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SOME FACTORS LEADING TO DEATH OF SMOOTH
BROMEGRASS (BROMUS INERMIS LEYYS.)
ON ORGANIC SOIL IN MICHIGAN

Thesis for the Degree of M. S.
MICHIGAN STATE UNIVERSITY

Kristen E. Myhr

1964



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Kristen E. Myhr

AN ABSTRACT

Submitted to
Michigan State University
in partial fulfillment of the requirements
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MASTER OF SCIENCE

Department of Crop Science

1964

Approved Milo B. Tesar.

ABSTRACT

SOME FACTORS LEADING TO DEATH OF SMOOTH BROMEGRASS (BROMUS INERMIS LEYYS.) ON ORGANIC SOIL IN MICHIGAN

by Kristen E. Myhr

Experiments were conducted in the field and under greenhouse conditions to determine why smooth brome grass dies on organic soil, and the effect of fungicide treatments, irrigation, and different levels of fertility and temperature on the time of death.

Irrigation depressed yields of brome grass and orchard grass approximately 16%. Irrigated plots of Lincoln brome grass treated with Dexon fungicide (a p-Dimethylaminobenzenediazo sodium sulfonate) yielded significantly more in the third harvest and exhibited a better stand late in the fall of the first harvest year than untreated plots.

Fusarium spp. and Pythium spp. were isolated from decayed brome grass roots in muck soil in the field. Fusarium poae was significantly associated with killing of roots and reduction of top growth of Lincoln brome grass under controlled conditions. Pythium spp. may have contributed to such death.

Dexon fungicide, which is highly specific against Pythium spp. but relatively ineffective for control of Fusarium spp. was not very effective in controlling root rot under greenhouse conditions. In unfertilized cultures

it depressed the yield significantly, but in fertilized soil the fungicide increased yields, although not significantly.

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TABLE OF CONTENTS

	Page
INTRODUCTION	1
LITERATURE REVIEW.	3
EXPERIMENT I. Field. Effect of fungicide and irrigation on established stands.	7
Materials and Methods.	7
Results.	10
EXPERIMENT II. Greenhouse. Effect of fungicide, fertilizer, and temperature on established grass	14
Materials and Methods.	15
Results.	17
EXPERIMENT III. Greenhouse. Evaluation of fungicide treatments . .	23
Materials and Methods.	23
Results.	24
EXPERIMENT IV. Greenhouse. Pathogenic effect of fungi isolated from infected bromegrass roots	27
Materials and Methods, General	27
Seedling-tests to screen "harmless" fungi.	28
Test of the fungal isolates on established plants.	30
Materials and Methods.	30
Results.	34
DISCUSSION	36
SUMMARY.	40
LITERATURE CITED	41

INTRODUCTION

With proper management, smooth brome grass (Bromus inermis Leyss.) may last indefinitely on mineral soil in Michigan. On organic soil, however, the stands start to die out after one year and are almost gone after four years. On muck soil the stand seems to be hurt during the summer time rather than during the winter. Careful examinations have revealed that roots and rhizomes often are heavily infected with micro-organisms.

Two ecotypes of Bromus inermis are recognized according to origin. The northern type is indigenous to Siberia and is now grown for seed mainly in Canada. The southern type was introduced from Hungary and is now grown for seed in the Midwest. The southern type is usually recommended for the Corn Belt area. It is more vigorous, earlier maturing, higher yielding, less competitive to the legume, and has fewer leaf diseases than the northern type. On organic soil, however, varieties of the southern type are first to be killed.

Difficulties in getting stands of brome grass established have been reported from Canada and several states in the Midwest. The seedlings are frequently killed by a root rot complex which usually is referred to as "damping off." The most active organisms in this complex have been Pythium spp., Helminthosporium spp. and Fusarium spp. In Michigan, "damping off" seldom causes poor establishment and has not affected the establishment of brome grass on the Michigan State University Muck Farm.

The experiments reported herein were designed to determine the cause of death of bromegrass after its establishment on muck soil, especially as it might be affected by 1) soil-borne fungi causing diseases, and 2) the interaction of water, temperature and fertilizer on diseases.

LITERATURE REVIEW

The present subject concerning death of established stands of smooth brome grass on organic soil has not been reported in the literature. The problem is identified, however, by Tesar and Shepherd (10) who evaluated forage species on organic soil in Michigan. Hay fields of eight grasses and four legumes in pure stand were compared for a four year period. Grimm alfalfa, Ladino clover, and Empire birdsfoot trefoil were unproductive after the first harvest year and were severely invaded by grasses and weeds in the second harvest year. Big trefoil did not survive the first winter. Failure in survival of legume species indicates that the forage species on muck soil necessarily have to be grasses. Reed canarygrass (4.28 tons) and tall fescue (3.89 tons) gave the greatest yields, but the authors did not recommend these species because they have not given good animal performance. Orchardgrass (3.43 tons) had resistance to invasion by weeds and other grasses. Canadian (2.80 tons) and Lincoln (2.42 tons) brome grass, representing the northern and southern ecotypes, respectively, were lower in yield but with excellent animal performance were preferable to other grass species in the Mid-West if more permanent. These results indicate that Lincoln produced a high yield the first harvest year (4.41 tons), but dropped gradually to (0.06 tons) in the fourth year. Canadian produced a more constant yield - 3.06 tons in the first and 1.74 tons in the four year - but even this yield is not considered satisfactory.

Vanterpool (12) who studied Pythium root rot of grasses in Saskatchewan, Canada, classifies four types or phases of Pythium damage to grasses:

1. Pre-emergence killing of the seedlings in which both roots and shoots are attacked;
2. Damping off or early seedling killing due to the rotting of the roots and the bases of the stems;
3. Lesioning of the larger roots and an invasion of many of the fine laterals in the late seedling stage while the plants are becoming established; and
4. Lesioning of the new roots of perennial grasses during subsequent growth periods.

