NO	NO	NO	NO
AT2G31230	35.16	0.60	22.22	0.41	-0.66	4.59E-03	NO	YES	NO	NO
AT2G31360	401.48	3.45	243.17	1.55	-0.72	1.83E-04	NO	NO	UP	NO
AT2G31840	70.59	1.42	50.54	0.54	-0.48	1.61E-02	NO	NO	NO	NO
AT2G33860	28.37	0.72	14.92	0.56	-0.93	1.91E-07	NO	YES	NO	NO
AT2G34650	9.26	0.46	5.41	0.14	-0.78	4.67E-03	NO	NO	NO	NO
AT2G34720	77.83	0.49	48.17	0.37	-0.69	2.51E-04	POSITIVE	YES	NO	NO
AT2G34850	5.32	0.17	1.80	0.11	-1.56	1.63E-03	NO	NO	NO	NO
AT2G35040	118.97	2.09	52.20	1.24	-1.19	0.00E+00	NO	NO	NO	NO
AT2G35700	21.89	0.61	6.63	0.41	-1.72	8.95E-06	NO	YES	NO	NO
AT2G35840	152.42	1.79	113.57	0.20	-0.42	6.53E-03	NO	NO	NO	NO
AT2G35960	85.68	1.59	60.07	1.26	-0.51	9.64E-03	NO	NO	UP	UP
AT2G36145	303.95	3.11	193.43	0.86	-0.65	2.30E-04	NEGATIVE	NO	NO	NO
AT2G36290	26.32	0.45	18.51	0.14	-0.51	2.14E-02	NO	NO	NO	NO
AT2G36390	100.24	1.80	50.23	0.53	-1.00	9.34E-09	NO	NO	NO	NO
AT2G36460	213.52	2.93	137.55	2.00	-0.63	2.49E-04	POSITIVE	NO	NO	NO
AT2G36470	24.80	0.29	14.94	0.34	-0.73	3.06E-04	NO	NO	NO	NO
AT2G37020	34.09	0.78	25.16	0.53	-0.44	1.77E-02	NO	NO	NO	NO
AT2G37600	127.87	1.66	82.01	1.93	-0.64	2.04E-04	NO	NO	NO	NO
AT2G37990	91.18	1.43	67.54	1.43	-0.43	3.15E-02	NO	NO	NO	NO
AT2G39470	170.35	0.85	112.01	0.33	-0.60	1.10E-04	NO	NO	NO	NO
AT2G40350	4.18	0.43	0.57	0.15	-2.87	1.15E-02	NO	YES	NO	NO
AT2G40840	109.73	1.47	49.12	0.61	-1.16	3.96E-11	NO	NO	NO	NO
AT2G41870	25.76	0.64	15.18	0.25	-0.76	1.82E-04	NO	NO	NO	UP
AT2G42130	144.72	1.35	87.94	1.79	-0.72	2.64E-10	NO	NO	NO	NO
AT2G42530	3092.92	21.48	1922.89	30.40	-0.69	8.67E-03	NO	NO	UP	UP
AT2G42540	12067.70	189.81	3281.28	31.62	-1.88	2.90E-08	NO	NO	UP	UP
AT2G43550	107.72	0.78	60.57	2.05	-0.83	1.59E-05	NEGATIVE	NO	NO	NO

Table A3.3 (cont'd)

AT2G44040	37.40	0.68	26.32	0.42	-0.51	1.42E-02	NO	NO	NO	NO
AT2G45560	29.52	0.25	13.52	0.61	-1.13	5.15E-11	NO	NO	NO	NO
AT2G46660	8.99	0.69	4.76	0.04	-0.92	4.05E-04	NO	NO	NO	NO
AT2G46680	81.38	2.39	44.11	1.95	-0.88	2.87E-07	POSITIVE	YES	NO	NO
AT2G47260	5.95	0.44	3.32	0.12	-0.84	9.12E-03	NO	YES	NO	NO
AT2G47420	48.41	0.75	31.99	0.74	-0.60	2.48E-03	NO	NO	NO	NO
AT2G47840	75.32	1.30	54.88	1.18	-0.46	2.99E-02	NO	NO	NO	NO
AT3G02020	76.86	1.20	55.74	1.34	-0.46	2.32E-02	NO	NO	NO	NO
AT3G02630	38.21	0.17	27.17	0.54	-0.49	1.53E-02	NO	NO	NO	NO
AT3G02730	303.82	0.78	224.60	2.28	-0.44	2.46E-02	NO	NO	NO	NO
AT3G03770	30.27	1.04	21.27	0.56	-0.51	3.92E-04	NO	NO	NO	UP
AT3G04860	7.36	0.22	4.14	0.28	-0.83	1.50E-02	NO	NO	NO	NO
AT3G05640	5.18	0.34	2.96	0.21	-0.81	3.24E-02	NO	NO	NO	NO
AT3G05730	822.32	4.21	458.72	6.35	-0.84	5.89E-06	NEGATIVE	NO	NO	NO
AT3G06035	82.79	1.81	59.35	1.30	-0.48	2.29E-02	NO	NO	NO	NO
AT3G06040	120.35	2.40	94.29	2.11	-0.35	3.35E-02	NO	NO	NO	NO
AT3G06510	103.37	0.41	78.63	0.56	-0.39	3.14E-03	NO	NO	NO	NO
AT3G08940	829.78	18.07	577.56	7.35	-0.52	2.85E-02	NO	NO	NO	NO
AT3G09540	18.48	0.87	8.35	0.52	-1.15	2.17E-07	NO	NO	UP	UP
AT3G09910	9.47	0.14	5.33	0.34	-0.83	2.26E-02	NO	NO	NO	NO
AT3G10920	493.76	4.17	346.63	2.61	-0.51	1.22E-02	NO	NO	NO	NO
AT3G11260	4.94	0.67	1.95	0.18	-1.34	3.93E-02	NO	YES	NO	NO
AT3G11410	58.31	1.19	31.89	1.05	-0.87	1.07E-06	NO	NO	NO	NO
AT3G11670	44.57	0.27	27.41	0.42	-0.70	6.02E-05	NO	NO	NO	UP
AT3G11720	18.99	0.67	13.54	0.27	-0.49	6.04E-03	NO	NO	NO	NO
AT3G12120	314.36	0.78	248.61	2.27	-0.34	2.71E-02	NO	NO	NO	NO
AT3G12490	638.39	4.59	403.22	5.61	-0.66	4.40E-05	POSITIVE	NO	NO	NO
AT3G12670	52.00	0.38	35.81	0.78	-0.54	4.39E-03	NO	NO	UP	NO
AT3G12990	19.86	0.36	12.46	0.47	-0.67	2.00E-03	NO	NO	NO	NO

Table A3.3 (cont'd)

AT3G13350	32.03	0.63	19.79	0.30	-0.69	5.90E-04	NO	YES	NO	NO
AT3G14200	50.43	0.68	37.16	0.72	-0.44	3.15E-02	NO	NO	NO	NO
AT3G14210	121.91	3.44	75.14	2.50	-0.70	1.17E-04	NO	NO	NO	NO
AT3G15140	13.31	0.97	7.44	0.21	-0.84	2.06E-03	NO	NO	NO	NO
AT3G15270	5.97	0.11	2.12	0.43	-1.49	5.11E-03	NO	YES	NO	NO
AT3G15357	1.59	0.04	0.21	0.03	-2.89	1.68E-02	NO	NO	NO	NO
AT3G15460	19.88	0.71	14.01	0.43	-0.50	4.51E-02	NO	NO	NO	NO
AT3G16140	858.71	6.55	562.03	5.84	-0.61	2.52E-03	NO	NO	NO	NO
AT3G16670	151.47	3.78	90.51	3.67	-0.74	7.43E-05	NO	NO	NO	NO
AT3G17120	17.75	0.13	11.82	0.62	-0.59	2.60E-03	NO	NO	NO	NO
AT3G17998	67.66	0.67	33.05	2.28	-1.03	1.13E-09	NO	NO	NO	NO
AT3G18090	1.72	0.02	0.62	0.06	-1.47	8.10E-04	NO	NO	NO	NO
AT3G18420	97.34	3.08	46.79	1.01	-1.06	1.59E-09	NO	NO	NO	NO
AT3G18490	204.13	1.77	146.43	1.98	-0.48	2.08E-02	NO	NO	NO	NO
AT3G18500	16.81	0.35	12.18	0.30	-0.47	2.93E-02	NO	NO	NO	NO
AT3G19800	60.07	0.55	36.03	0.96	-0.74	6.52E-06	NO	NO	DOWN	NO
AT3G19820	162.19	1.79	117.02	2.96	-0.47	7.85E-05	NO	NO	NO	NO
AT3G20300	13.84	0.93	9.15	0.27	-0.60	1.36E-02	NO	NO	NO	NO
AT3G20470	12.86	0.97	6.78	0.20	-0.92	1.44E-02	NO	NO	NO	NO
AT3G21870	47.52	0.39	31.29	0.51	-0.60	8.81E-03	NEGATIVE	NO	NO	NO
AT3G22150	14.86	0.43	9.59	0.27	-0.63	1.55E-03	NO	NO	NO	NO
AT3G22200	86.31	0.81	65.14	0.82	-0.41	5.30E-03	POSITIVE	NO	NO	NO
AT3G23570	35.98	0.54	26.35	0.56	-0.45	4.87E-02	NO	NO	NO	NO
AT3G23580	31.55	0.31	19.44	0.82	-0.70	3.93E-04	NO	NO	NO	NO
AT3G23640	24.94	0.34	17.78	0.42	-0.49	9.54E-04	NO	NO	NO	NO
AT3G23940	76.05	1.43	61.22	0.32	-0.31	3.96E-02	NO	NO	NO	NO
AT3G24420	20.37	2.34	11.95	0.81	-0.77	2.11E-03	NO	NO	NO	NO
AT3G24590	56.38	0.64	37.88	0.64	-0.57	4.77E-03	NO	NO	NO	NO
AT3G25770	495.10	1.10	352.86	7.04	-0.49	1.75E-02	NO	NO	NO	NO

Table A3.3 (cont'd)

AT3G26290	35.84	0.13	20.30	0.34	-0.82	1.07E-05	NO	NO	NO	NO
AT3G26300	58.94	0.52	41.05	0.14	-0.52	7.50E-03	NO	NO	NO	NO
AT3G26450	49.29	1.06	24.14	0.64	-1.03	5.97E-07	NO	NO	DOWN	NO
AT3G26900	54.10	0.68	39.69	0.63	-0.45	1.59E-02	NO	NO	NO	NO
AT3G27180	49.64	1.28	35.96	0.63	-0.47	1.92E-02	NO	NO	NO	NO
AT3G27690	622.51	11.88	312.38	7.42	-0.99	1.37E-07	NO	NO	NO	NO
AT3G29320	107.63	1.08	61.50	0.61	-0.81	1.24E-05	NO	NO	NO	NO
AT3G29390	9.87	0.21	5.84	0.04	-0.76	2.19E-03	NO	NO	NO	NO
AT3G46780	320.82	3.82	219.87	1.36	-0.55	9.05E-03	NO	NO	NO	NO
AT3G46970	413.39	5.52	211.20	0.97	-0.97	1.20E-05	NO	NO	NO	NO
AT3G47160	49.82	0.74	36.99	0.96	-0.43	4.32E-02	NO	NO	NO	NO
AT3G47860	255.85	0.78	136.79	1.99	-0.90	1.35E-07	NO	NO	NO	NO
AT3G48460	23.89	0.69	15.27	1.51	-0.65	3.40E-03	NO	NO	NO	NO
AT3G48610	23.50	0.46	15.44	0.65	-0.61	2.93E-03	NO	NO	NO	NO
AT3G48700	3.15	0.48	0.82	0.13	-1.93	1.83E-03	NO	NO	NO	NO
AT3G49560	171.74	0.51	123.01	2.72	-0.48	1.18E-02	NO	NO	NO	NO
AT3G49910	1235.69	9.19	887.91	12.03	-0.48	4.98E-02	NO	NO	NO	NO
AT3G50500	31.10	0.08	20.57	0.31	-0.60	2.41E-03	NO	NO	NO	NO
AT3G50740	19.33	1.25	13.74	0.42	-0.49	3.10E-02	NO	NO	NO	NO
AT3G52070	26.66	2.64	15.97	1.15	-0.74	3.34E-02	NO	NO	NO	NO
AT3G52390	34.31	0.74	20.61	0.11	-0.74	5.88E-05	NO	NO	NO	NO
AT3G53000	18.41	0.22	12.50	0.29	-0.56	2.17E-02	NO	NO	NO	NO
AT3G53530	17.90	0.42	11.74	0.93	-0.61	2.38E-02	NO	NO	NO	NO
AT3G54050	1147.94	11.42	836.04	5.08	-0.46	3.00E-02	NO	NO	NO	NO
AT3G55120	95.30	1.72	64.13	1.81	-0.57	4.25E-03	NO	NO	NO	NO
AT3G55580	87.91	1.88	40.95	0.93	-1.10	1.72E-10	NO	NO	UP	NO
AT3G55610	306.79	5.19	173.37	2.72	-0.82	3.00E-07	NO	NO	UP	NO
AT3G56490	131.05	0.44	96.96	1.71	-0.43	3.12E-02	NO	NO	NO	NO
AT3G57030	97.85	0.36	63.76	1.80	-0.62	7.39E-04	NO	NO	NO	NO

Table A3.3 (cont'd)

AT3G57062	51.13	1.30	35.45	1.02	-0.53	3.99E-02	NO	NO	NO	NO
AT3G58010	59.01	0.74	40.30	0.56	-0.55	5.71E-03	NO	NO	NO	NO
AT3G58070	21.20	0.51	13.84	0.64	-0.61	1.55E-02	NO	YES	NO	NO
AT3G59980	126.01	1.31	91.51	1.92	-0.46	1.76E-02	NEGATIVE	NO	NO	NO
AT3G60530	31.80	0.22	20.07	1.03	-0.66	2.12E-03	NO	YES	NO	NO
AT3G61080	41.04	0.22	20.25	0.31	-1.02	1.23E-07	NO	NO	NO	NO
AT3G62750	9.31	0.16	6.14	0.08	-0.60	4.31E-02	NO	NO	NO	NO
AT3G63160	4669.45	13.44	2926.92	14.12	-0.67	8.93E-03	NO	NO	NO	NO
AT4G00310	11.29	0.23	6.19	0.41	-0.87	1.60E-03	NO	NO	NO	NO
AT4G00670	0.32	0.07	0.05	0.03	-2.60	1.47E-02	NO	NO	NO	NO
AT4G00810	390.27	2.31	290.01	4.20	-0.43	2.63E-02	NO	NO	NO	NO
AT4G01130	18.25	0.13	6.22	0.28	-1.55	1.97E-09	NO	NO	NO	NO
AT4G01460	12.04	0.29	7.81	1.05	-0.63	2.20E-02	NO	YES	NO	NO
AT4G01670	17.36	0.08	9.26	0.26	-0.91	3.65E-04	NO	NO	NO	NO
AT4G01940	336.24	0.38	190.23	2.93	-0.82	2.64E-06	NO	NO	NO	NO
AT4G02920	396.75	3.16	265.80	1.68	-0.58	2.87E-04	NO	NO	NO	NO
AT4G04330	118.04	0.56	74.91	0.38	-0.66	3.64E-04	NO	NO	NO	NO
AT4G04470	53.27	0.63	39.34	0.30	-0.44	4.54E-02	NO	NO	NO	NO
AT4G04630	9.18	0.03	4.29	0.71	-1.10	1.98E-03	NO	NO	NO	NO
AT4G04840	14.78	0.57	8.56	0.80	-0.79	1.81E-02	NEGATIVE	NO	DOWN	NO
AT4G09970	28.64	1.28	20.16	0.93	-0.51	3.23E-02	NO	NO	NO	NO
AT4G10380	13.18	0.61	8.80	0.54	-0.58	2.69E-02	NO	NO	NO	NO
AT4G12000	19.93	0.49	11.64	0.56	-0.78	6.39E-04	POSITIVE	NO	UP	NO
AT4G12470	1853.39	67.10	1159.34	45.80	-0.68	8.50E-03	NO	NO	UP	UP
AT4G12600	273.29	5.11	190.84	3.16	-0.52	6.55E-03	NO	NO	NO	NO
AT4G12900	33.53	0.88	19.01	0.19	-0.82	1.09E-03	NO	NO	NO	NO
AT4G13150	33.31	0.05	20.37	0.19	-0.71	5.84E-04	NO	NO	NO	NO
AT4G13220	126.85	1.96	94.97	0.55	-0.42	4.24E-02	NEGATIVE	NO	NO	NO
AT4G13493	338.15	0.56	222.03	1.90	-0.61	4.49E-02	NO	NO	NO	NO

Table A3.3 (cont'd)

										DOW
AT4G13840	59.95	0.59	30.00	0.47	-1.00	7.31E-09	NO	NO	NO	N
AT4G14020	167.14	2.61	70.37	1.95	-1.25	1.79E-11	NO	NO	NO	NO
AT4G14105	1.06	0.84	0.55	0.38	-0.94	4.13E-06	NO	NO	NO	NO
AT4G14605	45.34	1.17	27.90	0.20	-0.70	1.68E-04	NO	NO	NO	NO
AT4G15510	91.04	1.31	69.27	0.47	-0.39	3.95E-02	NO	NO	NO	NO
AT4G16060	37.62	0.67	20.73	0.53	-0.86	2.11E-05	NO	NO	NO	NO
AT4G17090	1829.44	6.20	912.16	11.92	-1.00	1.04E-03	NO	NO	UP	NO
AT4G17550	60.35	1.96	34.18	0.87	-0.82	2.90E-06	NO	NO	UP	UP
AT4G17730	90.08	0.57	68.08	1.10	-0.40	4.27E-02	NO	NO	NO	NO
AT4G17770	34.77	1.66	16.44	0.76	-1.08	2.66E-10	NO	NO	NO	NO
AT4G18670	13.97	0.48	10.34	0.25	-0.44	4.68E-02	NEGATIVE	NO	NO	NO
AT4G18700	105.13	1.34	69.84	0.88	-0.59	1.67E-03	NO	NO	NO	NO
AT4G19120	123.77	0.93	85.47	1.30	-0.53	6.11E-04	NO	NO	NO	NO
AT4G19410	85.26	1.17	56.20	0.46	-0.60	6.08E-04	NO	NO	NO	NO
AT4G20170	33.27	0.09	18.15	0.18	-0.87	1.52E-06	NO	NO	NO	NO
AT4G21215	50.47	0.34	20.32	0.52	-1.31	1.53E-13	NO	NO	NO	NO
AT4G22890	240.03	1.93	182.99	2.69	-0.39	1.18E-03	NO	NO	NO	NO
AT4G23630	353.03	2.56	246.82	2.95	-0.52	1.87E-02	POSITIVE	NO	UP	UP
AT4G24010	4.40	0.07	2.06	0.16	-1.10	8.23E-04	NO	NO	NO	NO
AT4G24930	124.10	1.51	88.81	0.88	-0.48	1.01E-02	NO	NO	NO	NO
AT4G24960	78.24	1.44	51.39	1.56	-0.61	1.76E-03	NO	NO	UP	UP
AT4G25470	45.16	0.68	30.27	0.13	-0.58	8.06E-03	NO	YES	UP	UP
AT4G25910	83.37	0.41	58.94	1.60	-0.50	1.19E-02	NO	NO	NO	NO
AT4G26530	147.61	1.37	98.04	1.27	-0.59	7.07E-04	NO	NO	NO	NO
AT4G26555	35.33	0.47	22.77	0.79	-0.63	6.71E-03	NO	NO	NO	NO
AT4G26670	80.33	1.74	51.52	1.11	-0.64	5.13E-04	NO	NO	NO	NO
AT4G26950	19.08	0.69	11.05	0.85	-0.79	2.74E-02	NO	NO	NO	NO
AT4G27440	1757.23	14.33	765.72	10.88	-1.20	2.50E-06	NO	NO	NO	NO

Table A3.3 (cont'd)

AT4G27520	417.18	7.47	256.97	2.89	-0.70	3.76E-04	NO	NO	UP	NO
AT4G27560	26.69	0.49	19.70	0.70	-0.44	4.33E-02	NO	NO	NO	NO
AT4G27570	6.79	0.09	3.52	0.19	-0.95	4.41E-03	POSITIVE	NO	UP	NO
AT4G27800	135.76	1.46	103.14	0.39	-0.40	4.10E-02	NO	NO	NO	NO
AT4G28706	30.01	0.17	16.80	0.42	-0.84	1.79E-06	NO	NO	NO	NO
AT4G28750	902.84	5.77	658.46	5.73	-0.46	3.40E-02	NO	NO	NO	NO
AT4G29490	22.58	0.42	16.51	0.26	-0.45	4.23E-02	NO	NO	NO	NO
AT4G30710	7.60	0.21	4.02	0.10	-0.92	4.71E-05	NO	NO	NO	NO
AT4G31780	86.15	1.80	61.32	0.70	-0.49	1.25E-02	NO	NO	NO	UP
AT4G31810	29.04	0.38	21.09	0.55	-0.46	3.12E-02	NO	NO	NO	NO
AT4G32520	82.38	1.40	58.89	0.48	-0.48	1.07E-03	NO	NO	NO	NO
AT4G32590	123.17	1.51	98.43	1.10	-0.32	2.59E-02	NO	NO	NO	NO
AT4G32950	2.98	0.00	0.79	0.22	-1.91	7.88E-03	NO	NO	NO	NO
AT4G33480	67.68	1.35	47.50	0.31	-0.51	8.13E-03	NO	NO	NO	NO
AT4G33490	22.09	0.29	15.51	0.47	-0.51	6.67E-03	NO	NO	NO	NO
AT4G33520	65.85	0.59	33.09	0.57	-0.99	1.69E-13	NEGATIVE	NO	NO	NO
AT4G33905	5.26	0.37	1.70	0.34	-1.63	1.64E-03	POSITIVE	NO	NO	NO
AT4G33980	90.40	1.76	67.35	1.26	-0.42	2.81E-02	NO	NO	UP	NO
AT4G34560	10.01	0.46	5.55	0.50	-0.85	1.22E-02	NO	NO	NO	NO
AT4G34950	270.64	0.74	165.98	1.53	-0.71	3.14E-04	NO	NO	NO	NO
AT4G34990	17.70	0.31	7.96	0.27	-1.15	4.92E-05	NO	YES	UP	UP
AT4G35450	404.87	3.60	335.41	4.30	-0.27	8.71E-03	NO	NO	NO	NO
AT4G36240	12.02	0.14	3.79	0.68	-1.67	1.23E-05	NO	YES	NO	NO
AT4G36570	24.70	1.42	12.03	0.97	-1.04	3.28E-02	NO	NO	NO	NO
AT4G37320	25.27	0.44	8.77	0.19	-1.53	1.85E-14	NO	NO	UP	NO
AT4G37330	14.23	0.11	9.79	0.34	-0.54	2.89E-02	NO	NO	NO	NO
AT4G37470	112.63	1.04	72.40	1.12	-0.64	3.71E-04	NO	NO	NO	NO
AT4G37660	46.33	0.80	31.63	0.98	-0.55	1.57E-02	NO	NO	NO	NO
AT4G38050	4.99	0.10	3.05	0.07	-0.71	2.60E-02	NO	NO	NO	NO

Table A3.3 (cont'd)

AT4G39260	253.08	5.38	170.07	3.23	-0.57	1.07E-08	NO	NO	NO	NO
AT4G39510	16.21	0.42	11.05	1.06	-0.55	1.37E-02	NO	NO	NO	NO
AT4G39540	31.11	0.54	22.28	0.16	-0.48	2.50E-02	NO	NO	NO	NO
AT4G40065	63.05	0.41	31.93	0.50	-0.98	9.46E-08	NO	NO	NO	NO
AT5G01520	15.75	0.44	4.32	0.29	-1.87	7.72E-12	NO	NO	NO	NO
AT5G02190	11.45	0.66	7.93	0.28	-0.53	4.54E-02	NO	NO	NO	NO
AT5G02240	661.26	3.99	396.65	0.69	-0.74	1.37E-04	NO	NO	NO	NO
AT5G02790	206.73	0.43	143.53	0.10	-0.53	4.85E-03	NO	NO	NO	NO
AT5G03230	20.02	0.42	11.76	0.52	-0.77	6.75E-03	NO	NO	NO	NO
AT5G03670	6.61	0.19	3.65	0.26	-0.86	3.68E-03	NO	NO	NO	NO
AT5G06530	66.07	0.74	45.83	1.12	-0.53	3.16E-03	NO	NO	NO	NO
AT5G07580	53.96	0.80	37.87	1.23	-0.51	1.06E-02	NO	YES	NO	NO
AT5G07690	57.12	1.04	27.38	1.51	-1.06	1.81E-09	NO	YES	NO	NO
AT5G08030	3.70	0.28	1.13	0.13	-1.71	2.22E-03	NO	NO	NO	NO
AT5G08100	35.93	0.22	26.49	0.80	-0.44	4.09E-02	NO	NO	NO	NO
AT5G08260	137.93	0.92	95.57	1.34	-0.53	6.21E-03	NO	NO	NO	NO
AT5G08410	380.19	2.36	287.35	1.14	-0.40	4.39E-02	NO	NO	NO	NO
AT5G09672	5.57	1.39	2.65	0.59	-1.07	3.51E-04	NO	NO	NO	NO
AT5G10170	4.62	0.11	1.95	0.02	-1.24	9.25E-04	NO	NO	NO	NO
AT5G11110	12.76	0.72	8.00	0.14	-0.67	5.34E-04	NO	NO	UP	NO
AT5G11310	7.07	0.08	2.65	0.27	-1.41	1.06E-06	NO	NO	NO	NO
AT5G11450	255.86	0.78	182.63	2.88	-0.49	1.18E-02	NO	NO	NO	NO
AT5G11790	53.17	0.81	39.02	1.09	-0.45	2.93E-02	NO	NO	NO	NO
AT5G11840	44.16	0.24	31.62	0.38	-0.48	2.92E-02	NO	NO	NO	NO
AT5G13930	643.47	16.94	222.62	3.79	-1.53	0.00E+00	NO	NO	NO	NO
AT5G14090	9.91	0.74	4.73	0.30	-1.07	6.12E-04	NO	NO	NO	NO
AT5G14545	3469.79	48.24	606.23	4.05	-2.52	5.50E-07	NO	NO	NO	NO
AT5G14565	53.30	1.68	20.90	0.40	-1.35	0.00E+00	NO	NO	NO	NO
AT5G14570	32.90	0.35	21.95	0.28	-0.58	2.58E-03	NO	NO	UP	UP

Table A3.3 (cont'd)

AT5G15160	9.81	0.13	4.03	0.23	-1.28	1.85E-02	NO	NO	NO	NO
AT5G15190	19.10	1.07	9.97	0.18	-0.94	2.84E-03	NO	NO	NO	NO
AT5G15360	80.07	1.93	50.32	0.93	-0.67	7.02E-04	NO	NO	NO	NO
AT5G15650	517.49	9.35	340.35	2.74	-0.60	6.11E-03	NO	NO	NO	NO
AT5G16010	154.66	2.62	111.70	1.10	-0.47	1.81E-02	NO	NO	NO	NO
AT5G16290	43.91	0.94	30.25	0.44	-0.54	3.87E-03	NO	NO	NO	NO
AT5G17840	65.80	0.82	40.85	0.20	-0.69	7.49E-04	NO	NO	NO	NO
AT5G18680	18.33	0.14	12.50	0.13	-0.55	2.10E-02	NO	YES	NO	NO
AT5G19090	38.88	0.34	23.06	1.18	-0.75	1.30E-08	NO	NO	NO	NO
AT5G19150	31.04	0.05	23.37	0.27	-0.41	3.20E-02	NO	NO	NO	NO
AT5G19460	32.24	0.65	22.62	0.40	-0.51	1.44E-02	NO	NO	NO	NO
AT5G20280	129.33	2.10	86.86	0.54	-0.57	6.47E-03	POSITIVE	NO	NO	NO
AT5G20700	901.96	9.56	620.43	2.19	-0.54	2.36E-02	NO	NO	NO	NO
AT5G21020	196.73	1.04	127.91	1.72	-0.62	5.07E-04	NO	NO	NO	NO
AT5G21100	6.66	0.06	4.05	0.53	-0.72	1.39E-02	NO	NO	DOWN	NO
AT5G22020	27.41	0.63	20.13	0.34	-0.45	4.64E-02	NO	NO	NO	NO
AT5G22070	32.55	0.61	21.75	0.32	-0.58	2.63E-03	NO	NO	NO	NO
AT5G22650	254.73	2.29	187.60	3.36	-0.44	2.09E-02	NO	NO	NO	NO
AT5G22880	253.00	1.73	161.08	3.85	-0.65	4.14E-04	NO	NO	NO	NO
AT5G24300	157.63	1.66	108.50	0.63	-0.54	7.17E-03	NO	NO	NO	UP
AT5G24490	429.59	1.89	247.88	0.76	-0.79	1.80E-05	NO	NO	NO	NO
AT5G25890	22.52	0.63	14.95	0.10	-0.59	3.32E-02	NO	YES	NO	NO
AT5G26570	173.21	2.26	84.75	0.16	-1.03	8.40E-08	NO	NO	NO	NO
AT5G28150	28.30	0.82	14.78	0.37	-0.94	2.14E-06	NO	NO	NO	UP
AT5G28300	53.64	1.63	29.25	1.23	-0.87	4.43E-07	NO	YES	NO	NO
AT5G37360	52.59	0.34	31.46	0.51	-0.74	9.76E-05	NO	NO	NO	NO
AT5G39240	274.06	2.87	158.24	5.08	-0.79	1.25E-04	NO	NO	NO	NO
AT5G39350	4.44	0.23	2.50	0.14	-0.83	2.14E-02	NO	NO	NO	NO
AT5G42420	49.40	0.24	37.08	0.65	-0.41	4.47E-02	NO	NO	NO	NO

Table A3.3 (cont'd)

AT5G42530	3652.28	130.06	1728.50	9.29	-1.08	2.54E-06	NO	NO	NO	NO
AT5G42900	200.37	3.24	157.06	0.76	-0.35	5.63E-03	NO	NO	UP	NO
AT5G43150	70.15	1.13	33.67	0.91	-1.06	1.21E-06	NO	NO	NO	NO
AT5G43400	1.17	0.10	0.43	0.05	-1.44	3.44E-02	NO	NO	NO	NO
AT5G43850	101.90	0.30	70.42	1.53	-0.53	6.66E-03	NO	NO	NO	NO
AT5G44005	34.19	0.58	13.55	0.71	-1.34	7.59E-07	NO	NO	NO	NO
AT5G44530	12.75	0.29	8.85	0.16	-0.53	1.40E-02	NO	NO	NO	UP
AT5G45680	279.32	1.44	142.13	1.01	-0.97	7.27E-09	NO	NO	NO	NO
AT5G45930	178.18	1.66	93.94	1.35	-0.92	6.20E-08	NO	NO	NO	NO
AT5G46160	84.25	1.38	52.30	1.64	-0.69	7.77E-05	NO	NO	NO	NO
AT5G46600	13.77	0.30	7.64	0.29	-0.85	1.42E-04	NO	NO	NO	NO
AT5G46710	30.15	0.53	15.99	0.46	-0.91	1.74E-05	NO	YES	UP	NO
AT5G47060	66.36	1.15	29.78	0.89	-1.16	3.09E-10	NO	NO	UP	NO
AT5G47540	28.64	0.41	18.57	0.61	-0.63	1.93E-03	NEGATIVE	NO	NO	NO
AT5G47700	460.51	2.63	317.94	9.62	-0.53	3.40E-03	NO	NO	NO	NO
AT5G48630	37.21	0.85	26.28	0.41	-0.50	3.38E-02	NO	NO	NO	NO
AT5G48790	55.97	0.42	28.74	0.45	-0.96	1.83E-07	NO	NO	NO	NO
AT5G49190	3.15	0.26	1.55	0.03	-1.02	7.50E-03	NO	NO	NO	NO
AT5G50240	14.84	0.17	8.63	0.44	-0.78	8.58E-03	NO	NO	NO	NO
AT5G50250	179.70	1.35	130.69	2.00	-0.46	1.88E-02	NO	NO	NO	NO
AT5G50360	22.39	0.45	11.93	0.22	-0.91	7.07E-05	NO	NO	NO	NO
AT5G51820	150.21	1.44	89.79	0.62	-0.74	4.52E-05	NO	NO	NO	NO
AT5G54580	63.49	0.89	46.48	1.70	-0.45	3.67E-02	NO	NO	NO	NO
AT5G55280	177.75	2.58	125.73	1.13	-0.50	9.98E-03	NEGATIVE	NO	NO	NO
AT5G57345	205.84	2.27	147.91	0.85	-0.48	1.26E-02	NO	NO	NO	NO
AT5G57760	35.34	4.42	14.51	2.76	-1.28	2.01E-05	NO	NO	NO	NO
AT5G58490	128.35	1.49	71.78	0.87	-0.84	1.27E-06	NO	NO	NO	NO
AT5G58700	14.42	0.58	5.98	0.36	-1.27	2.18E-09	NO	NO	UP	NO
AT5G59080	20.23	0.26	9.75	0.59	-1.05	2.01E-04	NO	NO	NO	NO

Table A3.3 (cont'd)

AT5G59680	1.00	0.16	0.28	0.05	-1.82	9.25E-03	NO	NO	NO	NO
AT5G59700	2.08	0.08	0.97	0.08	-1.10	1.40E-02	NO	NO	NO	NO
AT5G59950	79.63	0.18	53.71	1.11	-0.57	1.48E-04	POSITIVE	NO	NO	NO
AT5G60540	94.80	0.50	51.24	0.66	-0.89	3.14E-07	NO	NO	NO	NO
AT5G61520	30.16	0.18	20.95	0.22	-0.53	6.38E-04	NO	NO	NO	NO
AT5G61660	145.45	1.28	106.65	2.18	-0.45	2.71E-02	NO	NO	NO	NO
AT5G61810	39.39	0.22	26.32	0.72	-0.58	2.24E-03	NO	NO	UP	UP
AT5G62360	149.35	2.66	97.20	3.60	-0.62	7.36E-04	NO	NO	UP	NO
AT5G62960	12.13	1.00	4.77	0.23	-1.35	2.43E-06	NO	NO	NO	UP
AT5G63810	44.33	0.39	31.87	0.39	-0.48	1.43E-02	NO	NO	NO	NO
AT5G64040	3250.16	81.42	1924.23	33.98	-0.76	5.00E-03	NO	NO	NO	NO
AT5G64240	11.61	0.28	7.14	0.08	-0.70	6.58E-03	NO	NO	DOWN	NO
AT5G64350	330.08	0.58	247.18	3.57	-0.42	3.42E-02	NO	NO	NO	NO
AT5G64460	104.70	0.94	76.87	0.87	-0.45	1.25E-03	NO	NO	NO	NO
AT5G64550	19.16	0.19	11.35	0.76	-0.76	7.49E-05	NO	NO	NO	NO
AT5G65220	1230.84	17.13	737.48	17.73	-0.74	4.90E-04	NEGATIVE	NO	NO	NO
AT5G65420	8.51	0.21	4.92	0.14	-0.79	1.49E-02	NO	NO	NO	NO
AT5G65480	95.46	1.35	65.95	0.29	-0.53	5.62E-03	NO	NO	NO	NO
AT5G66420	25.91	0.49	19.23	0.42	-0.43	1.99E-02	NO	NO	NO	NO
AT5G66530	271.43	2.45	192.21	2.00	-0.50	9.57E-03	NO	NO	NO	NO
AT5G66610	4.51	0.15	2.31	0.17	-0.97	6.22E-03	NO	YES	NO	NO
AT5G66720	118.40	0.94	81.66	1.59	-0.54	1.84E-03	NO	NO	NO	NO

Table A3.4. 1656 Category 1 FTD Candidate Genes with higher expression in IT at 2 weeks of cold-acclimation. Genes are ordered by their AGI numbers. FPKM values are the average of three biological replicates (n=3). Standard error (SE) values are also given for each FPKM value. Genes were considered significantly different between SW and IT under non-acclimated conditions if they had a Benjamini-Hochberg corrected p-value (FDR P-VALUE) of <0.05 and a minimum of ≥ 3 FPKM in IT. The differential expression of each gene is also represented by the logarithm (base=2) of the fold change (LOG2.FC). This table also denotes if a gene was previously described as positively or negatively correlating with freezing tolerance (HANNAH;(6)), a transcription factor (TF), a gene previously described as up- or down-regulated by 1 week of cold (COS, (25)), identified as an up- or down-regulated component of the CBF regulon (CBF REGULON; (25)).

AGI	SW.IT.2WK					FDR P-VALUE	HANNAH?	TF?	COS?	CBF REG?
	SW.2WK FPKM	SW.2WK SE	IT.2WK FPKM	IT.2WK SE	SW.IT.2WK LOG2FC					
AT1G01180	3.95	0.21	11.81	0.30	1.58	7.19E-07	NO	NO	NO	NO
AT1G01240	12.36	0.26	17.96	0.17	0.54	2.13E-02	NO	NO	NO	NO
AT1G01340	1.05	0.18	11.58	0.46	3.46	0.00E+00	NO	NO	NO	NO
AT1G01440	5.27	0.06	8.17	0.14	0.63	1.71E-02	NO	NO	NO	NO
AT1G01490	31.27	0.73	46.49	0.39	0.57	1.57E-03	NO	NO	NO	NO
AT1G01560	3.21	0.76	22.16	0.42	2.79	0.00E+00	NO	NO	NO	NO
AT1G02335	4.60	0.26	8.63	0.13	0.91	4.10E-02	NO	NO	NO	NO
AT1G02450	2.66	0.39	28.09	1.90	3.40	7.37E-13	NO	NO	NO	NO
AT1G02500	49.68	1.00	139.80	2.80	1.49	0.00E+00	NO	NO	NO	NO
AT1G02640	1.31	0.04	2.49	0.25	0.93	2.73E-02	NEGATIVE	NO	NO	NO
AT1G02920	113.78	4.47	449.52	0.57	1.98	0.00E+00	NO	NO	NO	NO
AT1G02930	207.21	14.24	1012.24	4.71	2.29	0.00E+00	NO	NO	NO	NO
AT1G03060	4.82	0.20	7.04	0.26	0.55	4.99E-03	NO	NO	NO	NO
AT1G03210	21.79	0.75	31.44	0.35	0.53	3.63E-02	NO	NO	NO	NO
AT1G03290	20.73	0.70	45.39	0.30	1.13	0.00E+00	POSITIVE	NO	NO	NO
AT1G03370	11.32	0.61	19.25	0.41	0.77	3.27E-05	NO	NO	NO	NO
AT1G03400	27.38	0.96	43.81	0.16	0.68	6.71E-04	NO	NO	NO	NO
AT1G03730	6.51	0.21	10.90	0.38	0.74	2.74E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G04140	18.15	0.43	23.81	0.27	0.39	1.35E-02	NO	NO	NO	NO
AT1G04425	30.21	0.54	63.27	3.12	1.07	5.43E-05	NO	NO	NO	NO
AT1G04530	34.88	0.58	77.34	1.48	1.15	1.85E-11	NO	NO	NO	NO
AT1G04770	8.61	0.44	21.16	0.86	1.30	4.52E-08	NO	NO	NO	NO
AT1G04780	20.89	0.32	39.76	0.39	0.93	7.32E-08	NO	NO	NO	NO
AT1G04980	8.28	0.38	32.65	0.84	1.98	0.00E+00	NO	NO	NO	NO
AT1G05000	3.05	0.34	6.33	0.65	1.05	7.42E-03	NO	NO	NO	NO
AT1G05010	96.25	0.92	193.56	0.98	1.01	3.09E-09	NO	NO	NO	DOWN
AT1G05230	6.76	0.24	9.59	0.39	0.50	1.06E-02	NO	YES	NO	NO
AT1G05570	12.02	0.43	20.02	0.31	0.74	2.55E-05	NO	NO	NO	NO
AT1G05630	5.93	0.24	10.10	0.23	0.77	1.44E-05	NO	NO	NO	NO
AT1G05880	0.33	0.14	5.50	0.14	4.07	0.00E+00	NO	NO	NO	NO
AT1G05910	14.38	0.27	20.19	0.23	0.49	1.20E-02	NO	NO	NO	NO
AT1G06410	26.90	0.39	43.52	0.29	0.69	1.08E-04	NO	NO	NO	NO
AT1G06430	55.48	0.90	86.22	1.44	0.64	4.06E-04	NO	NO	NO	NO
AT1G06620	1.91	0.33	5.83	0.24	1.61	1.27E-04	NO	NO	NO	NO
AT1G06645	8.20	0.40	15.11	0.19	0.88	6.60E-04	NO	NO	NO	NO
AT1G06650	52.94	0.09	75.50	0.52	0.51	6.95E-04	NO	NO	NO	NO
AT1G07000	7.43	1.16	22.05	0.53	1.57	1.93E-14	NO	NO	NO	NO
AT1G07128	2.12	0.11	5.22	0.14	1.30	2.64E-02	NO	NO	NO	NO
AT1G07135	9.88	1.10	24.64	0.26	1.32	6.79E-06	NO	NO	NO	NO
AT1G07590	141.59	1.64	208.87	1.46	0.56	1.16E-02	NO	NO	NO	NO
AT1G07600	4144.39	75.33	6055.84	37.14	0.55	1.62E-02	NO	NO	NO	NO
AT1G07620	5.92	0.72	18.54	0.49	1.65	0.00E+00	NO	NO	NO	NO
AT1G07640	5.78	0.28	10.15	0.08	0.81	2.26E-03	NO	YES	NO	NO
AT1G07650	19.03	0.32	31.75	0.41	0.74	1.02E-08	NO	NO	NO	NO
AT1G07900	3.75	0.97	9.84	1.08	1.39	1.11E-02	NO	YES	NO	NO
AT1G08050	1.68	0.17	13.21	0.54	2.98	0.00E+00	NO	NO	NO	NO
AT1G08060	3.04	0.08	4.42	0.32	0.54	5.85E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G08450	41.91	1.71	157.81	2.45	1.91	0.00E+00	NO	NO	NO	NO
AT1G08600	5.08	0.18	7.18	0.23	0.50	3.07E-03	NO	NO	NO	NO
AT1G08620	5.50	0.17	7.58	0.19	0.46	4.15E-02	NO	YES	NO	NO
AT1G08650	31.91	0.91	43.35	0.75	0.44	3.67E-02	NO	NO	NO	NO
AT1G08830	215.63	0.76	418.66	6.35	0.96	3.24E-08	NO	NO	NO	NO
AT1G08930	70.02	0.64	95.18	0.93	0.44	2.20E-02	NO	NO	NO	NO
AT1G08940	2.45	0.33	15.47	0.03	2.66	6.78E-12	NO	NO	NO	NO
AT1G09180	1.80	0.12	9.33	0.99	2.38	3.46E-06	NO	NO	NO	NO
AT1G09210	159.65	2.84	360.11	5.38	1.17	3.99E-10	NO	NO	NO	NO
AT1G09415	24.92	0.72	40.42	1.17	0.70	5.69E-03	NO	NO	DOWN	NO
AT1G09560	62.06	1.61	221.01	1.82	1.83	0.00E+00	NO	NO	NO	NO
AT1G09850	28.52	0.84	43.26	0.56	0.60	1.87E-03	NO	NO	NO	NO
AT1G09970	28.73	0.71	89.69	2.41	1.64	0.00E+00	NO	NO	NO	NO
AT1G10020	7.35	0.02	13.02	0.70	0.83	2.52E-04	NO	NO	NO	NO
AT1G10050	3.83	0.30	5.97	0.40	0.64	1.39E-02	NO	NO	NO	NO
AT1G10140	6.55	0.30	14.11	0.39	1.11	2.78E-03	NO	NO	NO	NO
AT1G10155	29.68	0.41	59.89	0.46	1.01	1.39E-05	NO	NO	NO	NO
AT1G10170	8.59	0.17	12.58	0.29	0.55	6.96E-03	NO	NO	NO	NO
AT1G10550	5.43	0.78	14.71	0.53	1.44	1.35E-06	NO	NO	NO	NO
AT1G10920	4.66	0.19	7.46	0.37	0.68	3.21E-03	NO	NO	NO	NO
AT1G10960	121.53	1.38	167.31	2.05	0.46	1.95E-02	NEGATIVE	NO	DOWN	NO
AT1G11050	3.59	0.37	11.14	0.56	1.63	1.16E-11	NO	NO	NO	NO
AT1G11190	0.93	0.22	3.11	0.63	1.74	2.16E-03	NO	NO	NO	NO
AT1G11310	35.64	0.34	66.00	0.67	0.89	5.82E-08	NO	NO	NO	NO
AT1G11410	2.23	0.12	3.74	0.23	0.75	2.66E-02	NO	NO	NO	NO
AT1G11790	15.75	0.69	21.96	0.05	0.48	2.99E-02	NO	NO	NO	NO
AT1G11905	27.86	0.87	41.86	1.08	0.59	2.10E-03	NO	NO	NO	NO
AT1G12110	25.71	0.17	35.47	0.43	0.46	2.09E-02	NO	NO	NO	NO
AT1G12140	18.41	0.08	28.13	0.67	0.61	2.27E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G12240	20.47	0.28	30.92	0.62	0.59	1.59E-03	NEGATIVE	NO	DOWN	NO
AT1G12290	2.66	0.17	5.24	0.29	0.98	1.73E-04	NO	NO	NO	NO
AT1G12360	24.86	0.12	39.12	0.14	0.65	3.36E-04	NO	NO	NO	NO
AT1G12420	8.25	0.23	17.87	0.30	1.12	1.14E-07	NO	NO	NO	NO
AT1G12440	144.56	1.06	209.85	1.45	0.54	3.32E-03	NO	NO	NO	NO
AT1G12520	37.47	0.20	60.71	0.78	0.70	3.88E-05	NO	NO	NO	NO
AT1G12990	13.24	0.33	24.86	0.25	0.91	1.82E-06	NO	NO	NO	NO
AT1G13210	16.94	0.87	44.47	0.86	1.39	0.00E+00	NO	NO	NO	NO
AT1G13245	47.19	1.42	107.41	2.25	1.19	2.44E-09	NO	NO	NO	NO
AT1G13260	61.71	1.05	92.94	1.26	0.59	1.32E-03	NO	YES	UP	NO
AT1G13950	2.37	0.25	6.15	0.28	1.38	1.53E-02	NO	NO	NO	NO
AT1G14010	33.14	0.33	64.06	0.45	0.95	5.13E-07	NO	NO	NO	NO
AT1G14330	4.07	0.19	9.15	0.38	1.17	3.19E-06	NO	NO	NO	NO
AT1G14360	18.38	0.48	50.23	1.00	1.45	0.00E+00	NO	NO	NO	NO
AT1G14370	7.13	0.35	17.52	0.60	1.30	2.38E-09	NO	NO	NO	NO
AT1G14820	40.45	0.40	54.23	0.57	0.42	3.67E-02	NO	NO	NO	NO
AT1G14870	1.17	0.34	8.73	0.38	2.91	5.19E-07	NO	NO	NO	NO
AT1G14890	4.94	0.11	9.45	0.16	0.94	1.56E-02	NO	NO	NO	NO
AT1G15020	8.43	0.12	11.66	0.16	0.47	4.70E-02	NO	NO	NO	NO
AT1G15340	52.61	0.62	82.58	1.26	0.65	3.16E-04	NO	NO	NO	NO
AT1G15430	5.88	0.48	9.48	0.40	0.69	4.31E-02	NO	NO	NO	NO
AT1G15520	2.98	1.38	76.00	2.25	4.67	0.00E+00	NO	NO	NO	NO
AT1G15670	1.05	0.22	4.62	0.57	2.14	2.25E-06	NO	NO	NO	NO
AT1G15750	32.47	0.41	43.96	0.27	0.44	3.40E-04	NO	NO	NO	NO
AT1G15860	44.88	0.67	65.95	0.99	0.56	3.90E-04	NO	NO	NO	NO
AT1G15950	54.65	0.85	101.87	0.98	0.90	1.58E-07	NO	NO	NO	NO
AT1G16090	3.95	0.15	11.99	0.69	1.60	7.17E-08	NO	NO	NO	NO
AT1G16110	3.08	0.31	4.95	0.24	0.69	3.19E-02	NO	NO	NO	NO
AT1G16260	6.19	0.12	9.59	0.33	0.63	5.35E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G16270	4.23	0.10	6.10	0.12	0.53	1.20E-02	NO	NO	NO	NO
AT1G16420	0.97	0.49	15.44	0.82	3.99	0.00E+00	NO	NO	NO	NO
AT1G16670	7.14	0.48	18.33	0.03	1.36	4.88E-10	NO	NO	NO	NO
AT1G17070	4.09	0.12	6.35	0.32	0.63	1.60E-02	NO	NO	NO	NO
AT1G17330	11.06	0.33	25.29	0.11	1.19	1.14E-06	NO	NO	NO	NO
AT1G17340	3.04	0.15	6.18	0.08	1.02	5.23E-05	NO	NO	NO	NO
AT1G17360	11.15	0.32	19.29	0.47	0.79	1.67E-05	NO	NO	NO	NO
AT1G17380	1.70	0.37	8.78	0.49	2.37	1.59E-08	NO	YES	NO	NO
AT1G17430	5.31	0.35	8.83	0.33	0.73	4.24E-02	NO	NO	NO	NO
AT1G17440	20.43	0.13	30.36	0.70	0.57	5.83E-05	NO	NO	NO	NO
AT1G17550	16.07	0.24	25.23	0.71	0.65	6.09E-04	NO	NO	NO	NO
AT1G17600	2.33	0.15	6.35	0.33	1.45	5.11E-09	NO	NO	NO	NO
AT1G17620	13.67	0.24	27.82	0.84	1.03	3.46E-06	NO	NO	NO	NO
AT1G17745	80.82	1.50	152.73	3.89	0.92	1.97E-07	POSITIVE	NO	NO	NO
AT1G18210	116.67	1.37	156.06	1.16	0.42	5.02E-03	NO	NO	NO	NO
AT1G18260	15.39	0.12	26.32	0.77	0.77	4.14E-05	NO	NO	NO	NO
AT1G18300	2.23	0.25	8.91	0.64	2.00	7.29E-07	NO	NO	NO	NO
AT1G18370	6.90	0.14	9.69	0.45	0.49	3.28E-02	NO	NO	NO	NO
AT1G18390	6.18	0.48	9.98	0.89	0.69	1.91E-03	NO	NO	NO	NO
AT1G18570	5.38	0.46	22.46	1.50	2.06	0.00E+00	NO	YES	NO	NO
AT1G19000	40.63	0.56	55.29	1.61	0.44	8.55E-03	NO	YES	DOWN	NO
AT1G19020	5.24	1.34	73.67	1.47	3.81	0.00E+00	NO	NO	NO	NO
AT1G19180	34.47	0.30	78.94	1.22	1.20	1.08E-12	NO	YES	UP	NO
AT1G19250	0.17	0.13	13.41	0.91	6.32	0.00E+00	NO	NO	NO	NO
AT1G19300	2.84	0.24	8.02	0.41	1.50	4.47E-06	NO	NO	NO	NO
AT1G19350	59.81	1.67	73.70	1.89	0.30	2.43E-02	NO	YES	NO	NO
AT1G19360	22.54	0.18	33.82	0.35	0.59	2.66E-03	NO	NO	NO	NO
AT1G19380	1.98	0.39	17.52	1.11	3.14	2.59E-11	NO	NO	NO	NO
AT1G19490	6.86	0.10	11.08	0.25	0.69	7.79E-03	NO	YES	NO	NO

