FAMILY IDENTIFICATION OF LEPIDOPTERA LARVAE WITH REFERENCE TO COMPUTER GENERATED KEYS

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ABSTRACT

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Ву

Peter John Martinat

An illustrated family key to immature Lepidoptera is developed, and brief diagnostic descriptions for larvae of the suborders and superfamilies are given. General conclusions on taxonomy and identification of immature Lepidoptera are also given in relation to specimens examined. The philosophy and history of key construction is reviewed, and the theoretical advantages of generating identification keys by computer are discussed. The key generating program of Dallwitz (1974) was investigated as an aid in developing the immature Lepidoptera key. The results from use of this program were unsatisfactory, and the reasons for its failure are discussed.

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Peter John Martinat

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INTRODUCTION

The only currently available family keys to Lepidoptera larvae are those by Fracker (1915), Forbes (1923), Capps (1939), Chu (1949)

Gerasimov (1952), and Peterson (1956). Recent advances in taxonomic knowledge of immature Lepidoptera have made these keys inadequate. In addition, they are based on outdated classifications (Comstock 1892, 1918) of the order. The keys by Capps and Chu are incomplete, excluding many Microlepidoptera and less common families. Forbes' key is similar to Fracker's and uses mixed chaetotaxy terminology. Peterson's key, although the most recent, contains many taxa that do not key properly. He also used Fracker's chaetotaxy terminology, now considered outdated (Hinton 1946). Gerasimov's key has only recently been translated from Russian (Gerasimov 1962). It is not generally available, uses a style to which North American workers are unaccustomed, and is based on eastern European fauna.

Recent investigations have made available a number of computer programs which automatically generate identification keys. The advantages of computer generated keys were discussed by Pankhurst (1970b), Morse (1971), Watson & Milne (1972), and Dallwitz (1974). In hopes of developing a more reliable key than Peterson's (1956), and avoiding many of the problems key writers have had, the most recent of these programs, Dallwitz' "KEY" was obtained. The objectives of this research were a) to write a new illustrated family key to Lepidoptera larwae, and b) to investigate KEY as an aid in developing this key.

This investigation is divided into 2 parts. The first is a general systematics section, in which a key, description of characters, diagnostic descriptions of the superfamilies, and observations on identification and taxonomy of immature Lepidoptera are given. The second reviews the historical development of keys, describes KEY, and discusses the results of its use. In conclusions, the results obtained from KEY are compared with the hand written version. The appendices include the classification of the Lepidoptera used in the systematics section, a table comparing the chaetotaxy nomenclatorial systems of different authors, and computer printouts obtained from KEY.

FAMILY SYSTEMATICS OF IMMATURE LEPIDOPTERA

A. METHODS AND PROCEDURE

Character Evaluation

All characters identified in a thorough search of the morphological and taxonomic literature were considered potentially useful, including those used at interspecific, intergeneric, as well as at interfamily levels (Fracker 1915, Heinrich 1916, McIndoo 1919, Jaywickreme 1940, Hinton 1946, 1947, 1952, 1956, Gerasimov 1952, MacKay 1964). Many new characters were added, based on preliminary examinations of larvae.

Measurements were taken of body and head features, such as the fronto-clypeus, labrum, head capsule, length and height of body segments, and distances between setae and ocelli. Sketches and notes were made on qualitative features. After about 300 specimens had been examined, these measurements were delineated into characters and character states such that useful taxonomic information would be obtained from the data. All characters were stated in qualitative terms but were given precise quantitative limits for each state within the character. For example, head shape was defined as the ratio of epicranial length to its greatest width. Four states were delineated as: A, longer than wide (ratio greater than 1.10); B, as long as wide (ratio between 0.90 and 1.10); C, between 0.50 as long as wide to nearly as long as wide (ratio between 0.50 and 0.90); and D, less than half as long as wide (ratio less than 0.50).