According to Vanterpool, pathogenic species of Pythium are most readily isolated from diseased roots of young plants up to the third week of June. Virulent species are not easily obtained in culture from the older roots of plants two or more years old. On the basis of several series of greenhouse experiments, Vanterpool concluded that the factors limiting yield were more effective in dry soil than in moist soil and that Pythium root rot was more severe at high than at low temperatures. He suggested fall planting of grasses as a means of obtaining better stands, and the use of phosphate fertilizers applied to naturally infested field soil as a means of increasing yields.

Hawk and Welch (6), Andrews (1), and Buchholtz (2), working in Iowa, Minnesota and South Dakota, respectively, all indicate the difficulties in obtaining a good stand when grasses are planted in the spring. Poor establishment was associated with several indigenous fungi. Hawk and Welch tested the resistance of several brome grass varieties to Pythium graminicola. Their results indicate that the southern ecotype was more tolerant of injury than the northern type. Injury was less pronounced at lower temperatures. Andrews isolated Helminthosporium sativum, Fusarium spp., Pythium graminicola, and Rhizoctonia solani from lesioned roots of grasses. The first three mentioned were pathogenic to seedlings of brome grass.

Nielsen, Dickson and Smith (8) report that varieties and seed treatment are factors in seedling establishment in Wisconsin. Brome grass treated with the commercial fungicides Captan, Ceresan, and Arasan had better stands than not treated. The southern ecotype exhibited slightly greater tolerance to Helminthosporium sativum, Fusarium roseum, and Pythium graminicola than the northern type of brome grass.

Thomason and Dickson (11) isolated root rot organisms from roots and crown tissue of brome grass in four locations in Wisconsin and one location in South Dakota. Of 2,788 isolations 40.7% proved to be Fusarium spp., 32.7% Pythium spp. and 16.7% Helminthosporium spp. In a factorial experiment, seedlings of Lincoln and Canadian brome grass were infected with the isolated organisms and allowed to grow at temperatures

of 16, 20, 24, and 28° C. Canadian brome grass showed better stands in the presence of Pythium graminicola and Helminthosporium sativum than Lincoln. The best stands were obtained at 16 and 20° C.

Kaufmann, Drolsom, and Nielsen (7) have reported differences in reaction to infection when seedlings of strains and experimental synthetics were inoculated with Pythium graminicola and Helminthosporium sativum. The variety Saratoga and four synthetics appeared significantly more tolerant than other strains.

Drolsom and Nielsen (3) have continued their efforts to develop new varieties of smooth brome grass resistant to root rot diseases detrimental to seedling establishment.

EXPERIMENT I

Field

Effect of fungicide and irrigation on established stands

The objective of this experiment was to determine the production and persistency of Lincoln and Canadian brome grass (Bromus inermis Leyss.) and commercial orchardgrass (Dactylis glomerata L.) in the first harvest year when treated with fungicide under irrigated and non-irrigated conditions. The research hypothesis was that the fungicide would by reducing pathogenic soil fungi such as Pythium spp. improve the production and longevity of brome grass on a muck soil.

Materials and Methods

The experiment was conducted on a well-drained Houghton muck soil on the Michigan State University Muck Farm. Lincoln brome grass, Canadian brome grass and commercial orchardgrass were each planted in pure stand on August 18, 1963. The seed was drilled one inch deep, in contact with 500 pounds per acre of 12-12-12 fertilizer in 7-inch rows in drill-width plots. Both brome grass varieties survived the winter without injury and had a 100% stand on May 2, 1964. Orchardgrass had a 50% stand due to considerable winter damage. The stand recovered after several weeks and by harvest time it had reached optimum density.



Figure 1. Orchardgrass, in the center, showed considerable winter damage while bromegrass, left and right, exhibited an excellent stand on May 2, 1964.

During the first harvest year no additional fertilizers were applied because the stand was very vigorous and earlier experiments had shown the soil released sufficient nitrogen for a satisfactory hay crop.

Dexon* (p-Dimethylaminobenzenediazo sodium sulfonate), a non-mercurial fungicide compound of possible protective value to germinating and growing plants from phycomycetons fungi, was applied to the stand on April 22-23, 1964, in the four following ways:

1. Check, no fungicide
2. 50 lbs. active Dexon per acre worked 4 inches deep into the soil between the rows with a tine rake.
3. 50 lbs. active Dexon per acre on the soil surface.
4. 100 lbs. active Dexon per acre - twice the recommended rate - on the soil surface.

One half of each replication was irrigated according to the following schedule:

Second cutting:

June 16	1"
" 18	2"
" 25	1"
" 30	1"
July 2	2"
Total	7"

Third cutting:

July 29	2"
Aug. 6	1"
" 7	1"
" 18	1"
" 19	1"
Sept. 3	1"
Total	7"

*obtained from Chemagro Corporation, Kansas City, Missouri

The experiment was designed as a split-plot, with 4 replications of 3 species, 4 fungicide treatments, and 2 water levels, giving a total of 96 subplots, each 8 x 18 feet in size. A strip 3 feet wide and 15 feet long was mowed from each plot for yield. Moisture samples were obtained from one replication. Yields are reported as tons per acre of hay containing 12% moisture.

Results

Bromegrass lodged heavily in the two first harvests. In the second cutting Lincoln had 50% lodging and Canadian had 30% lodging. Orchardgrass was not lodged. The experiment was free from weeds and volunteer grasses. The yield data are presented in Table 1.

Table 1. Tons of hay per acre for various treatments at each harvest on irrigated (I) and not irrigated (NI) muck soil.