Table A3.4 (cont'd)

AT1G19660	27.48	0.52	36.45	0.59	0.41	4.42E-02	NO	NO	NO	NO
AT1G19670	49.97	0.99	66.34	1.59	0.41	4.80E-02	NO	NO	DOWN	NO
AT1G19835	15.87	0.60	25.56	0.42	0.69	2.22E-06	NO	NO	NO	NO
AT1G20100	47.59	0.42	100.82	0.42	1.08	1.91E-10	NO	NO	NO	NO
AT1G20330	40.01	0.51	65.93	1.50	0.72	8.88E-05	NO	NO	NO	NO
AT1G20510	38.11	0.92	63.52	1.86	0.74	1.86E-05	NO	NO	NO	NO
AT1G20630	32.92	0.69	45.92	0.44	0.48	1.52E-02	NO	NO	NO	NO
AT1G20780	11.23	0.37	18.02	0.15	0.68	4.74E-04	NO	NO	NO	NO
AT1G20920	8.87	0.23	14.13	0.41	0.67	4.75E-05	NO	NO	NO	NO
AT1G21100	1.06	0.23	4.42	0.16	2.06	2.15E-05	NO	NO	NO	NO
AT1G21240	3.98	0.42	21.69	0.53	2.45	0.00E+00	NO	NO	NO	NO
AT1G21250	56.92	2.59	182.09	1.04	1.68	0.00E+00	NO	NO	NO	DOWN
AT1G21310	88.27	2.04	618.14	18.55	2.81	0.00E+00	NO	NO	NO	NO
AT1G21370	7.89	0.07	15.55	0.40	0.98	8.84E-06	NO	NO	NO	NO
AT1G21450	10.27	0.18	15.51	0.19	0.59	5.75E-03	NO	YES	NO	NO
AT1G21460	53.05	0.87	86.84	0.84	0.71	8.57E-05	NO	NO	NO	UP
AT1G21520	15.09	1.33	51.82	2.07	1.78	1.26E-12	NO	NO	NO	NO
AT1G21580	8.55	0.12	11.65	0.53	0.45	2.74E-02	NO	NO	NO	NO
AT1G21900	82.80	0.14	146.27	2.08	0.82	2.83E-06	NO	NO	NO	NO
AT1G21910	0.78	0.17	2.36	0.26	1.61	1.55E-02	NO	YES	UP	NO
AT1G21920	15.55	0.25	22.03	1.01	0.50	2.15E-02	NO	NO	NO	NO
AT1G22280	28.43	0.95	66.30	1.41	1.22	1.04E-13	NO	NO	NO	NO
AT1G22330	1.28	0.16	2.81	0.51	1.14	4.96E-02	NO	NO	NO	NO
AT1G22410	42.80	0.86	99.73	1.71	1.22	1.20E-13	NO	NO	NO	NO
AT1G22500	2.04	0.17	7.62	0.43	1.90	2.07E-07	NO	NO	NO	NO
AT1G22650	3.06	0.11	8.12	0.39	1.41	9.89E-07	NO	NO	NO	NO
AT1G22750	56.76	0.59	78.97	0.62	0.48	4.23E-04	NO	NO	NO	NO
AT1G22930	33.99	0.27	62.71	0.30	0.88	3.73E-13	NO	NO	NO	NO
AT1G23000	0.81	0.10	2.13	0.43	1.40	2.17E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G23020	0.92	0.32	2.07	0.14	1.17	8.78E-03	NO	NO	DOWN	NO
AT1G23030	5.88	0.05	8.87	0.42	0.59	2.30E-02	NO	NO	NO	NO
AT1G23140	21.76	0.69	37.15	0.38	0.77	3.25E-03	NO	NO	NO	NO
AT1G23170	2.25	0.05	5.09	0.20	1.18	1.23E-04	NO	NO	NO	NO
AT1G23390	15.44	0.57	41.67	0.29	1.43	0.00E+00	NO	NO	NO	NO
AT1G23440	47.02	0.54	92.47	0.93	0.98	1.64E-08	NO	NO	NO	NO
AT1G23480	4.12	0.15	6.96	0.35	0.75	4.12E-03	NO	NO	DOWN	NO
AT1G23710	6.60	0.24	13.41	0.92	1.02	2.89E-04	NO	NO	DOWN	NO
AT1G24100	32.76	0.29	63.52	0.79	0.96	4.34E-08	NEGATIVE	NO	NO	NO
AT1G24140	1.07	0.30	10.30	0.42	3.27	1.85E-14		NO	NO	NO
AT1G24147	29.36	3.05	149.55	1.42	2.35	0.00E+00	NO	NO	NO	NO
AT1G24150	5.42	0.43	14.65	0.63	1.43	2.05E-11	NO	NO	NO	NO
AT1G24265	1.77	0.10	3.71	0.12	1.07	1.88E-02	NO	NO	NO	NO
AT1G24340	4.26	0.07	9.23	0.26	1.12	1.75E-06	NO	NO	NO	NO
AT1G24350	19.19	0.26	25.97	0.19	0.44	3.92E-02	NO	NO	NO	NO
AT1G24460	11.27	0.43	15.81	0.64	0.49	1.11E-03	NO	NO	NO	NO
AT1G24530	17.87	0.37	26.44	0.49	0.57	9.37E-03	NO	NO	NO	NO
AT1G25220	7.66	0.28	21.53	0.21	1.49	1.62E-10	NO	NO	NO	NO
AT1G25280	17.68	0.36	25.03	0.34	0.50	2.21E-03	NO	YES	NO	NO
AT1G25390	7.64	0.13	16.16	0.49	1.08	2.00E-07	NO	NO	NO	NO
AT1G25400	1.95	0.06	6.52	0.16	1.74	3.64E-06	NO	NO	DOWN	NO
AT1G25550	17.05	0.99	24.01	0.77	0.49	3.11E-02	NO	YES	NO	NO
AT1G26130	5.90	0.18	10.52	0.33	0.84	4.15E-06	NO	NO	NO	NO
AT1G26270	18.16	0.46	29.43	0.65	0.70	1.62E-04	NO	NO	NO	NO
AT1G26380	2.49	0.81	10.94	0.67	2.13	2.81E-13	NO	NO	NO	NO
AT1G26390	2.41	1.65	41.54	1.24	4.11	0.00E+00	NO	NO	NO	NO
AT1G26420	0.58	0.21	5.66	0.16	3.27	1.66E-11	NO	NO	NO	NO
AT1G27020	18.94	1.51	37.27	1.25	0.98	6.62E-07	NO	NO	NO	NO
AT1G27100	19.70	0.04	30.91	0.70	0.65	5.28E-04	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G27290	16.41	0.51	35.53	0.16	1.11	8.73E-08	NO	NO	NO	NO
AT1G27340	27.18	0.64	36.64	0.15	0.43	3.86E-02	NO	NO	NO	NO
AT1G27730	9.11	0.77	39.18	0.28	2.11	0.00E+00	NO	YES	UP	NO
AT1G27770	14.85	0.17	33.85	0.60	1.19	1.85E-12	NO	NO	NO	NO
AT1G27980	16.07	0.55	23.30	0.24	0.54	1.12E-02	NO	NO	NO	NO
AT1G28010	1.48	0.06	2.97	0.11	1.00	1.04E-03	NO	NO	NO	NO
AT1G28240	8.42	0.08	11.70	0.29	0.47	4.92E-02	NO	NO	NO	NO
AT1G28280	27.00	0.39	48.95	0.84	0.86	4.86E-07	NO	NO	NO	NO
AT1G28370	5.35	0.72	9.51	0.61	0.83	3.00E-02	NO	YES	NO	NO
AT1G28380	22.00	0.43	34.86	0.81	0.66	3.79E-04	NO	NO	NO	NO
AT1G28480	1.52	0.54	29.91	0.99	4.30	0.00E+00	NO	NO	NO	NO
AT1G28680	12.05	0.37	19.91	0.33	0.72	8.04E-04	NO	NO	NO	NO
AT1G29290	0.77	0.37	6.16	0.29	2.99	7.65E-05	NO	NO	NO	NO
AT1G29310	32.29	0.45	65.61	0.77	1.02	7.40E-09	NO	NO	NO	NO
AT1G29690	5.04	0.09	11.90	0.58	1.24	3.42E-08	NO	NO	NO	NO
AT1G30040	1.26	0.02	3.65	0.22	1.54	9.63E-04	NO	NO	NO	NO
AT1G30420	1.43	0.11	4.00	0.21	1.49	2.15E-08	NO	NO	NO	NO
AT1G30450	8.68	0.05	11.61	0.22	0.42	1.55E-02	NO	NO	NO	NO
AT1G30470	15.22	0.13	24.99	0.50	0.72	6.31E-05	NO	NO	NO	NO
AT1G30590	8.21	0.38	13.06	0.30	0.67	2.54E-03	NO	NO	NO	NO
AT1G30720	0.73	0.11	3.14	0.64	2.11	2.01E-05	NO	NO	NO	NO
AT1G30755	6.87	0.32	13.91	0.64	1.02	2.76E-06	NO	NO	NO	NO
AT1G30810	6.01	0.17	9.25	0.19	0.62	5.66E-03	NO	YES	NO	NO
AT1G30850	0.09	0.03	4.89	0.52	5.78	1.04E-09	NO	NO	NO	NO
AT1G31130	39.25	0.33	87.80	0.46	1.16	3.59E-12	NO	NO	NO	NO
AT1G31710	2.61	0.33	6.21	0.46	1.25	2.03E-05	NO	NO	NO	NO
AT1G32100	1.01	0.16	2.84	0.37	1.49	1.74E-02	NO	NO	NO	NO
AT1G32230	87.88	0.67	132.58	0.41	0.59	3.61E-07	NO	NO	NO	NO
AT1G32460	26.02	0.27	47.11	0.56	0.86	2.06E-04	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G32640	20.26	0.09	28.10	0.51	0.47	2.17E-02	NO	YES	NO	NO
AT1G32700	18.62	0.32	46.25	0.18	1.31	4.90E-12	NO	YES	NO	NO
AT1G32750	5.36	0.17	7.84	0.29	0.55	7.40E-03	NO	NO	NO	NO
AT1G32928	1.00	0.24	3.72	0.35	1.90	1.99E-02	NO	NO	NO	NO
AT1G32960	1.74	0.63	37.61	1.51	4.43	0.00E+00	NO	NO	NO	NO
AT1G33030	1.34	0.40	11.21	0.45	3.06	2.96E-13	NO	NO	NO	NO
AT1G33240	12.97	0.18	21.71	0.28	0.74	8.57E-05	NO	YES	NO	NO
AT1G33360	7.30	0.19	12.56	0.10	0.78	2.66E-04	NO	NO	NO	NO
AT1G33560	3.29	0.15	10.91	0.01	1.73	1.86E-14	NO	NO	NO	NO
AT1G33700	0.73	0.06	4.13	0.07	2.51	0.00E+00	NO	NO	NO	NO
AT1G33811	6.68	0.54	12.26	0.76	0.88	1.15E-03	NEGATIVE	NO	DOWN	NO
AT1G33950	2.01	0.24	30.88	4.65	3.94	0.00E+00	NO	NO	NO	NO
AT1G34180	3.81	0.38	8.79	0.32	1.21	3.51E-06	NO	YES	NO	NO
AT1G34190	52.30	0.43	73.09	0.84	0.48	1.35E-02	NO	YES	NO	NO
AT1G34300	7.59	0.13	13.47	0.62	0.83	8.23E-05	NO	NO	NO	NO
AT1G34420	0.63	0.11	3.69	0.27	2.55	1.52E-10	NO	NO	NO	NO
AT1G34750	9.81	0.83	29.35	0.28	1.58	0.00E+00	NO	NO	NO	NO
AT1G35210	3.84	1.01	12.01	0.74	1.64	8.16E-05	NO	NO	NO	NO
AT1G35220	7.02	0.19	9.60	0.23	0.45	4.77E-02	NO	NO	NO	NO
AT1G35230	1.60	0.55	17.74	0.67	3.47	5.86E-11	NO	NO	NO	NO
AT1G35350	4.59	0.11	9.90	0.54	1.11	3.67E-06	NO	NO	NO	NO
AT1G35580	22.66	0.00	42.71	0.33	0.91	1.04E-08	NO	NO	NO	NO
AT1G35670	8.89	0.08	18.79	0.20	1.08	2.74E-07	NO	NO	NO	NO
AT1G36160	12.12	0.30	18.05	0.09	0.57	1.36E-04	NO	NO	NO	NO
AT1G36370	15.59	0.66	33.71	1.58	1.11	3.37E-10	NO	NO	NO	NO
AT1G37130	102.67	0.90	191.77	0.18	0.90	3.64E-06	NO	NO	NO	NO
AT1G42990	68.33	0.49	197.75	2.59	1.53	0.00E+00	NO	YES	NO	NO
AT1G43710	39.21	0.26	53.39	1.68	0.45	2.62E-02	NO	NO	NO	NO
AT1G47128	173.90	1.83	346.49	7.44	0.99	2.91E-07	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G48090	8.71	0.41	12.79	0.45	0.56	3.71E-03	NO	NO	NO	NO
AT1G48120	2.51	0.07	3.67	0.08	0.55	4.96E-02	NO	NO	NO	NO
AT1G48320	9.72	0.23	28.33	0.63	1.54	4.12E-09	NO	NO	NO	NO
AT1G48370	5.46	0.04	8.18	0.58	0.58	2.46E-02	NO	NO	NO	NO
AT1G48490	5.76	0.07	7.78	0.21	0.43	4.41E-02	NO	NO	NO	NO
AT1G48605	5.47	0.33	13.66	0.45	1.32	1.68E-03	NO	NO	NO	NO
AT1G49050	9.11	0.74	24.43	0.35	1.42	2.16E-13	NO	NO	NO	NO
AT1G49450	8.47	0.44	13.78	0.73	0.70	2.05E-03	NO	NO	NO	NO
AT1G49750	126.99	0.31	175.49	1.10	0.47	1.98E-02	NO	NO	NO	NO
AT1G50140	11.04	0.25	14.70	0.17	0.41	1.36E-02	NO	NO	NO	NO
AT1G50740	15.79	0.28	33.18	0.47	1.07	8.24E-06	NO	NO	NO	NO
AT1G51350	7.22	0.04	10.72	0.22	0.57	1.61E-02	NO	NO	NO	NO
AT1G51620	1.15	0.29	3.71	0.50	1.69	2.59E-04	NO	NO	NO	NO
AT1G51660	15.55	0.25	25.44	0.53	0.71	4.93E-04	NO	NO	NO	NO
AT1G51680	31.66	1.52	63.96	0.51	1.01	4.40E-11	NO	NO	NO	NO
AT1G51760	19.06	0.33	50.79	0.50	1.41	0.00E+00	NO	NO	NO	NO
AT1G51790	0.24	0.10	3.31	0.54	3.78	4.95E-13	NO	NO	NO	NO
AT1G51805	26.84	0.67	51.90	0.54	0.95	0.00E+00	NO	NO	NO	NO
AT1G51860	0.74	0.20	2.19	0.19	1.56	5.49E-04	NO	NO	NO	NO
AT1G52030	5.95	1.21	8.86	1.22	0.58	2.09E-02	NO	NO	NO	NO
AT1G52340	12.27	0.37	20.38	0.79	0.73	2.28E-03	NO	NO	DOWN	NO
AT1G52780	11.10	0.21	19.04	0.63	0.78	2.75E-05	NO	NO	NO	NO
AT1G52800	0.44	0.11	2.88	0.36	2.70	3.95E-04	NO	NO	NO	NO
AT1G53920	5.93	0.30	10.91	0.13	0.88	5.11E-03	NO	NO	NO	NO
AT1G54115	9.90	0.20	15.87	0.17	0.68	1.73E-03	NO	NO	NO	NO
AT1G55210	120.70	3.07	227.86	4.26	0.92	6.42E-09	NO	NO	NO	NO
AT1G55310	3.99	0.18	6.76	0.08	0.76	4.53E-02	NO	NO	NO	NO
AT1G55325	5.48	0.11	8.70	0.27	0.67	5.48E-06	NO	NO	NO	NO
AT1G55350	10.50	0.27	13.42	0.33	0.35	4.31E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G55450	36.73	1.10	73.87	0.94	1.01	3.64E-14	NO	NO	DOWN	DOWN
AT1G55850	7.76	0.27	19.33	0.56	1.32	1.35E-11	NO	NO	DOWN	NO
AT1G55910	20.07	1.18	48.57	0.66	1.28	1.02E-11	NO	NO	NO	NO
AT1G55915	3.87	0.07	6.74	0.05	0.80	2.00E-02	NO	NO	NO	NO
AT1G55960	12.85	0.27	23.11	0.44	0.85	2.60E-05	NO	NO	NO	NO
AT1G56060	1.83	0.54	32.49	1.92	4.15	9.76E-13	NO	NO	NO	NO
AT1G56120	3.27	0.37	10.13	0.08	1.63	1.84E-13	NO	NO	NO	NO
AT1G56145	3.41	0.12	6.81	0.20	1.00	1.41E-05	NO	NO	NO	NO
AT1G56150	3.44	0.12	7.98	0.46	1.22	4.58E-02	NO	NO	NO	NO
AT1G56330	212.73	1.10	308.75	4.47	0.54	6.42E-03	NO	NO	NO	NO
AT1G56340	112.05	1.76	246.84	3.89	1.14	0.00E+00	NO	NO	NO	NO
AT1G56460	19.98	0.24	26.83	0.40	0.43	3.95E-02	NO	NO	NO	NO
AT1G56550	0.45	0.15	4.16	0.36	3.20	7.62E-09	NO	NO	NO	NO
AT1G56610	10.03	0.36	14.14	0.16	0.50	2.81E-02	NO	NO	NO	NO
AT1G57990	2.84	0.30	11.19	0.32	1.98	3.41E-10	NO	NO	NO	NO
AT1G58070	0.69	0.11	2.37	0.27	1.78	8.54E-03	NO	NO	NO	NO
AT1G58440	10.41	0.39	15.72	0.39	0.59	8.72E-03	NO	NO	NO	NO
AT1G59580	10.95	0.59	16.37	0.30	0.58	7.49E-03	NO	NO	DOWN	NO
AT1G59620	4.73	0.36	12.79	0.53	1.43	6.19E-12	NO	NO	NO	NO
AT1G59710	17.03	0.20	38.13	0.46	1.16	1.02E-10	NO	NO	NO	NO
AT1G59870	166.74	1.91	315.07	5.44	0.92	1.85E-04	NO	NO	NO	NO
AT1G59910	5.40	0.12	7.73	0.32	0.52	3.68E-02	NO	NO	NO	NO
AT1G60270	3.30	0.13	5.60	0.32	0.76	4.17E-02	NO	NO	NO	NO
AT1G60420	25.83	0.04	34.77	0.40	0.43	3.74E-02	POSITIVE	NO	NO	NO
AT1G60490	15.97	0.19	27.19	0.40	0.77	2.54E-05	NO	NO	NO	NO
AT1G60610	2.30	0.74	4.72	0.00	1.04	1.21E-02	NO	NO	NO	NO
AT1G61100	12.10	0.29	17.64	0.23	0.54	6.67E-03	NO	NO	NO	NO
AT1G61120	0.25	0.19	4.64	0.33	4.18	0.00E+00	NO	NO	NO	NO
AT1G61140	5.83	0.23	8.57	0.39	0.56	7.28E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G61250	28.81	0.30	61.10	0.71	1.08	5.79E-10	NO	NO	NO	NO
AT1G61260	2.53	0.43	6.91	0.29	1.45	6.66E-05	NO	NO	NO	NO
AT1G61360	3.85	0.06	7.39	0.30	0.94	4.48E-05	NO	NO	NO	NO
AT1G61370	1.39	0.10	3.25	0.02	1.22	1.30E-03	NO	NO	NO	NO
AT1G61470	0.63	0.18	2.08	0.14	1.72	4.40E-02	NO	NO	NO	NO
AT1G61560	1.43	0.00	4.06	0.13	1.50	4.77E-06	NO	NO	NO	NO
AT1G61660	19.45	0.28	26.03	0.27	0.42	2.19E-02	NO	NO	NO	NO
AT1G62300	4.75	0.23	19.07	0.78	2.01	0.00E+00	NO	YES	NO	NO
AT1G62360	0.48	0.07	2.15	0.44	2.16	1.00E-03	NO	YES	NO	NO
AT1G62422	4.33	0.63	18.22	0.37	2.07	4.14E-10	NO	NO	NO	NO
AT1G62500	3.68	0.10	13.31	1.12	1.86	4.37E-11	NO	NO	NO	DOWN
AT1G62630	7.04	0.10	12.85	0.04	0.87	2.86E-05	NO	NO	NO	NO
AT1G62660	1.62	0.23	6.23	0.26	1.94	3.90E-10	NO	NO	NO	NO
AT1G62790	60.31	1.34	85.19	1.05	0.50	2.54E-03	NO	NO	NO	NO
AT1G62840	0.58	0.23	3.26	0.09	2.48	4.30E-04	NO	NO	NO	NO
AT1G63010	22.58	0.13	31.26	0.33	0.47	4.00E-04	NO	NO	NO	NO
AT1G63100	5.42	0.31	8.32	0.29	0.62	2.62E-02	NO	YES	NO	NO
AT1G63220	16.24	0.35	35.16	0.87	1.11	4.98E-06	NO	NO	NO	NO
AT1G63670	19.21	0.46	24.89	0.76	0.37	1.11E-02	NO	NO	NO	NO
AT1G63720	1.93	0.26	6.51	0.04	1.75	2.40E-06	NO	NO	NO	NO
AT1G63750	1.68	0.04	3.49	0.57	1.06	1.03E-04	NO	NO	NO	NO
AT1G63840	8.39	0.69	45.30	1.16	2.43	0.00E+00	NO	NO	NO	NO
AT1G64050	8.49	0.37	12.13	0.37	0.51	2.04E-02	NO	NO	NO	NO
AT1G64060	3.02	0.06	7.80	0.13	1.37	1.62E-10	NO	NO	NO	NO
AT1G64280	17.27	0.52	31.50	0.31	0.87	8.28E-07	NO	NO	NO	NO
AT1G64400	15.23	0.75	28.03	1.06	0.88	1.79E-06	NO	NO	NO	NO
AT1G64460	6.29	0.44	16.73	1.02	1.41	3.87E-07	NO	NO	NO	NO
AT1G64470	6.26	0.26	16.82	0.60	1.43	4.80E-07	NO	NO	NO	NO
AT1G64490	38.19	0.24	56.43	1.51	0.56	2.26E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G64600	7.71	0.30	11.88	0.13	0.62	1.40E-02	NO	NO	NO	NO
AT1G64620	1.51	0.14	3.22	0.20	1.09	2.43E-02	NO	YES	NO	NO
AT1G65040	13.10	0.18	37.17	1.55	1.50	0.00E+00	NO	NO	NO	NO
AT1G65200	2.08	0.04	3.73	0.00	0.85	4.84E-03	NO	NO	NO	NO
AT1G65481	0.48	0.24	10.67	1.25	4.47	7.37E-07	NO	NO	NO	NO
AT1G65800	5.50	0.59	12.33	0.15	1.17	2.47E-08	NO	NO	NO	NO
AT1G65820	220.36	3.01	302.72	2.46	0.46	1.74E-02	POSITIVE	NO	NO	NO
AT1G65845	13.41	1.52	76.46	3.60	2.51	0.00E+00	NO	NO	NO	NO
AT1G66090	0.50	0.15	6.32	0.20	3.67	3.27E-12	NO	NO	NO	NO
AT1G66330	45.08	0.34	62.31	0.73	0.47	1.28E-03	NO	NO	NO	NO
AT1G66760	5.72	0.22	9.08	1.29	0.67	1.33E-02	NO	NO	NO	NO
AT1G66840	11.73	0.26	16.27	0.41	0.47	3.91E-02	NO	NO	NO	NO
AT1G66850	0.00	0.00	6.95	3.60	Inf	2.55E-04	NO	NO	NO	NO
AT1G66910	3.01	0.08	8.41	0.48	1.48	1.12E-07	NO	NO	NO	NO
AT1G66920	4.87	0.06	12.22	1.06	1.33	8.40E-10	NO	NO	NO	NO
AT1G67195	28.24	0.46	62.11	1.61	1.14	3.52E-07	NO	NO	NO	NO
AT1G67260	2.66	0.11	5.37	3.57	1.02	2.01E-02	NO	NO	NO	NO
AT1G67490	4.47	0.10	6.44	0.08	0.53	4.23E-02	NO	NO	NO	NO
AT1G67560	5.96	0.14	13.05	0.01	1.13	1.63E-08	NO	NO	NO	NO
AT1G67800	23.19	0.31	36.85	0.80	0.67	4.73E-07	NO	NO	NO	NO
AT1G67865	8.39	1.27	45.30	2.21	2.43	1.10E-10	NO	NO	NO	NO
AT1G67900	12.04	0.25	20.96	0.29	0.80	1.47E-07	NO	NO	NO	NO
AT1G67920	9.36	0.94	30.74	0.59	1.71	6.60E-08	NO	NO	NO	NO
AT1G68290	1.28	0.17	5.30	0.84	2.05	4.01E-05	NO	NO	NO	NO
AT1G68440	5.40	0.07	23.20	1.57	2.10	0.00E+00	NO	NO	NO	NO
AT1G68670	9.16	0.71	14.72	0.92	0.68	4.28E-03	NO	YES	NO	NO
AT1G68690	0.98	0.07	4.93	0.04	2.33	1.96E-12	NO	NO	NO	NO
AT1G68710	2.96	0.06	4.91	0.10	0.73	3.17E-03	NO	NO	NO	NO
AT1G68840	14.66	0.31	23.43	1.32	0.68	2.01E-03	NO	YES	NO	NO

Table A3.4 (cont'd)

AT1G68910	11.17	0.24	17.68	0.10	0.66	2.01E-04	NO	NO	NO	NO
AT1G69160	9.16	0.08	19.64	0.83	1.10	1.07E-06	NO	NO	NO	NO
AT1G69440	1.83	0.14	3.48	0.12	0.92	3.56E-03	NO	NO	NO	NO
AT1G69520	3.39	0.40	9.06	0.57	1.42	1.55E-03	NO	NO	NO	NO
AT1G69570	6.71	0.08	12.19	0.52	0.86	4.95E-04	NO	YES	UP	NO
AT1G69572	9.00	0.15	14.98	0.65	0.74	1.13E-02	NO	NO	NO	NO
AT1G69780	7.58	0.10	12.68	0.80	0.74	5.68E-03	NEGATIVE	YES	NO	NO
AT1G69840	24.97	0.20	60.13	0.88	1.27	0.00E+00		NO	NO	NO
AT1G69900	0.99	0.16	3.16	0.24	1.67	2.01E-03	NO	NO	NO	NO
AT1G69920	0.56	0.29	2.96	0.85	2.40	1.33E-03	NO	NO	NO	NO
AT1G70140	0.83	0.10	3.27	0.19	1.99	1.10E-06	NO	NO	NO	NO
AT1G70160	60.58	0.30	92.33	0.98	0.61	1.12E-03	NO	NO	NO	NO
AT1G70180	7.76	0.09	12.89	0.06	0.73	3.89E-04	NO	NO	NO	NO
AT1G70250	8.15	0.40	14.29	0.93	0.81	1.45E-04	NO	NO	NO	NO
AT1G70290	4.07	0.27	7.71	0.24	0.92	6.30E-05	NO	NO	NO	NO
AT1G70520	12.40	0.45	29.39	0.63	1.25	7.05E-12	NO	NO	NO	NO
AT1G70530	10.56	0.45	19.52	0.37	0.89	1.55E-05	NO	NO	NO	NO
AT1G70620	12.58	0.16	17.19	0.55	0.45	3.03E-03	NO	NO	NO	NO
AT1G70700	35.34	0.46	82.14	1.30	1.22	6.92E-13	NO	YES	NO	DOWN
AT1G70710	4.54	0.15	7.86	0.59	0.79	6.88E-03	NO	NO	NO	NO
AT1G70740	3.87	0.25	7.69	0.21	0.99	6.68E-04	NO	NO	NO	NO
AT1G70782	42.74	0.80	78.56	2.38	0.88	1.05E-06	NO	NO	NO	NO
AT1G71010	7.52	0.10	12.16	0.17	0.69	2.62E-04	NO	NO	NO	NO
AT1G71030	61.57	0.94	100.03	0.90	0.70	9.97E-05	NO	YES	NO	NO
AT1G71140	3.76	0.32	7.41	0.55	0.98	2.33E-03	NO	NO	NO	NO
AT1G71220	25.23	0.68	41.06	0.72	0.70	1.06E-08	NO	NO	NO	NO
AT1G71390	0.10	0.06	4.21	0.55	5.33	5.10E-12	NO	NO	NO	NO
AT1G71840	36.83	0.82	48.79	0.97	0.41	4.99E-02	NO	NO	NO	NO
AT1G71970	9.75	0.20	15.27	0.77	0.65	3.41E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G72150	54.88	0.86	73.07	1.05	0.41	4.25E-02	NO	NO	NO	NO
AT1G72240	0.96	0.16	5.14	0.61	2.42	1.78E-03	NO	NO	NO	NO
AT1G72280	7.89	0.83	29.39	1.71	1.90	0.00E+00	NO	NO	NO	NO
AT1G72450	28.86	0.47	64.54	0.66	1.16	5.44E-11	NO	YES	NO	NO
AT1G72680	26.54	0.59	75.99	1.46	1.52	0.00E+00	NO	NO	NO	NO
AT1G72700	10.69	0.29	25.14	0.12	1.23	2.96E-13	NO	NO	NO	NO
AT1G72770	15.99	0.16	23.55	0.56	0.56	7.45E-04	NO	NO	NO	NO
AT1G72900	6.70	0.90	41.84	0.76	2.64	0.00E+00	NO	NO	NO	NO
AT1G72910	3.63	0.55	16.88	0.69	2.22	0.00E+00	NO	NO	NO	NO
AT1G72920	1.32	0.14	8.66	0.56	2.71	2.64E-09	NO	NO	NO	NO
AT1G72930	70.95	0.77	146.22	1.42	1.04	1.22E-10	NO	NO	DOWN	NO
AT1G72940	3.54	0.20	13.44	0.82	1.92	5.51E-12	NO	NO	DOWN	NO
AT1G73030	72.42	0.75	100.93	1.05	0.48	1.74E-02	NO	NO	NO	NO
AT1G73540	4.91	0.31	11.44	0.66	1.22	5.28E-04	NO	NO	NO	NO
AT1G73650	54.96	0.23	71.97	0.31	0.39	3.96E-03	NO	NO	NO	NO
AT1G73740	5.97	0.13	9.95	0.53	0.74	7.91E-03	NO	NO	NO	NO
AT1G74020	19.30	0.78	35.90	1.25	0.89	5.42E-06	NO	NO	NO	NO
AT1G74030	5.05	0.28	8.05	0.59	0.67	2.16E-02	NO	NO	NO	NO
AT1G74080	0.25	0.09	2.77	0.19	3.49	2.01E-06	NO	YES	NO	NO
AT1G74090	54.16	0.86	75.83	1.79	0.49	1.64E-02	NEGATIVE	NO	NO	NO
AT1G74100	38.77	0.58	96.33	1.64	1.31	1.88E-14	NO	NO	NO	NO
AT1G74210	5.24	0.24	13.56	0.39	1.37	1.94E-07	NO	NO	NO	NO
AT1G74360	2.32	0.24	9.32	0.06	2.01	0.00E+00	NO	NO	NO	NO
AT1G75000	1.07	0.06	5.23	1.00	2.28	9.82E-05	NO	NO	NO	NO
AT1G75130	6.35	0.02	9.87	0.25	0.64	2.35E-02	NO	NO	NO	NO
AT1G75220	5.01	0.33	11.36	0.05	1.18	7.79E-07	NEGATIVE	NO	DOWN	NO
AT1G75280	20.83	0.30	29.41	1.56	0.50	2.52E-02	NO	NO	NO	NO
AT1G75380	52.08	0.71	88.51	0.67	0.76	1.86E-07	NO	NO	NO	NO
AT1G75410	13.39	0.10	28.60	0.55	1.10	2.74E-10	NO	YES	NO	NO

Table A3.4 (cont'd)

AT1G75440	8.63	0.68	17.36	0.77	1.01	1.87E-03	NO	NO	NO	NO
AT1G75960	0.53	0.05	2.22	0.17	2.06	2.01E-04	NO	NO	NO	NO
AT1G76040	4.58	0.25	20.15	0.60	2.14	0.00E+00	NO	NO	NO	NO
AT1G76070	1.97	0.24	4.88	0.16	1.31	4.27E-03	NO	NO	NO	NO
AT1G76090	4.80	0.04	8.15	0.42	0.76	2.16E-02	NO	NO	NO	NO
AT1G76240	4.44	0.23	16.52	0.32	1.90	1.63E-10	NO	NO	NO	NO
AT1G76320	6.97	0.09	9.43	0.07	0.43	4.35E-02	NO	NO	NO	NO
AT1G76390	6.04	0.23	12.52	0.15	1.05	9.33E-09	NO	NO	NO	NO
AT1G76640	0.18	0.08	3.26	0.73	4.14	5.42E-03	NO	NO	NO	NO
AT1G76690	3.56	0.41	6.15	0.54	0.79	4.44E-02	NO	NO	NO	NO
AT1G76760	4.20	0.15	9.80	0.42	1.22	3.11E-03	NO	NO	NO	NO
AT1G76890	3.09	0.23	6.13	0.25	0.99	1.32E-03	NO	YES	NO	NO
AT1G76892	5.86	0.29	9.19	0.42	0.65	3.30E-02	NO	NO	NO	NO
AT1G76980	14.44	0.86	48.59	1.24	1.75	0.00E+00	NO	NO	NO	NO
AT1G77300	7.22	0.36	9.53	0.16	0.40	2.41E-02	NO	YES	NO	NO
AT1G77320	3.01	0.16	4.38	0.12	0.54	4.64E-02	NO	NO	NO	NO
AT1G77380	1.91	0.30	6.72	0.24	1.81	4.58E-08	NO	NO	NO	NO
AT1G77420	7.25	0.16	11.79	0.21	0.70	8.11E-03	NO	NO	NO	NO
AT1G77460	2.07	0.63	3.52	0.17	0.76	1.49E-04	NO	NO	NO	NO
AT1G77770	9.44	0.18	17.39	0.27	0.88	2.18E-04	NO	NO	NO	NO
AT1G77800	10.18	0.45	13.22	0.34	0.38	2.97E-02	NO	YES	NO	NO
AT1G77810	13.90	0.36	28.27	1.20	1.02	7.96E-10	NO	NO	NO	NO
AT1G77990	3.42	0.33	6.50	0.50	0.92	1.15E-03	NO	NO	NO	NO
AT1G78000	0.66	0.07	3.96	0.50	2.57	2.84E-11	NO	NO	NO	NO
AT1G78150	71.75	0.57	102.22	2.06	0.51	1.29E-04	NO	NO	NO	NO
AT1G78200	35.34	0.41	49.03	0.88	0.47	4.41E-03	NO	NO	NO	NO
AT1G78230	11.55	0.74	19.67	1.06	0.77	7.53E-05	NO	NO	NO	NO
AT1G78280	6.25	0.04	10.27	0.31	0.72	6.13E-04	NO	YES	NO	NO
AT1G78460	38.86	1.85	53.12	1.03	0.45	4.50E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT1G78510	48.67	0.00	80.65	0.00	0.73	4.70E-09	NO	NO	NO	NO
AT1G78850	51.94	1.45	80.70	0.76	0.64	4.07E-04	NO	NO	NO	DOWN
AT1G78920	22.55	0.29	36.65	0.89	0.70	1.05E-04	NO	NO	NO	NO
AT1G79000	8.19	0.32	11.65	0.33	0.51	5.87E-03	NO	YES	NO	NO
AT1G79380	27.31	0.36	43.40	0.34	0.67	3.25E-04	NO	NO	NO	NO
AT1G79680	0.55	0.20	4.61	0.20	3.06	1.14E-12	NO	NO	NO	NO
AT1G79830	27.19	0.45	33.59	0.35	0.30	6.42E-03	NO	NO	NO	NO
AT1G79990	52.87	0.77	69.11	0.86	0.39	4.45E-03	NO	NO	NO	NO
AT1G80020	8.31	0.22	11.29	0.17	0.44	4.85E-02	NO	NO	NO	NO
AT1G80280	8.66	0.30	16.01	0.44	0.89	5.01E-06	NO	NO	NO	NO
AT1G80420	10.74	0.52	19.24	0.38	0.84	1.09E-05	NO	NO	NO	NO
AT1G80440	1.71	0.35	7.94	0.57	2.21	3.05E-09	NO	NO	NO	NO
AT1G80460	21.73	0.33	52.88	0.58	1.28	1.87E-14	NO	NO	NO	NO
AT1G80490	7.49	0.10	11.39	0.26	0.61	3.81E-04	NO	NO	NO	NO
AT1G80510	7.73	0.03	11.07	0.14	0.52	4.71E-02	NO	NO	NO	NO
AT1G80610	1.61	0.16	4.11	0.08	1.35	8.15E-03	NO	NO	NO	NO
AT1G80840	2.27	0.57	30.31	1.33	3.74	0.00E+00	NO	YES	UP	NO
AT1G80960	1.53	0.16	4.80	0.13	1.65	1.88E-05	NO	NO	NO	NO
AT2G01180	4.94	0.35	10.49	0.16	1.09	1.94E-05	NO	NO	NO	NO
AT2G01190	6.00	0.20	12.54	0.42	1.06	4.32E-07	NO	NO	NO	NO
AT2G01450	29.04	0.39	55.21	0.69	0.93	4.43E-10	NO	NO	NO	NO
AT2G01650	11.99	0.14	19.87	0.25	0.73	7.01E-04	NO	YES	NO	NO
AT2G01680	11.26	0.23	17.92	0.29	0.67	1.42E-03	NO	NO	NO	NO
AT2G01850	34.08	1.42	47.71	0.71	0.49	1.79E-02	NO	NO	NO	NO
AT2G01860	14.53	0.84	22.19	0.38	0.61	3.30E-03	NO	NO	NO	NO
AT2G02010	1.38	0.37	4.93	0.57	1.84	3.56E-06	NO	NO	NO	NO
AT2G02220	6.33	0.34	16.64	0.72	1.39	2.00E-13	NO	NO	NO	NO
AT2G02810	13.92	0.36	50.41	1.65	1.86	0.00E+00	NO	NO	NO	NO
AT2G02870	15.85	0.30	25.73	0.48	0.70	7.78E-05	NO	NO	NO	NO

Table A3.4 (cont'd)

AT2G02930	48.69	2.21	157.33	3.60	1.69	0.00E+00	NO	NO	NO	DOWN
AT2G02990	2.27	1.61	6.85	1.18	1.59	5.34E-04	NO	NO	NO	NO
AT2G03120	106.40	1.10	199.87	1.76	0.91	2.55E-07	NO	NO	NO	NO
AT2G03730	19.71	0.45	26.76	0.12	0.44	3.25E-02	NO	NO	NO	NO
AT2G04160	16.32	1.86	31.34	1.51	0.94	4.98E-08	NO	NO	NO	NO
AT2G04390	34.13	0.63	48.88	0.91	0.52	3.32E-02	NO	NO	NO	NO
AT2G04400	40.68	0.67	81.01	0.75	0.99	5.44E-09	NO	NO	NO	NO
AT2G04430	5.79	0.28	29.57	0.35	2.35	0.00E+00	NO	NO	NO	NO
AT2G04515	0.44	0.11	5.32	0.47	3.60	6.56E-07	NO	NO	NO	NO
AT2G04650	2.87	0.09	8.95	0.21	1.64	2.03E-07	NO	NO	NO	NO
AT2G04780	19.58	1.00	31.59	1.26	0.69	3.75E-04	NEGATIVE	NO	NO	NO
AT2G05320	8.68	0.39	15.12	0.46	0.80	1.11E-03	NO	NO	NO	NO
AT2G06050	23.06	0.72	36.66	0.69	0.67	1.98E-04	NO	NO	NO	NO
AT2G07180	37.77	0.33	47.71	0.71	0.34	3.17E-02	NO	NO	NO	NO
AT2G11520	11.05	0.19	18.43	0.14	0.74	4.21E-04	NO	NO	NO	NO
AT2G13790	3.27	0.29	8.92	0.18	1.45	1.88E-08	NO	NO	NO	NO
AT2G14740	7.51	0.18	10.52	0.13	0.49	4.79E-02	NO	NO	NO	NO
AT2G15042	15.65	0.63	31.61	1.29	1.01	5.08E-08	NO	NO	NO	NO
AT2G15580	15.41	0.26	32.54	0.19	1.08	7.58E-07	NO	NO	NO	NO
AT2G15760	1.37	0.39	3.12	0.15	1.18	3.55E-02	NO	NO	NO	NO
AT2G16280	49.01	1.20	70.84	1.54	0.53	6.05E-03	NO	NO	NO	NO
AT2G16430	33.55	0.38	65.81	0.97	0.97	1.91E-08	NO	NO	NO	NO
AT2G16500	104.26	0.71	240.53	1.43	1.21	1.21E-10	NO	NO	NO	NO
AT2G16890	19.67	0.30	37.07	1.18	0.91	1.41E-07	NO	NO	UP	NO
AT2G16900	8.40	0.13	39.31	0.93	2.23	0.00E+00	NO	NO	NO	NO
AT2G17130	39.13	0.37	57.83	0.96	0.56	1.27E-03	NO	NO	NO	NO
AT2G17220	9.88	0.23	21.25	0.22	1.10	1.16E-10	NO	NO	NO	NO
AT2G17230	28.33	0.49	42.44	0.94	0.58	2.76E-03	NO	NO	NO	NO
AT2G17290	16.44	0.46	26.81	0.18	0.71	1.62E-04	NO	NO	NO	NO

Table A3.4 (cont'd)

AT2G17520	15.79	0.33	24.02	0.55	0.60	1.58E-03	NO	NO	NO	NO
AT2G17560	121.90	2.40	148.76	2.93	0.29	3.52E-02	NO	YES	NO	NO
AT2G17710	20.34	1.29	32.91	0.45	0.69	2.63E-03	NO	NO	NO	NO
AT2G17720	46.21	0.46	81.83	0.77	0.82	3.46E-06	NO	NO	NO	NO
AT2G17730	12.62	0.15	25.63	0.10	1.02	5.76E-07	NO	NO	NO	NO
AT2G17740	4.19	1.84	17.39	1.17	2.05	3.15E-10	NO	NO	NO	NO
AT2G17790	15.64	0.27	22.37	0.23	0.52	9.85E-03	NO	NO	NO	NO
AT2G17930	6.19	0.18	9.17	0.49	0.57	2.71E-03	NO	NO	NO	NO
AT2G18160	22.86	0.67	34.43	0.11	0.59	5.15E-03	NO	YES	NO	NO
AT2G18280	41.81	0.47	54.36	0.50	0.38	2.68E-02	NO	YES	NO	NO
AT2G18660	38.78	2.20	135.92	2.62	1.81	0.00E+00	NO	NO	NO	NO
AT2G19130	8.32	0.62	22.48	0.52	1.43	1.85E-14	NO	NO	NO	NO
AT2G19190	2.31	0.85	19.87	1.54	3.11	0.00E+00	NO	NO	NO	NO
AT2G19710	3.49	0.06	9.62	0.45	1.46	7.95E-12	NO	NO	NO	NO
AT2G20010	7.25	0.08	13.07	0.35	0.85	1.89E-07	NO	NO	NO	NO
AT2G20142	7.52	0.58	33.62	1.95	2.16	0.00E+00	NO	NO	NO	NO
AT2G20570	88.36	1.33	122.65	2.08	0.47	1.43E-02	NO	YES	NO	NO
AT2G20610	60.51	0.92	84.20	1.01	0.48	8.66E-03	NEGATIVE	NO	NO	NO
AT2G20720	1.86	0.08	3.38	0.44	0.86	2.90E-02		NO	NO	NO
AT2G20870	4.49	0.46	9.65	5.37	1.10	1.88E-02	NO	NO	NO	NO
AT2G20960	17.39	0.09	29.38	0.70	0.76	4.15E-05	NO	NO	NO	NO
AT2G21140	23.67	0.49	36.05	2.73	0.61	2.90E-03	NO	NO	NO	NO
AT2G21470	14.57	0.17	21.23	0.29	0.54	1.77E-03	NO	NO	NO	NO
AT2G21500	8.20	0.36	13.05	0.91	0.67	2.25E-03	NO	NO	NO	NO
AT2G22300	18.06	0.38	30.80	0.45	0.77	4.77E-09	NO	YES	NO	NO
AT2G22420	10.29	0.16	15.50	0.59	0.59	3.39E-02	NO	NO	NO	NO
AT2G22480	10.26	0.22	15.30	0.40	0.58	1.19E-02	NO	NO	NO	NO
AT2G22500	21.82	1.09	57.11	0.55	1.39	0.00E+00	NO	NO	UP	NO
AT2G22560	1.65	0.09	4.05	0.42	1.29	3.86E-05	NO	NO	NO	NO

Table A3.4 (cont'd)

AT2G23140	6.61	0.10	10.60	0.06	0.68	6.13E-04	NO	NO	NO	NO
AT2G23170	1.31	0.05	15.67	0.62	3.58	0.00E+00	NO	NO	NO	NO
AT2G23200	10.21	0.46	17.81	0.50	0.80	3.77E-05	NO	NO	NO	NO
AT2G23270	0.14	0.04	3.59	0.20	4.65	2.64E-04	NO	NO	NO	NO
AT2G23290	1.71	0.12	9.32	0.34	2.44	2.49E-09	NO	YES	NO	NO
AT2G23320	11.48	0.54	31.47	0.92	1.45	2.07E-12	NO	YES	NO	NO
AT2G23380	7.91	0.41	10.83	0.24	0.45	4.97E-02	NO	YES	NO	NO
AT2G23450	7.44	0.16	14.47	0.10	0.96	9.48E-08	NO	NO	NO	NO
AT2G23680	5.31	0.92	12.60	0.58	1.25	5.01E-05	NO	NO	NO	NO
AT2G23770	1.67	0.24	4.82	0.31	1.53	4.75E-05	NO	NO	NO	NO
AT2G23810	42.29	0.61	122.74	1.44	1.54	0.00E+00	NO	NO	NO	NO
AT2G24200	198.37	1.31	287.46	2.07	0.54	1.91E-04	NO	NO	NO	NO
AT2G24360	62.41	0.52	90.15	1.31	0.53	4.82E-03	NO	NO	NO	NO
AT2G24550	1.52	0.05	5.44	0.15	1.84	1.13E-04	NO	NO	NO	NO
AT2G24590	55.96	0.72	77.22	1.19	0.46	2.44E-02	NO	NO	NO	NO
AT2G24600	2.04	0.89	7.22	0.00	1.82	9.47E-13	NO	NO	NO	NO
AT2G24650	1.79	0.10	3.81	0.11	1.09	3.43E-04	NO	YES	NO	NO
AT2G25000	8.43	0.34	20.64	0.19	1.29	5.47E-08	NO	YES	NO	NO
AT2G25110	48.87	0.82	116.75	1.84	1.26	2.48E-13	NO	NO	NO	NO
AT2G25735	2.79	0.19	7.09	0.26	1.34	1.36E-02	NO	NO	NO	NO
AT2G26190	18.32	1.15	45.86	0.61	1.32	5.36E-14	NO	NO	NO	NO
AT2G26300	18.57	0.31	30.66	0.36	0.72	1.12E-04	NO	NO	NO	NO
AT2G26400	6.87	1.35	52.83	1.90	2.94	0.00E+00	NO	NO	NO	NO
AT2G26440	18.44	1.48	35.79	1.12	0.96	3.65E-07	NO	NO	NO	NO
AT2G26530	4.99	0.24	20.76	0.76	2.06	0.00E+00	NO	NO	NO	DOWN
AT2G26540	19.68	0.60	27.38	0.16	0.48	3.22E-02	NO	NO	NO	NO
AT2G26800	15.31	0.47	23.16	0.36	0.60	5.81E-04	NO	NO	NO	NO
AT2G26980	8.98	0.38	20.12	0.74	1.16	2.43E-12	NEGATIVE	NO	NO	NO
AT2G27200	12.49	0.03	18.07	0.19	0.53	1.69E-02	POSITIVE	NO	NO	NO