Characters were worded in a precise manner. For example, an important character in Peterson's key (1956, Pt. I, p. 69, B18) was:

setae kappa (L2) and eta (L1) remote on abdominal segments; or (B18a) setae kappa and eta adjacent on abdominal segments. This was changed to: L1 closer to the spiracle than it is to L2, or L1 & L2 closer to each other than either is to the spiracle.

An initial list of 65 characters was compiled. Only about half of these proved useful at the family level. Ten characters difficult to delineate into states until all available variation was known were added after all families had been examined. Fifteen more characters were added to 20 families of Microlepidoptera and 10 more to 26 families of Macrolepidoptera for separation of difficult groups in the computer generated key. A list of 100 characters was thus assembled (App. C, p. 99).

Specimen Evaluation

Specimens accompanied by collection locality, date, and biological information were chosen for examination. All specimens were assumed to be correctly identified. Considering the paucity of taxonomic knowledge on immature Lepidoptera, this is a dangerous assumption. However the goal of a key at the family level does not make correct identification to species a critical requirement.

Several assumptions directed the choice of specimens examined. A variable character (with more than 1 state) between species of the same genus will also likely be variable between genera within the family. Therefore, assuming that the classification is good, examination of specimens at the generic level will be sufficient for information on character variability at the family level. In constructing a key to

families the genus becomes the OTU (=Operational Taxonomic Unit, Sneath & Sokal 1972). Exceptions are large and complex genera such as Ethmia, Acronycta, Euxoa.

A second assumption was that a flawless key will be made only when larvae of all Lepidoptera genera are available. Given the present paucity of collections, one must take a sampling approach. Specimens were chosen which would show the widest range of biological and morphological diversity within each family.

Poor collections also require critical assumptions on the reliability of taxonomic data. If a family is a natural group (with high overall information content, Sneath & Sokal 1972) then general diagnostic statements can be made about it. Examination of a small percentage of the genera will be sufficient to get most of the diagnostic taxonomic information (what percentage is "sufficient" can not be determined empirically). Examination of additional genera will add decreasing amounts of new information (Fig. 15, curve A). On the other hand, if a family is merely a heterogeneous collection of genera (a "dumping ground"), then general diagnostic statements about the family are difficult, and examination of additional genera will add new information indefinitely (Fig. 15, curve B). If we are aware of the nature of a family (whether it is "natural"), then this argument allows us to make confident statements about a family when only a small number of specimens are available.

B. CHARACTERS USED IN THE KEY

Head. The caterpillar head is a sclerotized capsule with a very large occipital foramen strengthened dorsally by an inverted Y-shaped

internal ridge. Hinton's (1947) interpretation of the morphology of the head sutures is used in this thesis. Each anterior arm of the inverted Y-shaped ridge extends to the anterior margin of the head mesad of the dorsal articulation of the mandibles (Figs. 1 & 2). Externally, these arms are represented by the <u>lateral adfrontal sutures</u>. The stem, or midcranial part of the ridge extends to the <u>epicranial notch</u>. This part of the ridge is represented externally by the <u>median adfrontal suture</u>. In early instars the head capsule is shed whole at ecdysis. In later instars the head capsule splits along the dorsal lines of weakness, the <u>ecdysial lines</u>. The area between the ecdysial lines and the adfrontal sutures are the <u>adfrontal areas</u>. The triangular area between the lateral adfrontal sutures is a combined front and clypeus, or <u>fronto-clypeus</u>. Anterior to the fronto-clypeus is the <u>anteclypeus</u> which articulates with the labrum.