Dexon fungicide	Harvest date	Lincoln		Canadian		Orchard		Average	
		I	NI	I	NI	I	NI	I	NI
1. None, check	June 1	1.27	1.31	1.55	1.38	1.76	2.02	1.53	1.57
	July 23	1.30	1.32	1.00	1.45	1.67	1.75	1.32	1.50
	Sept. 25	0.51	0.85	0.52	0.71	0.85	1.04	0.62	0.86
	Total	<u>3.08</u>	<u>3.48</u>	<u>3.07</u>	<u>3.54</u>	<u>4.28</u>	<u>4.81</u>	<u>3.47</u>	<u>3.93</u>
2. 50 lbs/acre mixed into 4" depth	June 1	1.42	1.27	1.51	1.54	1.76	1.58	1.56	1.46
	July 23	1.29	1.29	1.17	1.38	1.49	2.01	1.32	1.56
	Sept. 25	0.52	0.83	0.62	0.64	1.02	1.01	0.72	0.83
	Total	<u>3.23</u>	<u>3.39</u>	<u>3.30</u>	<u>3.56</u>	<u>4.27</u>	<u>4.60</u>	<u>3.60</u>	<u>3.85</u>
3. 50 lbs/acre broadcast on surface	June 1	1.40	1.53	1.30	1.34	1.87	1.85	1.52	1.57
	July 23	1.34	1.38	1.11	1.43	1.51	1.89	1.32	1.56
	Sept. 25	0.62	0.83	0.40	0.63	1.14	0.97	0.72	0.81
	Total	<u>3.36</u>	<u>3.74</u>	<u>2.81</u>	<u>3.40</u>	<u>4.52</u>	<u>4.71</u>	<u>3.56</u>	<u>3.94</u>
4. 100 lbs/acre broadcast on surface	June 1	1.36	1.24	1.37	1.65	1.70	1.66	1.47	1.51
	July 23	1.03	1.23	1.21	1.33	1.40	1.88	1.21	1.48
	Sept. 25	0.85	0.85	0.64	0.73	0.83	1.09	0.77	0.89
	Total	<u>3.24</u>	<u>3.32</u>	<u>3.22</u>	<u>3.71</u>	<u>3.93</u>	<u>4.63</u>	<u>3.45</u>	<u>3.88</u>

5% LSD for 1st harvest = 0.33
2nd harvest = 0.24
3rd harvest = 0.22
total yield = 0.45

Irrigation has depressed total yield for all combinations of fungicide treatments and species. The effects of irrigation are shown in Table 2. No water was added prior to the first harvest and the yields were similar at the first cutting. In the next two harvests irrigation depressed yields approximately 16%.

Table 2. The average effect of irrigation in tons of hay per acre of Lincoln brome grass, Canadian brome grass, and orchardgrass on a muck soil.

Cutting	Cutting date	Irrigated	Not irrigated
First	June 1	1.52	1.53
Second	July 23	1.29	1.53
Third	Sept. 25	<u>0.71</u>	<u>0.85</u>
	Total	3.52	3.91

Table 3 shows total yield on irrigated plots for the last two harvests in percent of those not irrigated. On irrigated soil, the heaviest amount of fungicide resulted in greater yields of both brome grasses.

Table 3. Yield on irrigated plots in percent of those not irrigated for the two last harvests.

Dexon fungicide	Lincoln brome grass	Canadian brome grass	Orchard-grass
1. Check, no fungicide	83	70	90
2. 50 lbs./acre mixed in	81	88	83
3. 50 lbs./acre on surface	89	73	92
4. 100 lbs./acre on surface	90	90	75

The beneficial effects of the fungicide were not significant until the third harvest. The interaction fungicide x species x irrigation was significant in the second and third harvest. Under irrigated conditions; of primary significance was that the yield of Lincoln brome grass in the third harvest was significantly higher when the soil was treated with the 100-pound rate of fungicide, than when not treated or treated with the 50-pound rate, table 1. The significance of the 3rd-order interaction in the second harvest was mainly due to Orchardgrass. On irrigated land the heaviest amount of Dexon significantly depressed its yield.

The yields of Lincoln brome grass showed positive, but non-significant effects of the fungicide in the first harvest. Because no irrigation was applied before the first cutting, the data for irrigated and not irrigated plots were pooled. Treatment 3 - recommended fungicide rate broadcast - produced 0.18 tons more per acre than the untreated check, but in the first harvest the other treatments were similar to untreated plots. Twice the commended rate of the fungicide resulted in reduced yields in the second harvest under irrigated conditions.

Three persons, estimating independently, determined that the stand of irrigated Lincoln brome grass was significantly better on fungicide treated plots than on untreated plots at the end of the first harvest year, table 4.

Table 4. Percent stand on untreated and fungicide treated plots of 4 replications Lincoln brome grass on irrigated land on November 4, 1964, at the end of the first harvest year.

Treatment	Estimator 1	Estimator 2	Estimator 3	Average
Untreated	68	63	63	65
Dexon fungicide	76	69	73	73

5% LSD = 6

The effect of fungicide treatments on Canadian brome grass was somewhat different than on Lincoln brome grass. Treatment 3 - recommended fungicide rate broadcast - had lower production in both the first and third cutting compared with the untreated check, but the differences were not significant.

EXPERIMENT II

Greenhouse

Effect of fungicide, fertilizer, and temperature on established grass

A comparison was made of the performance of several grass species grown in untreated and fungicide treated muck soil under greenhouse conditions. It was assumed that it would be possible to get as many cuttings in the greenhouse in nine months as in two harvest years in the field. Assuming that brome grass is weakened to a certain degree after defoliation, and since many plants died during the second growing season in Tesar and Shepherd's experiment (10), there was reason to expect

thinning and killing during the latter part of the experimental period. The hypothesis was that brome grass grown in fungicide treated soil would yield more and live longer than in untreated soil.

Materials and Methods

The following variables were included in the experiment:

- 2 Fungicides - untreated and treated soil
- 2 Fertilizers - none and heavy
- 4 Species - Lincoln brome grass, Canadian brome grass, orchard-grass, and reed canarygrass
- 2 Night temperatures - 70°F and 80°F
- 4 Replications

The soil used in the experiment was obtained by compositing soil samples collected around the periphery of the field experiment. Only the upper 4 inch layer was used. After screening, 1850 grams of soil was mixed with fungicide and fertilizer, according to the design, and put into a 6-inch gallon can.

Fifty pounds of active Dexon fungicide per acre, and 2000 lbs. per acre of a 5-20-20 fertilizer was added to one half of the pots.

Fifty seeds of each of Lincoln brome grass, Canadian brome grass, commercial orchardgrass, and reed canarygrass (Phalaris arundinacea L.) were planted in separate pots one inch below the soil surface on December 23, 1963. After reseeding reed canarygrass on January 9, because of poor germination, and a few transplantations of brome grass on January 17, the

stand reached adequate density in all cultures. The first shoots from rhizomes on bromegrass appeared on February 11.