Table A3.4 (cont'd)

AT2G27310	4.95	0.27	10.14	0.16	1.03	2.31E-03	NO	NO	NO	NO
AT2G27350	20.11	0.15	28.01	0.19	0.48	3.80E-04	NO	NO	NO	NO
AT2G27510	15.98	0.53	23.88	1.07	0.58	4.88E-02	NO	NO	NO	NO
AT2G27690	1.11	0.06	3.52	0.38	1.67	1.11E-04	NO	NO	NO	NO
AT2G28190	280.95	1.46	413.77	6.23	0.56	4.68E-03	NO	NO	NO	NO
AT2G28210	0.38	0.07	1.74	0.24	2.19	3.39E-02	NO	NO	DOWN	NO
AT2G28290	8.69	0.36	11.69	0.40	0.43	1.32E-02	NO	NO	NO	NO
AT2G28570	1.71	0.33	10.48	0.69	2.62	2.62E-09	NO	NO	NO	NO
AT2G28760	2.25	0.69	9.05	0.46	2.01	5.59E-11	NO	NO	NO	NO
AT2G28890	13.21	0.72	22.90	0.44	0.79	3.57E-05	NO	NO	NO	NO
AT2G28910	104.84	0.50	141.36	0.78	0.43	2.77E-02	NO	NO	NO	NO
AT2G28950	29.43	0.82	50.30	1.83	0.77	2.62E-05	NO	NO	NO	NO
AT2G29070	11.22	0.27	16.14	0.44	0.52	3.61E-02	NO	NO	NO	NO
AT2G29110	0.84	0.19	8.37	1.01	3.31	0.00E+00	NO	NO	NO	NO
AT2G29460	5.67	1.46	87.94	0.69	3.96	0.00E+00	NO	NO	NO	NO
AT2G29510	12.13	0.17	16.77	0.44	0.47	2.57E-02	NO	NO	NO	NO
AT2G29720	15.84	1.43	24.09	0.74	0.60	5.73E-03	NO	NO	NO	NO
AT2G29990	11.48	0.19	26.53	0.57	1.21	3.27E-10	NO	NO	NO	NO
AT2G30040	3.00	0.32	5.65	0.29	0.91	1.78E-02	NO	NO	NO	NO
AT2G30140	9.33	0.83	44.26	0.37	2.25	0.00E+00	NO	NO	NO	NO
AT2G30250	15.25	0.75	53.08	0.64	1.80	0.00E+00	NO	YES	NO	NO
AT2G30395	0.54	0.07	2.74	0.08	2.34	3.20E-03	NO	NO	NO	NO
AT2G30520	38.44	0.74	71.03	1.21	0.89	3.56E-08	NO	NO	NO	NO
AT2G30640	1.79	0.03	3.65	0.12	1.03	5.81E-03	NO	NO	NO	NO
AT2G30720	14.64	0.11	21.65	0.40	0.56	8.61E-03	NO	NO	NO	NO
AT2G30750	3.00	0.98	30.63	1.26	3.35	0.00E+00	NO	NO	NO	NO
AT2G30840	0.88	0.15	2.21	0.11	1.32	4.45E-02	NO	NO	NO	NO
AT2G31090	22.99	0.51	41.51	0.62	0.85	4.77E-03	NO	NO	NO	NO
AT2G31110	2.78	0.08	10.16	0.52	1.87	2.47E-08	NO	NO	NO	DOWN

Table A3.4 (cont'd)

AT2G31260	5.08	0.17	7.76	0.15	0.61	9.51E-03	NO	NO	NO	NO
AT2G31490	232.93	0.66	313.37	4.02	0.43	3.24E-02	NO	NO	NO	NO
AT2G31570	99.84	0.20	197.37	2.32	0.98	3.36E-09	NO	NO	NO	NO
AT2G31750	16.83	0.75	27.57	1.80	0.71	3.02E-04	NO	NO	NO	NO
AT2G31865	5.46	0.71	30.21	1.57	2.47	0.00E+00	NO	NO	NO	NO
AT2G31870	4.44	0.40	6.89	0.15	0.63	3.12E-02	NO	NO	NO	NO
AT2G31970	5.06	0.26	8.96	0.25	0.82	3.92E-05	NO	NO	NO	NO
AT2G32020	0.62	0.17	2.91	0.04	2.22	8.20E-03	NO	NO	NO	NO
AT2G32030	1.56	0.44	5.87	0.15	1.91	4.97E-04	NO	NO	NO	NO
AT2G32130	0.61	0.17	8.33	0.35	3.78	1.95E-07	NO	NO	NO	NO
AT2G32190	1.39	0.60	18.06	0.32	3.69	1.99E-11	NO	NO	NO	NO
AT2G32210	6.41	0.45	17.48	0.63	1.45	3.46E-04	NO	NO	NO	NO
AT2G32400	4.96	0.14	8.14	0.23	0.72	1.55E-03	NO	NO	NO	NO
AT2G32800	3.94	0.33	12.58	0.17	1.68	0.00E+00	NO	NO	NO	NO
AT2G33170	9.79	0.23	14.35	0.64	0.55	6.15E-03	NO	NO	NO	NO
AT2G33580	1.28	0.23	6.50	0.21	2.34	6.53E-12	NO	NO	NO	NO
AT2G33620	12.12	0.41	16.70	0.77	0.46	3.49E-02	NO	NO	NO	NO
AT2G33770	29.24	0.35	39.34	0.53	0.43	3.46E-02	NO	NO	NO	NO
AT2G34250	203.22	2.69	323.41	5.27	0.67	1.33E-03	NO	NO	NO	NO
AT2G34300	6.92	0.24	12.49	0.27	0.85	1.66E-05	NO	NO	NO	NO
AT2G34780	4.19	0.12	6.11	0.11	0.54	6.21E-03	NO	NO	NO	NO
AT2G34810	3.30	0.64	7.20	2.19	1.13	2.04E-04	NO	NO	NO	NO
AT2G34940	1.23	0.24	5.37	0.22	2.12	1.69E-08	NO	NO	NO	NO
AT2G34960	4.97	0.29	8.13	0.24	0.71	1.19E-02	NO	NO	NO	NO
AT2G35020	5.21	0.25	11.05	0.15	1.09	1.24E-05	NO	NO	NO	NO
AT2G35070	0.71	0.12	3.15	0.15	2.14	4.36E-03	NO	NO	NO	NO
AT2G35110	8.43	0.14	12.54	0.35	0.57	3.63E-03	NO	NO	NO	NO
AT2G35930	1.33	0.23	10.61	0.88	3.00	0.00E+00	NO	NO	NO	NO
AT2G36080	3.47	0.31	13.32	0.33	1.94	8.21E-11	NO	YES	NO	NO

Table A3.4 (cont'd)

AT2G36310	20.22	0.13	32.47	0.73	0.68	6.52E-04	NEGATIVE	NO	NO	NO
AT2G36580	24.04	0.56	55.34	0.70	1.20	1.32E-12	NO	NO	NO	NO
AT2G36630	29.47	0.40	40.75	0.84	0.47	1.79E-02	NO	NO	NO	NO
AT2G36792	0.26	0.02	0.97	0.18	1.89	1.86E-02	NO	NO	NO	NO
AT2G36800	3.45	0.29	22.96	1.13	2.73	0.00E+00	NO	NO	NO	NO
AT2G36880	168.28	2.63	282.61	3.02	0.75	9.87E-05	NO	NO	NO	NO
AT2G36895	43.63	0.15	64.98	0.72	0.57	5.69E-04	NO	NO	NO	NO
AT2G37080	14.74	0.26	22.79	0.58	0.63	1.34E-03	NO	NO	NO	NO
AT2G37100	2.29	0.03	5.33	0.26	1.22	1.39E-02	NO	NO	NO	NO
AT2G37110	68.58	0.67	151.89	1.06	1.15	3.63E-12	NO	NO	NO	NO
AT2G37150	3.93	0.14	7.07	0.41	0.85	6.30E-04	NO	NO	NO	NO
AT2G37970	131.20	0.47	303.93	4.83	1.21	2.17E-13	NO	NO	NO	NO
AT2G38250	0.72	0.22	9.21	0.70	3.69	1.24E-10	NO	YES	NO	NO
AT2G38290	27.76	0.60	49.21	1.17	0.83	3.34E-06	NO	NO	NO	NO
AT2G38310	6.32	0.34	17.45	0.18	1.47	7.35E-08	NO	NO	NO	DOWN
AT2G38360	8.11	0.34	13.92	0.38	0.78	1.26E-02	NO	NO	NO	NO
AT2G38410	6.05	0.11	9.24	0.09	0.61	1.23E-02	NO	NO	NO	NO
AT2G38470	17.35	1.37	77.52	1.54	2.16	0.00E+00	NO	YES	UP	NO
AT2G38750	5.31	0.15	11.45	1.66	1.11	9.62E-05	NEGATIVE	NO	NO	NO
AT2G38790	13.08	0.81	29.23	0.84	1.16	1.41E-07	NO	NO	NO	NO
AT2G38860	32.96	1.10	106.55	0.48	1.69	0.00E+00	NO	NO	NO	NO
AT2G38940	4.27	0.80	9.96	0.57	1.22	2.64E-06	NO	NO	NO	NO
AT2G39200	0.92	0.29	5.03	0.58	2.45	5.89E-09	NO	NO	NO	NO
AT2G39250	12.96	0.13	21.81	1.01	0.75	1.08E-03	NO	YES	NO	NO
AT2G39350	0.86	0.07	3.34	0.46	1.96	1.19E-06	NO	NO	NO	NO
AT2G39360	4.85	0.11	9.62	0.14	0.99	8.57E-06	NO	NO	NO	NO
AT2G39420	17.28	0.87	35.19	0.76	1.03	3.41E-07	NO	NO	NO	NO
AT2G39518	1.81	1.38	7.65	1.07	2.08	3.43E-04	NO	NO	NO	NO
AT2G39570	6.28	0.15	11.83	0.52	0.91	2.22E-04	NO	NO	DOWN	NO

Table A3.4 (cont'd)

AT2G39660	9.61	0.41	21.59	0.36	1.17	1.15E-08	NO	NO	NO	NO
AT2G39770	139.53	0.48	191.92	4.48	0.46	1.58E-02	NO	NO	NO	NO
AT2G39870	5.40	0.16	9.86	0.63	0.87	9.48E-04	NO	NO	NO	NO
AT2G40140	35.89	0.65	93.13	1.01	1.38	0.00E+00	NO	YES	UP	NO
AT2G40330	1.60	0.23	7.25	0.52	2.18	1.29E-06	NO	NO	NO	DOWN
AT2G40520	16.28	0.23	29.18	0.21	0.84	2.38E-08	NO	NO	NO	NO
AT2G40540	17.17	0.43	27.62	0.24	0.69	1.26E-06	NO	NO	NO	NO
AT2G40570	4.69	0.06	8.78	0.27	0.90	4.65E-04	NO	NO	NO	NO
AT2G40610	3.34	0.24	10.47	1.40	1.65	2.35E-06	NO	NO	DOWN	NO
AT2G40750	16.88	2.29	48.35	1.61	1.52	0.00E+00	NO	YES	NO	NO
AT2G40940	29.33	0.05	40.32	0.33	0.46	1.94E-02	NO	NO	NO	NO
AT2G41100	0.00	4.11	234.53	7.88	3.38	0.00E+00	NO	NO	DOWN	DOWN
AT2G41110	71.91	1.07	186.57	3.05	1.38	0.00E+00	NO	NO	NO	NO
AT2G41220	6.27	0.10	10.50	0.12	0.74	7.66E-05	NO	NO	NO	NO
AT2G41250	153.48	1.75	203.76	1.22	0.41	4.59E-02	NO	NO	NO	NO
AT2G41410	42.98	1.35	137.98	1.80	1.68	0.00E+00	NO	NO	NO	NO
AT2G41430	342.70	7.21	447.09	2.18	0.38	4.67E-02	NO	NO	NO	NO
AT2G41700	6.15	0.14	11.99	0.90	0.96	1.43E-07	NO	NO	NO	NO
AT2G41730	2.37	0.31	26.37	2.36	3.48	5.57E-11	NO	NO	NO	NO
AT2G41835	4.00	0.29	9.50	0.08	1.25	1.56E-04	NO	YES	NO	NO
AT2G41850	1.65	1.03	7.44	2.86	2.18	8.32E-09	NO	NO	NO	NO
AT2G41880	6.02	0.25	10.35	0.05	0.78	4.89E-03	NO	NO	NO	NO
AT2G42330	2.79	0.24	4.24	0.31	0.61	3.73E-02	NO	NO	NO	NO
AT2G42360	2.46	0.38	12.73	0.92	2.37	6.41E-10	NO	NO	NO	NO
AT2G42380	1.45	0.10	3.88	0.48	1.42	8.73E-04	NO	YES	NO	NO
AT2G42490	16.22	0.19	23.33	0.43	0.52	7.23E-03	NO	NO	NO	NO
AT2G42610	20.53	0.54	30.95	1.34	0.59	4.23E-03	NO	NO	NO	NO
AT2G42840	105.50	1.30	143.75	5.27	0.45	2.31E-02	NO	NO	NO	NO
AT2G42890	11.08	0.25	15.79	0.23	0.51	1.52E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT2G43000	2.28	0.58	15.57	0.64	2.77	9.57E-12	NO	YES	NO	NO
AT2G43140	0.79	0.17	3.14	0.76	1.98	1.08E-03	NO	YES	NO	NO
AT2G43160	47.80	0.40	64.40	0.29	0.43	1.34E-04	NO	NO	NO	NO
AT2G43210	20.69	0.37	28.41	0.48	0.46	1.80E-02	NO	NO	NO	NO
AT2G43320	7.50	0.41	22.48	0.34	1.58	0.00E+00	NO	NO	NO	NO
AT2G43680	44.11	0.35	67.27	0.24	0.61	6.84E-07	NO	NO	NO	NO
AT2G44280	1.46	0.03	4.12	0.03	1.49	2.13E-05	NO	NO	NO	NO
AT2G44370	0.79	0.29	3.02	0.61	1.93	2.35E-03	NO	NO	NO	NO
AT2G44500	2.01	0.28	6.55	0.30	1.70	2.71E-09	NO	NO	NO	NO
AT2G45000	7.18	0.26	10.87	0.29	0.60	1.09E-02	NO	NO	NO	NO
AT2G45010	19.57	0.57	30.58	0.44	0.64	1.76E-03	NO	NO	NO	NO
AT2G45030	7.17	0.23	10.29	0.17	0.52	3.00E-02	NO	NO	NO	NO
AT2G45070	330.68	4.89	498.26	13.95	0.59	4.39E-07	NO	NO	NO	NO
AT2G45170	18.47	0.57	38.79	0.25	1.07	7.81E-06	NO	NO	NO	NO
AT2G45280	4.72	0.13	8.68	0.36	0.88	3.15E-03	NO	NO	NO	NO
AT2G45290	10.39	0.51	14.20	0.47	0.45	4.29E-02	NO	NO	NO	NO
AT2G45540	9.31	0.37	14.32	0.32	0.62	1.27E-05	NO	NO	NO	NO
AT2G45590	14.98	0.49	21.73	0.37	0.54	8.51E-03	NO	NO	NO	NO
AT2G45680	3.72	0.19	9.38	0.38	1.33	2.89E-06	NO	YES	NO	NO
AT2G45700	7.18	0.13	10.18	0.24	0.50	3.97E-02	NO	NO	NO	NO
AT2G45790	126.47	0.65	199.15	3.30	0.66	4.11E-04	NO	NO	NO	NO
AT2G45910	7.67	0.10	11.31	0.32	0.56	1.02E-02	NO	NO	NO	NO
AT2G45960	239.62	19.48	352.32	26.16	0.56	4.15E-03	NO	NO	NO	NO
AT2G46020	10.43	0.26	14.55	0.34	0.48	4.60E-04	NO	NO	NO	NO
AT2G46150	1.21	0.17	7.43	0.22	2.61	1.61E-07	NO	NO	NO	NO
AT2G46270	1.97	0.26	10.45	0.08	2.41	0.00E+00	NO	YES	NO	NO
AT2G46330	57.26	0.24	101.27	1.02	0.82	6.51E-06	NO	NO	NO	NO
AT2G46420	9.51	0.30	17.94	0.15	0.92	7.04E-06	NO	NO	NO	NO
AT2G46500	15.49	0.56	31.20	0.76	1.01	1.48E-10	NO	NO	NO	NO

Table A3.4 (cont'd)

AT2G46600	146.49	1.59	303.78	2.59	1.05	1.08E-09	NO	NO	NO	UP
AT2G47130	5.18	1.71	27.68	0.47	2.42	0.00E+00	NO	NO	NO	NO
AT2G47470	137.45	1.77	319.71	3.25	1.22	2.78E-11	NO	NO	NO	NO
AT2G47730	154.03	1.69	290.59	0.72	0.92	4.69E-07	NO	NO	NO	NO
AT2G47800	5.79	0.21	14.09	0.09	1.28	4.04E-13	NO	NO	NO	NO
AT2G48030	7.45	0.41	15.60	0.41	1.07	1.67E-06	NO	NO	NO	NO
AT3G01290	32.29	2.41	182.61	1.90	2.50	0.00E+00	NO	NO	DOWN	NO
AT3G01650	2.23	0.16	5.06	0.05	1.18	1.12E-03	NO	NO	NO	NO
AT3G01720	10.57	0.05	19.99	0.52	0.92	6.64E-07	NO	NO	NO	NO
AT3G01830	2.56	1.17	31.33	1.06	3.61	0.00E+00	NO	NO	NO	NO
AT3G02040	4.01	0.39	6.98	0.61	0.80	1.76E-02	NO	NO	NO	NO
AT3G02050	12.32	0.12	22.38	0.14	0.86	2.00E-06	NO	NO	NO	NO
AT3G02170	3.47	0.24	11.01	0.13	1.67	0.00E+00	NO	NO	NO	NO
AT3G02260	11.00	0.37	15.17	0.43	0.46	2.22E-02	NO	NO	NO	NO
AT3G02770	53.96	0.57	86.76	0.83	0.69	3.71E-04	NO	NO	NO	NO
AT3G02840	0.42	0.18	4.45	0.22	3.41	1.97E-08	NO	NO	NO	NO
AT3G02880	7.25	0.42	11.85	0.44	0.71	1.48E-03	NO	NO	NO	NO
AT3G02885	0.28	0.12	1.51	0.35	2.44	3.43E-02	NO	NO	NO	NO
AT3G03440	13.69	0.50	22.16	0.35	0.69	1.37E-03	NO	NO	NO	NO
AT3G03640	25.52	1.56	39.14	1.96	0.62	1.54E-03	NO	NO	UP	NO
AT3G03680	2.47	0.04	3.78	0.13	0.61	4.62E-02	NO	NO	NO	NO
AT3G03990	30.28	0.89	45.27	0.51	0.58	4.72E-03	NO	NO	NO	UP
AT3G04010	4.60	0.27	7.09	0.45	0.63	4.98E-02	NO	NO	UP	NO
AT3G04110	12.57	0.21	17.39	0.36	0.47	2.98E-02	NO	NO	NO	NO
AT3G04210	64.89	2.90	111.38	0.45	0.78	8.16E-06	NO	NO	DOWN	NO
AT3G04240	41.06	0.54	62.70	0.47	0.61	1.22E-03	POSITIVE	NO	NO	NO
AT3G04300	0.16	0.02	1.37	0.20	3.10	1.44E-02	NO	NO	NO	NO
AT3G04580	11.41	0.11	18.27	0.39	0.68	7.41E-06	NO	NO	NO	NO
AT3G04640	10.01	1.25	24.97	0.98	1.32	3.40E-05	NO	NO	UP	NO

Table A3.4 (cont'd)

AT3G04670	14.68	0.01	25.52	0.63	0.80	2.98E-05	NO	YES	NO	NO
AT3G04720	99.75	5.32	154.11	2.36	0.63	4.27E-04	NO	NO	NO	NO
AT3G05165	2.39	0.12	6.25	0.45	1.39	3.09E-05	NO	NO	NO	NO
AT3G05320	1.46	0.14	4.82	0.48	1.72	7.04E-05	NO	NO	NO	NO
AT3G05360	0.77	0.02	2.28	0.08	1.57	4.99E-04	NO	NO	NO	NO
AT3G05500	85.31	0.90	125.89	1.55	0.56	3.79E-03	NO	NO	NO	NO
AT3G05680	5.52	0.16	7.39	0.22	0.42	4.55E-02	NO	NO	NO	NO
AT3G05830	6.18	0.22	12.00	0.25	0.96	6.76E-04	NO	NO	NO	NO
AT3G05970	34.76	0.35	51.79	0.20	0.58	2.14E-03	NO	NO	NO	NO
AT3G06070	4.91	0.54	9.65	0.98	0.98	2.49E-02	NO	NO	NO	NO
AT3G06330	6.38	0.30	9.32	0.37	0.55	4.73E-02	NO	NO	NO	NO
AT3G06500	25.30	0.45	51.47	0.36	1.02	1.58E-09	NO	NO	NO	DOWN
AT3G06890	3.56	0.27	7.96	0.29	1.16	2.09E-02	NO	NO	NO	NO
AT3G07010	9.44	0.32	16.92	1.17	0.84	1.54E-04	NO	NO	NO	NO
AT3G07270	23.67	0.13	39.14	0.41	0.73	1.31E-06	NO	NO	NO	NO
AT3G07340	1.55	0.07	4.29	0.03	1.47	1.87E-04	NO	YES	NO	NO
AT3G07525	13.45	0.84	22.11	1.20	0.72	4.13E-03	NO	NO	NO	NO
AT3G07690	13.59	0.12	20.16	0.63	0.57	1.01E-02	NO	NO	NO	NO
AT3G07700	37.50	0.22	69.92	0.84	0.90	1.37E-13	NO	NO	NO	NO
AT3G07720	44.16	0.48	105.81	1.02	1.26	7.04E-14	NO	NO	NO	NO
AT3G08650	30.41	0.22	42.23	0.15	0.47	1.40E-03	NO	NO	NO	NO
AT3G08670	7.73	0.16	11.36	0.31	0.56	2.37E-02	NO	NO	NO	NO
AT3G08760	10.23	0.29	22.01	0.20	1.11	8.30E-09	NO	NO	NO	NO
AT3G08850	8.30	0.25	11.17	0.25	0.43	4.39E-02	NO	NO	NO	NO
AT3G09010	5.01	0.59	9.78	0.17	0.96	3.96E-04	NO	NO	NO	NO
AT3G09020	1.63	0.34	6.68	0.17	2.03	8.38E-07	NO	NO	NO	NO
AT3G09490	1.31	0.26	4.88	0.25	1.90	2.06E-04	NO	NO	NO	NO
AT3G09520	1.75	0.56	6.04	0.67	1.79	2.20E-07	NO	NO	NO	NO
AT3G09560	7.92	2.23	12.02	0.30	0.60	1.76E-04	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G09830	15.28	0.22	30.38	0.49	0.99	2.34E-09	NO	NO	NO	NO
AT3G09920	12.33	0.11	20.52	0.48	0.73	9.98E-08	NO	NO	NO	NO
AT3G10010	4.55	0.23	6.61	0.18	0.54	2.27E-02	NO	NO	NO	NO
AT3G10070	9.75	0.47	13.97	0.09	0.52	2.80E-02	NO	NO	NO	NO
AT3G10130	25.62	0.26	39.01	0.27	0.61	4.34E-03	NO	NO	NO	NO
AT3G10250	6.05	0.20	9.84	0.17	0.70	1.45E-02	NO	NO	NO	NO
AT3G10420	48.35	0.45	84.02	0.54	0.80	1.54E-06	NO	NO	NO	NO
AT3G10500	6.25	0.21	14.72	0.20	1.24	1.14E-08	NO	YES	NO	NO
AT3G10640	8.42	0.18	18.34	0.29	1.12	1.08E-05	NO	NO	NO	NO
AT3G10720	6.64	0.28	11.78	0.43	0.83	2.37E-04	NO	NO	NO	NO
AT3G10740	9.91	0.24	15.48	0.05	0.64	2.03E-03	NO	NO	NO	NO
AT3G10930	0.39	0.19	3.30	0.29	3.10	8.10E-04	NO	NO	UP	NO
AT3G10985	11.41	0.41	37.33	0.22	1.71	8.76E-14	NO	NO	NO	NO
AT3G11090	8.88	0.91	18.11	0.45	1.03	6.92E-03	NO	YES	DOWN	NO
AT3G11330	8.64	0.17	12.79	0.35	0.57	1.84E-02	NO	NO	NO	NO
AT3G11700	27.19	0.55	36.20	0.79	0.41	4.78E-02	NO	NO	NO	NO
AT3G11820	18.56	0.57	45.73	0.44	1.30	0.00E+00	NO	NO	NO	NO
AT3G11840	4.31	0.79	11.25	0.17	1.38	3.03E-07	NO	NO	NO	NO
AT3G12145	5.67	0.37	10.74	1.26	0.92	1.10E-03	NO	NO	NO	NO
AT3G12150	3.34	0.12	5.59	0.07	0.74	4.08E-02	NO	NO	NO	NO
AT3G12360	24.84	0.54	38.71	0.48	0.64	6.05E-04	NO	NO	NO	NO
AT3G12520	1.49	0.10	4.34	0.60	1.54	1.00E-05	NO	NO	NO	NO
AT3G12600	15.38	0.19	26.10	0.45	0.76	5.36E-04	NO	NO	NO	NO
AT3G12610	24.66	0.76	34.85	1.91	0.50	1.71E-02	NEGATIVE	NO	NO	NO
AT3G12710	1.99	0.27	5.32	0.27	1.42	7.53E-04	NEGATIVE	NO	NO	NO
AT3G12740	42.66	0.57	91.40	0.77	1.10	3.60E-11	NO	NO	NO	NO
AT3G12830	3.60	0.02	15.41	0.43	2.10	1.05E-09	NO	NO	NO	NO
AT3G13060	25.20	0.12	33.87	0.19	0.43	3.60E-02	NO	NO	NO	NO
AT3G13061	7.24	0.06	14.99	0.14	1.05	5.87E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G13062	32.12	0.55	47.36	0.28	0.56	2.58E-03	NO	NO	NO	NO
AT3G13080	14.02	0.14	57.64	0.39	2.04	0.00E+00	NO	NO	NO	NO
AT3G13430	9.49	0.26	13.46	0.64	0.50	4.90E-02	NO	NO	NO	NO
AT3G13437	0.55	0.25	2.60	0.04	2.26	4.92E-02	NO	NO	NO	NO
AT3G13445	8.14	0.22	12.02	0.19	0.56	4.32E-02	NO	NO	NO	NO
AT3G13670	41.70	0.56	55.39	0.32	0.41	4.26E-02	NO	NO	NO	NO
AT3G13720	18.66	0.87	27.56	0.99	0.56	2.33E-02	NO	NO	NO	NO
AT3G13810	23.49	0.60	33.83	0.38	0.53	6.62E-04	NO	YES	NO	NO
AT3G13910	34.83	0.91	63.83	1.13	0.87	1.33E-04	NO	NO	NO	NO
AT3G14050	14.65	0.54	24.76	0.17	0.76	5.00E-05	NO	NO	NO	NO
AT3G14060	2.63	0.75	9.81	0.47	1.90	8.58E-04	POSITIVE	NO	NO	NO
AT3G14067	39.58	0.89	65.62	0.58	0.73	3.83E-05	NO	NO	NO	NO
AT3G14090	5.31	0.06	8.95	0.17	0.75	2.48E-03	NO	NO	NO	NO
AT3G14230	97.59	0.95	132.75	0.65	0.44	7.01E-03	NO	YES	NO	NO
AT3G14270	3.95	0.17	6.82	0.23	0.79	7.26E-05	NO	NO	NO	NO
AT3G14470	2.08	0.09	5.85	0.33	1.49	7.81E-09	NO	NO	NO	NO
AT3G14610	1.52	0.08	4.09	0.08	1.42	4.78E-04	NO	NO	NO	NO
AT3G14770	7.69	0.11	12.28	0.05	0.68	7.57E-03	NO	NO	NO	NO
AT3G14790	16.84	0.46	30.71	0.23	0.87	1.51E-06	NO	NO	NO	NO
AT3G14840	16.57	0.87	45.40	0.44	1.45	0.00E+00	NO	NO	NO	DOWN
AT3G14920	7.72	0.06	11.54	0.18	0.58	1.81E-02	NO	NO	NO	NO
AT3G15353	1634.45	24.92	2244.20	9.07	0.46	3.74E-02	NO	NO	NO	NO
AT3G15400	2.19	0.18	7.90	3.71	1.85	1.92E-07	NO	NO	NO	NO
AT3G15430	24.66	0.22	37.49	0.60	0.60	1.81E-05	POSITIVE	NO	NO	NO
AT3G15540	1.88	0.12	8.58	0.21	2.19	6.78E-07	NEGATIVE	YES	NO	NO
AT3G15650	1.51	0.17	6.15	0.28	2.03	1.71E-05	NO	NO	NO	NO
AT3G15760	0.73	0.23	3.15	0.07	2.10	4.65E-03	NO	NO	DOWN	NO
AT3G15770	8.41	0.53	21.36	0.17	1.34	1.53E-06	NO	NO	NO	NO
AT3G15790	30.43	0.68	45.80	1.02	0.59	3.14E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G15810	36.24	1.14	77.21	0.71	1.09	1.01E-09	NO	NO	NO	NO
AT3G15820	4.77	0.33	17.65	0.13	1.89	9.76E-12	NO	NO	NO	NO
AT3G15880	14.80	0.28	18.71	0.33	0.34	2.85E-02	NO	NO	NO	NO
AT3G15980	22.16	0.48	47.71	2.17	1.11	0.00E+00	NO	NO	NO	NO
AT3G16230	12.74	0.35	17.54	0.10	0.46	2.29E-02	NO	NO	DOWN	NO
AT3G16370	115.85	0.52	168.84	5.24	0.54	4.27E-03	NO	NO	NO	NO
AT3G16460	2.03	0.22	5.50	0.53	1.44	3.29E-07	NO	NO	NO	NO
AT3G16720	2.85	0.32	11.68	0.56	2.03	1.26E-10	NO	NO	NO	NO
AT3G16857	8.09	0.24	12.85	0.05	0.67	3.17E-04	NO	YES	NO	NO
AT3G16910	12.17	0.25	19.30	0.57	0.67	1.56E-03	NO	NO	DOWN	NO
AT3G16950	3.72	0.15	5.87	0.24	0.66	2.78E-02	NO	NO	NO	NO
AT3G17000	26.38	0.06	39.01	0.50	0.56	6.32E-03	NO	NO	NO	NO
AT3G17050	1.54	0.19	5.40	0.22	1.81	1.02E-06	NO	NO	NO	NO
AT3G17205	12.33	0.20	18.11	0.24	0.55	1.01E-04	NO	NO	NO	NO
AT3G17240	93.11	1.26	141.89	1.73	0.61	1.08E-03	NO	NO	NO	NO
AT3G17420	5.33	0.11	11.65	0.48	1.13	2.52E-06	NO	NO	NO	NO
AT3G17609	75.03	1.66	147.59	0.82	0.98	6.47E-13	NO	YES	NO	NO
AT3G17700	7.52	0.29	15.78	0.60	1.07	7.74E-08	NO	NO	NO	NO
AT3G17770	10.87	0.63	17.19	0.31	0.66	1.28E-03	NO	NO	NO	NO
AT3G17800	81.99	1.04	159.86	1.29	0.96	1.14E-08	NO	NO	NO	NO
AT3G17820	43.96	0.43	64.82	1.68	0.56	3.66E-03	NO	NO	NO	NO
AT3G18280	46.02	1.13	104.69	2.34	1.19	1.21E-09	NEGATIVE	NO	NO	NO
AT3G18773	0.39	0.05	1.24	0.03	1.66	4.24E-02	NO	NO	NO	NO
AT3G18930	3.10	0.05	6.23	0.17	1.01	5.63E-04	NO	NO	NO	NO
AT3G19010	63.75	2.07	169.09	1.70	1.41	0.00E+00	NO	NO	NO	NO
AT3G19260	5.64	0.14	20.03	0.15	1.83	1.92E-14	NO	NO	NO	NO
AT3G19870	8.64	0.02	12.89	0.35	0.58	4.61E-03	NO	NO	NO	NO
AT3G19970	4.11	0.20	9.01	0.01	1.13	2.77E-05	NO	NO	NO	NO
AT3G20250	14.39	0.39	24.04	0.25	0.74	5.45E-05	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G20280	0.88	0.02	3.37	0.06	1.93	7.54E-08	NO	YES	NO	NO
AT3G20600	7.82	0.93	20.46	0.54	1.39	9.92E-07	NO	NO	NO	NO
AT3G20830	1.72	0.15	4.17	0.04	1.28	7.46E-03	NO	NO	NO	NO
AT3G21080	0.83	0.03	6.53	0.13	2.98	1.61E-08	NO	NO	NO	NO
AT3G21200	53.96	0.97	73.14	1.81	0.44	3.15E-02	NEGATIVE	NO	NO	NO
AT3G21230	1.82	0.18	7.79	0.49	2.10	6.97E-11	NO	NO	NO	NO
AT3G21520	0.31	0.14	5.02	0.56	4.00	4.86E-07	NO	NO	NO	NO
AT3G21560	73.70	0.41	106.05	0.28	0.52	5.21E-03	NO	NO	NO	NO
AT3G21630	11.90	0.25	18.51	0.25	0.64	2.38E-03	NO	NO	NO	NO
AT3G21780	2.70	0.48	9.07	0.26	1.75	6.53E-08	NO	NO	NO	NO
AT3G21810	6.90	0.26	13.47	0.17	0.96	2.37E-05	NO	YES	NO	NO
AT3G22060	18.17	2.02	87.21	3.19	2.26	0.00E+00	NO	NO	NO	NO
AT3G22160	6.19	0.13	20.09	0.76	1.70	9.72E-11	NO	NO	NO	NO
AT3G22170	16.38	0.07	23.31	0.38	0.51	7.79E-04	NO	NO	NO	NO
AT3G22190	28.77	0.32	41.33	0.62	0.52	4.79E-04	NO	NO	NO	NO
AT3G22400	1.01	0.08	6.33	0.09	2.65	0.00E+00	NO	NO	NO	NO
AT3G22420	9.98	0.32	26.13	0.27	1.39	0.00E+00	NO	NO	NO	NO
AT3G22460	0.99	0.14	2.66	0.17	1.43	2.46E-02	NO	NO	NO	NO
AT3G22550	9.48	0.41	14.25	1.06	0.59	2.02E-02	NEGATIVE	NO	NO	NO
AT3G22620	1.20	0.15	10.61	2.92	3.14	3.14E-10	NO	NO	NO	NO
AT3G22890	222.31	0.52	362.29	1.47	0.70	4.46E-04	NO	NO	NO	NO
AT3G23010	0.22	0.05	0.82	0.09	1.91	2.77E-02	NO	NO	NO	NO
AT3G23030	42.97	0.75	73.77	1.58	0.78	3.57E-05	NO	YES	NO	NO
AT3G23050	33.27	0.28	52.85	2.17	0.67	4.29E-05	NEGATIVE	YES	NO	NO
AT3G23110	0.94	0.04	2.89	0.30	1.62	7.62E-05	NO	NO	NO	NO
AT3G23250	0.38	0.27	2.93	0.30	2.94	1.89E-06	NO	YES	NO	NO
AT3G23280	63.10	0.35	80.83	0.69	0.36	9.98E-03	POSITIVE	NO	NO	NO
AT3G23530	4.24	0.11	6.43	0.10	0.60	1.62E-02	NO	NO	NO	NO
AT3G23750	12.27	0.35	16.91	0.14	0.46	2.78E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G24090	8.20	0.26	17.92	0.66	1.13	3.04E-08	NO	NO	NO	NO
AT3G24180	23.41	0.08	32.38	0.20	0.47	4.26E-03	NO	NO	NO	NO
AT3G24480	34.11	0.87	50.20	0.13	0.56	3.94E-03	NO	NO	NO	NO
AT3G24503	71.74	0.41	149.00	5.00	1.05	5.55E-10	NO	NO	NO	NO
AT3G24550	9.81	0.34	16.11	0.07	0.72	4.48E-04	NO	NO	NO	NO
AT3G24560	4.57	0.28	8.69	0.14	0.93	5.76E-05	NO	NO	NO	NO
AT3G25110	2.69	0.08	7.36	0.35	1.45	7.62E-06	NO	NO	NO	NO
AT3G25180	0.56	0.37	2.61	0.61	2.22	5.80E-05	NO	NO	NO	NO
AT3G25220	73.17	1.96	135.94	2.00	0.89	9.47E-07	NO	NO	NO	NO
AT3G25600	7.95	0.60	27.62	0.74	1.80	2.23E-09	NO	NO	NO	NO
AT3G25610	5.88	0.36	18.68	0.31	1.67	0.00E+00	NO	NO	NO	NO
AT3G25780	4.32	0.59	21.18	0.33	2.29	5.41E-14	NO	NO	NO	NO
AT3G25882	24.94	3.43	140.67	1.76	2.50	0.00E+00	NO	NO	NO	NO
AT3G25890	20.41	0.53	28.58	0.30	0.49	1.22E-02	NO	YES	NO	NO
AT3G26210	15.74	2.31	66.62	0.45	2.08	0.00E+00	NO	NO	NO	NO
AT3G26220	5.11	0.62	13.47	0.48	1.40	2.01E-08	NO	NO	NO	NO
AT3G26440	2.46	0.44	8.58	0.26	1.80	9.93E-11	NO	NO	NO	NO
AT3G26500	3.56	0.61	17.18	1.28	2.27	0.00E+00	NO	NO	NO	NO
AT3G26510	1.63	0.23	9.47	0.92	2.54	8.31E-12	NO	NO	DOWN	NO
AT3G26910	2.48	0.20	5.73	0.43	1.21	2.29E-05	NO	NO	NO	NO
AT3G27020	6.17	0.17	10.57	0.56	0.78	5.18E-04	NO	NO	DOWN	NO
AT3G28340	2.59	0.29	9.31	0.78	1.85	3.06E-09	NO	NO	NO	NO
AT3G28450	12.87	0.23	23.07	0.31	0.84	1.18E-05	NO	NO	NO	NO
AT3G28500	1.81	0.27	11.02	4.80	2.61	1.49E-06	NO	NO	NO	NO
AT3G28540	22.33	1.30	32.28	1.27	0.53	7.34E-03	NO	NO	NO	NO
AT3G28580	1.74	0.61	27.19	1.50	3.97	0.00E+00	NO	NO	NO	NO
AT3G28850	1.67	0.12	3.89	0.12	1.22	8.55E-03	NO	NO	NO	NO
AT3G28890	1.26	0.44	3.06	0.43	1.28	6.52E-04	NO	NO	NO	NO
AT3G28930	48.95	0.51	92.09	1.36	0.91	7.88E-07	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G29000	2.46	0.91	14.32	0.97	2.54	2.92E-07	NO	NO	NO	NO
AT3G29034	6.96	0.91	21.81	0.99	1.65	1.49E-05	NO	NO	NO	NO
AT3G29290	11.22	0.32	17.16	0.16	0.61	4.53E-03	NO	NO	NO	NO
AT3G29400	8.67	0.46	12.73	0.34	0.55	1.77E-02	NO	NO	NO	NO
AT3G44260	3.59	0.29	13.39	0.24	1.90	1.93E-08	NO	NO	DOWN	NO
AT3G44320	1.45	0.13	7.13	1.39	2.30	1.17E-08	NO	NO	NO	NO
AT3G44350	1.11	0.52	5.79	0.21	2.38	2.13E-04	NO	YES	NO	NO
AT3G44400	7.81	0.20	14.19	0.34	0.86	2.89E-06	NO	NO	NO	NO
AT3G44480	4.50	0.11	7.84	0.22	0.80	1.61E-04	NO	NO	NO	NO
AT3G44720	12.30	0.57	28.97	0.20	1.24	1.15E-10	NO	NO	NO	NO
AT3G45440	1.71	0.10	3.37	0.12	0.98	1.87E-02	NO	NO	NO	NO
AT3G45620	48.94	0.69	64.58	1.58	0.40	4.82E-02	POSITIVE	NO	NO	NO
AT3G45640	81.58	1.32	148.74	1.93	0.87	5.43E-07	NO	NO	NO	NO
AT3G45730	1.90	0.19	15.46	1.04	3.03	3.43E-09	NO	NO	NO	NO
AT3G45970	8.52	0.79	22.70	0.25	1.41	2.90E-08	NO	NO	DOWN	NO
AT3G46080	1.46	0.56	43.03	1.10	4.88	0.00E+00	NO	YES	NO	NO
AT3G46090	0.79	0.09	8.96	0.15	3.50	1.49E-07	NO	YES	NO	NO
AT3G46110	4.62	0.41	9.59	0.39	1.05	4.82E-04	NO	NO	NO	NO
AT3G46190	0.11	0.01	4.45	0.35	5.38	1.14E-06	NO	NO	NO	NO
AT3G46510	28.10	0.32	45.40	0.32	0.69	1.10E-04	NO	NO	NO	NO
AT3G46600	6.77	0.20	20.71	0.43	1.61	0.00E+00	NO	YES	NO	NO
AT3G46620	29.52	0.18	47.10	0.84	0.67	3.97E-04	NO	NO	NO	NO
AT3G46920	2.15	0.10	3.83	0.16	0.83	4.03E-03	NO	NO	NO	NO
AT3G47090	1.44	0.18	5.25	0.45	1.86	4.41E-10	NO	NO	NO	NO
AT3G47250	44.56	0.13	57.74	0.41	0.37	5.91E-03	NEGATIVE	NO	NO	NO
AT3G47480	36.53	2.04	257.43	2.68	2.82	0.00E+00	NO	NO	NO	NO
AT3G47590	23.80	0.34	34.67	0.96	0.54	1.06E-02	NO	NO	NO	NO
AT3G47780	2.36	0.15	7.42	0.25	1.65	7.42E-11	NO	NO	NO	NO
AT3G47800	56.55	0.11	121.47	1.05	1.10	6.15E-11	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G47820	7.28	0.50	15.19	0.22	1.06	2.82E-06	NO	NO	NO	NO
AT3G48090	29.43	1.33	61.61	0.83	1.07	2.01E-11	NO	NO	NO	NO
AT3G48100	5.82	0.20	10.52	0.51	0.85	3.88E-03	NO	NO	UP	NO
AT3G48310	11.26	0.13	24.78	0.15	1.14	7.33E-09	NO	NO	NO	NO
AT3G48320	21.30	0.05	36.37	0.86	0.77	4.80E-05	NO	NO	NO	NO
AT3G48360	0.71	0.15	1.82	0.21	1.36	4.12E-02	NO	NO	NO	NO
AT3G49060	6.09	0.11	9.46	0.20	0.64	4.77E-03	NO	NO	NO	NO
AT3G49120	220.37	6.97	441.30	8.66	1.00	3.12E-07	NO	NO	NO	NO
AT3G49210	2.25	0.31	8.29	0.19	1.88	1.61E-09	NO	NO	NO	NO
AT3G49350	7.77	0.29	13.02	0.35	0.74	8.31E-04	NO	NO	NO	NO
AT3G49530	11.31	0.17	30.26	0.88	1.42	1.04E-13	NO	YES	NO	NO
AT3G49800	24.10	0.20	41.05	0.62	0.77	3.26E-05	NO	NO	NO	NO
AT3G49970	1.42	0.25	3.79	0.41	1.41	1.79E-03	NO	NO	NO	NO
AT3G50340	2.90	0.23	5.57	0.07	0.94	1.09E-02	NO	NO	NO	NO
AT3G50880	3.62	0.21	9.75	0.28	1.43	6.00E-05	NO	NO	DOWN	NO
AT3G50910	14.65	0.45	23.50	0.24	0.68	6.84E-04	NO	NO	NO	NO
AT3G50930	2.83	0.86	31.13	0.51	3.46	0.00E+00	NO	NO	NO	NO
AT3G50950	28.96	1.32	63.52	0.51	1.13	0.00E+00	NO	NO	NO	NO
AT3G51000	35.15	0.44	48.79	0.48	0.47	2.18E-02	POSITIVE	NO	NO	NO
AT3G51130	18.64	0.49	27.82	0.45	0.58	5.71E-03	NO	NO	NO	NO
AT3G51250	23.60	0.24	32.83	0.60	0.48	2.19E-02	NO	NO	NO	NO
AT3G51330	4.80	0.56	23.84	0.33	2.31	0.00E+00	NO	NO	NO	NO
AT3G51370	65.38	0.89	95.13	0.34	0.54	3.14E-03	NO	NO	NO	NO
AT3G51440	3.76	0.25	15.37	0.25	2.03	6.28E-12	NO	NO	NO	NO
AT3G51450	15.39	1.50	31.51	0.84	1.03	4.42E-07	NO	NO	NO	NO
AT3G51550	39.41	0.45	54.75	0.40	0.47	1.62E-02	NO	NO	NO	NO
AT3G51630	28.76	0.29	36.08	0.10	0.33	3.73E-02	NO	NO	NO	NO
AT3G51670	19.49	0.28	38.29	1.33	0.97	1.10E-07	NO	NO	NO	NO
AT3G51770	6.97	0.08	9.32	0.31	0.42	3.74E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G51895	10.62	0.27	17.80	1.28	0.74	1.79E-04	NO	NO	NO	NO
AT3G51980	18.92	0.75	50.39	1.92	1.41	0.00E+00	NO	NO	NO	NO
AT3G52430	18.12	1.11	66.65	0.86	1.88	0.00E+00	NO	NO	NO	NO
AT3G52605	1.28	0.08	6.70	0.07	2.39	1.49E-07	NO	NO	NO	NO
AT3G52800	41.14	0.60	64.53	0.39	0.65	5.85E-04	NO	NO	NO	NO
AT3G52930	188.21	3.03	378.52	6.01	1.01	2.66E-07	NO	NO	NO	NO
AT3G53180	6.18	0.26	10.92	0.62	0.82	1.25E-04	NO	NO	NO	NO
AT3G53190	12.57	0.46	17.87	1.37	0.51	2.65E-02	NO	NO	NO	NO
AT3G53230	11.96	0.45	17.55	0.36	0.55	6.48E-03	NO	NO	NO	NO
AT3G53280	5.88	0.21	9.25	0.41	0.65	2.20E-02	NO	NO	NO	NO
AT3G53670	70.78	0.71	96.08	0.80	0.44	2.80E-02	NO	NO	NO	NO
AT3G53710	17.65	0.50	24.25	0.41	0.46	8.88E-03	NO	NO	NO	NO
AT3G54100	4.44	0.02	7.92	0.35	0.83	1.17E-03	NO	NO	NO	NO
AT3G54150	3.82	0.91	13.41	1.05	1.81	5.88E-09	NO	NO	NO	NO
AT3G54250	9.40	0.10	13.62	0.77	0.54	4.36E-02	NO	NO	NO	NO
AT3G54300	37.42	0.24	57.84	0.85	0.63	2.81E-04	NO	NO	NO	NO
AT3G54420	2.93	0.41	14.95	0.55	2.35	7.81E-10	NO	NO	NO	NO
AT3G54640	51.83	0.86	86.62	0.31	0.74	2.88E-05	NO	NO	NO	NO
AT3G54810	19.70	0.53	29.03	0.61	0.56	5.22E-03	NO	YES	NO	NO
AT3G54960	25.00	0.24	67.19	1.13	1.43	0.00E+00	NO	NO	NO	NO
AT3G55130	26.98	0.45	39.42	1.22	0.55	4.92E-03	NO	NO	NO	NO
AT3G55430	13.48	0.71	21.61	0.57	0.68	9.38E-04	NO	NO	NO	NO
AT3G55560	3.93	0.21	7.24	0.10	0.88	6.31E-03	NO	NO	NO	NO
AT3G55840	0.57	0.14	4.49	0.24	2.97	2.90E-10	NO	NO	NO	NO
AT3G55890	1.62	0.27	5.70	0.88	1.81	7.45E-03	NO	NO	NO	NO
AT3G55950	2.15	0.21	5.43	0.40	1.34	2.83E-06	NO	NO	NO	NO
AT3G55960	23.51	0.25	34.53	0.79	0.55	7.62E-03	NO	NO	NO	NO
AT3G55980	9.43	0.58	29.22	0.80	1.63	0.00E+00	NO	YES	DOWN	NO
AT3G56050	37.78	0.36	51.60	0.19	0.45	1.77E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G56060	3.07	0.10	5.10	0.32	0.73	3.35E-02	NO	NO	NO	DOWN
AT3G56170	15.84	0.14	34.15	0.48	1.11	5.92E-09	NO	NO	NO	NO
AT3G56190	39.81	0.04	52.32	0.84	0.39	9.06E-03	NO	NO	NO	NO
AT3G56200	11.08	0.47	36.20	0.88	1.71	0.00E+00	NO	NO	NO	NO
AT3G56410	4.90	0.22	8.90	0.56	0.86	5.35E-08	NO	NO	NO	NO
AT3G56590	18.37	0.18	25.89	0.33	0.49	1.98E-03	NO	NO	NO	NO
AT3G56710	107.71	5.21	201.16	1.55	0.90	9.86E-08	NO	NO	NO	NO
AT3G56880	33.82	0.24	57.32	0.90	0.76	8.68E-05	NO	NO	NO	NO
AT3G56891	0.45	0.11	6.02	0.37	3.76	2.61E-05	NO	NO	NO	NO
AT3G57300	6.90	0.36	9.93	0.22	0.53	9.58E-03	NO	NO	NO	NO
AT3G57330	14.69	0.87	35.97	0.64	1.29	3.64E-14	NO	NO	NO	NO
AT3G57450	9.23	0.20	19.55	0.74	1.08	1.06E-03	NO	NO	NO	NO
AT3G57530	16.58	0.06	27.41	0.50	0.73	1.20E-04	NO	NO	NO	NO
AT3G57550	22.63	0.27	44.64	0.56	0.98	7.68E-09	NO	NO	NO	NO
AT3G57630	11.29	0.52	17.43	0.29	0.63	1.32E-04	NO	NO	NO	NO
AT3G57880	9.03	0.26	13.39	0.13	0.57	6.67E-03	NO	NO	NO	NO
AT3G58110	21.79	0.47	29.36	0.11	0.43	3.15E-02	NO	NO	NO	NO
AT3G58640	16.97	0.05	23.73	0.30	0.48	1.03E-03	NO	NO	NO	NO
AT3G58840	19.01	0.56	25.79	0.09	0.44	4.29E-02	NO	NO	NO	NO
AT3G59080	2.24	0.17	6.16	0.15	1.46	1.84E-06	NO	NO	NO	NO
AT3G59570	3.25	0.08	7.15	0.18	1.14	7.32E-06	NO	NO	NO	NO
AT3G59660	7.97	0.42	18.72	0.31	1.23	6.55E-10	NO	NO	NO	NO
AT3G59700	1.61	0.07	3.61	0.13	1.16	2.76E-03	NO	NO	NO	NO
AT3G60260	10.80	0.41	18.88	0.76	0.81	1.06E-04	NO	NO	NO	NO
AT3G60420	68.26	2.09	254.55	6.20	1.90	0.00E+00	NO	NO	NO	NO
AT3G60520	123.54	0.31	185.59	2.33	0.59	1.40E-03	NO	NO	NO	UP
AT3G60600	39.89	0.52	55.89	0.72	0.49	9.07E-03	NO	NO	NO	NO
AT3G60690	10.04	0.12	21.01	0.54	1.07	1.73E-04	NO	NO	NO	NO
AT3G60860	11.46	0.14	15.87	0.47	0.47	1.79E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT3G61198	0.15	0.05	1.85	0.35	3.67	9.00E-03	NO	NO	NO	NO
AT3G61220	24.56	0.50	54.68	0.95	1.15	9.56E-12	NO	NO	NO	NO
AT3G61240	36.09	0.54	46.17	0.48	0.36	1.40E-02	NO	NO	NO	NO
AT3G61670	5.34	0.10	8.43	0.08	0.66	4.82E-03	NO	NO	NO	NO
AT3G62260	3.96	0.14	10.31	0.26	1.38	1.17E-07	NO	NO	NO	NO
AT3G62422	26.13	0.54	44.30	0.29	0.76	1.06E-04	NO	NO	NO	NO
AT3G62600	46.82	0.75	146.51	2.68	1.65	0.00E+00	NO	NO	NO	NO
AT3G62720	5.38	0.17	8.43	0.39	0.65	1.67E-02	NO	NO	NO	NO
AT3G62860	0.90	0.04	3.03	0.09	1.75	2.03E-03	NO	NO	NO	NO
AT3G63500	19.36	0.28	28.40	0.92	0.55	1.11E-03	NO	NO	NO	NO
AT4G00050	15.25	0.13	23.20	0.46	0.61	6.08E-03	NO	YES	NO	NO
AT4G00750	6.53	0.24	11.37	0.56	0.80	5.43E-04	NO	NO	NO	NO
AT4G00755	11.10	0.17	15.45	0.34	0.48	4.33E-02	NO	NO	NO	NO
AT4G01010	3.63	0.32	10.72	0.56	1.56	4.69E-10	NO	NO	NO	NO
AT4G01320	61.84	0.98	85.85	0.68	0.47	1.60E-02	NO	NO	NO	NO
AT4G01700	6.86	0.80	17.14	0.51	1.32	4.97E-06	NO	NO	NO	NO
AT4G01750	4.18	0.08	11.16	0.44	1.42	4.34E-07	NO	NO	NO	NO
AT4G01950	5.98	0.44	9.75	1.14	0.71	7.44E-03	NO	NO	NO	NO
AT4G02330	5.86	0.47	21.58	1.10	1.88	0.00E+00	NO	NO	UP	NO
AT4G02380	311.32	23.01	975.61	23.70	1.65	0.00E+00	NO	NO	UP	NO
AT4G02520	537.35	15.15	1742.96	9.96	1.70	0.00E+00	NO	NO	NO	NO
AT4G03080	15.06	0.23	24.62	0.28	0.71	1.10E-04	NO	NO	NO	NO
AT4G03110	10.74	0.38	17.16	0.02	0.68	1.23E-03	NO	NO	NO	NO
AT4G03190	5.06	0.09	8.15	0.29	0.69	1.08E-02	NO	NO	NO	NO
AT4G03390	6.37	0.07	13.63	0.23	1.10	9.18E-09	NO	NO	NO	NO
AT4G03400	39.83	0.31	74.63	0.77	0.91	1.79E-07	NO	NO	UP	NO
AT4G03550	12.50	0.35	17.49	0.46	0.49	1.26E-02	NO	NO	NO	NO
AT4G03820	3.78	0.25	9.31	0.26	1.30	1.99E-06	NO	NO	NO	NO
AT4G04220	3.05	0.13	6.56	0.24	1.10	4.57E-05	NO	NO	NO	NO