The orientation of the mouthparts is usually correlated with the relative lengths of the sutures. In the hypognathous head, typical of the Macrolepidoptera (Fig. 97), the mouthparts are directed downward, perpendicular to the long axis of the body, and the median adfrontal suture is very long relative to the length of the fronto-clypeus (Fig. 77). In the semi-prognathous head, typical of internal feeding Microlepidoptera the mouthparts are directed slightly more forward, at an obtuse angle with the long axis of the body (Fig. 32), and the fronto-clypeus is longer than the median adfrontal suture (Fig. 78). In the prognathous head, typical of leaf miners, the mouthparts are directed forward, parallel to the long axis of the body, and the apex of the fronto-clypeus reaches the epicranial notch, causing disappearance of the median adfrontal suture (Figs. 59, 63, 68). In some leaf miners the

severity of morphological change is such that the fronto-clypeus assumes a square or rectangular shape (Figs. 65, 67, 71, 72).

Eyes. Catepillars typically have 6 ocelli on each side of the head, adjacent to the antennal sockets. They are usually arranged with 5 in a semicircle (numbered 1-5 from top to bottom) with the 6th behind the others (Fig. 2). Reductions in the number of ocelli are usually found among leaf-mining families. The Nepticulidae, Eriocraniidae, and Opostegidae have only 1 ocellus on each side. Reductions in number are also found in the Heliozelidae, Oinophilidae and Gracillariidae. Many Prodoxinae, Lyonetiidae, and Tineidae have only 5 ocelli. Variations in the arrangement of ocelli are of 2 sorts. Ocelli 1 & 2 are sometimes close together and separated from the others (many Lyonetiidae, Gracillariidae, Elachistidae, and Tineidae; Fig. 85). Ocellus 5 is sometimes separated from the others as in most Macrolepidoptera (Fig. 88). All 6 ocelli are nearly equal in size, but in the Satyrinae, the 3rd ocellus is about twice as large as the others (Fig. 51).

Head Chaetotaxy. The terminology of the head setae used here is that of Hinton (1946), who named 22 primary setae on each side of the head (Fig. 1 & 2). Four minute setae (V1, V2, V3, G1) and 5 other setae (C1, C2, F1, AF1, AF2) on the frontal areas have not been used for taxonomic purposes. P1 and P2 are caudo-dorsad near the adfrontal sutures. A1, A2, A3, and L1 form a semi-circle cephalo-dorsad of the ocelli. A2 is usually closer to A1 than to A3 and is shorter than A1. A3 is usually closer to L1 than to A2 and is as long as A1. O1, O2, and O3 are closer to the ocelli. O1 is close to and caudad of ocellus 3. O2 is usually caudad of ocellus 1, and O3 is behind ocellus 6. O1 is as short as A2, O2 is usually as long as A1, and O3 is as short as or

shorter than 01. S01, S02, and S03 are below the ocelli. S01 is ventrad of ocelli 5 and 6, S02 and S03 are in the genal area near the base of the maxillae.

External Features of the Cuticle. Two types of structures may occur on the body of caterpillars; those associated with setae in origin, and those having an internal cavity connected with the body cavity. The simplest structure is the hair-like seta. Most setae are simple, but in many families plumose, knobbed, disc-like, or spatulate setae are found. A seta arises from a small sclerotized ring-like papilla on the ectocuticle and internally connects with at least one hypodermal cell. A sclerotized area occuring around the base of one or more setae is a pinaculum (Fig. 3, dotted circles). If the pinaculum is distinctly elevated and cone-shaped it is a chalaza (Fig. 53). If the chalaza is a large structure bearing many branching spines or setae, it is a scolus. Secondary setae may be grouped on non-sclerotized fleshy lobes, or flattened disc-like sclerotized verricules or convex sclerotized verrucae. Some structures such as the mid-dorsal horn or tubercle on A8* of sphingidae are thought to be modified setae (Gerasimov 1952). Other structures resembling horns, tubercles, and antlers are found in some Nymphalidae, Notodontidae, and Bombycidae. The anal comb found on many Oecophoridae, Gelechiidae, Tortricidae, Thyatiridae, and Hesperiidae probably originated from modification of cuticular granules (Gerasimov, 1952).