One half of the experiment was placed in a section of the greenhouse where the night temperature was approximately 80°F., the other half at a night temperature of approximately 70°F. Day temperature varied and during the first three harvests day temperature averaged about 10 degrees higher in the high temperature house. As the summer season approached the day temperature in the two houses was similar.

Water was added weekly to bring each container up to a weight of 2400 grams to adjust for uneven consumption. When necessary water was supplied daily.

Yields were obtained by cutting to a height of 3 cm. on the following days in the low temperature house:

February 29	July 1
March 30	August 12
May 18	September 30

The experiment was terminated after the first four harvests in the high temperature house. Yields are expressed in grams dry matter per culture. After each cutting all cultures were fertilized with a water solution of nitrogen at a rate of 100 lbs. N per acre.

The pots were grouped together in replications at all times, but the position of a particular pot, as well as the replication in which it occurred was systematically changed to avoid border effects in the experiment.

Statistical analysis of the data was carried out separately for each temperature section according to a factorial model. For the calculations of F-values and for interpretation of the data, all treatments were considered as fixed constants.

Results

The stand was healthy and vigorous during the three first cuttings in all pots, with the exception of the fungicide treated reed canarygrass. During the month of June, the plants began to suffer from unfavorable conditions. The bromegrass lost leaf turgor, luster and green color. Orchardgrass and reed canarygrass exhibited the same symptoms, although to a lesser extent. Close examinations did not reveal wilting symptoms were associated with the treatments. Because the experiment was inspected daily, deficient water supply was not a possible cause. A more reasonable explanation is that the extreme high temperatures which occurred in the greenhouse during hot sunny summer days were unfavorable for these cool season grasses. After the fourth and fifth cuttings, the plants showed the same wilting symptoms when they reached a height of 4-5 inches, and production was reduced.

The data for the low and high temperature section are presented separately in table 5 and table 6, respectively. Because the data do not correspond exactly, and there were two more harvestings on the low temperature material, the two parts were not pooled.

Table 5. Grams dry matter per pot for various treatments in a greenhouse maintained at a 70°F. night temperature.

Treatment	Harvest date	Lincoln brome-grass	Canadian brome-grass	Orchard grass	Reed canarygrass
NOT FERTILIZED					
No fungicide	Feb 29	6.24	7.55	5.66	5.83
	Mar 30	5.08	5.65	6.80	8.15
	May 18	11.15	10.86	13.28	16.75
	July 1	6.71	5.60	6.76	9.28
	Aug 12	2.54	2.26	3.35	3.48
	Sept 30	1.90	0.93	2.30	2.26
	Total	33.62	32.85	38.15	45.75
Fungicide	Feb 29	5.53	5.96	5.41	0.81
	Mar 30	4.94	4.74	6.51	3.98
	May 18	10.62	10.60	12.47	19.09
	July 1	5.18	5.62	5.51	15.14
	Aug 12	2.27	2.44	2.51	5.51
	Sept 30	1.93	1.29	2.24	3.05
	Total	30.47	30.65	34.65	47.58
FERTILIZED					
No fungicide	Feb 29	7.81	9.94	6.42	6.13
	Mar 30	8.40	8.61	9.41	9.85
	May 18	15.72	14.55	17.01	18.83
	July 1	12.91	13.03	12.64	15.83
	Aug 12	6.71	5.99	6.04	6.12
	Sept 30	2.93	2.28	4.30	3.43
	Total	54.48	54.40	55.82	60.19
Fungicide	Feb 29	6.73	8.78	7.06	1.89
	Mar 30	8.18	8.86	9.36	5.68
	May 18	17.63	16.82	18.52	20.93
	July 1	14.36	14.56	13.28	20.22
	Aug 12	5.97	5.74	3.48	9.08
	Sept.30	2.33	2.07	3.89	4.82
	Total	55.20	56.83	55.59	62.62

5% LSD for 1st harvest = 0.92 4th harvest = 1.38
2nd harvest = 0.74 5th harvest = 0.96
3rd harvest = 1.34 6th harvest = 0.68

total yields = 2.84

Table 6. Grams dry matter per pot for various treatments in a greenhouse maintained at 80°F. night temperature.

Treatment	Harvest date	Lincoln bromegrass	Canadian bromegrass	Orchard grass	Reed canary
NOT FERTILIZED					
No fungicide	Feb 29	6.22	7.62	6.35	7.09
	Mar 30	7.13	7.00	6.49	8.13
	May 18	8.62	6.65	8.63	12.62
	July 1	5.23	4.19	5.60	7.33
	Total	<u>27.20</u>	<u>25.46</u>	<u>27.07</u>	<u>35.17</u>
Fungicide	Feb 29	5.77	6.29	6.35	1.84
	Mar 30	6.84	6.95	7.82	7.69
	May 18	9.58	6.89	10.32	16.36
	July 1	4.73	3.24	5.02	9.56
	Total	<u>26.92</u>	<u>23.37</u>	<u>29.51</u>	<u>35.45</u>
FERTILIZED					
No fungicide	Feb 29	7.03	9.23	6.50	6.13
	Mar 30	9.51	9.95	8.38	8.32
	May 18	13.39	10.25	12.04	16.31
	July 1	12.41	10.12	10.21	13.69
	Total	<u>42.34</u>	<u>39.55</u>	<u>37.13</u>	<u>44.45</u>
Fungicide	Feb 29	6.02	7.58	6.35	3.52
	Mar 30	9.14	10.78	10.46	10.27
	May 18	14.19	9.54	13.31	19.23
	July 1	12.57	8.27	9.64	13.87
	Total	<u>41.92</u>	<u>36.17</u>	<u>39.76</u>	<u>46.89</u>

5% LSD for 1st harvest = 0.73
 2nd harvest = 1.14
 3rd harvest = 2.04

4th harvest = 1.50
 total yield = 3.70

The fungicide treatment caused a significant or near significant reduction in yield in the Lincoln brome grass in the first cutting and sometimes in the second cutting at both fertility and temperature levels. In the third and fourth cuttings under low night temperature, the fungicide significantly improved the yield of Lincoln in fertilized soil. For the two last cuttings on August 12 and September 30, the plants were not vigorous; the yields were low and slightly depressed in fungicide treated fertilized soil.