Table A3.4 (cont'd)

AT4G04500	1.07	0.25	4.79	0.43	2.16	3.77E-08	NO	NO	NO	NO
AT4G04540	0.88	0.35	9.25	0.37	3.39	0.00E+00	NO	NO	NO	NO
AT4G04570	20.78	0.52	41.11	0.59	0.98	2.08E-09	NO	NO	NO	NO
AT4G04610	53.48	0.91	83.24	2.23	0.64	5.21E-04	NO	NO	NO	NO
AT4G04870	9.22	0.45	14.42	0.54	0.65	1.28E-02	NO	NO	NO	NO
AT4G04880	5.81	0.11	9.88	0.20	0.77	1.32E-02	NO	NO	NO	NO
AT4G04960	4.55	0.10	8.74	0.17	0.94	1.24E-04	NO	NO	NO	NO
AT4G05020	29.12	0.44	50.99	1.31	0.81	2.45E-06	POSITIVE	NO	UP	NO
AT4G05390	7.45	0.23	12.24	0.48	0.72	6.03E-03	NO	NO	NO	NO
AT4G08180	7.21	0.00	9.99	0.01	0.47	2.04E-02	NO	NO	NO	NO
AT4G08480	5.71	0.26	8.11	0.11	0.50	4.44E-02	NO	NO	NO	NO
AT4G08555	2.46	0.22	18.58	1.60	2.92	3.96E-06	NO	NO	NO	NO
AT4G08850	25.71	0.34	71.63	1.69	1.48	0.00E+00	NO	NO	NO	NO
AT4G08870	120.96	1.75	167.30	5.08	0.47	9.11E-03	NO	NO	NO	NO
AT4G08950	4.34	0.10	12.59	0.55	1.54	2.71E-07	NO	NO	NO	NO
AT4G09030	9.13	0.67	27.77	0.48	1.60	1.28E-06	NO	NO	NO	NO
AT4G09670	40.62	0.51	65.48	0.55	0.69	1.45E-04	NO	NO	NO	NO
AT4G09770	0.27	0.15	1.93	0.13	2.82	1.14E-04	NO	NO	NO	NO
AT4G10270	1.53	0.27	6.20	0.49	2.02	9.53E-03	NO	NO	NO	NO
AT4G11280	9.28	0.99	29.42	0.50	1.67	0.00E+00	NO	NO	UP	NO
AT4G11290	0.63	0.17	3.08	0.26	2.29	2.31E-04	NO	NO	NO	NO
AT4G11300	4.43	0.54	9.92	0.20	1.16	1.05E-04	NO	NO	NO	NO
AT4G11350	2.71	0.16	7.59	0.31	1.48	1.00E-06	NO	NO	NO	NO
AT4G11360	16.91	0.31	32.72	0.47	0.95	1.33E-04	NO	NO	NO	NO
AT4G11370	2.79	0.68	9.37	1.00	1.75	3.51E-04	NO	NO	NO	NO
AT4G11530	0.80	0.10	3.10	0.17	1.95	2.17E-05	NO	NO	NO	NO
AT4G11800	5.03	0.15	8.90	0.55	0.82	1.20E-04	NO	NO	NO	NO
AT4G11820	21.47	0.20	32.03	0.28	0.58	3.10E-03	NO	NO	NO	NO
AT4G11850	8.66	0.30	20.28	0.33	1.23	1.00E-11	NO	NO	NO	NO

Table A3.4 (cont'd)

AT4G11890	11.80	2.10	106.43	2.84	3.17	0.00E+00	NO	NO	NO	NO
AT4G12010	4.93	0.15	9.85	0.52	1.00	1.03E-06	NO	NO	NO	NO
AT4G12120	7.43	0.16	21.02	0.46	1.50	0.00E+00	NO	NO	NO	NO
AT4G12490	31.26	6.22	61.22	0.62	0.97	1.97E-05	NO	NO	UP	NO
AT4G12500	14.79	5.04	63.43	3.93	2.10	0.00E+00	NO	NO	UP	NO
AT4G12720	26.23	1.12	76.56	2.34	1.55	0.00E+00	NO	NO	NO	NO
AT4G12730	17.01	0.41	28.94	1.38	0.77	1.01E-04	NEGATIVE	NO	NO	DOWN
AT4G12770	4.87	0.15	6.75	0.19	0.47	4.55E-02	NO	NO	NO	NO
AT4G13180	15.84	0.51	30.30	0.75	0.94	2.51E-05	NO	NO	NO	NO
AT4G13340	6.83	0.50	23.79	0.50	1.80	0.00E+00	NO	NO	NO	NO
AT4G13510	24.86	1.24	72.10	0.37	1.54	0.00E+00	NO	NO	NO	NO
AT4G13530	28.16	0.26	41.52	0.85	0.56	1.76E-03	NO	NO	NO	NO
AT4G13540	1.12	0.04	4.57	0.73	2.04	1.14E-03	NO	NO	NO	NO
AT4G14365	9.19	1.74	110.64	2.11	3.59	0.00E+00	NO	NO	NO	NO
AT4G14500	23.95	0.25	33.56	1.06	0.49	1.75E-02	NO	NO	NO	NO
AT4G14550	2.08	0.06	4.81	0.43	1.21	3.32E-03	NO	YES	NO	NO
AT4G15440	3.11	0.57	14.68	0.36	2.24	0.00E+00	NEGATIVE	NO	NO	NO
AT4G15620	6.50	0.05	14.32	1.17	1.14	1.07E-03	NO	NO	NO	NO
AT4G15680	3.62	0.25	8.93	0.58	1.30	1.13E-02	NO	NO	NO	NO
AT4G15920	4.43	0.41	9.63	0.60	1.12	2.26E-03	NO	NO	DOWN	NO
AT4G16660	49.20	0.23	128.57	1.81	1.39	0.00E+00	NO	NO	NO	NO
AT4G16780	3.33	0.21	9.86	0.49	1.56	5.84E-06	NO	YES	NO	NO
AT4G16790	3.66	0.28	7.67	0.25	1.07	4.45E-04	POSITIVE	NO	NO	NO
AT4G16990	83.84	1.22	121.59	1.58	0.54	3.45E-05	NO	NO	NO	NO
AT4G17070	12.04	0.52	24.63	0.65	1.03	1.28E-06	NO	NO	NO	NO
AT4G17230	17.69	0.69	29.15	0.63	0.72	1.59E-04	NO	YES	NO	NO
AT4G17245	9.88	0.51	20.65	0.97	1.06	1.92E-03	NO	NO	NO	NO
AT4G17250	3.94	0.33	6.39	0.57	0.70	1.17E-02	NO	NO	NO	NO
AT4G17330	13.24	0.30	25.52	0.27	0.95	2.89E-08	NO	NO	NO	NO

Table A3.4 (cont'd)

AT4G17420	11.41	0.32	19.50	0.21	0.77	4.81E-04	NO	NO	NO	NO
AT4G17490	0.67	0.13	2.31	0.42	1.80	1.30E-02	NO	YES	DOWN	NO
AT4G17500	5.11	0.32	24.78	0.34	2.28	0.00E+00	NO	YES	NO	NO
AT4G17870	8.67	0.35	14.74	0.45	0.77	2.23E-02	NO	NO	DOWN	NO
AT4G18280	25.30	1.07	45.12	1.78	0.83	2.25E-04	NO	NO	UP	UP
AT4G18430	0.95	0.27	4.87	0.32	2.35	1.07E-04	NO	NO	NO	NO
AT4G18630	8.69	0.31	12.96	0.40	0.58	2.07E-02	NO	NO	NO	NO
AT4G18760	4.08	0.20	7.92	0.23	0.96	5.55E-03	NO	NO	NO	NO
AT4G18880	22.63	0.83	36.35	0.50	0.68	2.65E-04	NO	YES	NO	NO
AT4G18950	11.91	0.25	18.81	0.37	0.66	1.64E-03	NO	NO	NO	NO
AT4G19170	77.25	1.37	119.25	1.83	0.63	6.74E-04	NO	NO	NO	NO
AT4G19180	5.69	0.16	8.50	0.14	0.58	1.71E-02	NO	NO	NO	NO
AT4G19530	4.74	0.42	7.36	0.51	0.64	4.76E-03	NO	NO	DOWN	NO
AT4G19660	6.44	0.37	14.11	0.41	1.13	1.62E-06	NO	NO	NO	NO
AT4G19860	22.14	0.24	30.13	0.23	0.44	3.44E-02	NO	NO	NO	NO
AT4G19880	33.11	0.15	42.31	0.40	0.35	4.13E-02	NO	NO	NO	NO
AT4G20000	0.33	0.06	3.66	0.37	3.49	2.33E-05	NO	NO	NO	NO
AT4G20070	26.44	0.56	36.50	0.44	0.47	2.12E-02	NO	NO	NO	NO
AT4G20110	5.29	0.45	17.43	0.23	1.72	0.00E+00	NO	NO	NO	NO
AT4G20320	1.59	0.09	4.39	0.30	1.47	5.64E-06	NO	NO	NO	NO
AT4G20830	32.08	1.70	101.59	2.34	1.66	0.00E+00	NO	NO	NO	NO
AT4G20860	7.87	0.70	21.89	0.48	1.48	2.80E-13	NO	NO	NO	NO
AT4G21380	4.14	0.59	11.48	0.19	1.47	5.86E-12	POSITIVE	NO	NO	NO
AT4G21400	0.32	0.04	5.55	0.48	4.09	0.00E+00	NO	NO	NO	NO
AT4G21850	1.40	0.32	33.36	1.99	4.58	0.00E+00	NO	NO	DOWN	NO
AT4G22305	5.53	0.48	12.90	0.08	1.22	1.01E-04	NO	NO	NO	NO
AT4G22530	3.01	0.46	10.20	0.64	1.76	1.03E-06	NO	NO	NO	NO
AT4G22590	8.10	0.07	19.09	0.74	1.24	3.00E-09	NO	NO	NO	NO
AT4G22980	2.82	0.43	5.25	0.22	0.90	1.30E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT4G23010	42.31	0.73	61.05	0.43	0.53	3.53E-03	NO	NO	NO	NO
AT4G23030	0.53	0.19	3.42	0.25	2.68	2.68E-06	NO	NO	NO	NO
AT4G23130	5.00	0.41	9.50	0.43	0.93	9.06E-05	NO	NO	NO	NO
AT4G23170	49.13	1.57	118.34	0.68	1.27	3.63E-14	NO	NO	NO	NO
AT4G23180	15.42	0.24	25.48	0.95	0.72	1.97E-04	NO	NO	DOWN	NO
AT4G23190	2.37	0.35	15.89	1.09	2.74	0.00E+00	NO	NO	NO	NO
AT4G23270	9.79	0.50	16.98	1.08	0.79	1.68E-04	NO	NO	NO	NO
AT4G23310	0.46	0.11	4.90	0.49	3.40	3.69E-14	NO	NO	NO	NO
AT4G23320	0.73	0.21	4.00	0.22	2.45	7.51E-06	NO	NO	NO	NO
AT4G23470	53.84	0.52	119.98	1.03	1.16	1.69E-13	NO	NO	NO	NO
AT4G23610	2.38	0.56	12.04	0.36	2.34	6.64E-08	NO	NO	NO	NO
AT4G23730	16.44	0.16	22.95	0.87	0.48	2.84E-02	NO	NO	NO	NO
AT4G23800	25.58	0.58	34.60	1.44	0.44	4.02E-02	NO	YES	NO	NO
AT4G23850	77.95	0.32	134.24	0.60	0.78	1.52E-05	NO	NO	NO	NO
AT4G24020	15.56	0.38	21.62	0.33	0.47	1.90E-02	NO	YES	NO	NO
AT4G24040	2.89	0.16	7.65	0.33	1.40	4.00E-07	NO	NO	NO	NO
AT4G24230	5.24	0.17	15.00	0.11	1.52	2.73E-11	NO	NO	NO	NO
AT4G24690	134.45	1.04	225.21	1.97	0.74	2.38E-04	POSITIVE	NO	NO	NO
AT4G24920	54.44	0.82	104.87	0.58	0.95	3.47E-06	NO	NO	NO	NO
AT4G24970	1.34	0.14	3.77	0.30	1.50	7.39E-05	NO	NO	NO	NO
AT4G25070	2.15	0.52	9.13	0.40	2.09	0.00E+00	NO	NO	NO	NO
AT4G25230	11.81	0.16	17.99	0.23	0.61	2.60E-03	NO	NO	NO	NO
AT4G25290	12.98	0.24	20.89	0.17	0.69	4.81E-04	NO	NO	NO	NO
AT4G25620	6.25	0.31	16.05	0.06	1.36	1.45E-10	NO	NO	NO	NO
AT4G25810	0.58	0.40	7.93	0.22	3.78	3.93E-10	NO	NO	NO	NO
AT4G25880	13.50	0.10	20.83	0.20	0.62	7.99E-06	NO	NO	NO	NO
AT4G25900	39.54	0.63	75.24	1.08	0.93	8.51E-08	NO	NO	NO	NO
AT4G26090	2.60	0.40	4.62	0.37	0.83	1.67E-03	NO	NO	NO	NO
AT4G26120	3.37	0.29	6.75	0.36	1.00	1.29E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT4G26190	7.52	0.21	11.87	0.08	0.66	1.11E-03	NO	NO	NO	NO
AT4G26400	21.94	0.51	29.62	0.25	0.43	1.86E-02	NO	NO	NO	NO
AT4G26470	0.96	0.33	5.40	0.20	2.49	5.19E-06	NO	NO	NO	NO
AT4G26630	49.19	0.61	65.77	0.01	0.42	1.44E-03	NO	NO	NO	NO
AT4G26690	12.53	0.38	21.37	0.08	0.77	3.61E-05	NO	NO	NO	NO
AT4G26910	27.91	0.36	43.80	0.25	0.65	1.71E-06	NO	NO	NO	NO
AT4G26970	34.88	0.31	60.60	0.21	0.80	6.01E-06	NO	NO	NO	NO
AT4G27020	4.01	0.27	9.84	0.26	1.29	9.92E-07	NO	NO	NO	NO
AT4G27040	7.64	0.05	14.12	0.31	0.89	3.14E-05	NO	NO	NO	NO
AT4G27280	7.30	1.08	45.45	1.26	2.64	0.00E+00	NO	NO	DOWN	NO
AT4G27450	1.31	0.26	8.27	1.05	2.66	4.33E-09	NO	NO	DOWN	NO
AT4G27470	7.82	0.37	15.60	0.58	1.00	3.19E-04	NO	NO	NO	NO
AT4G27745	8.23	0.21	14.44	0.40	0.81	2.11E-02	NO	NO	NO	NO
AT4G27830	15.73	0.17	25.60	0.57	0.70	6.24E-04	NO	NO	UP	NO
AT4G27860	2.21	0.39	9.17	1.02	2.06	0.00E+00	NO	NO	NO	NO
AT4G27870	12.68	0.26	18.96	0.22	0.58	4.01E-03	NO	NO	NO	NO
AT4G28300	29.77	0.33	56.43	0.46	0.92	4.25E-11	NO	NO	NO	NO
AT4G28390	5.62	0.48	27.38	0.08	2.29	0.00E+00	POSITIVE	NO	NO	NO
AT4G28395	0.00	0.00	3.72	1.30	Inf	5.84E-05	NO	NO	NO	NO
AT4G28400	17.63	0.20	40.98	0.47	1.22	9.54E-11	NO	NO	NO	UP
AT4G28490	6.52	0.53	14.29	0.53	1.13	5.95E-09	NO	NO	NO	NO
AT4G28570	7.90	0.20	18.11	0.11	1.20	2.10E-10	NO	NO	NO	NO
AT4G28710	4.74	0.25	7.24	0.29	0.61	3.83E-03	NO	NO	NO	NO
AT4G29160	36.01	0.51	74.53	0.74	1.05	3.29E-11	NO	NO	NO	NO
AT4G29210	9.94	0.44	14.81	0.65	0.57	6.00E-03	NO	NO	NO	NO
AT4G29380	7.71	0.06	11.63	0.35	0.59	2.77E-03	NO	NO	NO	NO
AT4G29440	4.49	0.08	8.85	0.39	0.98	2.82E-07	NO	NO	NO	NO
AT4G29740	3.33	0.31	12.73	0.90	1.93	1.03E-13	NO	NO	NO	NO
AT4G29750	23.65	0.27	31.40	0.41	0.41	4.46E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT4G29780	1.72	0.18	4.84	0.04	1.49	3.18E-05	NO	NO	DOWN	NO
AT4G29810	64.22	0.59	87.69	1.10	0.45	1.88E-02	NO	NO	NO	NO
AT4G29900	15.26	0.40	40.90	0.57	1.42	0.00E+00	NO	NO	NO	NO
AT4G29950	10.45	0.26	28.05	0.15	1.42	0.00E+00	NO	NO	NO	NO
AT4G30060	2.92	0.05	7.36	0.20	1.33	4.01E-06	NO	NO	NO	NO
AT4G30190	77.72	1.20	119.44	2.45	0.62	3.97E-05	NO	NO	NO	NO
AT4G30210	30.92	0.43	40.91	0.29	0.40	4.24E-02	NO	NO	NO	NO
AT4G30250	1.60	0.18	3.41	0.44	1.09	2.28E-02	NO	NO	NO	NO
AT4G30270	88.47	2.47	152.43	1.59	0.78	1.33E-05	NO	NO	NO	NO
AT4G30440	33.47	0.12	47.42	1.18	0.50	1.04E-02	NO	NO	NO	NO
AT4G30530	169.43	0.64	283.80	4.61	0.74	3.00E-05	NO	NO	NO	NO
AT4G30640	0.52	0.09	3.17	0.34	2.62	3.71E-04	NO	NO	NO	NO
AT4G31170	36.58	1.07	49.77	0.47	0.44	1.32E-03	NO	NO	NO	NO
AT4G31500	98.45	0.92	264.91	8.12	1.43	0.00E+00	NO	NO	UP	DOWN
AT4G31510	11.54	0.76	18.55	0.15	0.68	3.04E-02	NO	NO	NO	NO
AT4G31800	6.95	0.53	23.26	0.22	1.74	1.20E-13	NO	YES	UP	NO
AT4G31980	3.74	0.23	6.14	0.31	0.72	2.07E-02	NO	NO	NO	NO
AT4G32030	11.89	0.12	16.92	0.14	0.51	4.15E-02	NO	NO	NO	NO
AT4G32060	58.70	0.60	85.81	0.18	0.55	3.71E-03	NO	NO	NO	NO
AT4G32070	4.97	0.19	7.67	0.15	0.62	1.57E-02	NO	NO	NO	NO
AT4G32285	29.39	0.35	38.47	0.77	0.39	1.24E-02	NO	NO	NO	NO
AT4G32480	0.69	0.14	2.13	0.12	1.62	2.11E-02	NO	NO	NO	NO
AT4G32670	1.06	0.08	3.84	0.58	1.86	6.93E-08	NO	NO	NO	NO
AT4G32760	27.24	0.35	43.31	0.24	0.67	4.48E-07	NO	NO	NO	NO
AT4G33050	23.67	2.55	81.98	0.99	1.79	0.00E+00	NO	NO	NO	DOWN
AT4G33220	29.73	0.49	41.33	1.05	0.48	1.89E-02	NO	NO	NO	NO
AT4G33240	9.25	0.16	15.30	0.26	0.73	3.22E-10	NO	NO	NO	NO
AT4G33300	22.57	0.46	60.57	0.87	1.42	0.00E+00	NO	NO	NO	NO
AT4G33540	43.43	0.66	124.57	1.97	1.52	0.00E+00	POSITIVE	NO	NO	NO

Table A3.4 (cont'd)

AT4G33565	32.20	0.64	60.79	0.91	0.92	2.56E-07	NO	NO	NO	NO
AT4G33910	17.69	0.14	33.48	0.54	0.92	4.89E-06	NO	NO	NO	NO
AT4G33920	36.37	0.62	76.81	0.65	1.08	1.92E-10	NO	NO	UP	NO
AT4G34050	281.78	4.13	443.63	6.28	0.65	2.31E-07	NO	NO	NO	NO
AT4G34100	26.68	0.40	36.93	0.36	0.47	1.27E-02	NO	NO	NO	NO
AT4G34135	22.00	0.85	28.86	3.60	0.39	3.39E-02	NO	NO	NO	NO
AT4G34150	31.66	0.25	125.67	1.34	1.99	0.00E+00	NO	NO	NO	NO
AT4G34180	85.77	1.04	122.93	0.23	0.52	5.22E-03	NO	NO	NO	NO
AT4G34480	23.10	0.45	31.90	0.76	0.47	2.41E-02	NO	NO	NO	NO
AT4G34630	83.64	0.43	224.25	2.05	1.42	0.00E+00	NO	NO	NO	NO
AT4G35060	1.68	0.44	4.71	0.37	1.48	8.62E-03	NO	NO	NO	NO
AT4G35180	1.40	0.42	13.14	0.12	3.23	0.00E+00	NO	NO	NO	NO
AT4G35500	6.22	0.11	9.39	0.06	0.60	3.31E-02	NO	NO	NO	NO
AT4G35600	6.83	0.52	22.60	0.55	1.73	0.00E+00	NO	NO	NO	NO
AT4G35810	0.33	0.17	1.77	0.45	2.43	1.40E-02	NO	NO	NO	NO
AT4G36030	0.70	0.16	2.35	0.35	1.75	1.79E-04	NO	NO	NO	NO
AT4G36040	95.02	1.16	198.56	0.69	1.06	6.68E-11	NO	NO	NO	NO
AT4G36220	69.33	1.05	142.64	1.10	1.04	1.16E-09	NO	NO	NO	NO
AT4G36430	1.48	0.14	5.05	0.35	1.77	1.47E-04	NO	NO	NO	NO
AT4G36550	5.09	0.27	9.29	0.24	0.87	1.54E-04	NO	NO	NO	NO
AT4G36670	15.31	0.72	28.94	2.03	0.92	2.69E-06	NO	NO	NO	NO
AT4G36760	32.53	0.73	50.65	0.09	0.64	4.41E-04	NO	NO	NO	NO
AT4G36988	11.58	0.78	55.21	0.51	2.25	0.00E+00	NO	NO	NO	NO
AT4G37010	0.90	0.48	4.07	0.16	2.17	8.19E-04	NO	NO	NO	NO
AT4G37290	0.29	0.11	1.67	0.26	2.51	3.63E-02	NO	NO	NO	NO
AT4G37310	16.02	0.63	39.16	0.40	1.29	2.80E-13	NO	NO	NO	NO
AT4G37390	1.54	0.90	4.90	0.65	1.67	2.16E-06	NO	NO	NO	NO
AT4G37520	7.80	1.84	17.26	0.46	1.14	5.50E-06	NO	NO	NO	NO
AT4G37530	3.27	1.08	13.97	1.75	2.10	1.22E-11	NO	NO	NO	NO

Table A3.4 (cont'd)

AT4G37640	26.04	0.36	48.26	0.80	0.89	4.17E-07	NO	NO	NO	NO
AT4G37870	39.15	1.13	62.11	0.41	0.67	2.22E-04	NO	NO	NO	NO
AT4G37900	1.07	0.15	4.07	0.15	1.93	2.14E-07	NO	NO	NO	NO
AT4G38470	10.11	0.37	17.77	0.17	0.81	1.29E-04	NO	NO	NO	NO
AT4G38480	3.69	0.20	8.49	0.06	1.20	6.06E-05	NO	NO	NO	NO
AT4G38550	36.80	1.16	71.78	1.56	0.96	1.36E-08	NO	NO	NO	NO
AT4G38560	0.92	0.37	12.15	0.53	3.73	0.00E+00	NO	NO	NO	NO
AT4G38620	11.26	0.37	21.77	0.59	0.95	2.63E-05	NO	YES	NO	NO
AT4G39030	3.74	0.60	37.05	1.19	3.31	0.00E+00	NO	NO	NO	NO
AT4G39390	15.03	0.20	21.28	0.11	0.50	9.02E-03	NO	NO	NO	NO
AT4G39580	2.44	0.17	6.20	0.23	1.35	1.74E-03	NO	NO	NO	NO
AT4G39610	0.52	0.21	7.55	0.37	3.85	3.97E-11	NO	NO	NO	NO
AT4G39640	16.33	0.30	32.66	1.07	1.00	1.31E-10	NO	NO	NO	NO
AT4G39670	3.51	0.90	35.79	0.88	3.35	0.00E+00	NO	NO	NO	NO
AT4G39840	6.39	0.47	9.93	0.13	0.64	1.50E-02	NO	NO	NO	NO
AT4G39890	11.20	0.64	18.98	0.37	0.76	6.93E-03	NO	NO	NO	NO
AT4G39950	24.06	1.00	93.40	0.71	1.96	0.00E+00	NO	NO	NO	NO
AT4G40020	1.46	0.15	4.37	0.10	1.58	6.09E-05	NO	NO	NO	NO
AT5G01100	0.83	0.23	4.04	0.12	2.28	1.59E-08	NO	NO	NO	NO
AT5G01210	3.03	0.28	5.17	0.55	0.77	3.24E-02	NO	NO	NO	NO
AT5G01400	5.65	0.27	7.85	0.26	0.48	3.22E-02	NO	NO	NO	NO
AT5G01500	16.39	0.28	24.07	0.50	0.55	1.08E-02	NO	NO	NO	NO
AT5G01540	0.33	0.14	4.05	0.34	3.63	1.69E-13	NO	NO	NO	NO
AT5G01542	5.18	0.28	26.03	1.40	2.33	0.00E+00	NO	NO	NO	NO
AT5G01720	6.59	0.30	12.35	0.22	0.91	1.95E-05	NO	NO	NO	NO
AT5G01770	1.80	0.04	2.90	0.17	0.69	2.06E-02	NO	NO	NO	NO
AT5G01810	41.09	1.10	63.87	1.28	0.64	5.61E-04	NO	NO	NO	NO
AT5G01820	23.57	0.48	54.83	1.11	1.22	6.63E-13	NO	NO	NO	NO
AT5G01830	0.81	0.04	2.20	0.13	1.45	1.66E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G01900	0.30	0.09	2.07	0.24	2.79	1.24E-03	NO	YES	NO	NO
AT5G02230	6.94	0.55	14.12	1.00	1.03	2.06E-04	NO	NO	DOWN	NO
AT5G02270	35.04	1.02	53.66	0.38	0.61	6.75E-04	NO	NO	NO	NO
AT5G02290	18.83	0.71	45.03	0.49	1.26	6.63E-13	NO	NO	NO	NO
AT5G02310	8.72	0.13	11.97	0.31	0.46	2.34E-02	NO	NO	NO	NO
AT5G02620	17.09	0.28	28.86	0.43	0.76	8.23E-05	NO	NO	NO	NO
AT5G02780	0.92	0.39	16.01	1.73	4.12	0.00E+00	NO	NO	NO	NO
AT5G02830	48.41	0.46	64.85	0.31	0.42	3.68E-02	NO	NO	NO	NO
AT5G02880	9.31	0.24	14.38	0.40	0.63	9.97E-04	NO	NO	NO	NO
AT5G03040	38.90	0.43	61.13	0.17	0.65	1.11E-06	NO	NO	NO	NO
AT5G03120	22.36	0.60	45.91	1.73	1.04	5.77E-05	NO	NO	NO	NO
AT5G03160	23.51	0.41	56.61	0.92	1.27	4.04E-13	NO	NO	NO	NO
AT5G03260	1.36	0.12	2.64	0.25	0.95	4.83E-02	NO	NO	NO	NO
AT5G03555	25.22	0.66	40.46	0.55	0.68	2.32E-04	NO	NO	NO	NO
AT5G03610	9.87	0.79	28.49	0.61	1.53	7.08E-14	NO	NO	NO	NO
AT5G04020	5.30	0.18	8.18	0.73	0.63	3.26E-03	NO	NO	NO	NO
AT5G04040	7.65	0.26	11.80	0.20	0.62	2.56E-03	NO	NO	NO	NO
AT5G04160	5.62	0.23	11.18	0.36	0.99	6.17E-04	NO	NO	NO	NO
AT5G04550	24.32	0.26	35.97	0.31	0.56	2.82E-03	NO	NO	NO	NO
AT5G04560	6.42	0.31	10.86	0.50	0.76	4.29E-05	NO	NO	NO	NO
AT5G04720	11.94	0.55	42.09	0.52	1.82	0.00E+00	NO	NO	NO	NO
AT5G04930	19.45	0.86	40.98	0.42	1.07	4.04E-10	NO	NO	NO	NO
AT5G05090	7.02	0.51	11.44	0.20	0.70	1.87E-02	NO	YES	NO	NO
AT5G05100	16.21	0.39	23.58	0.23	0.54	1.21E-02	NO	NO	NO	NO
AT5G05190	3.62	0.18	10.31	0.25	1.51	1.20E-10	NO	NO	NO	NO
AT5G05340	1.26	0.89	4.68	0.60	1.89	2.48E-04	NO	NO	NO	NO
AT5G05410	9.06	0.42	13.13	0.15	0.54	4.27E-02	NO	YES	NO	NO
AT5G05460	6.51	0.33	9.31	0.11	0.52	4.75E-02	NO	NO	NO	NO
AT5G05570	6.69	0.05	10.58	0.33	0.66	1.31E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G05730	19.48	0.75	43.51	0.60	1.16	8.75E-12	NO	NO	NO	NO
AT5G06120	7.58	0.05	10.11	0.23	0.42	1.21E-02	NO	NO	NO	NO
AT5G06750	7.34	0.39	18.87	0.34	1.36	1.16E-10	NO	NO	NO	NO
AT5G07100	12.86	1.34	20.82	1.65	0.70	2.15E-03	NO	YES	NO	NO
AT5G07180	1.62	0.07	2.90	0.36	0.84	1.28E-02	NO	NO	NO	NO
AT5G07350	27.22	0.55	40.98	0.66	0.59	1.23E-05	NO	NO	NO	NO
AT5G07370	51.42	0.68	72.25	0.24	0.49	5.75E-03	NO	NO	NO	NO
AT5G07710	3.57	0.18	6.95	0.10	0.96	1.85E-03	NO	NO	NO	NO
AT5G07770	1.76	0.14	4.52	0.38	1.36	3.34E-06	NO	NO	NO	NO
AT5G07790	3.92	0.20	7.08	0.19	0.85	7.53E-04	NO	NO	NO	NO
AT5G07870	4.26	0.05	7.76	0.21	0.86	6.41E-03	NO	NO	NO	NO
AT5G08130	13.92	0.15	19.29	0.60	0.47	1.18E-03	NO	YES	NO	NO
AT5G08240	3.12	0.28	8.64	0.35	1.47	7.97E-06	NO	NO	NO	NO
AT5G08450	17.52	0.11	23.93	0.41	0.45	6.67E-04	NO	NO	NO	NO
AT5G08790	21.50	0.64	75.26	1.28	1.81	0.00E+00	NO	YES	NO	NO
AT5G09330	26.54	0.56	35.89	0.11	0.44	3.26E-03	NO	YES	NO	NO
AT5G09670	10.12	1.39	18.84	0.53	0.90	8.05E-07	NO	NO	NO	NO
AT5G10250	0.67	0.09	1.63	0.16	1.29	3.80E-02	NO	NO	NO	NO
AT5G10290	14.52	0.13	24.81	0.41	0.77	4.69E-05	NO	NO	NO	NO
AT5G10300	2.13	0.23	5.71	0.21	1.43	3.29E-03	NO	NO	NO	NO
AT5G10490	14.09	0.13	19.50	0.20	0.47	3.66E-03	NO	NO	NO	NO
AT5G10550	16.99	0.17	23.42	0.61	0.46	3.09E-02	NO	NO	NO	NO
AT5G10770	3.44	0.40	9.43	0.08	1.46	2.03E-07	NO	NO	NO	NO
AT5G11000	13.55	0.92	22.84	0.44	0.75	5.70E-04	NO	NO	NO	NO
AT5G11060	25.39	0.47	35.15	0.61	0.47	2.18E-02	NO	NO	NO	NO
AT5G11250	4.64	0.19	7.67	0.44	0.72	1.37E-03	NO	NO	NO	NO
AT5G11650	11.35	0.29	23.91	0.21	1.07	1.19E-07	NO	NO	NO	NO
AT5G11850	5.71	0.16	8.76	0.08	0.62	7.52E-03	NO	NO	NO	NO
AT5G11920	2.93	0.48	4.91	0.30	0.74	3.95E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G11970	16.93	0.84	40.76	0.80	1.27	2.15E-07	NO	NO	NO	NO
AT5G12010	18.35	0.68	32.23	0.39	0.81	1.56E-05	NO	NO	NO	NO
AT5G12300	4.48	0.14	12.83	0.62	1.52	1.93E-08	NO	NO	NO	NO
AT5G12480	24.60	0.43	37.33	0.20	0.60	1.42E-03	NO	NO	NO	NO
AT5G12850	10.89	0.43	16.58	0.50	0.61	2.55E-03	NO	YES	NO	NO
AT5G12940	2.86	0.26	6.46	0.39	1.17	2.26E-03	NO	NO	NO	NO
AT5G12950	6.06	0.38	8.86	0.25	0.55	2.01E-02	NO	NO	NO	NO
AT5G13080	1.46	1.01	21.46	1.40	3.88	1.88E-14	NO	YES	NO	NO
AT5G13190	29.01	0.51	69.41	0.85	1.26	1.76E-10	NO	NO	NO	NO
AT5G13370	24.41	0.27	32.76	0.32	0.42	4.28E-02	NO	NO	NO	NO
AT5G13420	51.84	1.35	71.57	0.35	0.47	1.78E-02	NO	NO	NO	NO
AT5G13490	4.15	1.09	16.33	1.39	1.98	1.89E-14	NO	NO	NO	NO
AT5G13500	8.17	0.09	13.00	0.13	0.67	3.54E-03	NO	NO	NO	NO
AT5G13820	1.96	0.18	4.40	0.13	1.17	2.59E-04	NO	NO	NO	NO
AT5G14120	6.66	0.18	11.77	0.44	0.82	1.74E-04	NO	NO	DOWN	NO
AT5G14780	150.00	1.98	282.61	4.38	0.91	4.57E-07	NO	NO	NO	NO
AT5G14930	18.39	0.55	33.47	0.84	0.86	7.25E-08	NO	NO	NO	NO
AT5G15850	76.38	1.47	106.37	0.88	0.48	1.31E-02	NO	YES	NO	NO
AT5G15870	10.05	0.15	26.15	1.00	1.38	1.53E-13	NO	NO	NO	NO
AT5G15980	9.13	0.16	12.87	0.10	0.50	2.93E-02	NO	NO	NO	NO
AT5G16170	1.28	0.21	4.81	0.27	1.92	3.22E-05	NO	NO	NO	NO
AT5G16190	1.17	0.05	2.61	0.26	1.16	4.14E-02	NO	NO	NO	NO
AT5G16230	6.70	0.23	14.29	0.83	1.09	1.74E-05	NO	NO	NO	NO
AT5G16510	37.64	0.72	51.90	1.05	0.46	1.72E-02	NO	NO	NO	NO
AT5G16800	11.32	0.26	16.09	0.18	0.51	3.19E-02	NO	NO	NO	NO
AT5G16840	94.87	0.37	117.78	2.23	0.31	3.41E-02	NO	NO	NO	NO
AT5G16880	46.73	0.44	78.62	1.12	0.75	5.51E-07	NO	NO	NO	NO
AT5G17020	14.44	0.28	18.28	0.29	0.34	4.60E-02	NO	NO	NO	NO
AT5G17300	13.53	0.34	28.34	0.44	1.07	1.35E-08	NO	YES	NO	NO

Table A3.4 (cont'd)