Structures not associated with setae in origin are usually fleshy lobes or protuberances. Eversible glands are found in the Lyonetiidae,

^{*}Standard abbreviations for the pro-, meso, and metathorax are TI, TII, & TIII; and for the abdominal segments Al, A2, A3.....AlO.

Yponomeutidae, Papilionidae, Lymantriidae, Notodontidae, and Noctuidae. In most groups the gland is cephalad of the prothoracic legs and may secrete noxious defensive fluids (Herrick & Detweiler, 1919), but the Papilionidae have an eversible mid-dorsal Y-shaped gland (osmeteria) on TI, and the Lymantriidae have mid-dorsal glands on A6 & A7. The Danainae and some Papilionidae, Noctuidae, and Geometridae have fleshy lobes on various segments.

Thoracic & Abdominal Setae.

Fracker (1915) distinguished 3 kinds of setae on the bodies of caterpillars. Primary setae are sensory in function and are present in the first and last instars of all but a few families. They are relatively constant in numbers and position throughout the order. Subprimary setae are like primaries in function except they do not appear until after the first molt of the larva. Primaries and subprimaries have been subject to nomenclatorial systems and their presence has been homologized in all groups throughout the order. Slight variations in position and numbers provide many useful taxonomic characters. For purposes of identification, primary and subprimary setae are all referred to as primaries.

Secondary setae are those varying in numbers, size, and position within as well as across taxa and do not appear until after the first instar. They usually occur in one of two patterns: as numerous small setae scattered over the cuticle (Figs. 36, 37, 46, 54); or as dense tufts or pencils on verrucae or verricules. Since primary setae occur on caterpillars throughout the Lepidoptera, their presence is probably a primitive condition (Hinton 1946). The appearance of secondary setae probably happened as an adaptation to an external feeding habit.

Verrucae and verricules usually occur in the same positions as the pinacula of primary setae on other larvae, suggesting that verrucae and verricules are homologous to pinacula. Small secondaries scattered over the cuticle such as are found in the Hesperioidea, Papilionoidea, Saturnioidea, Bombycoidea probably originated from modification of cuticular granules (Gerasimov 1952).

Secondary setae are extremely variable in their presence. Many
Tortricidae, Noctuidae, Lyonetiidae, Pterophoridae, and Oinophilidae have
spinules (cuticular spines which may represent intermediate conditions in
the development of secondary setae). Primaries are obscured by
secondaries in most butterfly larvae, but in the Megathymidae primaries
stand out above surrounding secondaries. Some Notodontidae, Noctuidae,
Ethmiidae, Scythridae, and Pterophoridae have extra setae on primary
pinacula. Since confusion might arise whether a structure should be
called a pinaculum or a verruca, an arbitrary criterion is used in the
key. If 6 or fewer setae are present on a sclerotized area, it is a
pinaculum. If more than 6 are present, it is a verruca. Many Cossidae,
Noctuoidea and Geometroidea have extra setae in fixed positions remote
from the primaries. These setae resemble primaries in size and have
occasionally been given names (for example, seta lambda of Fracker, 1915=
L4 of McGuffin 1958).

Body Chaetotaxy. Numerous systems for naming the primary setae have been proposed. The most commonly used are those of Dyar (1894), Fracker (1915), Heinrich (1916), Forbes (1923), and Hinton (1946). The systems of Gerasimov (1952) and Mutuura (1956) are less well known (App. B). Hinton's is considered the most reliable in the homologies it suggests and has been adopted by most contemporary workers.

Hinton was the first to recognize 2 kinds of primary setae:
microscopic or proprioreceptor (prefixed "M") which, owing to their
position, make contact with the segment immediately cephalad to their
position; and long tactile setae which function as receptors of external
stimuli. Because of their small size, the microscopic setae have little
taxonomic value and will not be discussed. The tactile setae are divided
into 6 groups based on their location (Fig. 3).

XD Group. Two XD setae are on the cephalic margin of the prothoracic shield. Both are nearly equal in length, are longer than D1, and as long as D2. XD2 is below XD1.