Canadian brome grass reacted similarly to Lincoln in the response to fungicide, except that it has been harmed more by the fungicide under high temperature.

Total yield of orchard grass was significantly depressed by the fungicide in unfertilized cultures at low temperatures. At high temperatures the orchard grass has given greater yields in fungicide treated cultures.

Reed canary grass reacted differently than the other two species. It gave a very small yield in the first and second cutting following fungicide treatment in the low temperature house. Also in the high temperature a similar depression is recorded at the first cutting. In subsequent harvests the yield was significantly increased, but differences in total yield were not significant.

The effect of fertilizer was highly significant for all cuttings. The fungicide x fertilizer interaction for total yield at low temperature is shown in table 7.

Table 7. Yield in grams per pot for combinations of fungicide and fertilizer for four grasses grown at night temperature of 70°F.

Treatment	Fungicide	No fungicide
Fertilizer	57.54	56.16
No fertilizer	35.80	37.56
Average	46.67	46.86

5% LSD = 1.42

The interaction indicates that the fungicide significantly depressed the yield in non-fertilized cultures. In those that were fertilized the fungicide tended to induce increased production.

In table 8 yields of different species in non-fertilized pots are expressed in percent of the production of those fertilized, for the two temperature levels separately. Data for the fifth and sixth cutting are not included in the table because of very small yields.

Table 8. Yield of the first four harvests of non-fertilized grass as percent of fertilized grass.

Species	Night temperature		
	Low	High	Average
Lincoln bromegrass	58	64	61
Canadian bromegrass	57	64	61
Orchardgrass	65	73	69
Reed canarygrass	75	77	76

Percentagewise, orchardgrass and reed canarygrass performed better on unfertilized soil than the bromegrass varieties.

After the last cutting the roots were washed, examined, dried and weighed. Rotted roots were found in both untreated and fungicide treated cultures. Visual estimation of the roots did not reveal any significant differences. The heaviest root-weights in Lincoln bromegrass were found in untreated soil, but since the data were variable as indicated by a large error-term, there was no significant difference in the effect of fungicide in root weights.

EXPERIMENT III

Greenhouse

Evaluation of fungicide treatments

This experiment was set up to determine the effectiveness of Dexon, the soil fungicide used in the two prior experiments, and propylene oxide in comparison to heat sterilization as methods of killing soil pathogens theorized as causing death of bromegrass on muck soil.

Materials and Methods

The following variables were used in greenhouse cultures at a night temperature of 70°F., 6 fungicide treatments:

- a. Check, no fungicide
- b. Dexon, 50 lbs. active ingredient per acre on top of soil
- c. Dexon, same rate as b, mixed into the upper 2" of soil
- d. Dexon, same rate as b, mixed into the entire soil volume
- e. Autoclaving for 1 hour, soil in pots; and
- f. Propylene oxide gaseous sterilization

3 species - Lincoln bromegrass, Canadian bromegrass and
orchardgrass

4 replications

Propylene oxide sterilization is described by Hansen and Snyder (5). Compared with heat sterilization this method causes little change in the physical-chemical nature of the soil.

The experiment was conducted in the same manner as the other greenhouse experiment. It was designed so that the fertilized Lincoln brome-grass, Canadian brome-grass, and orchardgrass of experiment II in the low temperature house were included. The two experiments had two treatments in common: 1) fertilized check, and 2) fertilized cultures with Dexon mixed into the entire soil volume.

Results

Good stands were obtained and maintained until after the third cutting in early summer at which time the grasses began to wilt and die slowly in the same way as described for experiment II. The data are presented in table 9.

Autoclaving was the only treatment which increased the yield of brome-grasses in the first and occasionally in the second harvest. Propylene oxide decreased the yield of the brome-grasses during the first two cuttings. Dexon fungicide applied on the surface or in the upper two inches had a similar harmful effect on all three species in the first two harvests.

In the last two cuttings the two brome-grass varieties but not orchard-grass yielded more when treated with propylene oxide than when not treated. Yields of brome-grass in Dexon treated soil and in autoclaved soil were similar to yields of non-treated brome-grass.

Table 9. Yield in grams per pot of Lincoln brome grass, Canadian brome grass, and Orchard grass when grown in untreated, fungicide treated, and sterilized soil.

Treatment	Harvest date	Lincoln brome grass	Canadian brome grass	Orchard grass	Average
Check, untreated	Feb 29	7.81	9.43	6.42	8.05
	Mar 30	8.40	8.61	9.41	8.80
	May 18	15.72	14.55	17.01	15.76
	July 1	12.91	13.63	12.64	12.86
	Aug 12	6.71	5.99	6.04	6.27
	Sept 30	2.93	2.28	4.30	3.17
	Total	<u>54.48</u>	<u>54.40</u>	<u>55.82</u>	<u>54.91</u>
Dexon on top	Feb 29	4.65	4.34	4.13	4.37
	Mar 30	5.77	5.55	7.60	6.31
	May 18	13.34	15.36	17.74	15.48
	July 1	10.87	14.26	12.42	12.52
	Aug 12	6.13	8.35	5.95	6.82
	Sept 30	3.44	3.59	3.96	3.67
	Total	<u>44.20</u>	<u>51.55</u>	<u>51.80</u>	<u>49.17</u>
Dexon in top 2 inches	Feb 29	5.23	7.13	5.32	5.89
	Mar 30	5.53	7.71	7.86	7.03
	May 18	13.24	15.83	17.21	15.42
	July 1	9.48	13.53	10.70	11.24
	Aug 12	6.35	6.66	5.01	6.00
	Sept 30	3.48	2.74	3.32	3.17
	Total	<u>43.31</u>	<u>53.60</u>	<u>49.42</u>	<u>48.75</u>
Dexon mixed in	Feb 29	6.73	8.78	7.06	7.52
	Mar 30	8.18	8.86	9.36	8.80
	May 18	17.63	16.82	18.52	17.65
	July 1	14.36	14.56	13.28	14.06
	Aug 18	5.97	5.74	3.48	5.06
	Sept 30	2.33	2.07	3.89	2.76
	Total	<u>55.20</u>	<u>56.83</u>	<u>55.59</u>	<u>55.85</u>