AT5G17640	5.52	0.08	12.44	0.28	1.17	8.58E-07	NO	NO	NO	NO
AT5G17910	4.82	0.08	8.37	0.30	0.80	4.01E-05	NO	NO	NO	NO
AT5G18480	25.23	0.12	46.82	0.80	0.89	3.45E-07	NO	NO	NO	NO
AT5G18490	7.01	0.44	11.20	0.28	0.68	5.90E-03	NO	NO	NO	NO
AT5G18650	11.27	0.35	19.12	0.49	0.76	6.37E-04	NO	NO	NO	NO
AT5G18670	4.26	0.13	14.71	0.78	1.79	3.67E-14	NO	NO	NO	NO
AT5G18780	3.20	0.38	6.74	0.50	1.08	7.82E-04	NO	NO	NO	NO
AT5G19120	1.13	0.06	3.57	0.05	1.66	8.07E-04	NO	NO	DOWN	NO
AT5G19230	0.80	0.37	4.47	0.52	2.49	2.74E-04	NO	NO	NO	NO
AT5G19290	32.36	0.88	44.93	1.36	0.47	2.33E-02	NO	NO	NO	NO
AT5G19320	19.14	0.41	29.78	0.38	0.64	8.63E-04	NO	NO	NO	NO
AT5G19780	41.64	0.68	58.19	0.85	0.48	1.31E-02	NO	NO	NO	NO
AT5G20350	24.08	0.20	32.74	0.37	0.44	3.09E-02	NO	NO	NO	NO
AT5G20400	10.76	0.66	18.82	0.53	0.81	9.58E-04	NO	NO	NO	NO
AT5G20730	11.55	0.15	16.51	0.47	0.52	4.30E-03	NO	YES	NO	NO
AT5G20960	1.40	0.14	6.67	0.08	2.26	0.00E+00	NO	NO	NO	NO
AT5G21170	6.73	0.55	10.90	0.13	0.70	1.70E-02	NO	NO	NO	NO
AT5G21326	21.10	0.31	31.81	0.24	0.59	2.45E-03	NO	NO	NO	NO
AT5G21940	7.19	0.55	14.14	0.59	0.98	4.89E-04	NO	NO	NO	NO
AT5G22060	86.47	0.05	117.46	2.01	0.44	2.62E-02	NO	NO	NO	NO
AT5G22090	32.27	0.71	54.09	0.43	0.75	4.46E-05	NO	NO	NO	NO
AT5G22380	0.88	0.16	11.33	0.79	3.70	4.20E-12	NO	YES	NO	NO
AT5G22430	0.04	0.01	14.26	3.68	8.41	2.18E-07	NO	NO	NO	NO
AT5G22460	0.38	0.09	3.06	0.37	3.02	2.60E-07	NO	NO	NO	NO
AT5G22510	25.19	0.26	35.74	0.27	0.50	8.71E-03	NO	NO	NO	NO
AT5G22555	0.47	0.21	6.02	1.08	3.67	6.89E-05	NO	NO	NO	NO
AT5G22570	1.27	0.63	7.83	0.49	2.62	2.22E-07	NO	YES	NO	NO
AT5G22580	59.10	1.15	89.04	2.02	0.59	3.85E-03	NO	NO	NO	DOWN
AT5G22850	5.51	0.05	8.15	0.04	0.57	4.53E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G23150	5.36	0.15	8.13	0.19	0.60	3.90E-03	NO	NO	NO	NO
AT5G23510	11.61	0.32	22.53	0.30	0.96	4.27E-06	NO	NO	NO	NO
AT5G23575	50.90	0.96	74.60	0.74	0.55	3.59E-03	NO	NO	NO	NO
AT5G23730	3.45	0.27	5.91	0.17	0.78	4.72E-02	NO	NO	NO	NO
AT5G23860	6.72	0.59	11.26	0.71	0.75	1.48E-03	NO	NO	NO	NO
AT5G24120	27.43	1.17	47.76	1.56	0.80	6.62E-06	NO	NO	NO	NO
AT5G24240	30.39	0.23	42.64	0.39	0.49	1.19E-02	NO	NO	NO	NO
AT5G24590	9.87	0.29	18.51	0.22	0.91	1.40E-05	NO	YES	NO	NO
AT5G24660	68.17	1.72	103.29	4.04	0.60	4.73E-03	NO	NO	NO	NO
AT5G24810	16.34	0.50	38.46	0.47	1.23	0.00E+00	NO	NO	NO	NO
AT5G25130	3.27	0.17	8.32	0.08	1.35	7.81E-06	NO	NO	NO	NO
AT5G25190	3.72	0.05	8.13	0.73	1.13	1.13E-02	NO	YES	NO	NO
AT5G25250	5.59	0.77	21.84	0.66	1.97	0.00E+00	NO	NO	NO	NO
AT5G25265	47.02	0.29	76.78	0.53	0.71	7.08E-05	NO	NO	NO	NO
AT5G25280	18.40	0.38	29.64	0.40	0.69	2.48E-04	NO	NO	NO	NO
AT5G25440	2.54	0.67	9.64	0.18	1.92	1.68E-07	NO	NO	NO	NO
AT5G25770	0.00	0.55	8.01	0.91	0.58	4.85E-02	NO	NO	DOWN	NO
AT5G25820	4.24	0.21	6.42	0.15	0.60	4.32E-02	NO	NO	NO	NO
AT5G25930	2.22	0.43	13.23	0.54	2.58	0.00E+00	NO	NO	NO	NO
AT5G26040	4.27	0.35	7.13	0.04	0.74	3.67E-02	NO	NO	NO	NO
AT5G26170	2.04	0.55	14.47	1.18	2.83	7.48E-07	NO	YES	NO	NO
AT5G26600	10.77	0.13	15.19	0.13	0.50	2.11E-02	NO	NO	NO	NO
AT5G26920	8.08	1.33	47.83	0.99	2.57	0.00E+00	NO	NO	NO	NO
AT5G27420	6.14	1.44	38.70	2.04	2.65	0.00E+00	NO	NO	NO	NO
AT5G27760	106.18	1.09	196.83	2.52	0.89	5.73E-07	NO	NO	NO	NO
AT5G28540	66.69	1.12	106.15	2.55	0.67	3.50E-04	NO	NO	NO	NO
AT5G28640	7.62	0.19	19.13	1.97	1.33	1.03E-07	NO	YES	NO	NO
AT5G28830	21.64	0.16	38.91	0.73	0.85	1.40E-05	NO	NO	NO	NO
AT5G34850	39.97	0.53	57.82	1.23	0.53	5.05E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G34940	3.32	0.32	8.24	0.14	1.31	3.32E-06	NO	NO	NO	NO
AT5G35080	17.10	0.05	33.66	0.83	0.98	1.63E-06	NO	NO	NO	NO
AT5G35200	19.06	0.36	27.50	0.27	0.53	7.60E-03	NO	NO	NO	NO
AT5G35370	1.32	0.10	2.41	0.09	0.86	3.46E-02	NO	NO	NO	NO
AT5G35735	52.00	0.89	101.16	0.93	0.96	2.92E-08	NO	NO	NO	NO
AT5G35950	1.23	0.04	3.53	0.19	1.52	2.96E-03	NO	NO	NO	NO
AT5G35970	46.00	0.41	80.10	0.64	0.80	1.05E-05	NO	NO	NO	NO
AT5G36160	36.84	0.63	50.20	0.09	0.45	2.89E-02	NO	NO	NO	NO
AT5G36880	30.56	0.44	47.60	0.49	0.64	6.57E-07	NO	NO	NO	NO
AT5G36970	0.36	0.16	5.43	1.74	3.92	2.25E-06	NO	NO	NO	NO
AT5G37260	62.78	1.03	87.60	1.12	0.48	1.55E-02	NO	YES	NO	NO
AT5G37540	1.98	0.09	5.50	0.53	1.48	1.04E-04	NO	NO	NO	NO
AT5G37600	173.68	2.54	265.55	1.91	0.61	1.73E-03	NO	NO	NO	NO
AT5G37770	11.78	0.57	24.39	1.62	1.05	1.47E-04	NO	NO	NO	NO
AT5G37830	13.66	0.29	20.05	0.29	0.55	3.39E-03	NO	NO	NO	NO
AT5G38210	5.38	0.46	12.11	0.70	1.17	1.46E-07	NO	NO	NO	NO
AT5G38212	2.88	0.15	7.82	0.51	1.44	4.11E-08	NO	NO	NO	NO
AT5G38310	0.84	0.18	3.85	0.26	2.20	6.13E-04	NO	NO	NO	NO
AT5G38410	194.55	3.68	276.48	4.42	0.51	7.43E-03	NO	NO	NO	NO
AT5G38650	60.49	0.19	81.91	2.33	0.44	3.61E-02	NO	NO	NO	NO
AT5G38900	18.06	2.04	90.89	2.33	2.33	0.00E+00	NO	NO	NO	NO
AT5G38990	16.12	0.50	29.03	0.30	0.85	2.57E-06	NO	NO	NO	NO
AT5G39080	3.48	0.21	9.86	0.57	1.50	5.40E-07	NO	NO	NO	NO
AT5G39110	0.93	0.29	3.06	0.75	1.71	4.15E-02	NO	NO	NO	NO
AT5G39660	11.05	0.37	16.87	0.37	0.61	1.56E-03	NO	YES	NO	NO
AT5G39860	2.32	0.33	6.00	1.28	1.37	1.91E-02	NEGATIVE	NO	NO	NO
AT5G40010	0.13	0.02	3.12	0.85	4.60	7.61E-09	NO	NO	NO	NO
AT5G40230	0.60	0.04	1.91	0.26	1.68	1.57E-02	NO	NO	NO	NO
AT5G40670	38.48	0.33	59.91	1.18	0.64	6.62E-04	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G40690	1.88	0.39	22.81	1.32	3.60	0.00E+00	NO	NO	NO	NO
AT5G40890	17.01	1.04	22.46	0.27	0.40	2.60E-02	NEGATIVE	NO	NO	NO
AT5G41761	24.20	1.79	42.72	1.14	0.82	1.69E-03		NO	NO	NO
AT5G42010	2.48	0.20	5.95	0.27	1.26	1.28E-05	NO	NO	NO	NO
AT5G42090	43.55	0.94	68.20	0.56	0.65	4.06E-04	NO	NO	NO	NO
AT5G42100	34.55	0.76	51.42	0.93	0.57	2.08E-04	NO	NO	NO	NO
AT5G42250	12.12	0.62	18.85	0.71	0.64	6.89E-03	NO	NO	NO	NO
AT5G42380	1.30	0.36	19.82	1.20	3.93	0.00E+00	NO	NO	NO	NO
AT5G42440	2.63	0.35	5.47	0.14	1.06	1.27E-02	NO	NO	NO	NO
AT5G42760	22.41	0.37	42.02	0.98	0.91	4.77E-07	NO	NO	NO	NO
AT5G42790	147.36	0.14	202.35	3.53	0.46	1.90E-02	NO	NO	NO	NO
AT5G42830	2.30	0.39	14.19	1.07	2.63	0.00E+00	NO	NO	NO	NO
AT5G42870	8.81	0.13	14.18	0.31	0.69	1.47E-05	NO	NO	NO	NO
AT5G42990	26.96	0.09	45.86	0.74	0.77	5.46E-04	NO	NO	NO	NO
AT5G43060	14.55	0.89	40.32	1.18	1.47	0.00E+00	NO	NO	NO	NO
AT5G43380	3.33	0.02	6.00	0.35	0.85	1.99E-02	NO	NO	NO	NO
AT5G43470	5.52	0.20	11.55	0.06	1.07	2.01E-09	NO	NO	NO	NO
AT5G43900	5.28	0.24	12.99	0.28	1.30	1.91E-14	NO	NO	NO	NO
AT5G44070	21.02	0.98	42.36	0.41	1.01	1.38E-08	NO	NO	NO	NO
AT5G44240	11.73	0.28	22.23	0.26	0.92	1.81E-07	NO	NO	NO	NO
AT5G44380	0.76	0.33	1.82	0.33	1.25	3.61E-02	NO	NO	NO	NO
AT5G44390	5.74	0.40	15.56	0.77	1.44	2.27E-10	NO	NO	NO	NO
AT5G44480	0.72	0.36	7.79	0.11	3.42	1.27E-11	NO	NO	NO	NO
AT5G44540	0.89	0.16	4.93	3.39	2.48	2.94E-02	NO	NO	NO	NO
AT5G44585	5.76	0.78	19.45	1.79	1.76	1.19E-03	NO	NO	NO	NO
AT5G44720	75.31	1.76	102.84	1.44	0.45	3.54E-03	NO	NO	NO	NO
AT5G44820	12.80	0.46	22.93	0.25	0.84	2.02E-04	NO	NO	NO	NO
AT5G44860	9.62	0.30	15.88	0.19	0.72	3.87E-03	NO	NO	NO	NO
AT5G44870	8.04	0.20	13.27	0.66	0.72	2.53E-04	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G44990	0.45	0.24	9.33	0.60	4.39	0.00E+00	NO	NO	NO	NO
AT5G45000	2.17	0.47	5.34	0.20	1.30	9.13E-04	NO	NO	NO	NO
AT5G45110	7.49	0.47	29.84	0.74	1.99	0.00E+00	NO	NO	NO	NO
AT5G46270	6.35	0.21	10.08	0.56	0.67	1.33E-03	NO	NO	NO	NO
AT5G46330	7.58	0.30	19.24	0.29	1.34	1.91E-14	NO	NO	NO	NO
AT5G46340	6.74	0.15	9.66	0.25	0.52	4.44E-02	NO	NO	NO	NO
AT5G46350	2.53	0.71	15.42	1.00	2.61	0.00E+00	NO	YES	NO	NO
AT5G46450	11.96	0.51	16.70	0.69	0.48	2.13E-02	NO	NO	NO	NO
AT5G46780	6.27	0.20	14.52	0.41	1.21	2.61E-05	NO	NO	NO	NO
AT5G46790	16.66	0.36	28.60	0.88	0.78	1.16E-03	NO	NO	NO	NO
AT5G47050	2.88	0.34	5.14	0.42	0.84	2.52E-02	NO	NO	NO	NO
AT5G47070	8.90	0.03	15.83	0.84	0.83	1.74E-04	NO	NO	NO	NO
AT5G47120	113.61	0.35	277.84	3.25	1.29	5.38E-14	NO	NO	NO	NO
AT5G47220	47.67	1.40	120.04	2.42	1.33	5.37E-14	NO	YES	NO	NO
AT5G47420	8.91	0.11	15.97	0.45	0.84	1.07E-03	NO	NO	NO	NO
AT5G47500	14.27	0.58	21.61	0.93	0.60	7.79E-03	NO	NO	NO	NO
AT5G47560	45.16	0.37	91.77	0.97	1.02	1.29E-09	NO	NO	NO	NO
AT5G47620	33.22	0.66	45.37	0.59	0.45	1.61E-03	NO	NO	NO	NO
AT5G47820	13.29	0.44	17.38	0.30	0.39	1.60E-02	NO	NO	NO	NO
AT5G47960	2.07	0.17	9.10	0.40	2.13	2.06E-06	NO	NO	NO	NO
AT5G48160	33.08	0.35	50.67	0.27	0.62	4.81E-04	NO	NO	NO	NO
AT5G48380	26.59	0.44	67.03	0.44	1.33	0.00E+00	NO	NO	NO	NO
AT5G48540	34.05	0.52	68.48	1.51	1.01	5.75E-08	NO	NO	NO	NO
AT5G48657	4.97	0.61	18.83	0.91	1.92	3.95E-10	NO	NO	NO	NO
AT5G48920	1.00	0.10	3.64	0.07	1.87	6.74E-03	NO	NO	NO	NO
AT5G48930	42.90	0.92	84.59	0.44	0.98	7.58E-09	NO	NO	NO	NO
AT5G49450	2.80	0.25	5.98	0.19	1.09	2.96E-02	NO	YES	NO	NO
AT5G49520	1.21	0.05	5.93	0.57	2.29	3.30E-09	NO	YES	NO	NO
AT5G49555	25.80	0.41	36.93	0.42	0.52	8.03E-03	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G49680	1.40	0.07	4.38	0.39	1.65	1.93E-14	NO	NO	NO	NO
AT5G49730	60.45	0.74	85.67	1.71	0.50	8.12E-03	NO	NO	NO	NO
AT5G49740	12.25	0.26	25.84	0.64	1.08	1.65E-09	NO	NO	NO	NO
AT5G49760	21.93	0.46	48.09	0.14	1.13	8.72E-12	NO	NO	NO	NO
AT5G49810	21.93	0.26	32.02	0.21	0.55	3.65E-03	NO	NO	NO	NO
AT5G50110	20.19	0.27	28.53	0.30	0.50	4.13E-02	NO	NO	NO	NO
AT5G50200	42.67	1.12	82.87	1.79	0.96	1.28E-07	NO	NO	NO	NO
AT5G50210	11.33	0.10	18.56	0.24	0.71	2.71E-04	NEGATIVE	NO	NO	NO
AT5G50460	160.23	4.09	257.73	4.63	0.69	2.77E-04	NO	NO	NO	NO
AT5G51070	35.89	0.55	65.38	0.80	0.87	4.15E-07	NO	NO	NO	NO
AT5G51190	1.21	0.01	4.10	0.14	1.76	5.33E-03	NO	YES	DOWN	NO
AT5G51290	10.64	0.34	18.05	0.37	0.76	1.48E-04	NO	NO	NO	NO
AT5G51630	2.22	0.09	6.54	0.51	1.56	1.89E-14	NO	NO	NO	NO
AT5G51770	7.79	0.30	12.94	0.13	0.73	1.53E-03	NO	NO	NO	NO
AT5G51830	45.33	1.43	84.19	1.52	0.89	2.74E-07	NO	NO	NO	NO
AT5G52160	0.00	0.00	3.03	1.77	Inf	2.67E-03	NO	NO	NO	NO
AT5G52250	0.45	0.10	2.14	0.22	2.26	7.58E-04	NO	NO	NO	NO
AT5G52390	2.13	0.13	18.68	3.91	3.13	0.00E+00	NO	NO	NO	NO
AT5G52540	39.48	0.39	60.71	0.36	0.62	6.92E-04	NO	NO	NO	NO
AT5G52730	0.56	0.23	4.87	0.48	3.11	6.34E-04	NO	NO	NO	NO
AT5G52760	3.72	1.12	59.88	1.50	4.01	0.00E+00	NO	NO	NO	NO
AT5G52810	10.61	0.57	27.76	0.85	1.39	5.32E-11	NO	NO	NO	NO
AT5G52870	4.59	0.05	8.44	0.06	0.88	8.73E-03	NO	NO	NO	NO
AT5G52900	5.43	0.27	15.64	0.42	1.53	1.20E-08	NO	NO	NO	NO
AT5G53110	2.65	0.33	9.21	0.25	1.80	2.15E-07	NO	NO	NO	NO
AT5G53170	30.48	0.40	41.70	0.35	0.45	2.23E-02	NO	NO	NO	NO
AT5G53370	63.42	1.14	121.27	1.21	0.94	5.31E-08	NO	NO	NO	NO
AT5G53550	29.86	0.38	61.63	1.26	1.05	0.00E+00	NO	NO	NO	NO
AT5G53760	2.34	0.03	4.10	0.08	0.81	1.27E-02	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G53830	2.58	0.23	5.33	0.24	1.05	4.56E-02	NO	NO	NO	NO
AT5G53870	0.84	0.34	3.34	0.52	2.00	3.25E-04	NO	NO	NO	NO
AT5G53890	2.71	0.08	4.17	0.14	0.62	3.27E-02	NO	NO	NO	NO
AT5G53970	18.88	0.56	28.31	0.79	0.58	4.40E-03	NO	NO	NO	NO
AT5G54080	11.61	0.04	16.49	0.16	0.51	2.86E-02	NO	NO	NO	NO
AT5G54100	10.82	0.35	19.21	0.66	0.83	1.59E-04	NO	NO	NO	NO
AT5G54160	45.05	1.97	184.13	0.49	2.03	0.00E+00	NO	NO	NO	NO
AT5G54380	4.77	0.23	9.85	0.24	1.05	1.55E-06	NO	NO	NO	NO
AT5G54470	59.49	0.39	96.58	0.22	0.70	1.03E-04	NO	NO	UP	NO
AT5G54490	6.35	1.01	33.22	0.94	2.39	2.37E-10	NO	NO	UP	NO
AT5G54510	7.54	0.22	11.11	0.74	0.56	2.05E-02	NO	NO	NO	NO
AT5G54690	0.44	0.05	1.22	0.10	1.46	3.65E-02	NO	NO	NO	NO
AT5G54710	26.47	0.48	86.02	1.90	1.70	0.00E+00	NO	NO	NO	NO
AT5G54860	8.76	0.24	29.24	1.00	1.74	0.00E+00	NO	NO	NO	NO
AT5G54920	4.63	0.09	7.21	0.12	0.64	2.36E-02	NO	NO	NO	NO
AT5G55100	11.82	0.16	16.67	0.50	0.50	2.41E-03	NO	NO	NO	NO
AT5G55300	13.31	0.38	17.75	0.20	0.42	2.75E-02	NO	NO	NO	NO
AT5G55520	4.46	0.15	6.81	0.29	0.61	1.43E-02	NO	NO	NO	NO
AT5G55600	12.12	0.24	16.49	0.28	0.44	9.12E-03	NO	NO	NO	NO
AT5G55730	12.41	0.90	18.23	1.41	0.55	7.60E-03	NEGATIVE	NO	NO	NO
AT5G55850	86.59	0.66	133.15	0.60	0.62	1.71E-06	NO	NO	NO	NO
AT5G55860	10.59	0.29	16.58	0.24	0.65	1.60E-03	NO	NO	NO	NO
AT5G56120	8.99	0.29	16.74	1.08	0.90	5.07E-04	NO	NO	NO	NO
AT5G56240	12.92	0.21	20.24	0.25	0.65	7.11E-06	NO	NO	NO	NO
AT5G56250	12.38	0.40	20.21	0.86	0.71	3.80E-06	NO	NO	NO	NO
AT5G56260	73.75	0.31	104.95	0.76	0.51	7.93E-03	NO	NO	NO	NO
AT5G56270	8.26	0.25	11.62	0.11	0.49	3.09E-02	NO	YES	NO	NO
AT5G56350	63.22	1.28	104.84	1.08	0.73	4.28E-05	NO	NO	NO	NO
AT5G56870	0.74	0.13	3.71	0.28	2.33	1.18E-08	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G57010	0.38	0.11	2.01	0.08	2.41	1.21E-04	NO	NO	NO	NO
AT5G57035	9.74	0.21	16.64	0.31	0.77	7.83E-05	NO	NO	NO	NO
AT5G57123	2.77	0.15	6.81	0.95	1.30	1.59E-02	NO	NO	NO	NO
AT5G57220	1.61	0.32	7.79	0.77	2.27	1.21E-10	NO	NO	NO	NO
AT5G57480	2.19	0.69	7.98	0.39	1.87	4.03E-08	NO	NO	NO	NO
AT5G57510	0.69	0.52	4.20	0.85	2.61	4.04E-02	NO	NO	NO	NO
AT5G57560	2.45	0.79	17.16	0.33	2.81	0.00E+00	NO	NO	DOWN	NO
AT5G57710	24.78	0.40	37.24	0.16	0.59	1.43E-03	NO	NO	NO	NO
AT5G57800	16.76	0.51	40.39	0.74	1.27	8.78E-14	NO	NO	NO	DOWN
AT5G58120	5.55	0.26	8.57	0.40	0.63	5.33E-03	NO	NO	NO	NO
AT5G58220	35.37	0.58	45.46	0.19	0.36	4.65E-02	NO	NO	NO	NO
AT5G58350	4.95	0.37	15.75	0.37	1.67	0.00E+00	NO	NO	NO	NO
AT5G58430	33.97	0.10	47.62	0.40	0.49	1.18E-02	NO	NO	NO	NO
AT5G58710	192.36	2.89	337.11	4.58	0.81	6.72E-06	NO	NO	NO	NO
AT5G58940	4.04	0.14	13.82	0.83	1.77	7.17E-12	NO	NO	NO	NO
AT5G59030	60.15	0.45	89.96	1.15	0.58	3.19E-03	NO	NO	NO	NO
AT5G59070	0.32	0.08	1.72	0.10	2.43	5.01E-04	NO	NO	NO	NO
AT5G59820	14.00	0.11	47.12	0.66	1.75	1.85E-14	NO	YES	NO	NO
AT5G59920	4.56	0.33	9.69	0.34	1.09	9.87E-06	NO	NO	NO	NO
AT5G59960	14.90	0.18	20.62	0.39	0.47	4.42E-02	NO	NO	NO	NO
AT5G60280	2.60	0.16	6.12	0.27	1.23	8.51E-05	NO	NO	NO	NO
AT5G60410	27.20	0.11	35.74	0.42	0.39	4.25E-02	NO	YES	NO	NO
AT5G60800	6.52	0.40	24.58	1.00	1.91	0.00E+00	NO	NO	NO	NO
AT5G60840	12.25	0.45	21.14	0.97	0.79	6.24E-03	NO	NO	NO	NO
AT5G60930	2.80	0.07	4.19	0.11	0.58	3.47E-02	NO	NO	NO	NO
AT5G60950	10.05	1.18	32.43	0.54	1.69	3.11E-13	NO	NO	NO	NO
AT5G61010	9.72	1.22	35.25	1.48	1.86	0.00E+00	NO	NO	NO	NO
AT5G61390	4.78	0.04	7.58	0.43	0.67	3.25E-02	NO	NO	NO	NO
AT5G61560	3.84	0.11	7.88	0.31	1.04	9.76E-07	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G61790	153.94	2.39	505.04	11.60	1.71	0.00E+00	NO	NO	NO	NO
AT5G61900	4.84	0.10	15.13	0.14	1.64	0.00E+00	NO	NO	NO	NO
AT5G62000	80.93	0.77	98.93	0.27	0.29	9.09E-03	NO	YES	NO	NO
AT5G62020	1.36	0.06	7.26	0.17	2.42	1.19E-09	NO	YES	NO	NO
AT5G62150	3.91	0.59	19.35	1.55	2.31	6.44E-07	NO	NO	NO	NO
AT5G62390	59.05	0.91	92.43	0.94	0.65	4.67E-04	NO	NO	NO	NO
AT5G62530	25.69	0.75	39.43	0.53	0.62	9.37E-04	NO	NO	NO	NO
AT5G62540	5.57	0.25	16.79	0.02	1.59	6.29E-07	NO	NO	NO	NO
AT5G62560	6.11	0.42	9.77	0.12	0.68	9.43E-03	NO	NO	NO	NO
AT5G62680	21.72	0.53	31.45	0.57	0.53	6.06E-03	NO	NO	NO	NO
AT5G62770	1.65	0.30	4.81	0.28	1.55	1.84E-03	NO	NO	NO	NO
AT5G63225	0.61	0.14	10.03	0.77	4.04	3.15E-08	NO	NO	NO	NO
AT5G63470	19.09	0.42	31.63	0.88	0.73	3.05E-04	NO	YES	NO	NO
AT5G63490	9.75	0.20	15.36	0.31	0.66	4.00E-03	NO	NO	NO	NO
AT5G63530	22.08	0.81	32.01	0.68	0.54	4.80E-03	NO	NO	NO	NO
AT5G63850	7.68	0.23	24.11	0.61	1.65	0.00E+00	NO	NO	NO	NO
AT5G63950	5.52	0.17	7.68	0.17	0.48	4.50E-02	NO	NO	NO	NO
AT5G63970	4.51	0.12	9.46	0.34	1.07	8.01E-05	NO	NO	NO	NO
AT5G64370	24.77	0.65	42.72	0.93	0.79	2.39E-05	NO	NO	NO	NO
AT5G64510	2.11	0.11	12.75	1.36	2.59	0.00E+00	NO	NO	NO	NO
AT5G64570	10.17	0.32	18.18	0.74	0.84	1.36E-05	NO	NO	DOWN	DOWN
AT5G64572	4.10	0.21	10.46	0.67	1.35	6.33E-06	NO	NO	NO	NO
AT5G64810	3.43	1.51	45.84	1.66	3.74	0.00E+00	NO	YES	NO	NO
AT5G64905	0.78	0.36	3.45	0.40	2.14	3.83E-02	NO	NO	NO	NO
AT5G65210	6.23	0.07	12.88	0.54	1.05	1.32E-07	NO	YES	NO	NO
AT5G65360	184.80	1.71	262.28	5.90	0.51	6.93E-03	NO	NO	NO	NO
AT5G65440	6.93	0.17	12.86	0.47	0.89	2.90E-10	NO	NO	NO	NO
AT5G65660	17.87	0.48	26.80	0.74	0.58	4.00E-02	NO	NO	NO	NO
AT5G66210	15.29	0.17	29.58	0.89	0.95	4.58E-12	NO	NO	NO	NO

Table A3.4 (cont'd)

AT5G66250	18.38	0.72	27.67	1.01	0.59	5.36E-04	NO	NO	NO	NO
AT5G66490	7.94	0.14	21.01	0.21	1.40	3.30E-04	NO	NO	NO	NO
AT5G66600	1.32	0.03	2.50	0.08	0.93	2.09E-02	NO	NO	NO	NO
AT5G66650	0.98	0.11	2.48	0.17	1.34	2.99E-02	NO	NO	NO	NO
AT5G66675	21.61	0.86	30.95	0.43	0.52	1.09E-02	NO	NO	NO	NO
AT5G66730	15.36	0.30	25.01	0.37	0.70	2.16E-04	NO	YES	NO	NO
AT5G66850	9.67	0.09	19.04	0.17	0.98	1.75E-07	NO	NO	NO	NO
AT5G66880	1.88	0.06	5.09	0.04	1.44	4.42E-04	NO	NO	DOWN	NO
AT5G66910	7.57	0.33	15.89	0.47	1.07	5.85E-08	NO	NO	NO	NO
AT5G67080	0.06	0.03	1.01	0.34	4.05	7.00E-03	NO	NO	NO	NO
AT5G67340	3.51	0.34	19.90	0.38	2.50	0.00E+00	NO	NO	NO	NO
AT5G67420	3.14	0.18	7.90	0.26	1.33	1.39E-05	NO	YES	NO	NO
AT5G67450	0.44	0.12	2.20	0.30	2.31	1.83E-03	NO	YES	NO	NO
AT5G67470	10.22	0.27	24.74	0.53	1.27	2.95E-13	NO	NO	NO	NO

Table A3.5. 294 Category 2 FTD Candidate Genes with higher expression in SW at 2 weeks of cold-acclimations. Genes are ordered by their AGI numbers. FPKM values are the average of three biological replicates (n=3). Genes were considered significantly different between SW and IT under non-acclimated conditions if they had a Benjamini-Hochberg corrected p-value (FDR P-VALUE) of <0.05 and a minimum of ≥ 3 FPKM in IT. The differential expression of each gene is also represented by the logarithm (base=2) of the fold change (LOG2.FC). This table also denotes if a gene was previously described as positively or negatively correlating with freezing tolerance (HANNAH;(6)), a transcription factor (TF), a gene previously described as up- or down-regulated by 1 week of cold (COS, (25)), identified as an up- or down-regulated component of the CBF regulon (CBF REGULON; (25)).

AGI	SW.1WK FPKM	SW.2WK FPKM	IT.1WK FPKM	IT.2WK FPKM	SW.IT.2WK LOG2FC	SW.IT.2 WK. FDR P-VALUE	HANNAH?	TF?	COS?	CBF REG?
AT1G01470	860.51	681.04	395.24	310.44	-1.13	4.71E-11	POSITIVE	NO	POSITIVE	UP
AT1G01670	4.42	4.79	1.48	1.77	-1.44	1.32E-03	NO	NO	NO	NO
AT1G02270	36.59	39.27	18.62	24.02	-0.71	1.62E-04	POSITIVE	NO	NO	NO
AT1G03420	54.53	87.54	6.17	6.24	-3.81	0.00E+00	NO	NO	NO	NO
AT1G04310	5.13	5.63	2.19	3.21	-0.81	4.32E-03	NO	NO	NO	NO
AT1G05320	0.00	12.32	7.15	6.21	-0.99	1.13E-06	NO	NO	NO	NO
AT1G05560	25.14	8.34	7.14	4.73	-0.82	5.34E-03	NO	NO	NO	NO
AT1G07702	0.00	319.82	646.35	278.55	-0.20	1.64E-02	NO	NO	NO	NO
AT1G09350	206.56	355.61	20.52	25.87	-3.78	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT1G10070	31.71	20.16	3.71	7.82	-1.37	1.67E-12	NO	NO	NO	NO
AT1G11580	115.76	35.72	30.41	16.33	-1.13	1.92E-10	NO	NO	NO	NO
AT1G11840	479.73	534.98	343.20	304.91	-0.81	0.00E+00	NO	NO	NO	NO
AT1G12350	39.94	41.62	18.57	20.08	-1.05	2.72E-08	NO	NO	NO	NO
AT1G12470	32.76	33.36	8.40	11.38	-1.55	0.00E+00	NO	NO	NO	NO
AT1G12550	11.72	12.29	4.21	5.05	-1.28	2.17E-05	NO	NO	NO	NO
AT1G13080	8.86	9.67	2.89	2.43	-1.99	3.65E-11	NO	NO	DOWN	NO

Table A3.5 (cont'd)

AT1G15125	90.96	117.61	20.60	55.00	-1.10	1.13E-09	NO	NO	NO	NO
AT1G16850	248.49	276.40	117.34	48.76	-2.50	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT1G20440	1586.75	1453.90	279.94	254.60	-2.51	0.00E+00	NO	NO	POSITIVE	UP
AT1G20450	758.00	712.70	196.22	182.55	-1.96	0.00E+00	NO	NO	POSITIVE	UP
AT1G20620	1910.93	1991.20	845.84	1107.69	-0.85	8.97E-05	NO	NO	NO	NO
AT1G20693	229.74	294.79	134.25	169.80	-0.80	2.94E-09	POSITIVE	YES	POSITIVE	NO
AT1G21130	74.08	94.42	31.10	54.55	-0.79	3.87E-06	NO	NO	NO	NO
AT1G22070	40.66	39.26	21.22	27.46	-0.52	9.18E-03	POSITIVE	YES	NO	NO
AT1G22090	11.46	4.70	0.06	0.00	#NAME?	1.01E-09	NO	NO	NO	NO
AT1G22360	14.46	15.20	6.76	8.43	-0.85	1.35E-04	NO	NO	NO	NO
AT1G22370	4.66	6.20	0.98	1.19	-2.38	2.29E-08	NO	NO	NO	NO
AT1G22403	25.06	22.12	11.54	9.38	-1.24	1.13E-07	NO	NO	NO	NO
AT1G23120	12.85	8.81	0.84	0.65	-3.76	2.99E-07	NO	NO	NO	NO
AT1G23800	30.15	19.22	8.99	9.74	-0.98	9.62E-07	NO	NO	NO	NO
AT1G23950	11.82	11.84	0.91	0.93	-3.68	0.00E+00	NO	NO	NO	NO
AT1G24370	91.85	88.30	0.11	0.24	-8.54	0.00E+00	NO	NO	NO	NO
AT1G24388	266.65	258.07	0.08	0.51	-8.99	0.00E+00	NO	NO	NO	NO
AT1G27200	31.50	49.45	10.57	14.74	-1.75	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT1G28330	40.21	50.56	11.34	14.56	-1.80	2.09E-12	NO	NO	NO	NO
AT1G28570	5.72	6.05	1.92	2.52	-1.26	8.58E-04	NO	NO	NO	NO
AT1G29050	19.81	14.62	8.38	8.27	-0.82	8.25E-04	NO	NO	NO	NO
AT1G29395	274.36	455.70	59.17	81.93	-2.48	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT1G33170	19.38	25.16	10.34	9.93	-1.34	1.53E-13	NO	NO	NO	NO
AT1G33970	65.18	68.70	39.41	47.81	-0.52	4.51E-04	NO	NO	NO	NO
AT1G36180	3.01	2.76	0.48	0.58	-2.25	0.00E+00	NO	NO	NO	NO
AT1G42705	4.43	4.66	0.02	0.03	-7.35	6.20E-11	NO	NO	NO	NO
AT1G46768	17.71	24.76	6.34	9.53	-1.38	3.00E-07	NO	YES	POSITIVE	UP
AT1G48100	17.68	21.78	8.46	7.98	-1.45	4.19E-13	NO	NO	NO	UP
AT1G48360	16.64	15.13	3.81	3.85	-1.97	0.00E+00	NO	NO	NO	NO

Table A3.5 (cont'd)

AT1G52720	13.55	14.24	4.47	6.57	-1.12	2.33E-03	NO	NO	DOWN	NO
AT1G53230	63.17	61.66	32.14	43.05	-0.52	6.50E-03	NO	YES	NO	NO
AT1G53580	61.84	51.09	27.46	29.98	-0.77	1.92E-07	POSITIVE	NO	NO	NO
AT1G54160	9.68	9.02	3.41	4.19	-1.11	2.71E-04	NO	YES	NO	NO
AT1G54890	5.08	0.86	0.30	0.11	-2.95	2.20E-02	NO	NO	NO	NO
AT1G56650	12.38	10.34	3.80	3.04	-1.77	2.03E-05	POSITIVE	YES	NO	NO
AT1G58300	3.47	8.24	0.52	2.37	-1.79	2.14E-04	NO	NO	NO	NO
AT1G58602	14.28	12.14	4.84	8.79	-0.47	1.91E-02	NO	NO	NO	NO
AT1G59950	16.46	5.90	0.38	0.37	-3.99	5.27E-07	NO	NO	NO	NO
AT1G60730	62.87	38.04	12.23	20.07	-0.92	9.68E-11	NO	NO	NO	NO
AT1G61800	93.26	112.94	35.70	48.16	-1.23	7.05E-14	NO	NO	NO	NO
AT1G62560	50.21	31.44	19.12	9.55	-1.72	0.00E+00	NO	NO	NO	NO
AT1G62570	49.71	82.13	14.01	13.92	-2.56	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT1G63350	4.30	3.67	0.01	0.01	-8.37	2.54E-07	NO	NO	NO	NO
AT1G64660	23.05	17.76	5.64	11.60	-0.61	5.86E-03	NO	NO	NO	NO
AT1G64760	11.68	15.90	5.21	5.68	-1.49	2.04E-11	NO	NO	NO	NO
AT1G64780	8.52	6.00	2.43	1.58	-1.93	1.49E-07	NO	NO	NO	NO
AT1G64890	17.73	21.38	7.90	11.83	-0.85	2.63E-05	NO	NO	POSITIVE	UP
AT1G65980	236.37	270.97	105.14	134.32	-1.01	8.23E-10	NO	NO	NO	NO
AT1G67265	3.90	4.83	0.87	1.67	-1.54	3.73E-02	NO	NO	NO	NO
AT1G68500	44.40	56.74	15.61	11.79	-2.27	1.04E-13	NO	NO	POSITIVE	UP
AT1G68600	15.27	20.41	2.30	6.53	-1.64	0.00E+00	NO	NO	NO	NO
AT1G69720	23.25	14.10	1.43	1.32	-3.41	0.00E+00	NO	NO	NO	NO
AT1G69890	21.94	20.69	4.40	6.55	-1.66	1.41E-11	NO	NO	NO	NO
AT1G70260	4.97	10.70	0.88	2.85	-1.91	1.81E-09	NO	NO	NO	NO
AT1G70645	14.14	14.74	0.00	0.00	#NAME?	0.00E+00	NO	NO	NO	NO
AT1G70830	159.30	82.01	59.25	51.81	-0.66	1.27E-02	NO	NO	NO	NO
AT1G71340	23.51	25.02	2.21	2.47	-3.34	0.00E+00	NO	NO	NO	NO
AT1G71360	13.66	17.15	7.05	9.28	-0.89	6.96E-06	NO	NO	NO	NO

Table A3.5 (cont'd)

AT1G72830	8.37	6.31	1.76	2.88	-1.13	3.94E-04	NO	YES	NO	NO
AT1G75370	50.02	63.64	25.50	36.75	-0.79	9.35E-06	NO	NO	POSITIVE	UP
AT1G76180	811.66	855.55	369.49	390.72	-1.13	0.00E+00	NO	NO	NO	NO
AT1G76520	111.41	68.90	34.18	45.27	-0.61	6.78E-05	NO	NO	NO	NO
AT1G76530	3.57	2.97	0.89	1.28	-1.22	2.82E-02	NO	NO	NO	NO
AT1G76580	16.90	20.00	6.47	8.75	-1.19	1.13E-11	NO	NO	POSITIVE	UP
AT1G76590	47.44	51.12	18.99	19.72	-1.37	1.04E-13	POSITIVE	YES	POSITIVE	UP
AT1G76820	2.76	3.58	0.16	0.23	-3.95	0.00E+00	NO	NO	NO	NO
AT1G77120	381.80	355.69	150.04	111.48	-1.67	0.00E+00	NO	NO	POSITIVE	NO
AT1G78070	117.52	140.91	45.46	43.89	-1.68	0.00E+00	NO	NO	POSITIVE	UP
AT1G80130	293.11	554.44	153.50	213.76	-1.38	3.67E-14	POSITIVE	NO	POSITIVE	UP
AT1G80570	3.70	4.21	0.45	0.98	-2.10	4.21E-07	NO	NO	NO	NO
AT2G05380	2196.17	2680.92	529.44	1140.20	-1.23	4.18E-09	POSITIVE	NO	NO	NO
AT2G05510	25.45	8.77	3.06	0.87	-3.34	5.61E-08	NO	NO	NO	NO
AT2G12190	6.12	9.06	0.78	0.93	-3.29	1.87E-14	NO	NO	NO	NO
AT2G15050	98.77	122.57	23.52	17.40	-2.82	0.00E+00	NO	NO	DOWN	NO
AT2G15970	1564.76	1562.47	590.27	517.44	-1.59	0.00E+00	NO	NO	POSITIVE	UP
AT2G16700	140.34	151.38	42.67	37.97	-2.00	0.00E+00	NO	NO	NO	UP
AT2G16790	4.99	3.53	0.84	0.91	-1.96	3.37E-03	POSITIVE	NO	NO	NO
AT2G17280	25.98	31.01	11.91	13.10	-1.24	3.14E-09	NO	NO	NO	NO
AT2G17840	143.48	201.80	76.61	111.57	-0.86	1.10E-06	POSITIVE	NO	POSITIVE	UP
AT2G18110	88.39	80.23	19.27	18.10	-2.15	0.00E+00	NO	NO	NO	NO
AT2G19810	27.19	60.23	11.08	14.26	-2.08	0.00E+00	NO	YES	NO	NO
AT2G21620	384.16	262.42	144.60	155.70	-0.75	2.19E-05	POSITIVE	NO	POSITIVE	NO
AT2G21830	1.05	1.03	0.25	0.19	-2.46	3.46E-03	NO	NO	NO	NO
AT2G21860	26.96	28.21	12.70	15.56	-0.86	8.85E-06	NO	NO	NO	NO
AT2G22590	7.67	5.03	1.30	0.78	-2.68	1.13E-07	NO	NO	NO	NO
AT2G22660	39.60	42.29	14.10	21.58	-0.97	6.09E-09	NO	NO	NO	NO
AT2G25510	8645.94	10643.80	25.11	40.26	-8.05	0.00E+00	NO	NO	NO	NO

Table A3.5 (cont'd)

AT2G26010	154.12	237.06	5.34	66.62	-1.83	0.00E+00	NO	NO	NO	NO
AT2G27360	16.91	13.50	5.04	5.83	-1.21	3.39E-06	NO	NO	NO	NO
AT2G28550	124.43	155.52	79.16	77.01	-1.01	0.00E+00	NO	YES	NO	NO
AT2G28900	983.81	1399.48	424.69	418.81	-1.74	0.00E+00	NO	NO	POSITIVE	UP
AT2G29420	69.20	29.94	16.50	16.50	-0.86	2.25E-04	NO	NO	NO	NO
AT2G33830	770.26	840.49	310.33	457.27	-0.88	4.66E-07	NO	NO	NO	NO
AT2G34655	15.00	13.25	1.64	3.16	-2.07	1.31E-07	NO	NO	NO	NO
AT2G36750	18.28	13.69	6.88	6.50	-1.08	2.88E-05	NO	NO	NO	NO
AT2G39850	18.21	5.47	9.57	2.53	-1.11	8.11E-05	NO	NO	DOWN	NO
AT2G40010	19.08	15.02	0.08	0.04	-8.39	0.00E+00	NO	NO	NO	NO
AT2G41010	21.54	23.95	8.92	15.99	-0.58	2.54E-02	NO	NO	NO	NO
AT2G43500	29.93	32.48	7.46	8.71	-1.90	0.00E+00	NO	YES	NO	NO
AT2G46440	10.37	12.40	0.12	0.17	-6.18	0.00E+00	NO	NO	NO	NO
AT2G46640	13.72	11.32	5.53	5.66	-1.00	5.32E-04	NO	NO	NO	NO
AT2G47180	78.66	46.04	32.40	33.14	-0.47	1.80E-02	NO	NO	POSITIVE	NO
AT2G47890	55.55	54.56	25.76	26.51	-1.04	1.43E-09	NO	YES	POSITIVE	UP
AT3G05660	12.07	17.35	4.81	8.32	-1.06	9.44E-09	POSITIVE	NO	POSITIVE	UP
AT3G05880	500.06	397.98	255.71	207.90	-0.94	5.80E-08	NO	NO	NO	NO
AT3G07320	29.83	36.52	12.44	16.28	-1.17	3.73E-11	NO	NO	NO	NO
AT3G09030	10.86	11.55	4.69	7.64	-0.60	2.10E-02	NO	NO	NO	NO
AT3G12500	88.88	14.23	2.97	2.32	-2.62	3.08E-12	NO	NO	NO	NO
AT3G13310	139.98	99.33	73.66	67.80	-0.55	4.37E-03	NO	NO	NO	NO
AT3G14150	26.04	33.95	9.28	7.60	-2.16	0.00E+00	NO	NO	NO	NO
AT3G14940	2.95	3.54	0.76	0.44	-3.02	6.29E-12	NO	NO	NO	NO
AT3G17130	17.49	20.53	5.51	4.50	-2.19	1.29E-06	NO	NO	POSITIVE	UP
AT3G19000	10.79	9.13	0.19	0.11	-6.32	7.52E-13	NO	NO	DOWN	NO
AT3G20100	50.62	49.57	9.41	13.66	-1.86	0.00E+00	NO	NO	NO	NO
AT3G21500	5.26	2.57	1.11	1.21	-1.09	1.66E-02	NO	NO	NO	NO
AT3G22121	284.13	267.25	127.46	138.18	-0.95	7.73E-09	NO	NO	NO	NO

Table A3.5 (cont'd)

AT3G22142	62.20	68.17	15.80	15.85	-2.10	0.00E+00	NO	NO	NO	NO
AT3G22840	290.33	502.14	155.08	275.06	-0.87	9.46E-07	NO	NO	POSITIVE	NO
AT3G26200	4.37	4.70	1.26	2.37	-0.99	1.09E-02	NO	NO	NO	NO
AT3G26280	21.27	30.61	4.24	5.35	-2.52	0.00E+00	NO	NO	NO	NO
AT3G26460	8.02	14.62	1.96	0.56	-4.71	2.35E-10	NO	NO	NO	NO
AT3G26740	395.85	342.98	210.41	232.47	-0.56	1.99E-03	NO	NO	NO	NO
AT3G27210	61.39	59.15	33.67	31.14	-0.93	5.83E-07	NO	NO	POSITIVE	UP
AT3G28050	38.00	36.73	15.92	18.24	-1.01	4.87E-08	NO	NO	DOWN	NO
AT3G28310	66.01	102.25	8.89	19.27	-2.41	5.94E-10	NO	NO	NO	NO
AT3G28320	66.81	100.01	8.45	20.41	-2.29	0.00E+00	NO	NO	NO	NO
AT3G28740	51.78	22.67	13.17	7.37	-1.62	1.86E-14	NO	NO	NO	NO
AT3G28880	3.16	2.37	0.66	0.55	-2.12	2.98E-05	NO	NO	NO	NO
AT3G28950	21.18	22.38	3.62	6.36	-1.82	8.41E-10	NO	NO	NO	NO
AT3G32230	1.01	0.86	0.07	0.14	-2.66	2.85E-05	NO	NO	NO	NO
AT3G42658	67.13	104.06	32.11	42.18	-1.30	5.38E-14	NO	NO	NO	NO
AT3G42670	4.84	6.40	1.84	2.54	-1.34	3.09E-09	NO	NO	NO	NO
AT3G43270	17.69	14.11	8.23	9.50	-0.57	1.07E-02	NO	NO	NO	NO
AT3G43520	139.84	124.49	67.75	58.99	-1.08	1.97E-10	NO	NO	NO	NO
AT3G44430	56.06	48.19	23.73	15.08	-1.68	0.00E+00	NO	NO	NO	NO
AT3G44620	49.51	51.93	17.68	27.22	-0.93	5.26E-08	NO	NO	NO	NO
AT3G44670	11.85	10.44	2.07	3.25	-1.69	0.00E+00	NO	NO	NO	NO
AT3G45600	195.08	188.10	67.28	62.81	-1.58	0.00E+00	NO	NO	NO	UP
AT3G46370	5.16	5.55	0.02	0.03	-7.76	3.32E-12	NO	NO	NO	NO
AT3G46490	6.08	5.35	0.64	0.33	-4.01	3.80E-07	NO	NO	NO	DOWN
AT3G46900	11.73	16.32	0.87	3.25	-2.33	7.17E-07	NO	NO	NO	NO
AT3G49130	6.46	0.39	0.08	0.00	#NAME?	4.62E-02	NO	NO	NO	NO
AT3G50660	15.34	20.44	6.46	5.65	-1.85	0.00E+00	NO	NO	NO	NO
AT3G50970	1609.21	1188.91	341.83	260.26	-2.19	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT3G52180	189.78	255.58	105.04	85.09	-1.59	0.00E+00	POSITIVE	NO	NO	NO

Table A3.5 (cont'd)

AT3G52340	33.54	40.29	19.25	19.78	-1.03	3.11E-13	NO	NO	NO	NO
AT3G53990	279.86	311.84	114.72	174.88	-0.83	1.09E-06	NO	NO	POSITIVE	UP
AT3G55760	41.67	43.58	13.09	11.50	-1.92	0.00E+00	NO	NO	POSITIVE	UP
AT3G55940	9.05	11.01	2.48	3.51	-1.65	7.06E-10	POSITIVE	NO	NO	UP
AT3G57460	3.48	1.36	0.05	0.06	-4.39	6.99E-04	NO	NO	NO	NO
AT3G57470	6.32	6.45	0.01	0.01	-9.02	6.31E-11	NO	NO	NO	NO
AT3G61010	5.86	5.94	0.04	0.02	-8.57	1.16E-07	NO	NO	NO	NO
AT3G62740	11.79	8.31	2.01	1.13	-2.88	2.66E-11	NO	NO	POSITIVE	UP
AT4G01390	27.40	11.43	2.02	1.18	-3.27	7.05E-10	NO	NO	NO	NO
AT4G01500	3.12	3.28	0.47	0.37	-3.16	2.24E-06	NO	YES	NO	NO
AT4G01883	31.74	28.09	14.55	11.71	-1.26	7.97E-09	NO	NO	NO	NO
AT4G01985	43.10	39.78	22.05	25.57	-0.64	8.44E-04	NO	NO	NO	NO
AT4G04020	140.38	104.74	64.25	46.93	-1.16	2.33E-12	NO	NO	NO	NO
AT4G04340	59.98	72.58	35.33	33.33	-1.12	7.37E-13	NO	NO	NO	NO
AT4G04426	1.37	1.62	0.02	0.02	-6.36	1.32E-05	NO	NO	NO	NO
AT4G04925	35.49	48.77	10.47	14.77	-1.72	7.59E-12	NO	NO	NO	NO
AT4G05090	42.87	49.63	23.20	25.25	-0.97	4.92E-08	NO	NO	NO	NO
AT4G06746	39.58	62.79	7.68	17.34	-1.86	3.68E-14	NO	YES	NO	NO
AT4G08390	366.40	269.16	178.17	195.65	-0.46	2.69E-03	NO	NO	NO	NO
AT4G09020	119.34	208.91	52.11	63.04	-1.73	0.00E+00	POSITIVE	NO	NO	NO
AT4G09760	42.74	51.58	20.32	23.53	-1.13	1.89E-14	NO	NO	NO	NO
AT4G10955	0.18	0.85	1.88	0.17	-2.36	3.35E-03	NO	NO	NO	NO
AT4G11410	14.71	15.88	6.93	9.76	-0.70	5.67E-03	NO	NO	NO	NO
AT4G14270	73.23	97.47	22.41	34.73	-1.49	0.00E+00	NO	NO	NO	NO
AT4G14580	49.50	69.76	10.86	13.06	-2.42	0.00E+00	NO	NO	NO	NO
AT4G15130	51.50	63.98	24.88	27.19	-1.23	3.10E-12	NO	NO	POSITIVE	NO
AT4G15450	41.96	43.04	16.95	19.21	-1.16	3.94E-10	NO	NO	NO	NO
AT4G15910	324.05	169.33	57.83	28.33	-2.58	0.00E+00	NO	NO	NO	NO
AT4G16146	71.67	76.17	35.84	33.41	-1.19	2.38E-09	NO	NO	NO	NO

Table A3.5 (cont'd)

AT4G16770	6.40	6.99	1.47	1.52	-2.20	2.16E-06	NO	NO	NO	NO
AT4G16860	10.83	11.75	3.69	5.46	-1.11	1.81E-08	NO	NO	NO	NO
AT4G16890	15.95	17.33	8.26	12.80	-0.44	3.18E-02	NO	NO	NO	NO
AT4G18070	78.00	87.26	44.90	40.40	-1.11	0.00E+00	NO	NO	NO	NO
AT4G18422	432.66	441.74	214.21	128.37	-1.78	0.00E+00	NO	NO	NO	NO
AT4G19500	7.58	8.92	0.29	0.28	-5.01	0.00E+00	NO	NO	NO	NO
AT4G19510	9.68	10.76	0.56	0.82	-3.71	0.00E+00	NO	NO	NO	NO
AT4G20310	6.39	6.12	3.00	2.46	-1.31	1.40E-05	NO	NO	NO	NO
AT4G23600	306.42	161.04	139.73	76.29	-1.08	0.00E+00	NO	NO	NO	NO
AT4G23670	218.14	153.99	116.61	37.05	-2.06	0.00E+00	NO	NO	DOWN	NO
AT4G23920	23.15	27.67	9.43	7.47	-1.89	0.00E+00	NO	NO	POSITIVE	UP
AT4G24220	123.54	159.79	58.60	113.38	-0.49	9.56E-03	NO	NO	NO	UP
AT4G25433	24.84	27.81	4.75	3.80	-2.87	5.41E-13	NO	NO	NO	NO
AT4G26255	13.33	12.39	1.06	1.28	-3.28	1.68E-13	NO	NO	NO	NO
AT4G27050	61.39	51.93	17.22	13.57	-1.94	0.00E+00	NO	NO	NO	NO
AT4G27070	21.11	17.59	6.40	7.24	-1.28	5.37E-10	NO	NO	NO	NO
AT4G27300	38.86	42.74	17.91	22.41	-0.93	3.01E-07	NO	NO	NO	NO
AT4G27820	26.99	25.91	9.04	6.87	-1.91	0.00E+00	NO	NO	NO	NO
AT4G27900	36.63	36.02	14.78	19.58	-0.88	6.66E-07	NO	YES	NO	NO
AT4G27970	7.02	7.58	2.42	2.70	-1.49	7.74E-06	NO	NO	NO	NO
AT4G28220	40.47	50.86	22.04	26.26	-0.95	2.72E-08	NO	NO	NO	NO
AT4G30830	16.39	28.21	2.86	2.86	-3.30	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT4G31620	2.97	3.92	0.92	1.17	-1.74	9.52E-05	NO	YES	NO	NO
AT4G33070	34.90	33.52	6.76	7.35	-2.19	0.00E+00	NO	NO	POSITIVE	NO
AT4G33180	12.63	11.48	4.53	4.75	-1.27	2.32E-05	NO	NO	NO	NO
AT4G34530	7.88	10.98	3.13	1.24	-3.14	4.19E-13	NO	YES	NO	NO
AT4G35300	73.80	100.64	42.49	48.10	-1.07	0.00E+00	POSITIVE	NO	POSITIVE	NO
AT4G36010	24.92	24.96	12.24	9.93	-1.33	1.82E-11	NO	NO	POSITIVE	UP
AT4G37000	51.43	60.46	25.14	27.52	-1.14	4.72E-10	NO	NO	NO	NO

Table A3.5 (cont'd)

AT4G38580	141.00	144.43	67.73	78.34	-0.88	6.67E-07	NO	NO	POSITIVE	NO
AT4G39210	27.59	45.44	12.45	18.03	-1.33	0.00E+00	POSITIVE	NO	NO	NO
AT4G39800	406.75	514.02	199.33	216.45	-1.25	3.00E-10	NO	NO	POSITIVE	NO
AT5G01600	335.37	344.75	93.33	147.02	-1.23	3.32E-12	NO	NO	NO	NO
AT5G02130	29.98	29.53	13.36	13.82	-1.10	1.95E-08	POSITIVE	NO	NO	NO
AT5G03350	180.08	302.99	56.15	109.48	-1.47	0.00E+00	NO	NO	NO	NO
AT5G05270	96.18	104.41	56.56	46.33	-1.17	0.00E+00	NO	NO	NO	NO
AT5G07010	21.10	36.05	2.52	2.44	-3.88	0.00E+00	NO	NO	NO	NO
AT5G08640	94.23	86.83	42.40	35.48	-1.29	3.61E-14	NO	NO	NO	NO
AT5G10140	145.95	86.18	41.81	28.39	-1.60	0.00E+00	NO	YES	NO	NO
AT5G13750	34.18	18.68	12.68	11.19	-0.74	9.00E-05	POSITIVE	NO	NO	NO
AT5G14550	51.44	98.97	32.49	43.51	-1.19	0.00E+00	NO	NO	NO	NO
AT5G14920	299.20	292.91	136.81	112.41	-1.38	0.00E+00	NO	NO	POSITIVE	NO
AT5G17040	4.53	5.41	0.84	1.17	-2.21	2.76E-06	NO	NO	NO	NO
AT5G17190	166.31	157.65	75.06	96.36	-0.71	7.05E-05	NO	NO	NO	NO
AT5G17460	126.59	138.84	44.65	37.01	-1.91	0.00E+00	NO	NO	NO	NO
AT5G17700	20.09	20.54	5.80	5.38	-1.93	0.00E+00	NO	NO	NO	NO
AT5G20750	4.35	4.06	0.01	0.02	-7.84	1.84E-10	NO	NO	NO	NO
AT5G20830	281.83	314.66	75.39	107.11	-1.55	0.00E+00	POSITIVE	NO	POSITIVE	NO
AT5G23240	25.83	35.71	11.63	25.31	-0.50	1.41E-02	NO	NO	NO	NO
AT5G23820	50.35	23.88	15.00	10.08	-1.24	4.12E-06	NO	NO	DOWN	NO
AT5G24160	17.92	34.00	1.27	3.63	-3.23	0.00E+00	NO	NO	DOWN	NO
AT5G24210	69.59	52.95	7.63	26.16	-1.02	2.18E-08	NO	NO	NO	DOWN
AT5G25110	184.09	334.24	11.96	13.19	-4.66	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT5G26770	18.56	18.11	8.91	11.04	-0.71	4.10E-04	NO	NO	NO	NO
AT5G26970	15.23	18.89	2.70	4.84	-1.96	7.92E-03	NO	NO	NO	NO
AT5G26980	1.48	1.97	0.37	0.68	-1.53	6.74E-03	NO	NO	NO	NO
AT5G27930	28.95	31.49	15.44	14.84	-1.09	1.87E-11	NO	NO	POSITIVE	UP
AT5G28913	11.63	12.76	0.04	0.04	-8.32	3.05E-07	NO	NO	NO	NO

Table A3.5 (cont'd)

AT5G30495	50.14	49.94	28.33	19.95	-1.32	3.11E-13	NO	NO	NO	NO
AT5G32460	4.56	2.68	0.26	0.20	-3.77	1.74E-06	NO	NO	NO	NO
AT5G35935	43.62	53.82	0.12	0.13	-8.66	0.00E+00	NO	NO	NO	NO
AT5G38120	13.96	12.38	5.53	5.41	-1.19	8.37E-07	NO	NO	NO	NO
AT5G38250	2.46	1.91	0.12	0.12	-4.03	8.56E-06	NO	NO	NO	NO
AT5G39340	31.94	37.62	15.21	16.11	-1.22	8.04E-08	NO	NO	NO	NO
AT5G39610	14.34	16.65	4.04	8.68	-0.94	2.08E-04	NO	YES	NO	NO
AT5G40390	33.15	39.46	16.34	17.26	-1.19	1.55E-12	NO	NO	POSITIVE	NO
AT5G41700	135.29	140.04	62.15	71.14	-0.98	1.19E-11	NO	NO	NO	NO
AT5G41740	3.18	3.23	0.15	0.43	-2.92	7.04E-14	NO	NO	NO	NO
AT5G42570	107.31	101.15	56.93	72.73	-0.48	2.02E-02	POSITIVE	NO	POSITIVE	NO
AT5G42825	51.11	100.46	18.94	27.90	-1.85	0.00E+00	NO	NO	NO	NO
AT5G43950	8.42	11.05	3.02	3.72	-1.57	3.12E-10	NO	NO	NO	NO
AT5G44400	41.21	26.99	11.16	10.51	-1.36	9.32E-13	NO	NO	NO	NO
AT5G44565	298.94	294.16	54.54	63.70	-2.21	0.00E+00	NO	NO	NO	NO
AT5G44590	22.35	57.70	8.09	17.95	-1.68	0.00E+00	NO	NO	NO	NO
AT5G45470	10.46	13.21	5.33	9.74	-0.44	4.53E-02	NO	NO	NO	NO
AT5G45490	40.54	63.06	0.12	0.25	-7.97	0.00E+00	NO	NO	NO	NO
AT5G45500	6.48	9.78	2.13	4.96	-0.98	2.95E-06	NO	NO	NO	NO
AT5G45510	47.41	65.79	8.21	13.44	-2.29	0.00E+00	NO	NO	NO	NO
AT5G47130	19.94	15.92	0.79	0.87	-4.19	6.92E-09	NO	NO	NO	NO
AT5G47810	9.73	10.00	0.62	1.04	-3.26	0.00E+00	NO	NO	NO	NO
AT5G48490	550.57	566.92	280.57	166.41	-1.77	0.00E+00	NEGATIVE	NO	NO	NO
AT5G48530	5.30	2.85	0.77	0.63	-2.18	1.24E-03	NO	NO	NO	NO
AT5G48670	3.90	5.05	0.69	1.10	-2.20	5.75E-05	NO	YES	NO	NO
AT5G48880	73.63	64.90	35.35	33.05	-0.97	5.36E-14	NO	NO	NO	NO
AT5G49120	3.78	6.46	0.72	2.12	-1.61	8.65E-03	NO	NO	NO	NO
AT5G49330	36.66	42.76	16.80	12.22	-1.81	0.00E+00	NO	YES	NO	NO
AT5G49630	42.45	50.73	11.09	26.13	-0.96	3.09E-08	NO	NO	NO	NO

Table A3.5 (cont'd)

AT5G49650	52.13	57.42	23.78	29.79	-0.95	3.32E-08	NO	NO	NO	NO
AT5G50950	18.14	21.79	8.84	8.85	-1.30	1.35E-11	NO	NO	NO	NO
AT5G52310	1276.59	1702.55	232.93	238.95	-2.83	0.00E+00	POSITIVE	NO	POSITIVE	UP
AT5G53120	34.48	25.92	16.92	17.75	-0.55	2.62E-03	NO	NO	NO	NO
AT5G53420	114.21	167.27	40.23	60.66	-1.46	0.00E+00	NO	YES	NO	NO
AT5G54190	75.17	158.76	18.54	29.45	-2.43	0.00E+00	NO	NO	POSITIVE	NO
AT5G54960	95.77	129.97	41.06	76.58	-0.76	1.27E-05	NO	NO	NO	NO
AT5G55250	5.55	6.28	1.91	1.94	-1.69	2.09E-05	NO	NO	NO	NO
AT5G57110	62.10	71.56	33.91	38.11	-0.91	2.43E-07	NO	NO	POSITIVE	NO
AT5G57380	1.90	3.70	0.48	1.45	-1.36	4.74E-04	NO	NO	NO	NO
AT5G59130	6.94	5.31	1.56	0.99	-2.42	4.11E-11	NO	NO	NO	NO
AT5G59320	12.57	15.11	38.55	7.70	-0.97	1.20E-02	NO	NO	NO	NO
AT5G59670	5.70	9.46	2.44	3.32	-1.51	2.96E-11	NO	NO	NO	NO
AT5G59732	3.72	3.84	0.29	0.67	-2.51	4.11E-06	NO	NO	NO	NO
AT5G65060	40.27	37.23	9.41	9.32	-2.00	0.00E+00	NO	YES	NO	NO
AT5G66400	77.83	11.85	15.22	5.96	-0.99	5.83E-03	POSITIVE	NO	NO	NO
AT5G66480	3.41	2.61	0.87	0.93	-1.48	6.49E-03	NO	NO	NO	NO

Table A3.6. 504 Category 2 FTD Candidate Genes with higher expression in IT at 2 weeks of cold-acclimations. Genes are ordered by their AGI numbers. FPKM values are the average of three biological replicates (n=3). Genes were considered significantly different between SW and IT under non-acclimated conditions if they had a Benjamini-Hochberg corrected p-value (FDR P-VALUE) of <0.05 and a minimum of ≥ 3 FPKM in IT. The differential expression of each gene is also represented by the logarithm (base=2) of the fold change (LOG2.FC). This table also denotes if a gene was previously described as positively or negatively correlating with freezing tolerance (HANNAH;(6)), a transcription factor (TF), a gene previously described as up- or down-regulated by 1 week of cold (COS, (25)), identified as an up- or down-regulated component of the CBF regulon (CBF REGULON; (25)).

AGI	SW.1WK FPKM	SW.2WK FPKM	IT.1WK FPKM	IT.2WK FPKM	SW.IT.2WK.		HANNAH?	TF?	COS?	CBF REG?
					SW.IT.2WK LOG2FC	FDR P- VALUE				
AT1G01480	8.34	0.32	1.91	2.39	2.91	2.24E-07	NO	NO	NO	NO
AT1G01620	42.30	57.78	123.45	111.61	0.95	2.17E-08	NEGATIVE	NO	NO	NO
AT1G03870	11.15	9.32	27.33	20.77	1.16	4.74E-06	NO	NO	NO	NO
AT1G03905	6.63	4.45	16.31	19.80	2.15	3.64E-14	NO	NO	NO	NO
AT1G03940	0.28	0.09	1.36	0.74	3.01	3.42E-03	NO	NO	NO	NO
AT1G04250	15.91	13.52	33.42	24.95	0.88	1.22E-04	NO	YES	NO	NO
AT1G04680	15.13	17.36	33.66	27.90	0.68	3.65E-04	NO	NO	NO	NO
AT1G05550	0.60	0.33	2.30	2.38	2.85	2.94E-05	NO	NO	NO	NO
AT1G05680	27.57	0.28	1.84	4.26	3.92	1.85E-09	NO	NO	NO	NO
AT1G07570	14.32	14.56	28.20	31.08	1.09	1.53E-13	NO	NO	NO	NO
AT1G07610	23.26	13.71	96.25	59.98	2.13	9.30E-12	NO	NO	NO	NO
AT1G08610	4.53	3.76	11.30	7.03	0.90	1.37E-03	NO	NO	NO	NO
AT1G09420	2.62	2.00	15.93	16.47	3.04	0.00E+00	NO	NO	NO	NO
AT1G09932	10.59	3.04	47.32	52.92	4.12	0.00E+00	NO	NO	NO	NO
AT1G10340	0.04	0.04	3.04	9.81	8.12	6.32E-13	NO	NO	NO	NO
AT1G10470	45.17	41.35	85.36	65.58	0.67	3.68E-04	NO	NO	NO	NO
AT1G10585	31.14	2.18	7.09	9.90	2.18	5.22E-06	NO	YES	NO	NO

Table A3.6 (cont'd)

AT1G11175	0.05	0.00	6.19	9.83	Inf	2.31E-11	NO	NO	NO	NO
AT1G12220	0.02	0.01	4.47	5.55	8.94	4.47E-08	NO	NO	NO	NO
AT1G13350	1.96	1.54	11.14	10.91	2.82	0.00E+00	NO	NO	NO	NO
AT1G13470	23.26	23.68	79.09	164.60	2.80	0.00E+00	NO	NO	NO	NO
AT1G13650	0.03	0.00	2.10	0.85	Inf	3.36E-03	NO	NO	DOWN	NO
AT1G14700	0.54	0.55	6.19	4.43	3.02	3.28E-09	NEGATIVE	NO	DOWN	NO
AT1G14880	191.11	286.21	548.89	980.29	1.78	0.00E+00	NO	NO	NO	NO
AT1G15010	7.71	0.94	1.47	5.08	2.44	4.41E-03	NO	NO	NO	NO
AT1G15870	3.63	2.49	12.68	6.98	1.48	8.34E-04	NO	NO	NO	NO
AT1G15960	0.44	0.89	2.06	3.23	1.86	1.74E-05	NO	NO	NO	NO
AT1G15980	64.43	79.74	121.81	118.60	0.57	1.89E-03	NO	NO	NO	NO
AT1G16510	4.82	5.13	19.10	22.54	2.13	5.36E-12	NO	NO	NO	NO
AT1G16820	0.08	0.27	2.50	4.25	4.00	1.92E-03	NO	NO	NO	NO
AT1G17170	315.23	64.23	71.61	137.50	1.10	1.94E-10	POSITIVE	NO	NO	NO
AT1G17180	14.44	0.44	2.03	4.20	3.24	1.72E-05	NO	NO	NO	NO
AT1G17190	4.46	8.13	18.90	17.12	1.07	2.03E-04	NEGATIVE	NO	NO	NO
AT1G17665	20.11	20.86	39.85	35.92	0.78	1.20E-04	NO	NO	NO	NO
AT1G17880	132.66	115.11	303.81	185.93	0.69	7.84E-05	NO	NO	NO	NO
AT1G18330	0.26	0.27	1.78	1.73	2.67	1.93E-04	NEGATIVE	YES	NO	NO
AT1G18910	1.28	1.02	5.61	10.00	3.30	0.00E+00	NO	NO	NO	NO
AT1G20160	1.77	1.00	7.05	5.82	2.55	1.04E-13	NO	NO	NO	NO
AT1G20190	16.22	27.07	40.89	47.79	0.82	1.66E-05	NO	NO	NO	NO
AT1G20350	0.04	0.05	4.15	1.85	5.32	3.62E-03	NO	NO	NO	NO
AT1G21270	1.19	1.76	40.11	69.59	5.30	0.00E+00	NO	NO	NO	DOWN
AT1G21750	153.45	119.07	255.57	386.01	1.70	0.00E+00	NO	NO	NO	NO
AT1G22400	14.75	2.31	6.24	21.78	3.24	0.00E+00	NO	NO	NO	NO
AT1G23410	14.20	10.85	41.72	19.68	0.86	1.83E-02	NO	NO	NO	NO
AT1G23850	1.37	0.14	11.26	16.17	6.82	0.00E+00	NO	NO	NO	NO
AT1G27330	71.88	64.60	146.63	205.94	1.67	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT1G28670	0.56	1.06	3.04	3.42	1.69	9.07E-04	NO	NO	DOWN	NO
AT1G30700	19.20	2.23	2.83	8.76	1.98	3.35E-10	NO	NO	NO	NO
AT1G30730	3.43	0.96	0.84	2.90	1.60	1.38E-03	NO	NO	NO	NO
AT1G30900	8.43	5.76	20.12	25.78	2.16	0.00E+00	NO	NO	NO	NO
AT1G31540	5.28	4.94	14.14	23.47	2.25	0.00E+00	NO	NO	NO	NO
AT1G32920	30.56	7.65	9.64	26.23	1.78	6.51E-06	NO	NO	DOWN	NO
AT1G32940	24.51	4.85	8.65	17.27	1.83	0.00E+00	NO	NO	NO	NO
AT1G32950	4.44	0.13	0.89	1.24	3.28	1.69E-05	NO	NO	NO	NO
AT1G33610	1.57	0.97	12.98	18.93	4.29	0.00E+00	NO	NO	NO	NO
AT1G33960	74.61	17.50	18.95	220.93	3.66	0.00E+00	NO	NO	NO	NO
AT1G34575	0.06	0.09	22.63	25.60	8.17	0.00E+00	NO	NO	NO	NO
AT1G35710	9.96	5.45	39.22	53.32	3.29	0.00E+00	NO	NO	NO	NO
AT1G36622	13.65	2.44	51.22	73.32	4.91	0.00E+00	NO	NO	NO	NO
AT1G43910	12.36	9.22	5.52	26.96	1.55	0.00E+00	POSITIVE	NO	NO	NO
AT1G44350	12.96	2.31	5.40	8.98	1.96	2.32E-11	NO	NO	NO	NO
AT1G44750	15.10	15.20	30.81	38.79	1.35	0.00E+00	POSITIVE	NO	NO	NO
AT1G45201	16.08	5.38	53.25	26.96	2.32	0.00E+00	NO	NO	NO	NO
AT1G47370	0.05	0.07	1.52	1.98	4.82	1.25E-04	NO	NO	NO	NO
AT1G47510	16.58	1.04	3.00	3.72	1.83	2.24E-04	NO	NO	NO	NO
AT1G47760	0.08	0.09	7.72	5.50	5.98	6.93E-04	NO	YES	NO	NO
AT1G48260	0.83	0.80	3.71	4.54	2.51	6.56E-08	NO	NO	NO	NO
AT1G48460	20.68	16.50	45.61	22.96	0.48	4.56E-02	NO	NO	NO	NO
AT1G48750	13.04	11.81	58.82	40.33	1.77	4.61E-09	NO	NO	NO	NO
AT1G50110	1.20	1.50	7.93	4.44	1.57	7.73E-04	NO	NO	NO	NO
AT1G50520	0.02	0.03	4.99	10.09	8.59	4.26E-10	NO	NO	NO	NO
AT1G51270	0.08	0.08	6.76	17.01	7.78	0.00E+00	POSITIVE	NO	NO	NO
AT1G51430	3.33	4.41	16.09	14.93	1.76	2.87E-07	POSITIVE	NO	NO	NO
AT1G51890	4.82	1.05	10.50	17.17	4.03	0.00E+00	NO	NO	NO	NO
AT1G52040	47.33	50.83	162.86	233.88	2.20	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT1G52200	176.46	78.73	79.71	124.75	0.66	2.38E-04	NO	NO	NO	NO
AT1G52290	6.40	4.56	17.61	21.66	2.25	0.00E+00	NO	NO	NO	NO
AT1G52342	3.58	3.12	14.61	19.68	2.66	7.26E-08	NO	NO	NO	NO
AT1G52400	329.62	218.65	1020.56	974.36	2.16	0.00E+00	NO	NO	NO	NO
AT1G53480	8.94	5.43	83.13	61.00	3.49	0.00E+00	NO	NO	NO	NO
AT1G55330	56.02	42.22	128.20	71.21	0.75	4.96E-03	NO	NO	DOWN	NO
AT1G56510	0.19	0.17	2.21	7.04	5.34	0.00E+00	NO	NO	NO	NO
AT1G56520	0.08	0.07	4.64	7.70	6.80	0.00E+00	NO	NO	NO	NO
AT1G56660	6.04	4.80	20.21	48.80	3.35	0.00E+00	NO	NO	NO	NO
AT1G57630	0.00	0.00	1.28	4.69	Inf	5.67E-04	NO	NO	NO	NO
AT1G57650	0.12	0.09	2.63	7.97	6.51	0.00E+00	POSITIVE	NO	NO	NO
AT1G57660	30.17	28.98	105.56	69.70	1.27	8.39E-10	NO	NO	NO	NO
AT1G58225	43.95	12.15	187.81	104.62	3.11	0.00E+00	NO	NO	NO	NO
AT1G58380	21.08	16.63	61.75	39.80	1.26	1.84E-10	NO	NO	NO	NO
AT1G58420	5.45	1.60	16.96	12.78	3.00	2.39E-09	NO	NO	NO	NO
AT1G60190	7.31	15.05	17.37	20.99	0.48	2.17E-02	NO	NO	NO	NO
AT1G61820	3.18	0.41	0.72	1.65	2.03	1.34E-03	NO	NO	NO	NO
AT1G61980	4.57	4.07	13.74	8.60	1.08	3.17E-04	NO	NO	NO	NO
AT1G61990	8.42	7.38	23.26	16.90	1.20	3.80E-08	NO	NO	NO	NO
AT1G62355	0.00	0.00	10.98	5.62	Inf	3.39E-13	NO	NO	NO	NO
AT1G62540	0.06	0.10	0.86	0.77	2.88	4.11E-03	NO	NO	NO	NO
AT1G63160	11.31	9.71	28.43	20.83	1.10	1.59E-06	NO	NO	NO	NO
AT1G63780	32.67	21.83	62.18	31.93	0.55	1.24E-02	NO	NO	NO	NO
AT1G63860	0.31	0.30	4.47	7.27	4.62	0.00E+00	NO	NO	NO	NO
AT1G65150	0.35	0.34	3.18	1.32	1.97	1.67E-02	NO	NO	NO	NO
AT1G65490	49.55	62.83	178.33	229.20	1.87	0.00E+00	NO	NO	NO	UP
AT1G65500	34.66	23.58	134.03	294.68	3.64	0.00E+00	NO	NO	NO	NO
AT1G65790	6.35	5.29	18.60	20.61	1.96	0.00E+00	NO	NO	NO	NO
AT1G66725	10.13	9.69	43.58	88.10	3.19	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT1G66880	14.14	8.80	31.95	48.11	2.45	0.00E+00	NO	NO	NO	NO
AT1G66960	0.02	0.05	7.84	10.15	7.80	0.00E+00	NO	NO	NO	NO
AT1G67365	0.00	0.05	6.89	11.59	7.96	0.00E+00	NO	NO	NO	NO
AT1G67590	0.27	0.44	5.65	5.86	3.73	3.79E-11	NEGATIVE	NO	NO	NO
AT1G67980	40.86	11.03	16.60	43.41	1.98	0.00E+00	NO	NO	NO	NO
AT1G68620	8.64	0.87	22.47	24.40	4.81	0.00E+00	NO	NO	NO	NO
AT1G69530	19.91	34.29	88.43	97.64	1.51	0.00E+00	NO	NO	DOWN	DOWN
AT1G69880	19.88	2.06	2.93	7.52	1.87	1.81E-03	NO	NO	NO	NO
AT1G70090	22.24	14.81	40.78	28.19	0.93	2.41E-08	NO	NO	NO	NO
AT1G70200	62.82	75.02	129.16	106.71	0.51	9.22E-03	NO	NO	NO	NO
AT1G70420	18.09	22.79	79.36	58.81	1.37	3.66E-14	NO	NO	NO	NO
AT1G70985	2.74	2.78	10.64	8.66	1.64	2.90E-03	NO	NO	NO	NO
AT1G71880	26.41	21.58	67.52	62.98	1.54	0.00E+00	NO	NO	NO	NO
AT1G72130	4.75	3.86	12.48	12.73	1.72	1.04E-13	NO	NO	NO	NO
AT1G72430	8.93	2.64	36.31	30.88	3.55	0.00E+00	NO	NO	DOWN	DOWN
AT1G72520	8.00	3.85	3.01	7.99	1.06	4.87E-06	NO	NO	NO	NO
AT1G72970	11.58	12.72	26.39	20.79	0.71	2.86E-04	NEGATIVE	NO	NO	NO
AT1G73260	24.81	24.99	72.52	97.23	1.96	0.00E+00	NO	NO	NO	NO
AT1G73805	10.37	12.01	27.81	86.07	2.84	0.00E+00	NO	NO	NO	NO
AT1G74670	14.88	8.58	70.40	51.51	2.59	0.00E+00	NO	NO	DOWN	NO
AT1G74710	6.67	6.29	14.45	27.71	2.14	0.00E+00	NO	NO	NO	NO
AT1G75030	2.70	2.13	9.56	6.90	1.69	3.81E-04	NEGATIVE	NO	NO	NO
AT1G75040	1396.09	359.53	8818.64	8119.78	4.50	0.00E+00	NO	NO	POSITIVE	NO
AT1G76470	30.54	0.06	0.08	0.53	3.13	3.81E-02	NO	NO	NO	NO
AT1G76650	20.07	4.03	8.36	27.88	2.79	0.00E+00	NO	NO	NO	NO
AT1G76930	461.90	155.62	122.20	624.22	2.00	0.00E+00	NO	NO	NO	NO
AT1G76955	22.27	27.04	55.22	43.05	0.67	9.31E-03	NO	NO	NO	NO
AT1G76970	8.86	6.20	20.65	23.37	1.92	0.00E+00	NO	NO	NO	NO
AT1G77510	64.03	31.34	213.11	281.48	3.17	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT1G77630	6.48	4.41	15.64	9.92	1.17	1.64E-05	NO	NO	NO	NO
AT1G77760	23.40	20.68	42.37	43.76	1.08	7.77E-11	NO	NO	NO	NO
AT1G78080	14.51	18.21	27.88	39.48	1.12	3.76E-10	NO	YES	NO	NO
AT1G78290	20.46	23.90	49.12	60.90	1.35	0.00E+00	NO	NO	NO	NO
AT1G78410	16.77	6.68	43.36	106.49	4.00	0.00E+00	NO	NO	NO	NO
AT2G01670	28.44	13.42	79.08	68.28	2.35	0.00E+00	NO	NO	NO	NO
AT2G01890	1.94	0.81	15.20	9.88	3.60	1.91E-14	NO	NO	NO	NO
AT2G03750	15.05	20.08	54.35	47.16	1.23	4.26E-11	NO	NO	DOWN	NO
AT2G04040	27.39	3.36	13.81	23.64	2.82	0.00E+00	NO	NO	NO	NO
AT2G04450	38.95	17.47	14.49	112.95	2.69	0.00E+00	NO	NO	NO	NO
AT2G04495	3.49	1.14	15.43	14.71	3.69	0.00E+00	NO	NO	NO	NO
AT2G07750	0.06	0.06	4.89	3.62	5.99	0.00E+00	NO	NO	NO	NO
AT2G10940	641.34	427.76	1405.64	689.21	0.69	1.39E-03	NO	NO	NO	NO
AT2G14580	2.21	2.81	17.58	7.73	1.46	7.67E-03	NO	NO	NO	NO
AT2G14610	1484.66	1091.57	4492.87	15144.50	3.79	0.00E+00	NO	NO	NO	NO
AT2G15128	1.25	0.97	383.42	430.07	8.79	4.88E-08	NO	NO	NO	NO
AT2G16676	0.02	0.00	1.37	1.91	Inf	6.92E-07	NO	NO	NO	NO
AT2G16680	0.01	0.00	0.99	1.62	Inf	7.01E-12	NO	NO	NO	NO
AT2G17040	3.95	3.31	11.63	24.66	2.90	0.00E+00	NO	YES	NO	NO
AT2G17250	19.27	12.76	38.86	20.78	0.70	4.67E-04	NO	NO	NO	NO
AT2G18193	3.54	0.49	0.38	1.42	1.54	2.51E-02	NO	NO	NO	NO
AT2G18300	46.62	43.56	69.80	64.40	0.56	5.40E-06	NO	YES	NO	NO
AT2G18560	2.17	1.67	7.20	6.35	1.93	6.26E-08	NO	NO	NO	NO
AT2G18680	3.92	0.05	12.71	9.74	7.59	7.89E-08	NO	NO	NO	NO
AT2G22330	19.52	18.36	44.94	30.13	0.72	2.14E-04	NO	NO	NO	NO
AT2G22980	3.01	2.88	11.91	11.57	2.01	0.00E+00	NO	NO	NO	NO
AT2G23000	1.92	1.64	6.58	4.63	1.50	3.93E-04	NO	NO	NO	NO
AT2G23110	0.36	0.07	4.54	5.90	6.42	2.31E-04	NO	NO	NO	NO
AT2G23600	23.61	23.39	119.48	91.89	1.97	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT2G24850	23.62	4.61	141.32	133.81	4.86	0.00E+00	NO	NO	NO	NO
AT2G25440	0.25	0.10	2.92	5.06	5.68	3.68E-14	NO	NO	NO	NO
AT2G27420	1.88	0.93	7.85	2.98	1.67	3.05E-03	NO	NO	NO	NO
AT2G27660	1.09	0.45	6.04	10.82	4.60	0.00E+00	NO	NO	NO	NO
AT2G29310	6.91	6.48	19.69	13.33	1.04	1.16E-03	NEGATIVE	NO	NO	NO
AT2G29350	72.51	15.31	176.40	300.97	4.30	0.00E+00	NO	NO	NO	NO
AT2G29470	30.68	3.33	8.74	25.58	2.94	0.00E+00	NO	NO	NO	NO
AT2G30010	26.75	27.35	60.60	56.11	1.04	3.90E-09	NEGATIVE	NO	NO	DOWN
AT2G30400	8.45	0.42	1.38	2.53	2.60	6.79E-04	NO	NO	NO	NO
AT2G30550	11.60	8.20	26.05	32.67	1.99	0.00E+00	NO	NO	NO	NO
AT2G30770	45.39	2.42	16.80	13.56	2.49	0.00E+00	NO	NO	NO	NO
AT2G31420	4.01	0.12	0.60	1.38	3.56	2.54E-02	NO	NO	NO	NO
AT2G31880	45.23	25.29	132.42	190.79	2.92	0.00E+00	NO	NO	NO	NO
AT2G32530	0.13	0.06	1.37	0.96	3.90	4.23E-05	NO	NO	NO	NO
AT2G32560	8.98	8.27	22.32	19.90	1.27	2.86E-08	NO	NO	NO	NO
AT2G32680	9.55	3.84	25.99	35.79	3.22	0.00E+00	NO	NO	NO	NO
AT2G32870	1.51	0.17	10.11	2.84	4.02	2.69E-08	NO	NO	NO	NO
AT2G32880	2.03	0.35	9.76	3.80	3.46	1.65E-07	NO	NO	DOWN	NO
AT2G33400	2.52	5.10	7.12	10.05	0.98	3.97E-04	NO	NO	NO	NO
AT2G33710	3.87	0.19	1.11	1.20	2.63	4.83E-03	NO	YES	NO	NO
AT2G34070	25.41	10.98	10.84	18.19	0.73	1.60E-03	NO	NO	NO	DOWN
AT2G35035	0.60	1.07	3.46	4.60	2.11	9.23E-05	NO	NO	NO	NO
AT2G36780	3.82	0.59	0.64	2.02	1.78	2.76E-03	NO	NO	NO	NO
AT2G36790	2.66	0.45	0.29	1.23	1.46	4.72E-02	NO	NO	NO	NO
AT2G36830	34.90	29.60	127.65	74.42	1.33	5.37E-14	NEGATIVE	NO	DOWN	NO
AT2G37710	60.27	51.54	175.02	280.31	2.44	0.00E+00	NO	NO	NO	NO
AT2G37760	48.93	17.62	20.27	23.51	0.42	4.69E-02	NO	NO	NO	NO
AT2G37770	37.03	7.05	16.65	24.94	1.82	1.86E-14	NO	NO	NO	NO
AT2G38240	31.81	3.82	9.22	15.75	2.04	1.53E-11	NO	NO	NO	NO

Table A3.6 (cont'd)

AT2G38340	4.95	0.62	1.28	4.92	2.98	3.48E-06	NO	YES	NO	NO
AT2G39030	120.09	6.08	16.25	16.77	1.46	1.43E-06	NO	NO	NO	NO
AT2G39210	15.16	10.10	30.93	47.34	2.23	0.00E+00	NO	NO	NO	NO
AT2G39330	24.36	5.93	11.79	10.93	0.88	8.43E-04	NO	NO	NO	NO
AT2G39390	181.59	172.65	345.16	231.96	0.43	3.18E-02	NO	NO	NO	NO
AT2G39710	16.28	17.94	43.86	36.63	1.03	2.55E-08	NO	NO	POSITIVE	NO
AT2G40000	29.40	5.70	11.23	22.99	2.01	0.00E+00	NO	NO	DOWN	NO
AT2G40205	2047.97	2835.29	8695.17	6573.37	1.21	5.05E-05	NO	NO	NO	NO
AT2G40740	0.92	0.43	3.94	3.31	2.93	1.24E-03	NO	YES	NO	NO
AT2G41120	3.40	3.64	18.19	21.68	2.57	0.00E+00	NO	NO	NO	NO
AT2G42170	0.30	0.36	10.52	7.22	4.31	3.64E-14	NO	NO	NO	NO
AT2G43150	228.88	58.20	77.25	103.87	0.84	1.13E-06	NO	NO	NO	NO
AT2G43530	96.10	36.88	220.75	189.27	2.36	0.00E+00	NO	NO	NO	NO
AT2G43535	11.04	4.35	59.24	36.47	3.07	0.00E+00	NO	NO	NO	NO
AT2G43570	99.62	10.79	737.05	418.54	5.28	0.00E+00	NO	NO	NO	NO
AT2G43820	69.95	4.43	11.09	17.38	1.97	0.00E+00	POSITIVE	NO	NO	NO
AT2G44240	0.06	0.09	30.21	55.03	9.29	0.00E+00	NO	NO	NO	NO
AT2G44290	5.60	7.85	60.45	74.46	3.25	0.00E+00	NO	NO	NO	NO
AT2G44790	27.68	2.66	7.65	8.86	1.74	3.10E-05	NO	NO	NO	NO
AT2G45570	27.02	1.64	4.69	7.43	2.18	8.59E-10	NO	NO	NO	NO
AT2G45660	25.94	34.57	64.51	89.14	1.37	0.00E+00	NO	YES	POSITIVE	NO
AT2G45710	31.00	24.68	92.21	46.41	0.91	4.71E-04	NO	NO	NO	NO
AT2G46430	10.76	11.01	27.25	46.16	2.07	0.00E+00	NO	NO	NO	NO
AT2G46710	5.21	6.62	19.78	16.23	1.29	4.92E-09	NEGATIVE	NO	NO	NO
AT2G47000	14.07	3.81	2.86	9.76	1.36	5.29E-12	NO	NO	NO	NO
AT2G47950	4.51	0.27	0.85	1.56	2.53	3.52E-02	NO	NO	NO	NO
AT3G02790	9.84	1.94	57.45	17.65	3.19	5.75E-10	NO	YES	DOWN	NO
AT3G03030	1.41	1.65	5.78	3.91	1.25	2.16E-03	NO	NO	NO	NO
AT3G03450	3.81	3.74	9.49	8.81	1.24	1.87E-06	NO	YES	NO	NO

Table A3.6 (cont'd)

AT3G03850	1.39	1.14	8.42	4.20	1.89	1.14E-02	NO	NO	NO	NO
AT3G04480	8.82	13.11	17.47	19.68	0.59	3.69E-03	NO	NO	NO	NO
AT3G06145	2.58	2.47	16.48	9.97	2.01	2.38E-05	NO	NO	NO	NO
AT3G06200	31.59	26.41	62.95	44.21	0.74	1.03E-04	NO	NO	NO	NO
AT3G06530	17.72	10.29	29.68	13.63	0.41	3.48E-03	NO	NO	POSITIVE	NO
AT3G07520	4.41	3.62	12.42	14.34	1.98	0.00E+00	NO	NO	NO	NO
AT3G08870	1.25	0.73	6.59	10.34	3.82	0.00E+00	NO	NO	NO	NO
AT3G08970	3.41	3.13	10.83	23.42	2.90	0.00E+00	NO	NO	NO	NO
AT3G09270	70.18	6.61	8.98	22.97	1.80	1.16E-09	NO	NO	NO	NO
AT3G09940	2.43	0.38	17.79	33.00	6.43	0.00E+00	NO	NO	NO	NO
AT3G11000	0.08	0.05	1.87	0.81	3.88	8.26E-05	NO	NO	NO	NO
AT3G11010	0.09	0.04	6.18	9.82	7.88	0.00E+00	NO	NO	NO	NO
AT3G11080	0.98	0.81	11.97	16.48	4.35	0.00E+00	NO	NO	NO	NO
AT3G11280	14.36	10.47	42.95	33.84	1.69	0.00E+00	NO	YES	NO	NO
AT3G11340	13.22	0.49	3.40	9.67	4.29	0.00E+00	NO	NO	NO	NO
AT3G13130	2.29	1.08	8.94	8.64	3.00	2.21E-08	NO	NO	NO	NO
AT3G13433	0.92	0.67	19.18	21.88	5.02	1.53E-10	NO	NO	NO	NO
AT3G13560	17.13	15.52	34.68	34.00	1.13	3.42E-13	NEGATIVE	NO	NO	NO
AT3G13610	28.30	1.63	12.07	50.87	4.96	0.00E+00	NO	NO	NO	NO
AT3G13750	18.94	24.75	43.02	57.22	1.21	2.17E-13	NEGATIVE	NO	DOWN	NO
AT3G14990	181.64	53.71	86.39	135.78	1.34	0.00E+00	NO	NO	NO	NO
AT3G15536	8.13	2.70	72.57	73.19	4.76	0.00E+00	NO	NO	NO	NO
AT3G16240	25.22	31.18	67.21	54.88	0.82	1.18E-05	NO	NO	DOWN	NO
AT3G16470	139.59	35.68	53.09	55.60	0.64	3.12E-04	NO	NO	NO	NO
AT3G17510	14.85	14.91	37.44	40.54	1.44	0.00E+00	NO	NO	NO	NO
AT3G18830	86.28	49.21	40.80	66.53	0.44	2.70E-02	NO	NO	NO	NO
AT3G19515	4.82	5.09	22.10	23.36	2.20	0.00E+00	NO	NO	NO	NO
AT3G19520	10.49	9.28	23.00	27.82	1.58	5.37E-14	POSITIVE	NO	NO	NO
AT3G21150	2.99	3.14	10.78	20.55	2.71	1.92E-14	NO	NO	NO	NO

Table A3.6 (cont'd)

AT3G22370	55.20	22.25	27.15	49.59	1.16	9.80E-11	NO	NO	NO	NO
AT3G22930	0.29	0.13	2.64	4.95	5.30	7.52E-07	NO	NO	NO	NO
AT3G23080	10.53	9.99	20.20	19.10	0.94	2.09E-06	NO	NO	DOWN	NO
AT3G23120	1.82	1.87	6.77	9.93	2.41	0.00E+00	NO	NO	NO	NO
AT3G23240	7.29	0.86	1.77	2.93	1.77	9.62E-03	NO	YES	NO	NO
AT3G23480	0.93	0.91	6.09	6.18	2.76	3.67E-11	NO	NO	NO	NO
AT3G23620	88.22	68.95	204.42	94.61	0.46	2.30E-02	NO	NO	POSITIVE	NO
AT3G23730	7.47	10.56	21.25	18.52	0.81	1.75E-03	NO	NO	NO	NO
AT3G23890	2.31	4.21	4.84	6.38	0.60	1.71E-03	NO	NO	NO	NO
AT3G24516	2.27	1.54	273.52	216.80	7.13	2.15E-05	NO	NO	NO	NO
AT3G24517	0.47	0.41	74.93	50.31	6.93	0.00E+00	NO	NO	NO	NO
AT3G24518	1.08	1.54	15.14	9.72	2.65	1.36E-11	NO	NO	NO	NO
AT3G25010	1.46	0.65	8.62	9.03	3.80	0.00E+00	NO	NO	NO	NO
AT3G25020	0.07	0.06	6.74	9.49	7.36	0.00E+00	NO	NO	NO	NO
AT3G25250	5.77	0.49	1.43	4.01	3.05	6.66E-08	NO	NO	NO	NO
AT3G25572	0.77	0.88	3.52	2.78	1.67	1.04E-03	NO	NO	NO	NO
AT3G26170	2.97	1.95	8.83	10.78	2.47	1.91E-14	NO	NO	NO	NO
AT3G26240	0.04	0.04	2.61	2.42	6.07	3.73E-13	NO	NO	NO	NO
AT3G26520	46.81	45.98	235.68	208.37	2.18	0.00E+00	NEGATIVE	NO	NO	NO
AT3G26830	119.27	10.37	42.52	76.07	2.88	0.00E+00	NO	NO	NO	NO
AT3G28270	0.19	0.16	9.21	1.85	3.50	3.46E-07	NO	NO	DOWN	NO
AT3G28510	0.26	0.11	3.30	7.75	6.11	0.00E+00	NO	NO	NO	NO
AT3G29240	41.24	43.43	79.69	110.08	1.34	0.00E+00	NO	NO	NO	NO
AT3G29575	3.71	2.33	13.02	16.42	2.82	0.00E+00	NO	NO	NO	NO
AT3G29590	2.27	1.42	6.97	4.02	1.51	1.71E-03	POSITIVE	NO	NO	NO
AT3G43670	10.19	11.54	20.39	23.48	1.02	3.15E-08	NO	NO	NO	NO
AT3G43800	37.23	42.60	100.26	116.26	1.45	0.00E+00	NO	NO	NO	NO
AT3G44215	0.02	0.01	7.10	10.24	9.75	0.00E+00	NO	NO	NO	NO
AT3G44300	144.08	24.36	28.96	79.80	1.71	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT3G44310	114.22	128.73	280.91	332.51	1.37	0.00E+00	NO	NO	NO	NO
AT3G44970	0.04	0.05	2.68	1.20	4.57	3.53E-04	NO	NO	NO	NO
AT3G45030	161.26	131.07	307.10	186.51	0.51	1.06E-02	NO	NO	NO	NO
AT3G45290	5.34	3.34	17.72	9.90	1.57	8.92E-08	NO	NO	NO	NO
AT3G45390	0.36	0.73	1.88	2.63	1.86	2.56E-04	NO	NO	NO	NO
AT3G45443	1.73	2.37	7.43	7.13	1.59	3.13E-04	NO	NO	NO	NO
AT3G45446	0.02	0.02	2.33	4.81	8.18	5.25E-07	NO	NO	NO	NO
AT3G45860	20.46	3.93	71.63	51.93	3.72	0.00E+00	NO	NO	NO	NO
AT3G46650	0.46	0.35	4.56	1.42	2.01	8.76E-03	NO	NO	NO	NO
AT3G46668	0.00	0.00	3.18	2.83	Inf	1.64E-03	NO	NO	NO	NO
AT3G47560	4.22	4.25	12.31	12.45	1.55	8.82E-07	NO	NO	DOWN	NO
AT3G48080	4.12	8.01	11.64	24.86	1.63	0.00E+00	NO	NO	NO	NO
AT3G48640	23.78	14.19	66.49	218.10	3.94	0.00E+00	NO	NO	NO	NO
AT3G48850	12.06	0.70	3.25	4.78	2.78	1.30E-06	NO	NO	NO	NO
AT3G49180	15.81	13.64	37.59	21.10	0.63	4.15E-03	NO	NO	NO	NO
AT3G49340	0.50	0.16	47.82	34.60	7.74	0.00E+00	NO	NO	NO	NO
AT3G49780	59.43	26.52	168.86	171.89	2.70	0.00E+00	NO	NO	NO	NO
AT3G51230	0.68	1.92	5.41	8.82	2.20	3.46E-07	NO	NO	NO	NO
AT3G51860	25.18	5.37	87.42	68.76	3.68	0.00E+00	NO	NO	NO	NO
AT3G53150	3.12	0.44	11.31	8.86	4.32	0.00E+00	NO	NO	NO	NO
AT3G54390	1.69	1.39	5.95	4.50	1.69	2.53E-04	NO	YES	NO	NO
AT3G54830	10.87	7.84	45.73	27.79	1.83	0.00E+00	NO	NO	NO	NO
AT3G56400	22.07	20.16	52.03	125.10	2.63	0.00E+00	NO	YES	NO	NO
AT3G57040	7.44	9.34	17.78	14.46	0.63	1.73E-02	NEGATIVE	NO	NO	NO
AT3G57260	552.31	123.31	3942.03	2454.12	4.31	0.00E+00	NO	NO	NO	NO
AT3G57490	39.33	25.39	74.37	35.80	0.50	2.77E-02	NO	NO	NO	NO
AT3G58120	4.42	3.29	11.94	9.16	1.48	8.90E-07	NO	YES	NO	NO
AT3G59140	14.00	4.31	4.15	6.20	0.53	2.41E-02	NO	NO	NO	NO
AT3G60140	19.06	2.17	1.51	19.34	3.16	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT3G60170	0.01	0.01	1.66	2.80	7.70	4.35E-10	NO	NO	NO	NO
AT3G60500	16.25	13.62	27.72	18.73	0.46	2.09E-02	NO	NO	NO	NO
AT3G60540	9.28	5.86	29.19	36.38	2.63	3.36E-12	NO	NO	NO	NO
AT3G62890	2.87	2.37	9.15	5.30	1.16	8.80E-04	NO	NO	NO	NO
AT4G00700	4.63	1.01	15.88	16.67	4.05	0.00E+00	NO	NO	NO	NO
AT4G01490	0.03	0.02	8.29	10.52	9.03	0.00E+00	NO	NO	NO	NO
AT4G01525	0.51	0.56	146.37	247.03	8.79	0.00E+00	NO	NO	NO	NO
AT4G01720	2.37	1.42	6.86	8.32	2.55	5.56E-13	NO	YES	NO	NO
AT4G01870	388.35	46.12	85.28	137.91	1.58	0.00E+00	NO	NO	NO	NO
AT4G03320	3.39	0.43	0.92	1.32	1.61	5.00E-02	NO	NO	NO	NO
AT4G03450	5.12	2.70	12.63	22.77	3.08	0.00E+00	NO	NO	NO	NO
AT4G04510	0.77	0.32	3.89	6.23	4.28	0.00E+00	NO	NO	NO	NO
AT4G05050	98.34	83.62	204.45	315.81	1.92	0.00E+00	NO	NO	NO	NO
AT4G08470	5.82	6.68	21.66	33.85	2.34	0.00E+00	NO	NO	NO	NO
AT4G08770	42.17	0.59	6.17	8.50	3.84	1.07E-12	NO	NO	NO	NO
AT4G08780	11.41	0.12	1.85	2.89	4.57	1.74E-06	NO	NO	NO	NO
AT4G10120	23.44	39.04	42.77	65.83	0.75	1.74E-09	NO	NO	NO	NO
AT4G10500	3.49	0.55	50.01	41.30	6.24	0.00E+00	NO	NO	NO	NO
AT4G10860	0.81	0.37	20.01	81.53	7.78	0.00E+00	NO	NO	NO	NO
AT4G11000	1.02	0.83	14.62	26.50	4.99	0.00E+00	NO	NO	NO	NO
AT4G11170	0.07	0.06	2.91	7.09	6.98	0.00E+00	NO	NO	NO	NO
AT4G11211	3.02	3.01	13.03	17.52	2.54	1.36E-06	NO	NO	NO	NO
AT4G11650	8.31	2.14	2.75	4.55	1.09	4.10E-02	NO	NO	NO	NO
AT4G12320	12.34	10.46	35.67	38.43	1.88	0.00E+00	NO	NO	NO	NO
AT4G12420	22.21	28.47	41.77	39.89	0.49	1.37E-02	NO	NO	NO	NO
AT4G12735	24.95	1.97	3.93	8.14	2.05	1.97E-03	NO	NO	NO	NO
AT4G12880	40.19	42.77	82.31	69.19	0.69	1.06E-04	NEGATIVE	NO	NO	NO
AT4G13810	0.65	0.49	3.13	5.05	3.35	8.78E-14	NO	NO	NO	NO
AT4G13820	0.31	0.16	2.12	2.50	3.93	2.53E-11	NO	NO	NO	NO

Table A3.6 (cont'd)