Table continues on next page

Table 9. continued

Treatment	Harvest	Lincoln	Canadian	Orchard	Average
Autoclaved soil	Feb 29	10.52	13.11	9.33	10.99
	Mar 30	9.44	8.84	9.28	18.19
	May 18	16.39	14.94	18.20	16.51
	July 1	12.15	12.12	12.68	12.32
	Aug 18	6.03	6.21	6.76	6.33
	Sept 30	3.85	3.52	4.58	3.99
	Total	<u>58.38</u>	<u>58.74</u>	<u>60.83</u>	<u>68.33</u>
Propylene oxide	Feb 29	5.08	6.91	5.47	5.82
	Mar 30	6.06	6.85	8.75	7.22
	May 18	14.97	17.80	18.59	17.12
	July 1	13.25	15.52	12.35	13.71
	Aug 18	7.06	8.55	6.27	7.29
	Sept 30	4.17	4.20	4.38	4.25
	Total	<u>50.59</u>	<u>59.83</u>	<u>55.81</u>	<u>55.41</u>

5% LSD for 1st harvest = 1.74
 2nd harvest = 1.56
 3rd harvest = 1.94

4th harvest = 2.08
 5th harvest = 1.70
 6th harvest = 1.42
 total yield = 4.30

Lincoln brome grass appeared to be more sensitive to both Dexon fungicide and propylene oxide than Canadian brome grass.

After the last harvest the roots were washed, examined, dried and weighed, but because of a large experimental error no differences between treatments were obtained. The roots were partially decayed in all pots.

EXPERIMENT IV

Greenhouse

Pathogenic effect of fungi isolated from infected brome grass roots

The objective of this study was to determine, under controlled conditions, how brome grass is affected by the fungi isolated from decaying roots of brome grass on muck soil.

Materials and Methods, General

Roots of Lincoln brome grass were brought in from the Muck Farm on June 18, July 9, and August 18. The roots were carefully washed in running water for 3 hours. Partially lesioned sections of young white roots were then selected and surface sterilized in a 1% chlorazene solution for 2 minutes. A section of root 5 mm long was obtained from the border zone between healthy and decayed tissue, and placed on sterile water agar in a petri dish for three days at room temperature.

Approximately 40 such transplants were made on each date. In most of the dishes fungus mycelia grew out radially from the root sections and spread over the agar. Some small bits of mycelial growth from the periphery of various colonies were transferred to potatodextrose agar in test tubes for further experimentation.

The major interest in this investigation was to inoculate established bromegrass with isolates of the more pathogenic fungi. In order to screen out the less harmful organisms, however, seedling tests were conducted first. The assumption made was that pathogenic fungi would harm a young seedling rapidly and more severely than an established plant.

Seedling tests to screen "harmless" fungi

One hundred and twenty cc of vermiculite and 50 cc of a standard potato juice (200 gm potato extract in a liter of water) was put into 1 pint, widemouth Mason jars, covered with a half petri dish and autoclaved. Mycelial cultures were then transferred aseptically from the test tubes into the vermiculite and allowed to establish for a few days before bromegrass seeds, surface-sterilized with chlorazene were planted at a rate of 20 per jar. Seedlings were counted after 14 days. To make comparative evaluations of damage caused by some known pathogenic species of Pythium, pure cultures of a few identified isolations were obtained from the Department of Botany and Plant Pathology.

The number of seedlings developed in the presence of the most harmful isolates is shown in table 10. The data for the different inoculums are given as percentage of the check (non-inoculated).

Table 10. Bromegrass seedlings developed in sterile and inoculated vermiculite medium 14 days after planting. Stands given as percent of check.

Inoculum	Test	Test July 9		Test
	June 18 Lincoln bromegrass	Lincoln bromegrass	Canadian bromegrass	Aug 18 Lincoln bromegrass
Check, sterile medium	100	100	100	100
<u>Pythium ultimum</u>	55	72	85	
<u>Pythium irregulare</u>	71	47	80	
<u>Culture 18</u>	31	6	20	
<u>Fusarium</u> , culture L3				28
<u>Fusarium</u> , culture L8				23
<u>Fusarium</u> , culture L23				23
<u>Fusarium</u> , culture L32				50
<u>Fusarium</u> , culture L33				32

In addition to the 8 inoculums recorded in table 10, 43 others were tested. Number of developed seedlings in those ranged from 60 to 100 percent of check.

Fusarium spp. and to a lesser extent, Pythium spp. reduced the number of seedlings significantly. Canadian bromegrass was more resistant than Lincoln bromegrass to the fungi tested July 9. Two (L8 and L23) of the Fusarium isolations made on August 18 were identified as Fusarium poae (Pk.) Wr., Gilman (4). Three others recorded in table 10

(L3, L32, and L33) were probably the same species with similar red-brown mycelia. Culture 18 which was isolated on June 18 and tested twice was difficult to purify and identify because it was an inextricably mixed combination of a Pythium species and a Fusarium species with similar growth habits. Its effect on brome grass in these tests is illustrated in Fig. 2.

Seeds, seedlings, and the length of the longest root were examined when the experiments were terminated. The lower numbers of seedlings in the inoculated jars were due to pre-emergence killing of the seedlings. Both root ~~primor~~dium and coleoptile had been attacked and severed by the fungus. The average root length in the checks in table 10, was 20mm. Inoculated seedlings had much shorter roots ranging from 2 to 5 mm.

Test of the fungal isolates on established plants

Materials and Methods

On June 17, surface sterilized seeds of Lincoln brome grass were planted in 4-inch circular peat pots which previously had been filled with muck soil, embedded in flats and autoclaved. When these plants had passed the seedling stage and had been cut once, the cultures were taken out of the flats, cleaned carefully outside, transferred to 6-inch gallon cans, and embedded in vermiculite. This vermiculite had previously been moistened with potato juice, autoclaved, and inoculated with some of the fungus isolates which had previously proved most detrimental to brome grass-seedlings, table 10.