AT4G13890	0.22	0.11	8.62	12.83	6.87	0.00E+00	NO	NO	NO	NO
AT4G14090	7.42	4.04	20.43	15.37	1.93	1.85E-14	NO	NO	NO	NO
AT4G14390	0.16	0.16	30.89	43.04	8.06	0.00E+00	NO	NO	NO	NO
AT4G14400	75.26	86.66	196.22	281.37	1.70	0.00E+00	NO	NO	POSITIVE	NO
AT4G14640	1.09	0.98	6.83	7.44	2.93	1.58E-05	NO	NO	NO	NO
AT4G14750	1.40	2.61	6.10	8.91	1.77	4.16E-08	NO	NO	NO	NO
AT4G15248	6.30	4.53	24.70	16.45	1.86	7.17E-05	NO	NO	NO	NO
AT4G15530	81.86	47.74	47.85	81.65	0.77	0.00E+00	NO	NO	NO	NO
AT4G16000	1.31	0.12	9.07	4.30	5.18	1.45E-04	NO	NO	NO	NO
AT4G16760	61.24	29.43	28.21	38.33	0.38	4.35E-02	NO	NO	NO	NO
AT4G17340	1.92	0.72	18.70	9.50	3.73	5.12E-12	NO	NO	DOWN	NO
AT4G18010	19.10	11.47	31.58	50.12	2.13	0.00E+00	NO	NO	NO	NO
AT4G18170	15.76	2.40	4.86	7.16	1.58	5.15E-05	NO	YES	NO	NO
AT4G18970	4.08	4.94	15.09	12.78	1.37	3.76E-07	NO	NO	NO	NO
AT4G19370	2.91	0.79	10.29	17.89	4.51	0.00E+00	NO	NO	NO	NO
AT4G19420	8.35	8.56	16.19	19.29	1.17	7.18E-10	NO	NO	NO	NO
AT4G19630	0.32	1.07	11.99	8.38	2.96	1.37E-06	NO	NO	NO	NO
AT4G19990	4.98	5.53	9.94	14.76	1.42	0.00E+00	NO	NO	NO	NO
AT4G20960	16.54	18.25	36.04	36.11	0.98	1.80E-07	NEGATIVE	NO	NO	NO
AT4G21390	3.99	0.88	1.37	2.88	1.72	1.51E-05	NO	NO	NO	NO
AT4G21640	0.00	0.02	2.05	1.98	7.03	2.98E-05	NO	NO	NO	NO
AT4G21680	25.71	2.15	1.19	4.43	1.04	2.78E-03	NO	NO	NO	NO
AT4G21750	10.66	11.42	19.72	22.57	0.98	7.15E-09	NO	YES	NO	NO
AT4G21870	1.30	1.55	19.68	15.79	3.35	1.63E-10	NO	NO	DOWN	NO
AT4G22485	22.65	15.17	47.77	42.46	1.48	0.00E+00	NO	NO	NO	NO
AT4G22505	23.62	15.78	46.45	32.43	1.04	4.58E-08	NO	NO	NO	NO
AT4G22570	24.92	33.82	80.97	60.33	0.84	1.86E-05	NEGATIVE	NO	NO	NO
AT4G23150	8.73	2.59	35.84	43.22	4.06	0.00E+00	NO	NO	NO	NO
AT4G23160	2.48	0.81	11.42	15.01	4.22	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT4G23200	1.57	0.18	5.09	10.00	5.83	0.00E+00	NO	NO	NO	NO
AT4G23210	0.97	0.47	4.55	16.91	5.16	0.00E+00	NO	NO	NO	NO
AT4G23220	0.13	0.16	22.00	33.28	7.73	0.00E+00	NO	NO	NO	NO
AT4G23230	0.37	0.13	4.68	6.03	5.56	0.00E+00	NO	NO	NO	NO
AT4G23260	8.05	8.04	16.25	31.92	1.99	0.00E+00	NO	NO	NO	DOWN
AT4G23810	6.69	3.47	26.46	34.17	3.30	0.00E+00	NO	YES	NO	NO
AT4G23880	13.60	9.62	38.28	24.62	1.36	1.67E-06	NO	NO	NO	NO
AT4G24780	22.92	36.43	52.97	55.53	0.61	1.13E-03	NO	NO	DOWN	NO
AT4G25000	2.39	1.11	34.11	16.94	3.93	0.00E+00	NO	NO	NO	NO
AT4G25110	0.33	0.30	8.48	17.10	5.85	0.00E+00	NO	NO	NO	NO
AT4G26760	4.22	6.75	9.89	11.74	0.80	4.03E-04	NO	NO	NO	NO
AT4G29520	49.07	32.64	96.39	118.75	1.86	0.00E+00	NO	NO	NO	NO
AT4G29700	3.35	1.29	9.68	12.68	3.29	0.00E+00	NO	NO	NO	NO
AT4G29905	55.47	48.47	148.11	71.06	0.55	1.33E-02	NO	NO	NO	NO
AT4G30720	35.61	41.66	191.54	154.76	1.89	0.00E+00	NO	NO	POSITIVE	NO
AT4G31790	28.25	16.79	49.15	25.75	0.62	1.89E-03	NO	NO	NO	NO
AT4G31870	0.15	0.10	8.52	18.24	7.48	1.76E-09	NO	NO	NO	NO
AT4G32340	52.99	63.93	104.36	105.77	0.73	7.26E-05	NEGATIVE	NO	NO	NO
AT4G33355	0.35	1.30	4.66	4.65	1.84	1.93E-03	NO	NO	NO	NO
AT4G33360	1.50	1.27	28.27	35.06	4.79	0.00E+00	NO	NO	NO	NO
AT4G33560	2.57	3.57	16.73	22.73	2.67	5.96E-09	NO	NO	NO	NO
AT4G34131	10.82	2.79	2.27	4.75	0.77	4.29E-02	NO	NO	NO	NO
AT4G34760	20.41	30.15	50.28	46.20	0.62	9.34E-03	NO	NO	NO	NO
AT4G34930	0.30	0.28	1.68	1.20	2.10	7.51E-03	NO	NO	NO	NO
AT4G36130	168.11	135.32	340.21	189.28	0.48	1.31E-02	NO	NO	NO	NO
AT4G37150	0.37	0.33	8.36	7.53	4.53	2.38E-10	NO	NO	NO	NO
AT4G37370	21.47	4.72	7.09	19.85	2.07	0.00E+00	NO	NO	NO	NO
AT4G37800	13.58	16.03	28.25	26.50	0.73	1.36E-03	NO	NO	DOWN	NO
AT4G37990	54.06	1.88	6.82	13.51	2.84	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT4G38540	45.54	19.59	15.45	27.95	0.51	1.83E-02	NO	NO	NO	NO
AT4G38840	44.11	46.52	90.62	77.15	0.73	7.53E-04	NEGATIVE	NO	DOWN	NO
AT4G38850	2.78	5.66	14.53	18.83	1.73	1.05E-04	NO	NO	NO	NO
AT4G39830	3.48	2.22	10.73	20.43	3.20	0.00E+00	NO	NO	NO	NO
AT4G40060	4.53	4.79	22.83	23.79	2.31	0.00E+00	NO	YES	NO	NO
AT5G01290	9.47	7.84	21.66	16.99	1.12	1.53E-08	NO	NO	NO	NO
AT5G01550	0.41	0.16	2.78	4.75	4.91	3.63E-14	NO	NO	NO	NO
AT5G02020	35.83	0.66	3.40	2.87	2.13	7.64E-04	NO	NO	NO	NO
AT5G03380	9.37	5.61	26.68	36.04	2.68	0.00E+00	NO	NO	NO	NO
AT5G03760	28.57	24.06	54.54	35.95	0.58	2.22E-03	NEGATIVE	NO	NO	NO
AT5G05600	34.73	6.61	13.01	16.77	1.34	4.96E-08	NO	NO	NO	NO
AT5G06860	111.41	24.82	24.92	36.68	0.56	8.38E-03	POSITIVE	NO	POSITIVE	NO
AT5G06870	6.34	2.96	31.45	33.70	3.51	0.00E+00	NO	NO	NO	NO
AT5G07440	95.06	24.31	29.15	33.46	0.46	3.45E-03	NO	NO	NO	NO
AT5G08760	37.63	27.22	141.56	191.60	2.82	0.00E+00	NO	NO	NO	NO
AT5G09290	0.10	0.07	4.08	10.26	7.25	3.40E-11	NO	NO	NO	NO
AT5G10380	28.68	11.78	77.88	95.21	3.01	0.00E+00	NO	NO	NO	NO
AT5G10572	98.89	110.74	760.83	397.41	1.84	2.04E-02	NO	NO	NO	NO
AT5G10760	176.34	39.35	736.26	639.62	4.02	0.00E+00	NO	NO	NO	NO
AT5G11580	22.24	27.12	47.35	49.09	0.86	1.27E-06	NO	NO	NO	NO
AT5G11930	1.44	2.45	6.86	9.30	1.92	1.11E-04	NO	NO	NO	NO
AT5G13330	18.68	1.39	2.58	5.29	1.93	9.54E-05	NO	YES	NO	NO
AT5G14800	53.65	58.66	134.11	113.66	0.95	1.61E-08	NO	NO	NO	NO
AT5G15350	24.99	24.67	50.17	39.89	0.69	2.26E-03	NO	NO	NO	NO
AT5G15948	2.17	1.90	7.38	7.70	2.02	8.79E-10	NO	NO	NO	NO
AT5G15950	209.86	167.81	524.89	521.42	1.64	0.00E+00	NO	NO	POSITIVE	NO
AT5G16250	9.47	16.30	28.16	38.53	1.24	1.49E-08	NO	NO	NO	NO
AT5G16960	10.89	0.52	1.16	2.74	2.40	1.45E-04	NO	NO	NO	NO
AT5G17220	13.42	10.00	31.84	20.41	1.03	5.70E-04	NO	NO	NO	NO

Table A3.6 (cont'd)

AT5G17330	0.19	0.09	1.44	3.30	5.17	3.60E-10	NO	NO	NO	NO
AT5G17760	27.54	20.50	13.33	31.07	0.60	1.72E-03	NO	NO	NO	NO
AT5G17860	9.24	2.12	3.47	7.35	1.80	8.77E-09	NO	NO	NO	NO
AT5G18130	42.01	17.91	22.50	30.90	0.79	4.64E-05	NO	NO	NO	NO
AT5G18470	26.63	16.32	100.75	112.36	2.78	0.00E+00	NO	NO	NO	NO
AT5G19190	11.17	9.65	34.82	32.59	1.76	1.42E-09	NO	NO	NO	NO
AT5G19240	144.36	94.13	348.53	448.98	2.25	0.00E+00	NO	NO	NO	NO
AT5G19880	0.40	0.20	3.05	6.69	5.06	1.03E-10	NO	NO	NO	NO
AT5G20480	5.04	3.18	2.10	4.76	0.58	4.25E-02	NO	NO	NO	NO
AT5G21060	8.49	11.15	38.30	35.90	1.69	0.00E+00	NO	NO	NO	NO
AT5G22300	62.12	19.15	12.16	29.39	0.62	3.25E-03	POSITIVE	NO	NO	NO
AT5G22320	12.95	11.74	27.48	21.53	0.87	9.98E-06	NO	NO	NO	NO
AT5G22330	27.69	22.99	55.09	38.23	0.73	8.43E-05	NO	NO	NO	NO
AT5G22390	11.93	15.89	36.79	28.81	0.86	7.94E-05	NO	NO	NO	NO
AT5G22520	0.00	0.15	4.27	25.53	7.38	6.83E-11	NO	NO	NO	NO
AT5G23010	254.95	112.20	598.88	236.47	1.08	1.77E-08	NO	NO	NO	NO
AT5G24110	6.01	1.92	26.95	22.99	3.58	0.00E+00	NO	YES	NO	NO
AT5G24150	0.18	0.19	7.90	6.41	5.08	0.00E+00	NO	NO	DOWN	NO
AT5G24200	16.84	5.63	2.09	10.66	0.92	1.53E-03	NO	NO	NO	NO
AT5G24530	36.09	12.93	267.38	257.60	4.32	0.00E+00	NO	NO	NO	NO
AT5G24655	1.12	1.74	14.39	32.58	4.22	0.00E+00	NO	NO	NO	NO
AT5G24770	1.54	0.57	6.75	3.00	2.40	3.65E-05	NO	NO	NO	NO
AT5G25180	0.03	0.01	3.54	3.95	8.41	0.00E+00	NO	NO	NO	NO
AT5G25220	14.99	15.06	38.23	41.50	1.46	0.00E+00	NO	NO	NO	NO
AT5G25460	29.36	38.13	0.00	70.33	0.88	3.72E-07	NEGATIVE	NO	NO	NO
AT5G26340	83.69	28.78	33.14	59.47	1.05	4.10E-10	NO	NO	NO	NO
AT5G28770	14.11	10.86	26.57	18.54	0.77	3.20E-04	NO	YES	NO	NO
AT5G33290	32.79	19.84	15.41	26.90	0.44	3.94E-02	POSITIVE	NO	NO	NO
AT5G35790	28.51	33.15	55.80	54.01	0.70	8.49E-05	NO	NO	DOWN	NO

Table A3.6 (cont'd)

AT5G36930	0.03	0.04	9.88	14.82	8.58	0.00E+00	NO	NO	NO	NO
AT5G38150	2.82	3.04	7.40	10.21	1.75	6.41E-10	NO	NO	NO	NO
AT5G38430	502.18	303.86	1114.11	450.18	0.57	5.81E-03	NO	NO	NO	NO
AT5G38940	2.44	1.36	8.85	5.87	2.11	2.71E-04	NO	NO	NO	NO
AT5G39050	39.29	14.02	17.57	24.86	0.83	5.97E-05	NO	NO	NO	UP
AT5G39100	4.64	2.45	15.28	13.82	2.50	1.86E-09	NO	NO	NO	NO
AT5G40340	9.70	8.56	18.04	18.67	1.13	7.32E-10	NO	NO	NO	NO
AT5G41590	8.78	6.79	30.90	28.87	2.09	7.83E-13	NO	NO	NO	NO
AT5G42020	49.68	38.33	97.39	165.70	2.11	0.00E+00	NO	NO	NO	NO
AT5G43440	5.34	4.46	19.91	27.97	2.65	0.00E+00	NO	NO	NO	NO
AT5G43450	45.30	28.89	20.52	42.35	0.55	6.21E-03	NO	NO	NO	NO
AT5G43870	4.66	5.91	11.83	14.23	1.27	1.37E-07	NEGATIVE	NO	NO	NO
AT5G44020	14.42	18.81	68.46	53.72	1.51	0.00E+00	NO	NO	DOWN	NO
AT5G44420	247.91	152.80	38.75	439.65	1.52	8.73E-14	NO	NO	NO	DOWN
AT5G44430	65.29	16.67	13.26	143.59	3.11	0.00E+00	NO	NO	NO	NO
AT5G44568	7.92	7.74	91.63	213.40	4.78	0.00E+00	NO	NO	NO	NO
AT5G44570	1.01	1.66	70.24	205.43	6.95	0.00E+00	NO	NO	NO	NO
AT5G45030	7.76	9.69	15.09	20.65	1.09	5.39E-14	NO	NO	NO	NO
AT5G45100	14.53	12.18	32.52	33.33	1.45	9.92E-13	NO	NO	NO	NO
AT5G45380	6.08	5.31	14.72	24.19	2.19	0.00E+00	NO	NO	NO	NO
AT5G45670	6.08	7.28	16.26	16.66	1.19	3.23E-07	NEGATIVE	NO	NO	NO
AT5G45820	5.13	3.95	24.64	14.53	1.88	1.30E-12	NO	NO	NO	NO
AT5G45950	19.00	19.65	37.98	31.20	0.67	1.05E-03	NO	NO	NO	NO
AT5G46050	10.26	4.17	40.63	49.94	3.58	0.00E+00	NO	NO	NO	UP
AT5G46260	4.72	5.35	10.60	18.57	1.80	0.00E+00	NO	NO	NO	NO
AT5G46490	0.06	0.03	3.09	5.71	7.41	0.00E+00	NO	NO	NO	NO
AT5G46500	0.38	0.30	30.99	56.23	7.57	0.00E+00	NO	NO	NO	NO
AT5G47630	49.93	41.40	96.89	59.28	0.52	2.95E-02	NO	NO	NO	NO
AT5G48110	0.73	1.84	8.17	13.05	2.83	0.00E+00	NO	NO	NO	NO

Table A3.6 (cont'd)

AT5G48620	4.61	4.51	10.10	14.20	1.66	0.00E+00	NO	NO	NO	NO
AT5G48960	4.94	3.71	32.53	23.14	2.64	0.00E+00	NO	NO	NO	NO
AT5G49360	0.96	1.14	4.55	6.88	2.60	0.00E+00	NO	NO	DOWN	NO
AT5G50915	3.20	2.03	8.64	6.95	1.77	7.91E-07	NO	YES	NO	NO
AT5G51460	3.26	3.50	6.92	7.05	1.01	4.99E-04	NO	NO	NO	NO
AT5G52750	18.24	12.67	47.35	76.53	2.59	0.00E+00	NO	NO	NO	NO
AT5G53460	50.53	36.75	87.25	63.80	0.80	1.38E-07	NO	NO	NO	NO
AT5G54610	10.49	10.41	65.06	95.39	3.20	0.00E+00	NO	NO	NO	NO
AT5G54720	3.54	3.20	17.20	37.96	3.57	0.00E+00	NO	NO	NO	NO
AT5G55450	429.86	232.46	1212.13	1217.23	2.39	0.00E+00	NO	NO	NO	NO
AT5G55460	21.38	17.30	50.62	48.44	1.48	1.09E-07	NO	NO	NO	NO
AT5G55510	17.65	15.66	45.22	27.38	0.81	1.27E-03	NO	NO	NO	NO
AT5G57180	20.19	19.02	55.08	41.30	1.12	0.00E+00	NO	YES	NO	NO
AT5G57700	2.23	3.73	6.77	9.50	1.35	1.92E-06	NO	NO	NO	NO
AT5G60900	7.85	6.90	16.51	30.42	2.14	0.00E+00	NO	NO	NO	NO
AT5G62630	14.13	16.33	35.41	40.32	1.30	3.66E-14	NO	NO	NO	NO
AT5G63120	16.93	12.95	36.74	26.45	1.03	4.00E-09	NO	NO	NO	NO
AT5G64000	10.02	2.43	23.80	17.27	2.83	0.00E+00	NO	NO	NO	NO
AT5G65730	39.44	44.38	79.15	68.68	0.63	1.16E-03	NEGATIVE	NO	NO	NO
AT5G66052	0.05	0.05	10.64	13.49	8.05	8.75E-07	NO	NO	NO	NO
AT5G66053	4.51	0.00	120.50	146.67	Inf	3.87E-03	NO	NO	NO	NO
AT5G66630	0.08	0.14	2.07	4.36	4.97	0.00E+00	NO	YES	NO	NO
AT5G66690	0.38	0.12	10.26	9.50	6.28	0.00E+00	NO	NO	NO	NO

Table A3.7. Primer pairs used for qRT-PCR and sequencing. QRT-PCR Primers used in this study were designed with Primer Express 3.0.

EQTL PRIMERS

AGI	NAME	FW	RV
AT1G27730	ZAT10	TCGAGCACTGGACAAAGGGTAAGC	CCTGAGTGAGGTTTTGGTGGTGGA
AT1G29390	COR314	CCAGGTGAACTTGAGCTTCCA	TGATATGGCGCCACAATCA
AT2G42540	COR15A	CGTTGATCTACGCCGCTAAAG	TGGCCTCGTTGAGGTCATC
AT3G55980	STZ1	TGTCCCCTGTCCCGAGTTC	CGTGAGCATACTCGCAAGAATC
AT5G59820	ZAT12	GTGCGAGTCACAAGAAGCCTAACA	GCGACGACGTTTTACCTTCTTCA
AT1G20440	COR47	CGGTACCAGTGTCTGGAGAGT	ACAGCTGGTGAATCCTCTGC
AT1G20450	ERD10	TTCCAGGCCACAGCAAGAA	CAACCAGCGGTGTGGTGTT
AT1G75040	PR5	AACGGCGGCGGAGTTC	CGCCATCGCCTACTAGAGTGA
AT4G15910	DI21	TGGCCGCTCGTTCACTCT	TGCGGCGGAGCAAAGA
AT5G52310	COR78	GAAAGGAGGAGGAGGAATGG	AACCAGCCAGATGATTTTGG
AT3G02780	IPP2	ATTTGCCCATCGTCCTCTGT	GAGAAAGCACGAAAATTCGGTAA
AT2G41100	TCH3	CAATATCATCGCTTCGCCAAT	ATAACAGCGCTTCGAACAAATCT

SEQUENCING PRIMERS

AGI	NAME	FW	RV
AT4G25490	CBF1	GGGATTGCTTCGCTATGTACTATACACGTGTC	TCCACATATTTTGCCTTGAAAGCAACTACAATATGAC
AT4G25470	CBF2	ATCCAAATATCTTCCGGCCAATATAAACAGCAAGCT	AGCAGAAAGGCAGAAACTGTTGCTATCAACCAAC
AT4G25480	CBF3	CCGCCATTATAAAACAGCATGCTCTCACTCC	GCTGAATCGGTTGTTTCGGTTTTATACTCTAATTTAAAC
AT2G42540	COR15A	GGCCTGAAAAGAACGAACAGAACTCGGTAATG	GGTACAACAATGTATCATACCAATTGATGTCATAC
AT1G20440	COR47	GTTACCTCCGCGTTGGCCTGGACCCTTCTC	CCTAAGAAGCCAGATGGCGAAATATTGAAATATTG
AT5G52310	COR78	CCCTCCTCTGTTTTACTCACAAATATGCAAACTAG	CTTTAACGATAGAGACAACACCTCAACAAGTCAC

Table A3.8. Number of reads passing the Illumina purity filter for each RNA-seq sample. The lane name represents the lane in which each sample was run. ‘PF clusters’ indicates reads passing Illumina purity filters, while ‘Raw Clusters’ denotes all reads. For the sample names: ‘SW’ indicates the Swedish accession; ‘IT’ indicates the Italian accession; ‘NONACC’ indicates the samples not cold-acclimated, ‘1WK’ indicates samples cold-acclimated for 1 week; ‘2WK’ indicates samples cold-acclimated for 2 weeks; the last number in the sample name indicates biological replicate number.

Lane	Sample	Raw Clusters	PF Clusters	Sample % PF
1	SW-NONACC-1	29,958,443	24,697,162	82.4%
4	SW-NONACC-1	29,918,355	24,249,872	81.1%
	SW-NONACC-1 Total	59,876,798	48,947,034	81.7%
1	SW-NONACC-2	25,139,472	20,662,571	82.2%
4	SW-NONACC-2	25,085,948	20,259,178	80.8%
	SW-NONACC-2 Total	50,225,420	40,921,749	81.5%
1	SW-NONACC-3	24,918,522	20,505,290	82.3%
4	SW-NONACC-3	24,880,981	20,140,545	80.9%
	SW-NONACC-3 Total	49,799,503	40,645,835	81.6%
1	IT-NONACC-1	28,636,037	23,420,339	81.8%
4	IT-NONACC-1	28,552,106	22,949,660	80.4%
	IT-NONACC-1 Total	57,188,143	46,369,999	81.1%
1	IT-NONACC-2	24,842,595	20,467,125	82.4%
4	IT-NONACC-2	24,815,959	20,088,847	81.0%
	IT-NONACC-2 Total	49,658,554	40,555,972	81.7%
1	IT-NONACC-3	31,151,487	25,609,805	82.2%
4	IT-NONACC-3	31,118,564	25,143,267	80.8%
	IT-NONACC-3 Total	62,270,051	50,753,072	81.5%
2	SW-1WK-1	28,952,674	23,350,344	80.7%
5	SW-1WK-1	28,715,485	22,777,897	79.3%
	SW-1WK Total	57,668,159	46,128,241	80.0%
2	SW-1WK-2	26,129,644	21,279,472	81.4%

Table A3.8 cont'd

5	SW-1WK-2	25,897,791	20,772,596	80.2%
	SW-1WK-2 Total	52,027,435	42,052,068	80.8%
2	SW-1WK-3	29,760,111	24,072,758	80.9%
5	SW-1WK-3	29,565,270	23,523,986	79.6%
	SW-1WK-3 Total	59,325,381	47,596,744	80.2%
2	IT-1WK-1	29,677,878	24,017,917	80.9%
5	IT-1WK-1	29,401,956	23,424,329	79.7%
	IT-1WK-1 Total	59,079,834	47,442,246	80.3%
2	IT-1WK-2	29,822,085	24,213,801	81.2%
5	IT-1WK-2	29,615,504	23,659,830	79.9%
	IT-1WK-2 Total	59,437,589	47,873,631	80.5%
2	IT-1WK-3	23,782,918	19,711,843	82.9%
5	IT-1WK-3	23,506,197	19,220,083	81.8%
	IT-1WK-3 Total	47,289,115	38,931,926	82.3%
3	SW-2WK-1	28,637,922	23,936,002	83.6%
6	SW-2WK-1	28,377,699	23,775,237	83.8%
	SW-2WK-1 Total	57,015,621	47,711,239	83.7%
3	SW-2WK-2	25,168,550	21,001,016	83.4%
6	SW-2WK-2	24,935,016	20,853,600	83.6%
	SW-2WK-2 Total	50,103,566	41,854,616	83.5%
3	SW-2WK-3	25,484,480	21,280,683	83.5%
6	SW-2WK-3	25,236,381	21,125,228	83.7%
	SW-2WK-3 Total	50,720,861	42,405,911	83.6%
3	IT-2WK-1	32,233,028	26,752,432	83.0%
6	IT-2WK-1	31,955,554	26,583,643	83.2%
	IT-2WK-1 Total	64,188,582	53,336,075	83.1%
3	IT-2WK-2	26,054,424	21,749,785	83.5%
6	IT-2WK-2	25,813,016	21,599,147	83.7%

Table A3.8 cont'd

	IT-2WK-2 Total	51,867,440	43,348,932	83.6%
3	IT-2WK-3	28,738,532	23,962,836	83.4%
6	IT-2WK-3	28,468,237	23,789,534	83.6%
	IT-2WK-3 Total	57,206,769	47,752,370	83.5%
	Grand Total	994,948,821	814,627,660	81.9%

Table A3.9. Summary of SW and IT RNA-seq samples. ‘Average PF Clusters’ indicates the average number of reads passing the Illumina purity filters for each sample. ‘# Genes’ indicates the number of genes in each sample. ‘Average R2’ indicates the average correlation of genes for each biological replicate when all comparisons are made (i.e. replicate 1 to 2, replicate 2 to 3, and replicate 1 to 3) \pm the standard error. While calculating the correlation (R^2) for each sample genes that were outliers were removed, none of the outliers removed from cold samples were genes found to be cold regulated and none of the outliers removed from non-acclimated samples were included in the list of genes differentially regulated between SW and IT. The total number of genes removed from calculating correlation was small (18 genes total) in comparison to the number of genes analyzed (‘# Genes’). Genes removed were: AT1G15800, AT1G26240, AT1G50310, AT1G71070, AT2G19790, AT2G23740, AT2G30370, AT3G13061, AT4G08109, AT4G15020, AT4G15620, AT5G07720, AT5G08480, AT5G20620, AT5G27450, AT5G35900, AT5G47450, and AT5G58787.

Sample	Average PF Clusters	# GENES	Average R2	Maximum FPKM
SW-NONACC Average	43,504,873	23,127	0.90 \pm 0.015	28,114
IT-NONACC Average	45,893,014	23,429	0.90 \pm 0.032	24,553
SW-1WK Average	45,259,018	23,303	0.88 \pm 0.022	42,983
IT-1WK Average	44,749,268	23,123	0.91 \pm 0.012	18,584
SW-2WK Average	43,990,589	22,818	0.90 \pm 0.015	35,927
IT-2WK Average	48,145,792	23,785	0.90 \pm 0.013	20,597

LITERATURE CITED

LITERATURE CITED

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CHAPTER FOUR

INTERESTING OBSERVATIONS AND FUTURE DIRECTIONS

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SUMMARY

The following represents results and preliminary results from several experiments that were conducted during the course of research described in Chapters Two and Three. These experiments produced interesting results that require further research and may be of value to continue as full focused projects. Preliminary results for additional mechanisms of circadian and gated cold-induced expression are described for the *CBFs* and the CBF-independent gene, *COR27*. Temperature-entrained circadian and gated cold-induced expression of *CBF* genes are also examined in this section, as well as the disruption of the circadian clock by extended cold temperatures. Lastly, morphological differences between SW and IT ecotypes used in Chapter Three are documented, and future construction of a low-temperature network using eQTL and RNA-seq data is discussed.

RESULTS AND DISCUSSION

Regulation of CBF genes by the REVEILLES, CK2, and CAMTAS, and circadian regulation of CBF-independent gene, COR27. In Chapter Two of this study, we show that EE-binding transcription factors CCA1 and LHY are major positive regulators of *CBF* genes under both circadian and gated (circadian with cold-treatment) conditions (Fig 2.1 and 2.4). ChIP experiments seem to suggest that this interaction is direct. *CBF*-target gene expression is also affected by mutations in CCA1 and LHY and electrolyte leakage assays indicate that these central clock genes impact freezing tolerance. CCA1 and LHY are thus components of the *CBF* pathway upstream of the *CBFs*. Doherty et al. 2009 determined that CAMTA3 is a transcription factor that also positively regulates *CBF* genes (1). In Chapter Two we present a model that suggests, CCA1, LHY and CAMTA transcription factors (1) work synergistically to increase expression of *CBF* genes (Fig 2.5).

The CAMTAs are hypothesized to be a connection between calcium and *CBF* signaling (1). Cellular calcium levels also oscillate with circadian rhythms (2). Therefore, the CAMTAs may be a second connection between the clock and *CBF* genes. As a first step CAMTA mutants were tested to see if they affect basal circadian and gated cold-induction of *CBF* genes. There are a total of six CAMTAs in *Arabidopsis*, some of which have been shown to have overlapping function ((1) and Kim and Thomashow, unpublished). Preliminary results indicate that peak expression of *CBF1* and *2* is reduced under circadian conditions in a *camta3/camta2* double mutant (mutant constructed by Kim and Thomashow, unpublished), which is consistent with the

CAMTAs acting as activators of *CBF1* and *2* (1). This also suggests that CAMTAs influence basal expression of *CBF* genes. The reduction of *CBF* expression in *camta3/camta2* was not as severe as in *cca1-11/lhy-21* mutants. Gated cold-induced expression of *CBF1-3* also appears to be reduced though the reduction is greater for *CBF1* and *CBF2* than *CBF3*. The reduction of gated *CBF* expression in *camta3/camta2* was not as severe as in *cca1-11/lhy-21* mutants. This result needs to be repeated, but does support the model proposed in Chapter Two.

Although *CBF1* and *CBF3* expression is essentially knocked-out under both circadian and gated conditions in the *cca1-11/lhy-21* double mutant, there is some residual (~20%) expression of *CBF2* (Fig. 2.1 and 2.4). In Chapter Two we suggest that the CCA1-like REVEILLE (RVE) genes may play a role in the remaining induction of *CBF2*. Like CCA1 and LHY, the RVEs bind to the EE and are circadian regulated. Preliminary results indicate that *rve2*, *rve3*, *rve4*, and an additional EE-binding *tcp2* mutant do not significantly affect *CBF* expression, under both circadian and gated conditions, though higher order mutants would need to be constructed since there may be redundancy between these genes.

Preliminary results show that the *rve8-1* mutant may have altered *CBF* expression in comparison to wild-type. However, *CBF* expression appears to be elevated in *rve8-1* rather than reduced. This would suggest that RVE8 functions as a repressor rather than an activator of *CBF* expression like CCA1 and LHY. *RVE8* transcripts peak at the same time as *CCA1* and *LHY*, but Rawat et al. 2011 demonstrated that the RVE8 protein does not accumulate until after subjective noon (3),

when *CBF* expression levels drop off (Fig. 2.1). Therefore, circadian oscillations of RVE8 are consistent with RVE8 functioning as a repressor of *CBF* expression. Interestingly, RVE8 was recently found to promote expression of *PRR5* (3), which has previously been implicated as a repressor of *CBF* expression (4). Therefore, RVE8 may potentially acts as a direct or indirect regulator of *CBF* expression.

CCA1 and LHY are controlled by transcriptional as well as post-translational mechanisms (5, 6). Phosphorylation of CCA1 and LHY by CASEIN KINASE 2 (CK2) is necessary for normal clock function and can affect DNA binding (6, 7). Therefore, CK2 was tested to determine if it is necessary for induction of the *CBF* genes. Preliminary time-course experiments using the *ck2 β 3* mutant indicate that circadian expression, as well as gated cold-induction of *CBF1-3*, and *CBF*-target genes are not strongly affected. This suggests that post-translational modification of CCA1 and LHY is not necessary for binding to the *CBF* locus, but experiments need to be repeated.

The CBF regulon only comprises 12% of cold-regulated genes (8). Therefore, it is of interest to determine if CCA1 and LHY also regulate CBF-independent pathways of cold-signaling. The cold regulated gene, *COR27*, is independent of the CBF pathway, and also gated by the circadian clock (9). Promoter analysis of *COR27* demonstrated that the EE has a role in the cold-induction of *COR27* (9). However, the transcription factors mediating cold-induction through the *COR27*-EEs have yet to be identified (9). EE-binding transcription factors CCA1 and LHY seem like especially good candidates because of their role in the induction of *CBF* genes (10).

Under circadian conditions, peak *COR27* expression in the *cca1-11/lhy-21* double mutant is not reduced nearly as much as that of the *CBF* genes (Fig 4.1; Fig A4.1; Fig. 2.1). Although more biological replicates may be necessary, five independent biological replicates of ChIP experiments did not find statistically significant enrichment of CCA1 on the *COR27* promoter or coding region (Fig 4.2). Gated cold-induced expression of *COR27* is also not strongly reduced in *cca1-11/lhy-21* in comparison to wild-type (Fig. 4.1). Therefore, CCA1 and LHY do not appear to play a major role in circadian regulation or gated cold-induction of *COR27* (Fig. 4.1). This suggests that there is specificity for CCA1 and LHY in the regulation of *CBF* signaling and that there are multiple mechanisms for gating cold gene expression.

Contrary to previous reports that *CBF* and *COR27* genes have similar peaks and troughs of expression (9), data from this study suggests that *COR27* peaks in expression a few hours after *CBF* genes (Fig 4.1; Fig 2.1 and 2.4). The shift in peak expression is most likely due to the greater number of time-points used in this study. The difference in expression pattern between *CBF* and *COR27* seems significant and may be the foundation for differing modes of circadian regulation. Further work must be conducted in order to determine the major mechanism of circadian and gated cold-expression of *COR27*. The previously mentioned RVE genes are known to bind to the EE (11). Many of these genes peak in expression before *COR27*, and thus are good first candidates for regulating *COR27* expression.

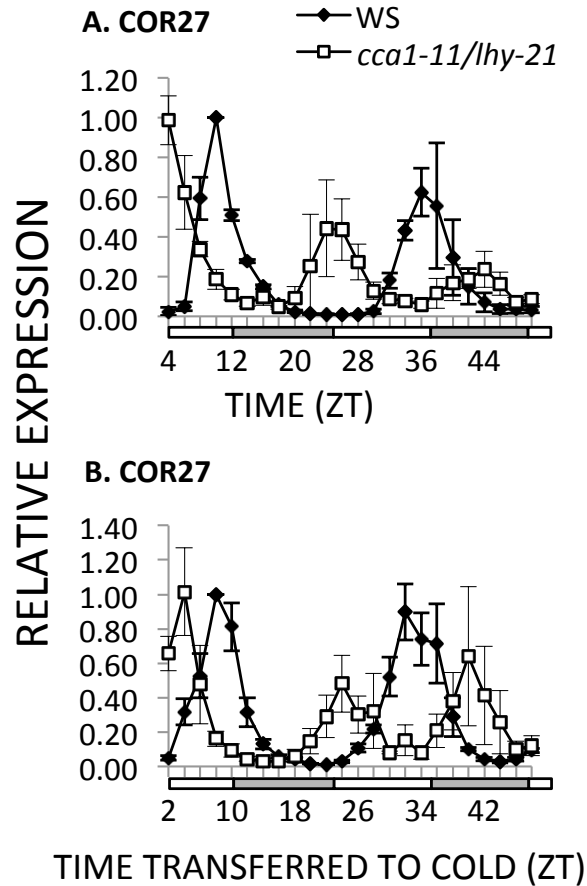


Figure 4.1. Effects of the *cca1-11/lhy-21* double mutation on circadian and gated cold-induction of *COR27*. Expression for *COR27* was relative to one wild-type sample (WS) set to a value of 1 in each biological replicate. Values are averages from three independent experiments (n=3); error bars indicate \pm SEM. (A) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were harvested every 2 h and the transcript levels for *COR27* were determined by qRT-PCR. Gene expression was normalized to IPP2 for each sample. (B) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were transferred to cold temperature (4°C) for 2 h, every 2 h, for 48 h at the start of constant light conditions and the transcript levels for *COR27* were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were dropped to cold temperature. Gene expression was normalized to UBQ10 for each sample. See Table A4.1 for primer pair sequences.

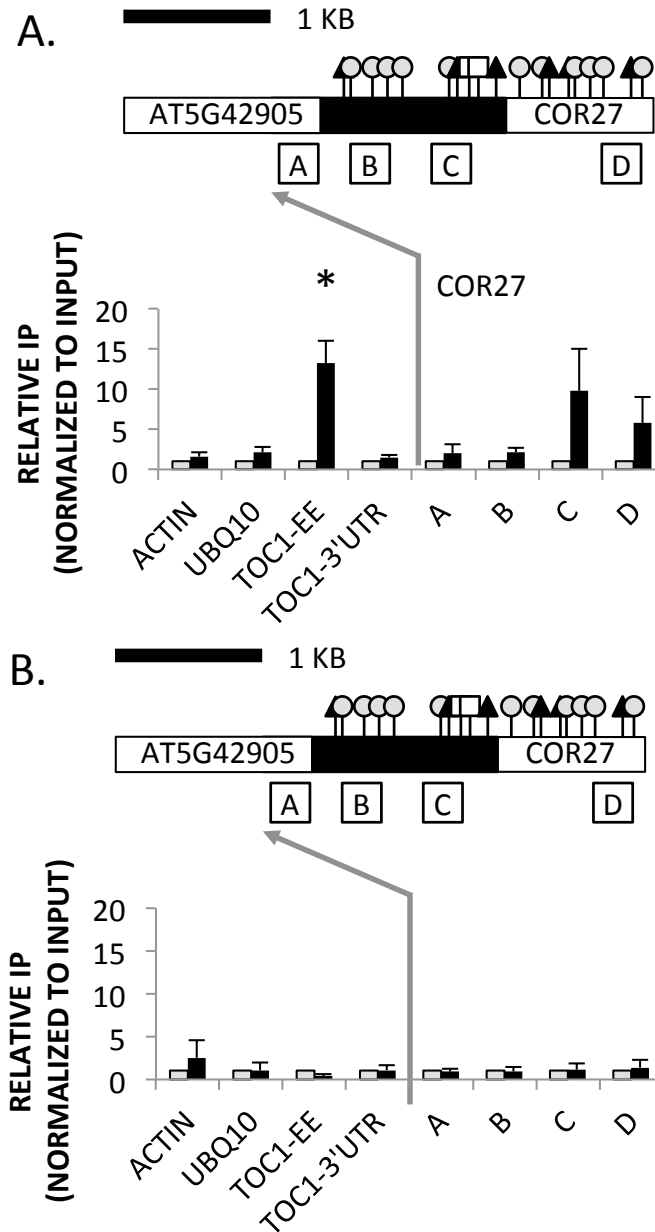


Figure 4.2. Binding of CCA1 on *COR27*. *cca1-1* and *cca1-1 CCA1p:CCA1-GFP* plants were grown at 22°C under a 12 h photoperiod to the four leaf stage. Tissue was fixed at ZT4 and ChIP was performed using an anti-GFP (A) or mock IgG (B) antibody. Immunoprecipitated DNA was quantified by qRT-PCR with primers on *COR27* (boxes A through D). Levels of immunoprecipitated DNA were normalized to input DNA. Immunoprecipitation in *cca1-1 CCA1p:CCA1-GFP* plants (black bars) is relative to *cca1-1* plants (gray bars) set to 1. Primers TOC1-3'UTR, *ACTIN7* (*ACTIN*) and *UBQUITIN10* (*UBQ10*) were negative controls. TOC1-EE was a positive control. Values are the average of five independent experiments (n=5). Error bars indicate \pm SEM. *= $p < 0.05$ using a paired, one-tailed t-test. In the *COR27* diagram, the transcribed regions are indicated with white boxes and the approximate positions of CBS (AATCT), and EE (AAAATATCT) motifs are indicated by gray circles, and white squares respectively.

Temperature-entrained circadian regulation and gated cold-induction of CBF genes. Both light and temperature cues entrain the circadian clock to its external environment (12, 13). *CBF2*, *CBF3* and the CBF-independent cold-regulated gene, *COR27*, all oscillate under temperature-entrained circadian conditions (Fig. 4.3). Temperature-entrained circadian oscillations of *CBF1* do not appear regular and therefore are not as robust as under light-entrained circadian conditions (Fig 4.3; Fig 2.1). This is interesting and may suggest that *CBF1* is distinctively regulated by light in comparison to *CBF2* and *CBF3*. In Chapter Two we establish that CCA1 and LHY play a major role in the induction of *CBF* genes under light-entrained circadian conditions. Consequently, it is of interest to determine if CCA1 and LHY also play a major role in the induction of *CBF* genes under temperature-entrained circadian conditions.

Peak expression of *CBF1-3* is significantly reduced in the *cca1-11/lhy-21* double mutant under temperature-entrained circadian conditions (Fig 4.3). CBF-target genes, *COR15A*, *COR47* and *COR78* also have reduced peak expression in *cca1-11/lhy-21* in comparison to wild-type (Fig 4.4). This suggests that CCA1 and LHY play a major role in the basal induction of *CBF* genes under temperature-entrained circadian conditions. This also would suggest a similar mechanistic model for circadian regulation under temperature-entrained and light entrained conditions (Fig 2.5). However, analysis of clock and cycling genes reveals that there may be a more widespread reduction of rhythmicity in *cca1-11/lhy-21* under temperature-entrained circadian conditions in comparison to light entrained conditions.

COR27 expression is significantly reduced in *cca1-11/lhy-21* under temperature-entrained conditions (Fig 4.3), which is interesting considering that *COR27* expression is not strongly reduced in *cca1-11/lhy-21* under light-entrained circadian conditions (Fig. 4.1 and 4.3). Moreover, cycling control gene, *LHCB1.4*, also has significantly reduced peak expression in the *cca1-11/lhy-21* under temperature-entrained conditions (Fig. 4.4), which is not the case under light-entrained conditions (Fig A2.3). Therefore expression of clock components, *TOC1*, *PRR3*, *PRR5*, *PRR7* and *PRR9*, were further tested in *cca1-11/lhy-21* under temperature-entrained circadian conditions (Fig. 4.5). Previous literature has shown that the *cca1-11/lhy-21* mutant has robust oscillations of *TOC1* under light-entrained conditions (14). Under temperature-entrained circadian conditions *TOC1*, *PRR3*, *PRR5*, and *PRR7* have significantly reduced peak expression in *cca1-11/lhy-21* in comparison to WT (Fig 4.5). Conversely, *PRR9* maintains robust oscillations in *cca1-11/lhy-21* under temperature-entrained conditions (Fig. 4.5). Previous literature has shown robust cycling of clock-output genes *CAB2* (15, 16) and *CCR2* (14) in *cca1-11/lhy-21*, under light-entrained conditions. Therefore, expression of cycling control genes, *CAB2* and *CCR2*, were tested under temperature-entrained circadian conditions. *CAB2* and *CCR2*, did show robust oscillations in *cca1-11/lhy-21*, with peak levels similar to wild-type (WS-2) under temperature-entrained conditions (Fig 4.4). Therefore, not all oscillations are knocked-out in *cca1-11/lhy-21* under temperature-entrained conditions. Consequently, there is some specificity for the reduction in *CBF* gene expression under temperature-entrained conditions by CCA1 and LHY, in support of a mechanistic model similar to light-entrained conditions (Fig

2.5). But a more general clock question also seems to emerge from this data. To what extent to do CCA1 and LHY function differently under light- and temperature-entrained circadian conditions? To determine the extent of these differences, light and temperature cycling time-course microarrays or RNA-seq data would have to be performed for *cca1-11/lhy-21* and wild-type (WS-2) plants. This type of data for *cca1-11/lhy-21* does not appear to be publicly available, though there are light and temperature-entrained arrays publically available for wild-type plants.

Significant progress has been made in determining the targets of light input into the clock (17-21), and temperature entrainment of the circadian clock is well established (12). However, mechanisms underlying temperature entrainment are still largely unknown (13, 22). Previous literature suggests that PRR7 and PRR9 are important for temperature-entrainment (22). This was shown through arrhythmic *CCA1* and *TOC1* expression as well as arrhythmic hypocotyl movement in *prr7-3/prr9-1* under temperature-entrained conditions (22). The *cca1-11/lhy-21* mutant also fails to maintain circadian oscillations of several clock genes, as well as cycling control gene *LHCB1.4* under temperature-entrained conditions. CCA1 and LHY have been previously connected to temperature compensation of the clock (23). But data from this study suggests that CCA1 and LHY are also important for temperature entrainment of the circadian clock. This hypothesis should be further tested using other clock-output measurements such as hypocotyl movement.

Fowler et al. 2005 demonstrated that cold-induced expression of *CBF* genes is gated by a light-entrained circadian clock (24). Therefore cold-induced gene expression

was tested under temperature-entrained circadian conditions to see if it is gated as well. Indeed, cold-induced expression of *CBF1-3*, CBF-target genes and *COR27* is gated under temperature-entrained circadian conditions (Fig 4.6 and 4.7). In Chapter Two we show that CCA1 and LHY play a major role in the gated cold-induced expression of *CBF* genes. Therefore, it is of interest to determine if CCA1 and LHY influence gated cold-induction of *CBF* genes under temperature-entrained circadian conditions. The peak cold-induced expression of *CBF1-3* and CBF-target genes is significantly reduced in *cca1-11/lhy-21* under temperature-entrained conditions (Fig. 4.6 and 4.7), similar to light-entrained conditions (Fig 2.4), which suggests that CCA1 and LHY also are vital for the gated cold-induction of *CBF* genes under temperature-entrained conditions and might imply a similar gating mechanism to the one proposed for gated cold-induced *CBF* expression under light-entrained circadian conditions (Fig. 2.5). However, there again appears to be a general reduction in rhythmicity as well as expression level in *cca1-11/lhy-21* under gated temperature-entrained conditions.

Under both basal temperature-entrained circadian conditions and gated temperature-entrained conditions the *cca1-11/lhy-21* double mutant appears to have a more generalized reduction in gene expression in comparison to light-entrained conditions. This sensitivity seems to be intensified with the addition of a cold treatment (gated conditions) since *COR27* and all cycling control genes had significantly reduced peak expression in comparison to wild-type (Fig 4.7). Clock genes *TOC1*, *PRR3*, *PRR5*, and *PRR7*, are arrhythmic in *cca1-11/lhy-21* under gated temperature-entrained circadian conditions, similar to basal temperature-entrained circadian conditions.

However, *PRR9* is also arrhythmic under gated temperature-entrained circadian conditions (Fig 4.8), which was not the case under basal temperature-entrained circadian conditions (Fig. 4.5). *CCA1* and *LHY* have previously been shown to be important for temperature compensation of the clock under light-entrained conditions (23). The increased arrhythmia seen with the addition of a cold treatment under temperature-entrained conditions may be due to the role of *CCA1* and *LHY* in temperature compensation of the clock under temperature-entrained conditions. However, further experiments would need to be conducted to support this claim.

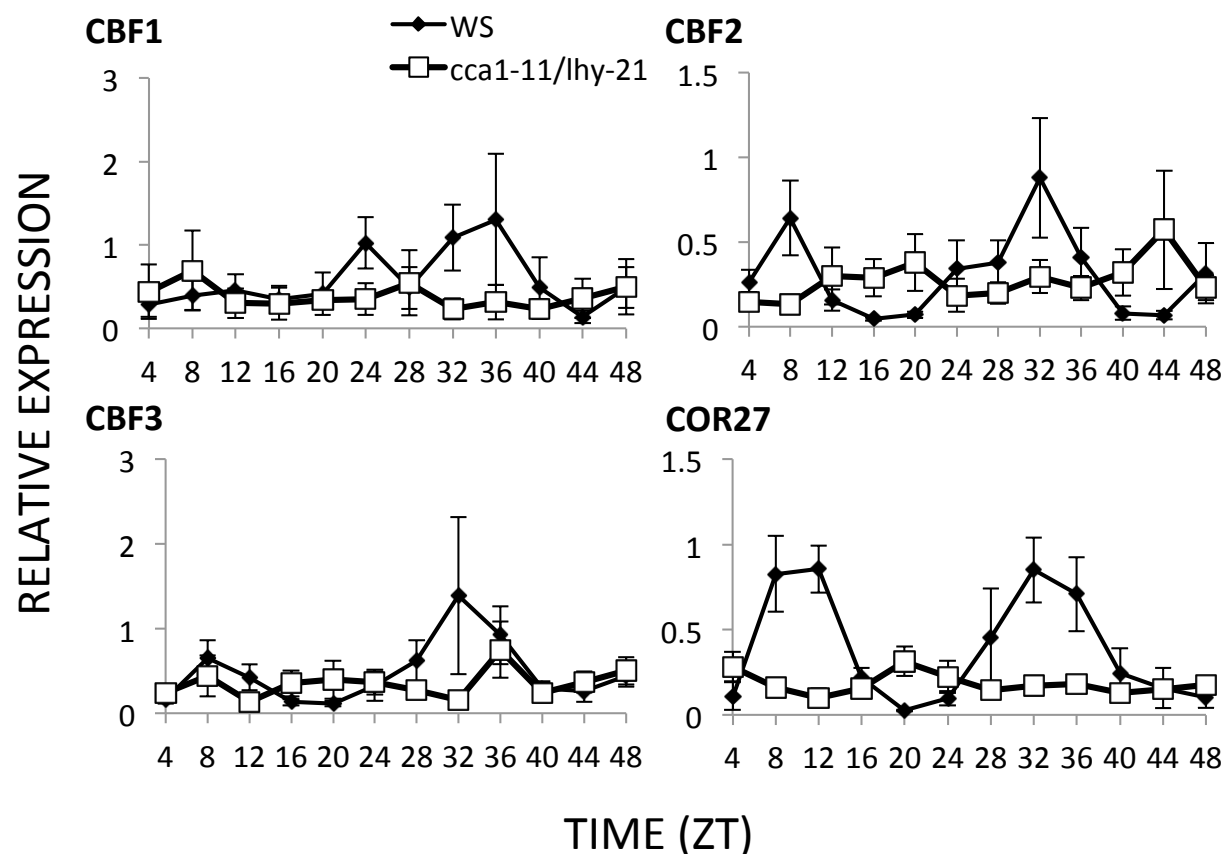


Figure 4.3. Effects of the *cca1-11/lhy-21* double mutation on temperature-entrained circadian regulation of *CBF1*, *CBF2*, *CBF3* and *COR27*. Wild-type *Ws-2* (WS) and *cca111/lhy-21* double mutant plants were grown at constant light under a 12 h temperature period (22°C day/ 12°C night) to the four leaf stage and transferred to constant temperature (22°C) at ZT0. Plants were harvested every 4 h and the transcript levels for the indicated genes were determined by qRT-PCR. Gene expression was normalized to *IPP2* for each sample. Gene expression is relative to one wild-type sample set to a value of 1 for each biological replicate. Values are averages from four independent biological experiments (n=4). Error bars indicate \pm SEM. Please see Table A4.1 for primer pair sequences.

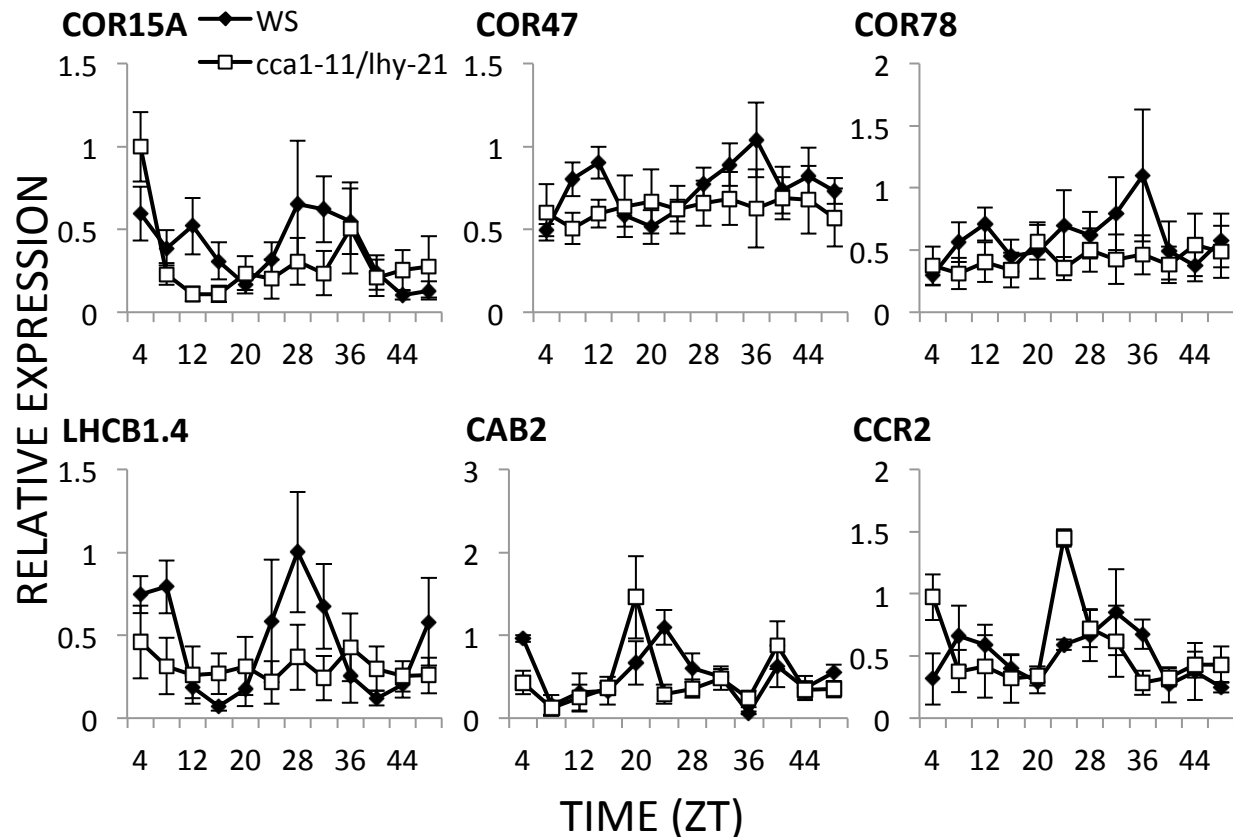


Figure 4.4. Effects of the *cca1-11/lhy-21* double mutation on temperature-entrained circadian regulation of CBF target genes, *COR15A*, *COR47* and *COR78*, and cycling control genes *LHCB1.4*, *CAB2* and *CCR2*. Wild-type Ws-2 (WS) and *cca111/lhy-21* double mutant plants were grown at constant light under a 12 h temperature period (22°C day/ 12°C night) to the four leaf stage and transferred to constant temperature (22°C) at ZT0. Plants were harvested every 4 h and the transcript levels for the indicated genes were determined by qRT-PCR. Gene expression was normalized to IPP2 for each sample. Gene expression is relative to one wild-type sample set to a value of 1 for each biological replicate. Values are averages from four independent biological experiments (n=4). Error bars indicate \pm SEM. Please see Table A4.1 for primer pair sequences.

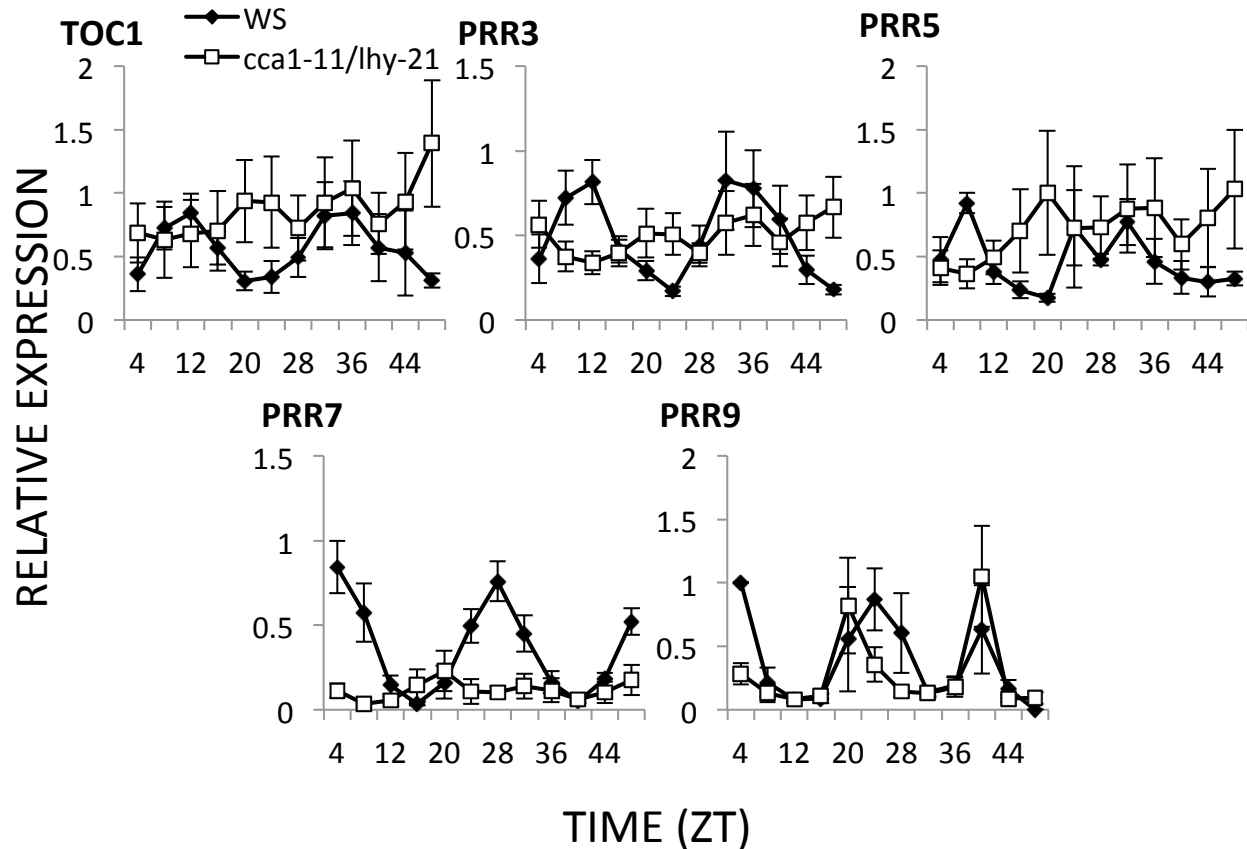


Figure 4.5. Effects of the *cca1-11/lhy-21* double mutation on temperature-entrained circadian regulation of clock genes *TOC1*, *PRR3*, *PRR5*, *PRR7* and *PRR9*. Wild-type Ws-2 (WS) and *cca111/lhy-21* double mutant plants were grown at constant light under a 12 h temperature period (22°C day/ 12°C night) to the four leaf stage and transferred to constant temperature (22°C) at ZT0. Plants were harvested every 4 h and the transcript levels for the indicated genes were determined by qRT-PCR. Gene expression was normalized to IPP2 for each sample. Gene expression is relative to one wild-type sample set to a value of 1 for each biological replicate. Values are averages from four independent biological experiments (n=4). Error bars indicate \pm SEM. Please see Table A4.1 for primer pair sequences.

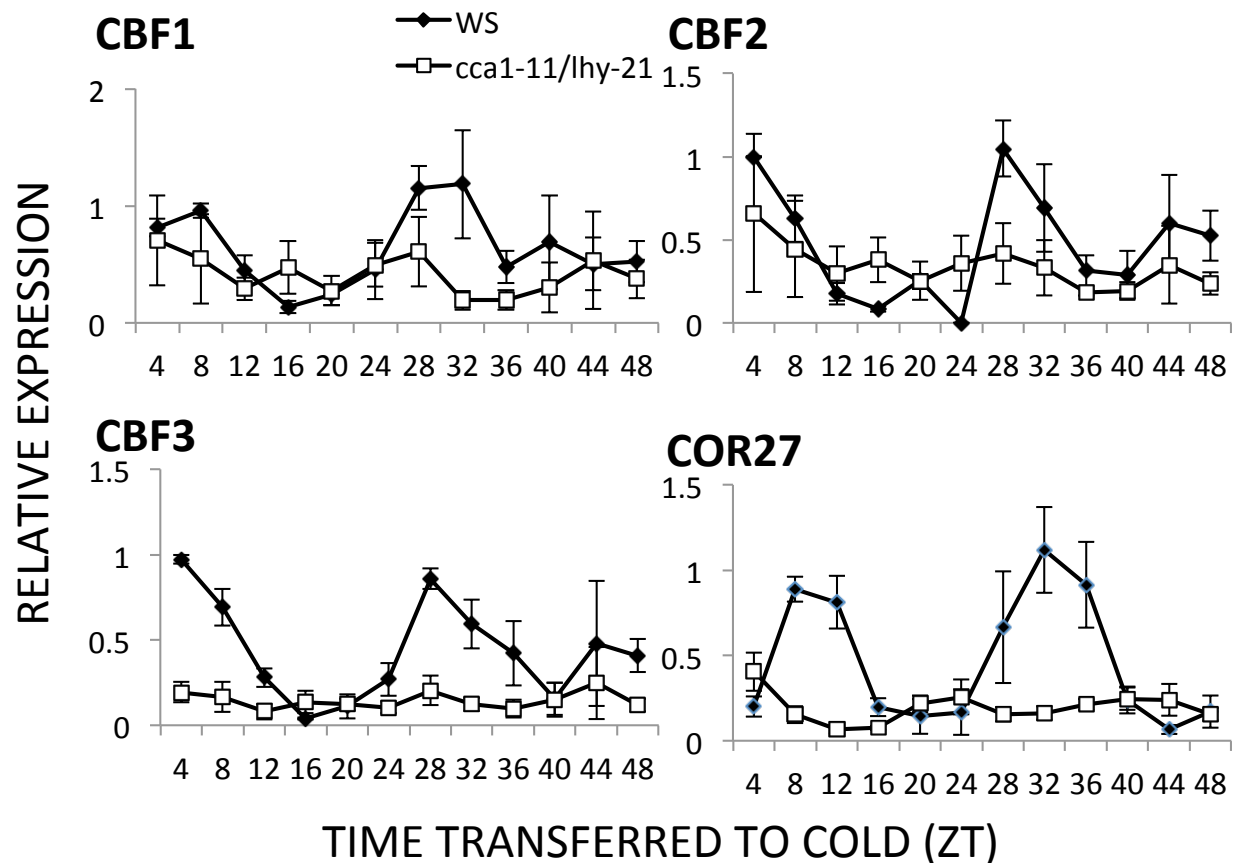


Figure 4.6. Effects of the *cca1-11/lhy-21* double mutation on temperature-entrained cold-induction of *CBF1*, *CBF2*, *CBF3* and *COR27*. Wild-type Ws-2 (WS) and *cca1-11/lhy-21* double mutant plants were grown under constant light with a 12 h temperature period (22°C day/ 12°C night) to the four leaf stage and transferred to constant light at ZT0. Plants were transferred to cold temperature (4°C) for 2 h, every 4 h at the start of constant temperature conditions. Transcript levels for the indicated genes were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were transferred to cold temperature. Gene expression was normalized to UBQ10 for each sample. Gene expression is relative to one wild-type sample set to a value of 1 for each biological replicate. Values are averages from four independent biological experiments (n=3). Error bars indicate \pm SEM. Please see Table A4.1 for primer pair sequences.

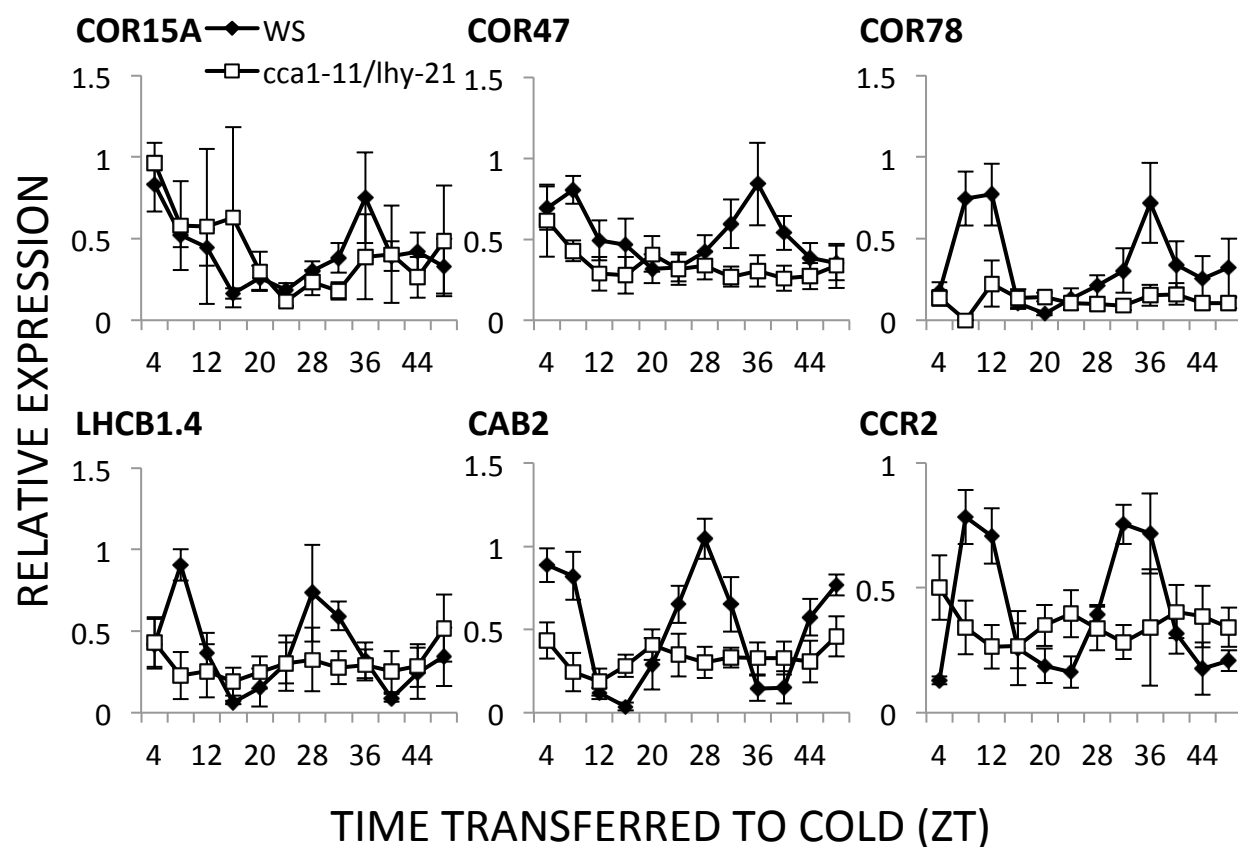


Figure 4.7. Effects of the *cca1-11/lhy-21* double mutation on temperature-entrained cold-induction of CBF target genes, *COR15A*, *COR47* and *COR78*, and cycling control genes *LHCB1.4*, *CAB2* and *CCR2*. Wild-type Ws-2 (WS) and *cca1-11/lhy-21* double mutant plants were grown under constant light with a 12 h temperature period (22°C day/ 12°C night) to the four leaf stage and transferred to constant light at ZT0. Plants were transferred to cold temperature (4°C) for 2 h, every 4 h at the start of constant temperature conditions. Transcript levels for the indicated genes were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were transferred to cold temperature. Gene expression was normalized to UBQ10 for each sample. Gene expression is relative to one wild-type sample set to a value of 1 for each biological replicate. Values are averages from four independent biological experiments (n=3). Error bars indicate \pm SEM. Please see Table A4.1 for primer pair sequences.

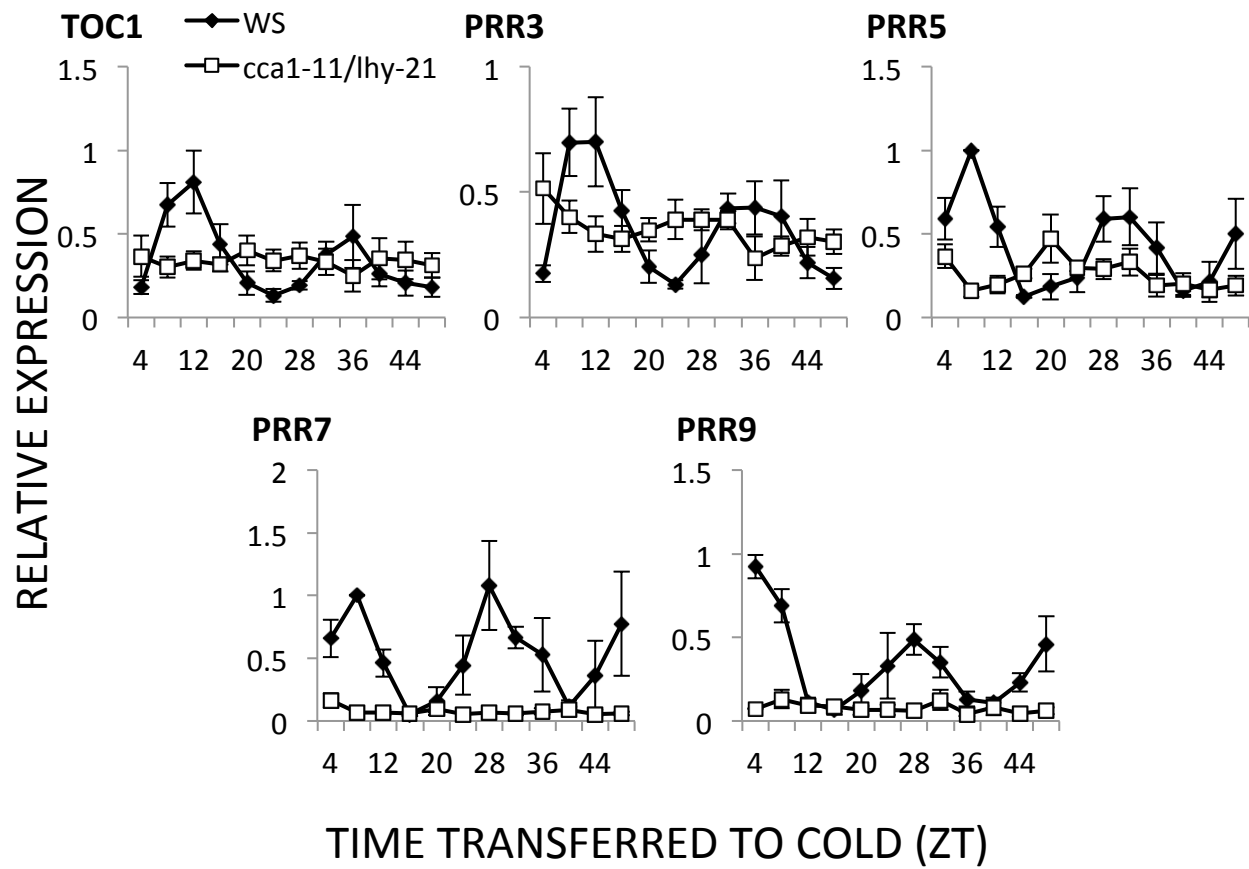


Figure 4.8. Effects of the *cca1-11/lhy-21* double mutation on temperature-entrained cold-induction of PRR clock genes *TOC1*, *PRR3*, *PRR5*, *PRR7* and *PRR9*. Wild-type Ws-2 (WS) and *cca1-11/lhy-21* double mutant plants were grown under constant light with a 12 h temperature period (22°C day/ 12°C night) to the four leaf stage and transferred to constant light at ZT0. Plants were transferred to cold temperature (4°C) for 2 h, every 4 h at the start of constant temperature conditions. Transcript levels for the indicated genes were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were transferred to cold temperature. Gene expression was normalized to UBQ10 for each sample. Gene expression is relative to one wild-type sample set to a value of 1 for each biological replicate. Values are averages from four independent biological experiments (n=3). Error bars indicate \pm SEM. Please see Table A4.1 for primer pair sequences.

Disruption of the circadian clock by low-temperature. In 2005, Ramos et al. concluded that the chestnut tree circadian clock was disrupted with extended cold treatment (25). Bieniawska et al. 2008 further explored the disruption of circadian and diurnal oscillations by extended cold in *Arabidopsis* (26). They showed that extended cold treatment disrupts circadian and diurnal oscillations of all clock output genes (*CCR1*, *CCR2*, *CAB2*, *CAT3*), clock components (*PRR3*, *PRR5*, *PRR7*, *PRR9*, *ELF3*, *ELF4*, *GI*, *TOC1*, *CCA1*, *LHY*), and cold-genes (*CBF1-3*, *COR15A*, *COR47*, *COR78*) tested (26). These results are very interesting, but the reported induction pattern of *CBF1-3* genes was also surprising (26). Bieniawska et al. 2008 show *CBF* expression quickly increasing within 2 h of cold treatment then staying at this maximal level for over 2 days (26). In previous experiments performed under constant-light conditions in constant-light grown plants, *CBF* genes also consistently reached maximal levels of cold-induced expression by 2 h (27-29). However, this maximum *CBF* expression level is then reduced significantly (though still higher than warm expression levels) by 24 h in cold (27-29) and remains down for at least 3 weeks (28). Experiments were conducted similar to Bieniawska et al. 2008 to determine if cold-induced *CBF* kinetics in plants grown under entrained conditions is significantly different from previously reported constant-light experiments (27-29). Preliminary experiments (2 biological reps; Fig. 4.9) indicate very different cold-induced *CBF* kinetics from Bieniawska et al. 2008 (26), but similar kinetics to previous constant-light experiments (27-29). There are some differences experimental set-up in comparison to Bieniawska et al. 2008. For example, plants were entrained under a 12 h photoperiod rather than long day condition (26)

before release into constant light conditions. Plants were also transferred to cold temperature at ZT4, rather than at ZT14 (26). These differences in experimental set up could be significant so future experiments should be conducted exactly as they were described in Bieniawska et al. 2008. Yet, more significant might be technical differences in the amount of RNA used for qRT-PCR. Bieniawska et al. 2008, used 2500ng of RNA to synthesize cDNA for expression analysis of *CBF* genes by qRT-PCR. The 25ng-200ng of RNA, used in this study, fell well into the dynamic range of *CBF* genes, whereas 2500ng did not. Therefore, it is possible that differences seen in kinetics of *CBF* genes are the result of technical rather than biological differences, but further experiments are required.

Ramos et al. 2005 used northern blots to show that circadian oscillations of clock genes, *TOC1* and *LHY*, were disrupted with extended cold treatment in chestnut trees (25). These experiments used plants grown in natural winter conditions with both cycling light and temperature (25). Consequently, Ramos et al. concluded that circadian clocks do not function during winter months (25). Although this may be the case in chestnut trees, *Arabidopsis* carrying null mutations in either *TOC1* (30) or *LHY*(31) can continue to have robust rhythms. Furthermore, *Arabidopsis* carrying null mutations in both *TOC1* and *LHY* also have continued rhythmicity (32). In a more extensive analysis of clock gene disruption by cold, Bieniawska et al. 2008 also saw interrupted oscillations of many clock genes (26). However, clock component *LUX* continued to cycle under diurnal conditions (26). This indicates that there is continued rhythmicity under extended cold and thus the possibility of a functioning 'low-temperature' oscillator.

Furthermore, *LUX* may not be the only cold oscillating gene in the *Arabidopsis* transcriptome (26). Therefore, the extent to which genes oscillate under these low-temperature conditions should continue to be examined. Moreover, Ramos et al. 2005 sampled plants in natural conditions with both diurnal cycles and changing day and night temperature, whereas Bieniawska et al. 2008 sampled plants only under circadian and diurnal conditions (25, 26). Both light and temperature entrain the circadian clock and temperature oscillations could be especially important under low-temperature conditions. Accordingly, it is of interest to determine if the disruption of clock gene oscillations by cold also occurs in plants sampled under temperature cycles (comparable to a diurnal treatment; Fig. 4.10).

In preliminary experiments, robust rhythms of central clock component, *CCA1*, were observed in samples without cold treatment ('WARM'), similar to Bieniawska et al. 2008. Disrupted rhythmicity of *CCA1* with extended cold treatment ('COLD') was also observed, similar to Bieniawska et al. 2008 (Fig 4.10). Interestingly, plants sampled under thermocycles ('TEMP CYCLING'), continued to show regular though widened oscillations (Fig. 4.10). However one caveat for these experiments is that plants were grown under 12 h photoperiods then transferred to 12 h thermocycles (9°C day and 4°C night) for sampling (Fig. 4.10). Previously, 8°C treatments have been used for chilling-stress in peppers (33), 7°C treatments for wheat (34), 6°C treatments for soybean (35), and 5°C treatments for tomato (36). Consequently, future experiments should be set up using colder temperature cycles such as a 6°C day with 2°C night, or 4°C day with 0°C night. Nevertheless, preliminary results suggest the possibility that the circadian clock

functions under low-temperature conditions, which should be further explored in the future.

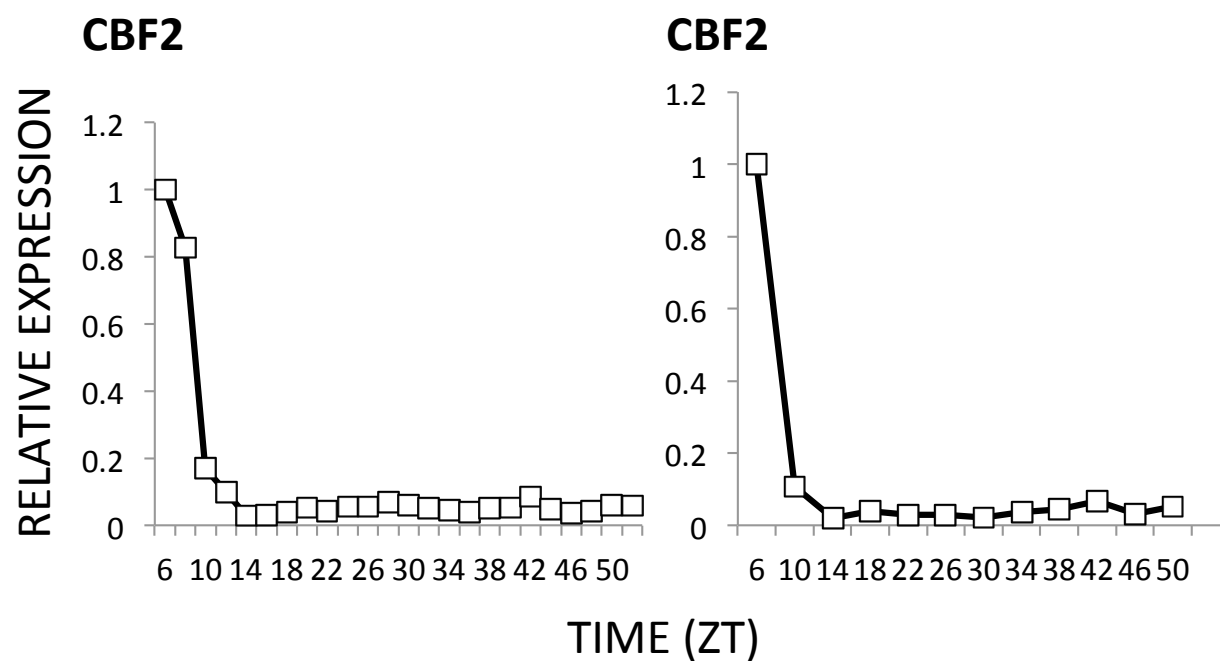


Figure 4.9. Cycling of *CBF2* expression in plants grown under light-entrained conditions then sampled in constant-cold conditions. Wild-type, Ws-2, plants were grown at 22°C with a 12 h photoperiod to the four leaf stage and then transferred to constant light and constant cold (4°C) conditions at ZT4. Transcript levels for the indicated genes were determined by qRT-PCR. Gene expression was normalized to UBQ10 for each sample. Gene expression is relative to one sample set to a value of 1 in each experiment. Two biological replications are shown (left graph and right graph). Please see Table A4.1 for primer pair sequences.

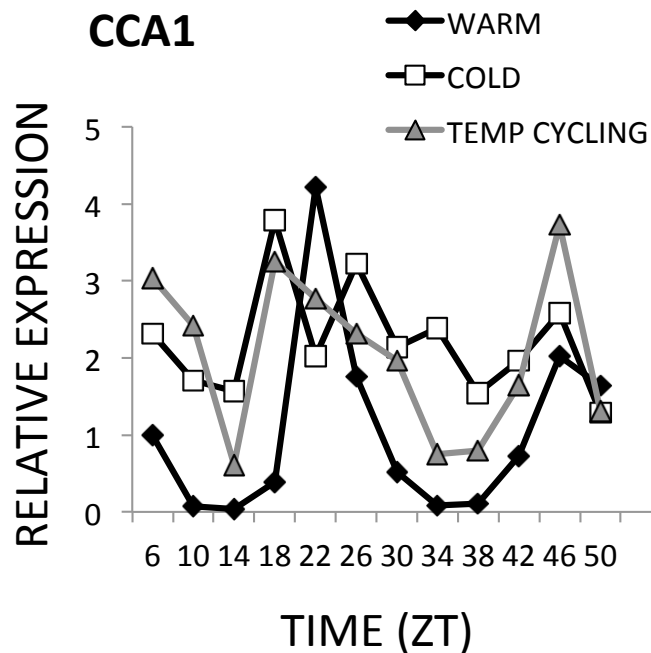


Figure 4.10. Cycling of *CCA1* under warm, temperature cycling or constant-cold conditions. Wild-type, Ws-2, plants were grown at 22°C with a 12 h photoperiod to the four leaf stage and then transferred to one of three conditions at ZT4. 'WARM' plants were transferred to constant light and constant temperature (22°C) conditions. 'TEMP CYCLING' plants were transferred to constant light with a 12 h cold temperature period (9°C day/ 4°C night). 'COLD' plants were transferred to constant light and constant cold (4°C) conditions. Transcript levels for *CCA1* were determined by qRT-PCR. Gene expression was normalized to *UBQ10* for each sample. Gene expression is relative to one sample set to a value of 1 for each condition. Values are from one experiment (n=1). Please see Table A4.1 for primer pair sequences.

Morphological differences between SW and IT. Morphological differences between the SW and IT ecotypes, used in Chapter Three of this study, were observed but not quantified. SW leaf edges were rounded, whereas IT leaf edges were variegated. SW and IT plants also appear to grow at different rates, since IT has more leaves than SW at the same number of days in age. SW leaves seem to be thicker than IT at all time-points examined (non-acclimated, 1 week, 2 weeks and 3 weeks cold-acclimated). Interestingly, plants overexpressing *CBF* genes have thicker leaves than wild-type plants and are constitutively freezing tolerant, (37). There is also a strong positive correlation between leaf thickness and photosynthetic capacity (38). Increased photosynthetic capacity is also associated with cold-acclimation (39). This is especially interesting because RNA-seq data in Chapter Three shows that the majority of differentially expressed photosynthesis genes have higher expression in SW (Fig. 3.33). SW leaf thickness also seemed to increase with cold, which is a change associated with cold-acclimation (40-43) and cold developed leaves (43). However, these morphological changes and differences between SW and IT will have to be measured experimentally.

Low-temperature network construction using SW and IT RILs. In Chapter Three of this study, eQTL mapped for genes associated with differences in freezing tolerance (FTD genes), which also fall under fitness QTL, were detailed. The gene hypothesized to be under the FTD eQTL on chromosome 4, which overlaps with a fitness QTL, is *CBF2*. The FTD eQTL on chromosome 5, overlapping with a fitness

QTL, may be a cis-eQTL, implicating COR78 as the underlying gene. Alternatively, the eQTL on chromosome 5 may be a trans-eQTL, which is postulated to be CBF4. As stated in Chapter Three, near-isogenic lines and complementation lines will need to be made for candidate genes underlying these eQTL, to establish their importance for differential expression of cold-regulated genes in SW and IT. These lines will also be employed to ascertain the role of these genes in freezing tolerance and fitness.

There were also eQTL mapped for genes associated with differences in freezing tolerance that did not fall under fitness QTL (Table 3.18 and 3.19). These genes are not included in the CBF-regulon and for most of these genes specific roles in cold-acclimation are yet to be established. Many of the eQTL were located trans- to the gene they were mapped for and thus potentially represent regulatory loci for CBF-independent cold-regulated genes (Table 3.18 and 3.19), which is especially interesting because relatively little is known about CBF-independent pathways of cold acclimation (44).

Hundreds of genes underlie these trans-eQTL. However, RNA-seq data described in detail in Chapter Three can be used to define a smaller set of candidate regulatory genes. For example, SALT-INDUCIBLE ZINC FINGER 1 (SZF1) is a cold-regulated transcription factor previously described as down-regulated at 1 week of cold-acclimation in *Arabidopsis* (29). Indeed, RNA-seq data from this study also shows SW and IT both down-regulating SZF1 with 1 week of cold-acclimation (Fig 4.11). But SW and IT have significantly different *SZF1* expression at 2 weeks of cold-acclimation, when eQTL were mapped (Fig. 4.11). The trans-eQTL mapped for *SZF1* on chromosome 5

encompasses 805 genes. Of these 805 genes, 108 are differentially regulated between SW and IT at 2 weeks of cold-acclimation, which is when *SZF1* expression is significantly different between SW and IT (Fig. 4.11). There are limitations to this candidate gene list, for instance it would not include genes that have differences in post-transcriptional regulation between SW and IT. Nevertheless, these 108 genes seem like a good starting point for regulation of *SZF1*. Since this is a trans-eQTL, the transcription factors in this list of 108 genes were determined. There were a total of 11 transcription factors among these 108 candidate regulatory genes (Table 4.1). Using the RNA-seq data from all time points, expression patterns for *SZF1* and the 11 transcription factors can be compared (Fig. 4.11). Based on these data, AT5G26170 (WRKY50; Table 4.1) is the transcription factor with an expression pattern that best parallels *SZF1* expression and analysis of the *SZF1*-500 bp promoter reveals two known WRKY binding sites (Fig. 4.11). Again, near-isogenic lines and complementation lines will need to be made for WRKY50 to establish its importance in regulation of *SZF1* expression. These lines would also be tested for differences in freezing tolerance. This method of network construction is a rather tedious process, especially for a single gene that may or may not largely contribute to differences in freezing tolerance. Consequently, future efforts for low-temperature network construction should include transcriptomic analysis of the RILs used for mapping.

It is fiscally daunting to do microarray or RNA-seq eQTL analysis on 544 RILs. However, the minimum number of randomly selected RILs necessary to get similar eQTL results to 544 RILs can be determined through analysis of eQTL mapped for the

10 FTD genes in 544 RILs (described in Chapter Three). The same RIL RNA samples used in eQTL analysis can be submitted for transcriptomic analysis by either RNA-seq or Affymetrix arrays. Genome-wide eQTL would thus be mapped for this subset of RILs and trans-regulatory hotspots (45-48) can be identified at a time-point where there are differences in freezing tolerance (2 weeks of cold-acclimation; Fig 3.2). Accordingly, efforts to find the genes responsible for these trans-eQTL will be focused on major regulatory hubs. It is clear that there is much future effort needed to determine the genes responsible for differences in freezing tolerance between the SW and IT ecotypes, especially for CBF-independent pathways. Nevertheless, data from this study provides a necessary backbone for future studies.

Table 4.1. Candidate cold-regulated transcription factors for SZF1 trans-QTL on chromosome 5 (between RIL markers 20.0 and 36.7). The SeqViewer tool on TAIR (www.arabidopsis.org) was used to determine within SZF1 trans-QTL on chromosome 5. From this list SW and IT cold-regulated transcription factors were identified using merge function in R. The comparison of SW and IT expression at 2 weeks of cold-acclimation for each gene, was measured by RNA-seq and is represented by the logarithm (base=2) of the fold change (LOG2FC). The FDR corrected P-value is given for each comparison. The results presented are based on averages from three independent experiments (n=3). The shaded transcription factor is located closest to the peak RIL marker for the SZF1-QTL (marker 22.0).

AGI	SHORT DESCRIPTION	2WK.SW.IT	2WK.LOG2FC	FDR P-VALUE
AT5G24110	WRKY DNA-BINDING PROTEIN 30	ITHIGH	3.58	0.00E+00
AT5G24590	TCV-INTERACTING PROTEIN	ITHIGH	0.91	1.40E-05
AT5G25190	INTEGRASE-TYPE DNA-BINDING	ITHIGH	1.13	1.13E-02
AT5G25390	INTEGRASE-TYPE DNA-BINDING	SWHIGH	-1.20	5.15E-05
AT5G25810	INTEGRASE-TYPE DNA-BINDING	SWHIGH	-1.92	2.21E-03
AT5G25890	INDOLE-3-ACETIC ACID INDUCIBLE 28	SWHIGH	-0.59	3.32E-02
AT5G26170	WRKY DNA-BINDING PROTEIN 50	ITHIGH	2.83	7.48E-07
AT5G27610	DIRP; MYB-LIKE DNA-BINDING DOMAIN	ITHIGH	0.74	2.60E-02
AT5G28300	SUPERFAMILY PROTEIN	SWHIGH	-0.87	4.43E-07
AT5G28640	SSXT FAMILY PROTEIN	ITHIGH	1.33	1.03E-07
AT5G28770	BZIP TRANSCRIPTION FACTOR FAMILY	ITHIGH	0.77	3.20E-04

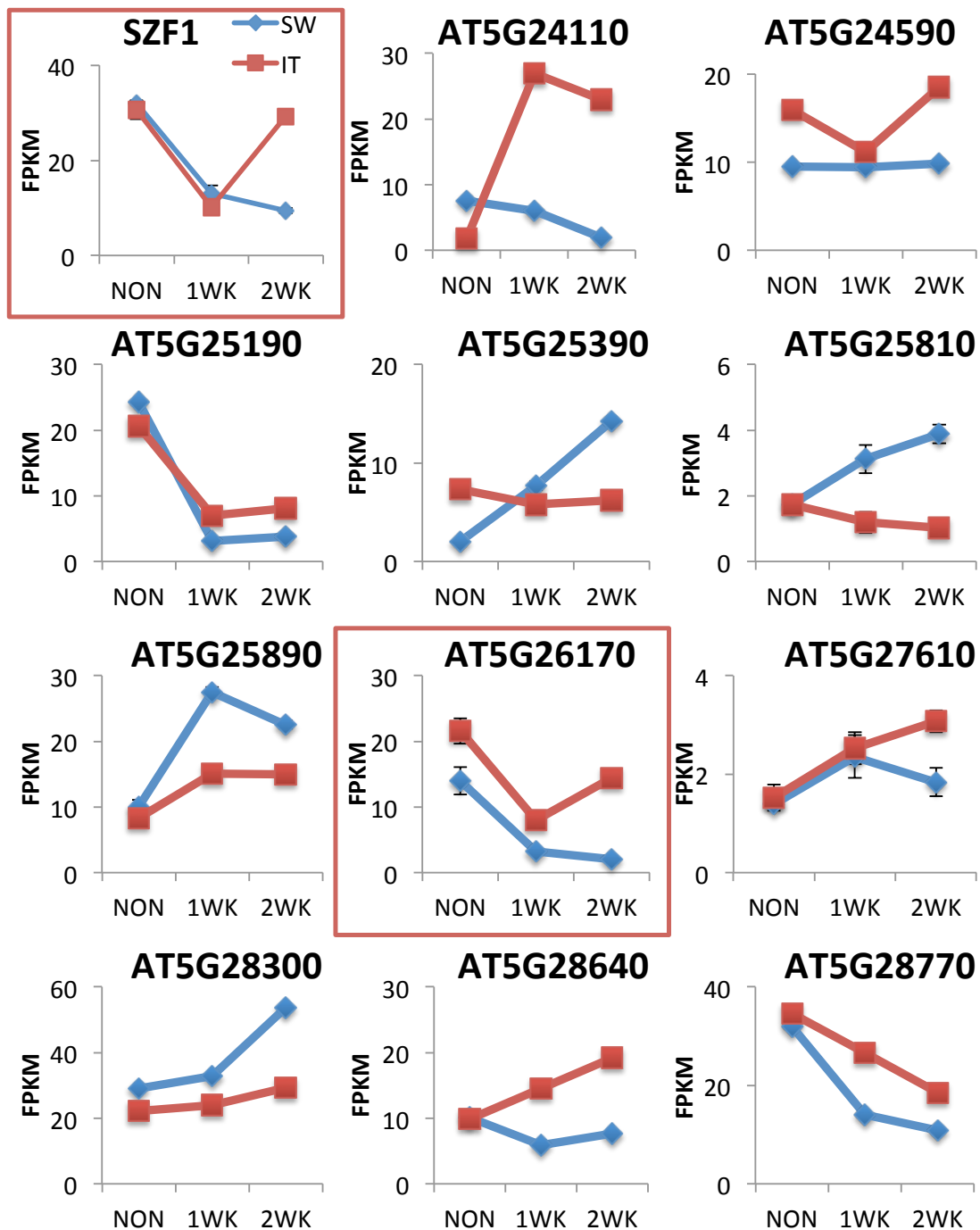


Figure 4.11. *SZF1*-regulation candidate gene expression. IT samples are shown in red, and SW in blue. Gene expression was measured by RNA-seq (FPKM). The results presented are average values from three independent experiments (n=3). Error bars indicate \pm SEM. Plants were grown at 22°C under a 12 h photoperiod and sampled directly or transferred to 4°C for 1 or 2 weeks under a 12 h photoperiod then sampled.

MATERIALS AND METHODS

Regulation of CBF genes by the REVEILLES, CK β 3, and CAMTAS and circadian regulation of CBF-independent gene, COR27: Plants were grown and experiments were conducted as described in Chapter Two of this study. Please see Table A4.1 and A4.2 for primers.

Temperature-entrained circadian regulation and gated cold-induction of CBF genes: Plants were grown and experiments were conducted as described in Chapter Two of this study, except that instead of entrainment with a 12 h photoperiod, plants were entrained with 12 h thermocycles (22°C day/ 12°C night) under constant light, then sampled under constant light and constant temperature conditions (22°C). Please see Table A4.1 for primers.

Disruption of the circadian clock by low-temperature: Experiments were conducted as described in Chapter Two of this study. Wild-type Ws-2 plants were grown at 22°C with a 12 h photoperiod to the four leaf stage and then transferred to one of three conditions at ZT4: 'WARM' plants were transferred to constant light and constant temperature (22°C) conditions. 'TEMP CYLING' plants were transferred to constant light with a 12 h cold temperature period (9°C day/ 4°C night). 'COLD' plants were transferred to constant light and constant cold (4°C) conditions. Please see Table A4.1 for primers.

Low-temperature network construction using SW and IT RILs: Experiments were conducted as described in Chapter Three of this study.

APPENDIX

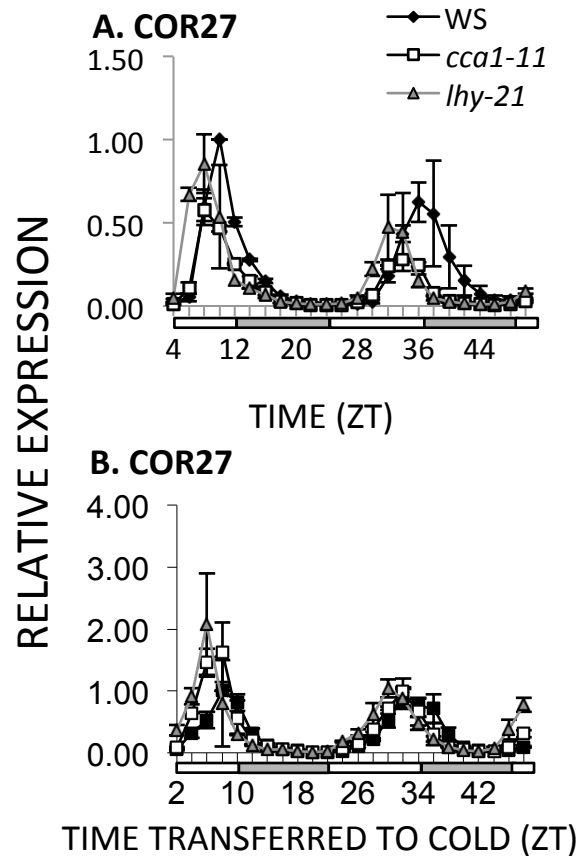


Figure A4.1. Effects of the *cca1-11* and *lhy-21* single mutants on circadian and gated cold-induction of *COR27*. Expression for *COR27* was relative to one wild-type sample WS-2 (WS) set to a value of 1 in each biological replicate. Values for WS-2 are averages from three independent experiments (n=3). Values for *cca1-11* and *lhy-21* are averages from two independent experiments (n=2). Error bars indicate \pm SEM. (A) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were harvested every 2 h and the transcript levels for *COR27* were determined by qRT-PCR. Gene expression was normalized to IPP2 for each sample. (B) Plants were grown with a 12 h photoperiod at 22°C to the four leaf stage and transferred to constant light at ZT0 (subjective day and night are indicated by white and gray bars, respectively). Plants were transferred to cold temperature (4°C) for 2 h, every 2 h, for 48 h at the start of constant light conditions and the transcript levels for *COR27* were determined by qRT-PCR. Horizontal axis represents time (ZT) when plants were dropped to cold temperature. Gene expression was normalized to UBQ10 for each sample. Please see Table A4.1 for primer pair sequences.

Table A4.1. Primer pairs used for qRT-PCR. Primers used in this study were designed with Primer Express 3.0.

qRT-PCR Primers

Name	Forward	Reverse	ATG
CAB2	CCGGAAGGCTGTGAACCT	CACACGGCCGCTTCCA	AT1G29920
CBF1	GGAGACAATGTTTGGGATGC	CGACTATCGAATATTAGTAACTCC	AT4G25490
CBF2	CGACGGATGCTCATGGTCTT	TCTTCATCCATATAAAACGCATCTTG	AT4G25470
CBF2	CGACGGATGCTCATGGTCTT	TCTTCATCCATATAAAACGCATCTTG	AT4G25470
CBF3	TTCCGTCCGTACAGTGGAAT	AACTCCATAACGATACGTCGTC	AT4G25480
CCA1	CCGCAACTTTCGCCTCAT	GCCAGATTTCGGAGGTGAGTTC	AT2G46830
CCR2	TAGGGCGACGTTATTGATTCC	CCCTCAATCGCATCCTTCA	AT1G06820
COR15A	GAAAAAAACAGTGAAACCGCAGAT	CCACATACGCCGCAGCTT	AT2G42540
COR27	GCTCTGGCTCAGCCTCTAGTCT	GGTCGTGGTCACGCGAAT	AT5G42900
COR47	CGGTACCAAGTGTGCGAGAGT	ACAGCTGGTGAATCCTCTGC	AT1G20440
COR78	GAAAGGAGGAGGAGGAATGG	AACCAGCCAGATGATTTTGG	AT5G52310
IPP2	ATTTGCCCATCGTCCTCTGT	GAGAAAGCACGAAAATTCGGTAA	AT3G02780
LHCB1.4	GCCTTCGCTACCAACTTCGTC	AACCGGATACACAACTCGATC	AT2G34430
PRR3	GTGGGAGTAGTGGTGGTTTGAGTA	TTTGTCCAAGAACTCTGAGTTCCA	AT5G60100
PRR5	CGAGAAGCCGCTTTAACCAA	CGGCTCTCGTAACGAACCTT	AT5G24470
PRR7	CCACGAGCGGTATCTCTATGG	ACTTGAAACTCAGGGTTAGAA	AT5G02810
PRR9	GCCAGAGAGAAGCTGCATTGA	CCTGCTCTGGTACCGAACCTT	AT2G46790
TOC1	TCTTCGCAGAATCCCTGTGAT	GCTGCACCTAGCTTCAAGCA	AT5G61380
UBQ10	GGCCTGTATAATCCCTGATGAATAAG	AAAGAGATAACAGGAACGGAAACATAGT	AT4G05320

Table A4.2. Primer pairs used for ChIP qRT-PCR. Primers used in this study were designed with Primer Express 3.0.

ChIP PRIMERS

Name	Forward	Reverse	ATG
ACTIN2	CGTTTCGCTTTCCTTAGTGTTA	AGCGAACGGATCTAGAGACTC	AT3G18780
COR27-A	TTCGCATGGGAGAAGAAGGT	CCACTATCGCCTCTGAATCCA	AT5G42900
COR27-B	AGTTACAACAATCAATAAGCA	GAGAAGGGTGAGTAATTATGTGAATATACAA	AT5G42900
COR27-C	TCTTCCAAAAGAGACATGTAATAGTCAA	TGGGACGATTCACTTTTCTTATTAGA	AT5G42900
COR27-D	GCTCTGGCTCAGCCTCTAGTCT	GGTCGTGGTCACGCGAAT	AT5G42900
TOC1	TTTTATGGCCTGCACTTTTTATTG	GGTGGGACTTGGGATATTTTAGG	AT5G61380
TOC1 3'UTR	GCTACAGCCAAAAAACATCGA	GAGCCGCAAGAGCCAACAT	AT5G61380
UBQ10	TCCAGGACAAGGAGGTATTCCTCCG	CCACCAAAGTTTTACATGAAACGAA	AT4G05320

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