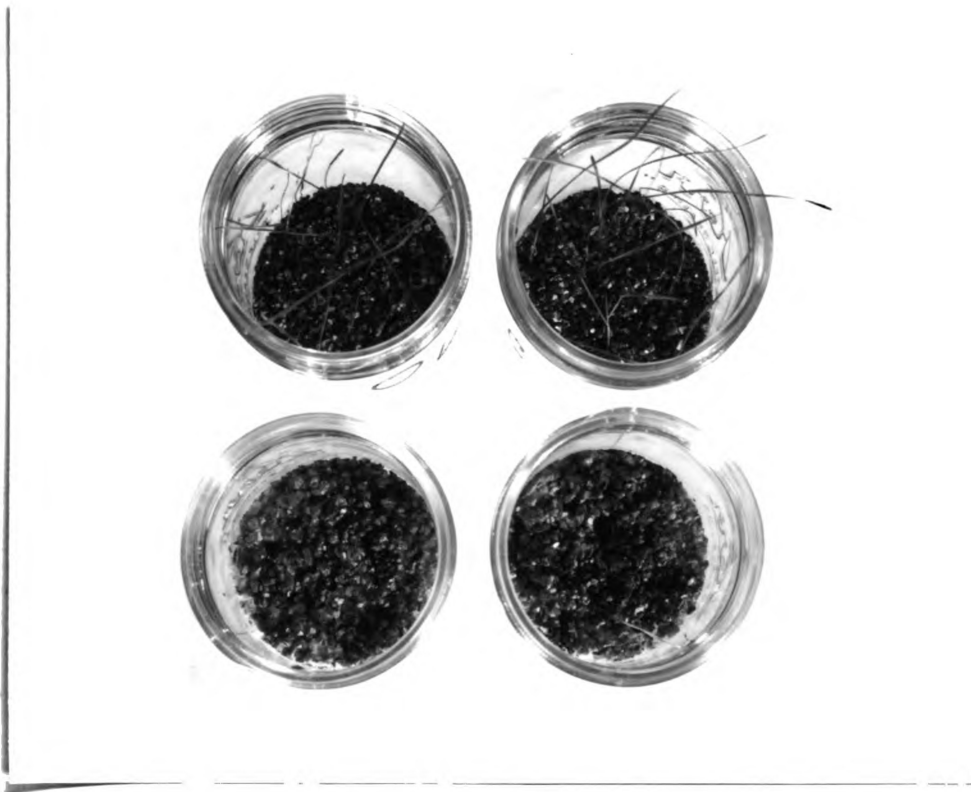


Figure 2. Relative survival of Lincoln (left) and Canadian bromegrass (right) inoculated with fungus culture 18 isolated June 18 from decayed roots in the field (lower row) compared with check (upper row).

On August 2, eight bromegrass cultures were inoculated with each of the following treatments separately:

1. Check. No inoculum
2. Pythium irregulare
3. Culture 18
4. Culture T50

On September 29 four cultures of each of the following treatments were added:

5. Check. No inoculum
6. Fusarium, culture L3
7. Fusarium, culture L8
8. Fusarium, culture L33

Culture T50 was included in this experiment as a form of control treatment because it had produced a heavy mycelium without, however, visibly harming the small bromegrass plants in the seedling test.

When the experiment was terminated on October 29, the tops were severed from the roots, dried and weighed. Root weights had to be determined indirectly because the roots had infiltrated the vermiculite and could not be separated by washing, figure 3. The roots were cut close to the wall outside the peat pot, and then the roots and vermiculite together were dried and weighed. The dry weight of vermiculite added initially was subtracted to get dry root weight.



Figure 3. Method of growing bromegrass to determine susceptibility of established plants to fungi.
From left to right:
1. The gallon pots in which the peat pots were embedded
2. Peat pots with vermiculite
3. Roots and vermiculite after separation from peat pot
4. Peat pots after roots and vermiculite were removed
5. Peat pots after the tops were removed

Results

Relative dry weights of tops and roots on October 29 are reported in table 11.

Table 11. Relative top and root weights for Lincoln brome grass on October 29 after being established June 17 in sterile muck soil and inoculated with fungus on August 2 and September 29.

Inoculum	Top weight	Root weight
Check	100	100
1) Group 1: <u>Pythium irregulare</u>	85	66
Culture 18	83	68
Culture T50	86	69
5% LSD	N.S.	N.S.
2) Group 2: <u>Fusarium</u> , culture L3	72	57
<u>Fusarium</u> , culture L8	74	66
<u>Fusarium</u> , culture L33	65	64
5% LSD	19	25

- 1) 88 days experimental period (August 2 - October 29)
- 2) 30 days experimental period (September 29 - October 29)

The Fusarium isolations in Group 2 significantly reduced both top-growth and root weights, compared with the check. The fungi in Group 1, did not reduce top or root growth significantly because of extreme variability.

The data strongly suggest, however, that an actual reduction in top growth and especially root growth, resulted when Lincoln brome grass was inoculated with these fungi.

DISCUSSION

The yields of both brome-grass and orchardgrass in the field were approximately 16% lower when irrigated during a hot and dry season. That result would ordinarily not be expected. Whether this depression was due to an increase in or an activation of the parasites in the soil or to harmful side effects (such as oxygen deficiency) of the irrigation itself was not determined. Before the third cutting Lincoln brome-grass lacked vigor and luster, and the production was obviously depressed on irrigated land. The dry matter percent was higher on irrigated plots - Lincoln brome-grass had 22.1% dry matter on the irrigated plots and 16.8% on non-irrigated plots in the third harvest. The higher dry matter percent on the wetter plots suggests that the parasites in the soil may have severed the roots and in that way limited the water supply for the plants during the last three weeks before cutting when no irrigation was provided.

All the roots sampled from the field for isolation of pathogenic organisms on August 18 were obtained from irrigated plots of Lincoln brome-grass. Of 35 isolations 5 proved to be pathogenic Fusarium spp. Two were later identified as Fusarium poae. The other three tested, and an additional three which were discarded because of contaminations were probably all of the same species. These data indicate that Fusarium spp. are very frequent parasites on brome-grass roots in wet soil. Inoculations of brome-grass plants in the greenhouse showed Fusarium poae to be very harmful to Lincoln brome-grass.

Dexon fungicide was selected for use because it was believed that Pythium spp. were a major cause of the root rot. If Fusarium spp. had been initially considered the major fungus causing root death, another fungicide would preferably have been selected. Dexon is highly specific against Pythium spp. but is relatively ineffective for control of Fusarium spp. It significantly improved the stand of Lincoln brome grass, however, for the third cutting on the irrigated plots in the field.

In experiment II and III conducted in the greenhouse only small differences between fungicide treatments and the check were obtained. Dexon fungicide and propylene oxide gaseous sterilization were detrimental to yield until after the second cutting. Plant nutrients not utilized in these first two periods may have contributed to later yield increases. Heat sterilization gave increased production for the first two harvests, a response which may have been due to released nitrogen, as mentioned by Vanterpool (12). After the experiments were terminated rotted roots were found in all cultures, regardless of treatment, indicating that none of the fungicide treatments used gave completely protection. Recontamination with spores and propagules of the pathogenic fungi could later produce root infection in the sterilized soil. Such recontaminations may come from dust, splashing water or handling. Once begun it can be more severe than the original infestation present before sterilization.

Testing for pathogenic effects of fungi on roots of established plants by the method described in experiment IV has several drawbacks.

The major objection is that the vermiculite cannot be washed away from the roots when the experiment is terminated. Therefore a fixed weight of vermiculite must be put into the pots initially so the root weights finally can be determined by subtraction. In experiment IV dry vermiculite weighed 200 grams per pot while the root averaged only 20 gram. Under such conditions great care must be exercised if results are to be obtained.

This investigation has not finally solved the problem why brome grass is killed on organic soil. The results obtained, however, substantiate the hypothesis that fungi reduce the growth and longevity of Lincoln brome grass. The data indicate that, especially under wet conditions, Fusarium poae may well be one of the principal fungi causing root death and dying of Lincoln brome grass as early as the latter part of the first harvest year. Plant feeding nematodes were not recovered from either soil or from decaying brome grass roots in samples from the Muck Farm, examined by the Michigan State University Nematode Laboratory, and may therefore be excluded as a possible cause of root death of brome grass on organic soil.

Further elucidation of the conditions favoring disease and possibly synergistic relationships with other soil fungi should be attempted.

Fusarium poae has been considered a weak parasite, Sprague (9), but may prove to be more adapted to parasitism in a highly organic environment, such as in muck soil, than on mineral soils. It is intriguing that the

disease appears to be of much less consequence on mineral soils where most of the experience with Fusarium poae has been gained.

Results cited by Tesar and Shepherd (10) show that the dying of bromegrass is not readily detected by yield differences until in the second harvest year. Results in the present experiment indicate root death and decreased stands may be evidenced as early as the end of the first harvest year.

SUMMARY

Investigations were conducted in the field and under controlled conditions to determine why smooth brome grass dies on organic soil during the growing season, and its reaction to fungicide treatments, and different levels of water supply, fertility, and temperature.

1. Isolations of fungi from decayed brome grass roots in the field show that Fusarium spp. are frequent and apparently harmful parasites in muck soil.

2. Fusarium poae was particularly identified with killing of Lincoln brome grass roots in the greenhouse. Pythium spp. appeared to be associated with similar injury.

3. Dying of Lincoln brome grass in the field was detected in the third cutting near the end of the first harvest year.

4. Dexon fungicide selected for its specific action against Pythium spp. was not as helpful as desired in reducing the dying of roots later attributed to Fusarium poae. Dexon improved Lincoln brome grass yields in third cutting and stands after the end of the first year in the field.

5. Dexon fungicide increased yields of brome grass in fertilized, but not in unfertilized cultures in the greenhouse.

6. Yields were reduced approximately 16% as a result of relatively high irrigation on a muck soil.

LITERATURE CITED

1. Andrews, E. A. Seedling blight and root rot of grasses in Minnesota. *Phytopathology* 33:234-239. 1943.
2. Buchholtz, W. F. A comparison of grass pathogenic effects of Pythium graminicola, Pythium debaryanum, and Helminthosporium sativum on seedlings of crested wheatgrass. *Phytopathology* 39:102-116. 1949.
3. Drolsom, P. N., and Nielsen, E. L. Selection for seedling disease tolerance in smooth brome grass. *Crop Science* 3:283-285. 1963.
4. Gilman, J. C. A manual of Soil Fungi. pp 358. Iowa State Coll. Press. Ed. 2, 1957.
5. Hansen, H. N., and Snyder, W. C. Gaseous sterilization of biological materials for use in culture media. *Phytopathology* 37:369-371. 1947.
6. Hawk, V. B., and Welch, A. W. Tolerance of varieties of smooth brome grass, Bromus inermis, Leyss., to injury by root rot, Pythium graminicola. *Subr. Journ. Amer. Soc. Agron.* 40:809-817. 1948
7. Kaufmann, M. J., Drolsom, P. N., and Nielsen, E. L. Reaction of smooth brome grass to seedling pathogens. *Agron. Journ.* 53:77-80. 1961.
8. Nielsen, E. L., Dickson, J. G., and Smith, D. C. Strain and seed treatment as factors in germination and seedling growth of smooth brome grass. *Phytopathology* 49:8-12. 1959.
9. Sprague, R. Diseases of Cereals and Grasses in North America. pp. 347-348. 1950.
10. Tesar, M. B., and Shepherd, L. N. Evaluation of forage species on organic soil. *Agron. Journ.* 55:131-134. 1963.
11. Thomason, T. F. and Dickson, J. G. Influence of soil temperature on seedling blight of smooth brome grass. *Phytopathology* 50:1-7. 1960.
12. Vanterpool, T. C. Pythium root rot of grasses. *Sci. Agr.* 22:674-687. 1942.

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