Dorsal (D) Group. Two D setae are usually present on all segments close to the dorsal midline. Dl is shorter than D2. On T1, both are on the prothoracic shield, and Dl is usually dorsad of D2. On TII & TIII, Dl is above D2, and they are usually closer together than D2 is to the next seta below. On A1-8, Dl is cephalad of D2. On A9, D2 is dorsad, caudo-dorsad, or caudad of D1.

Subdorsal (SD) Group. Two SD setae are present on all segments except A9. SD1 is longer than SD2. On TI they are on the prothoracic shield below D1, D2, XD1 and XD2. SD1 is usually ventrad of XD2 and cephalad of SD2. Sometimes (in many Noctuoidea) SD1 and SD2 are very close together and excluded from the prothoracic shield. On TII & TIII, SD1 is ventrad of SD2, and D1, D2, SD2, SD1 from a nearly straight vertical line with D1 & D2 on one pinaculum and SD2 & SD1 on another. On A1-8, SD2 is usually minute and cephalad of the spiracle, and SD1 is usually dorsad of the spiracle. On A9 SD2 is usually absent and SD1 is ventrad of D1.

Lateral (L) Group. Three L setae are usually present on all segments except TI and A9. On TI they are cephalad of the spiracle, usually on a single pinaculum. L1 is the longest, and L2 the next longest. L2 is usually cephalad of L1, and L3 is closest to the spiracle. In many Copromorphoidea, Noctuoidea and Pyraloidea L3 is missing on TI. On TII & TIII L1 is primary, L2 & L3 are subprimary. L3 is caudo-dorsad of L1, and L2 is cephalo-ventrad of L1. L1 & L2 usually share a pinaculum, and L3 has its own. On A1-8 L1 & L2 are close together and ventrad of the spiracle in most Gelechioidea, Tortricoidea, and Pyraloidea, but are far apart with L1 often caudad of the spiracle in most Tineoidea, Yponomeutoidea, Geometroidea, and Noctuoidea. L3 is caudo-ventrad and farther from the spiracle than L1 and L2. On A9 the L group is often unisetose or bisetose. If all 3 are present they are usually arranged vertically with L1 in the middle, and with L3 below L1.

Subventral (SV) Group. The SV group is usually bisetose on TI and unisetose (bisetose in Thyrididae, Pterophoridae, some Tineidae) on TII & TIII. They are dorsad of the coxa and are often on large pinacula. On Al-8, the SV group may be unstable in number, and may vary even on opposite sides of the same segment. In most Microlepidoptera and cutworm-type Macrolepidoptera, the SV group is uni-, bi-, or trisetose on Al, 2, 7 & 8, and trisetose on the basal part of ventral prolegs. In many Ethmidae, Oecophoridae, Pterophoridae, Scythridae, secondary setae may be present in the SV group. In most external feeding Macrolepidoptera, the ventral prolegs have many secondary setae, while other abdominal segments may have a normal SV group, or an SV verruca. In other families or the SV group may be obscured by numerous secondaries.

Ventral (V) Group. A single V seta is present on all segments. It is nearer to the ventral meson than the SV group. On TI-TIII it is on the underside of the coxa. On A3-6 it is on the underside of the proleg.

Caudal Abdominal Segment. Little taxonomic value has been found in the four pairs of setae on the anal shield. Hinton did not discuss them, but MacKay (1959) homotyped them with setae on A1-9. They are D1, D2, SD1, and SD2. There are usually 9 setae below the transverse anal slit arranged in 3 rows around each caudal proleg. In some Gelechiidae, Cosmopterygidae, Oecophoridae, Ethmiidae, Stenomidae, and Notodontidae, more than 9 setae are present in this area.

Pinacula and verrucae. Pinacula are sometimes named according to the primary setae occuring on them. Seta D1 is on pinaculum D1. If D1 & D2 setae are on a single pinaculum, that pinaculum is D1+D2. Verrucae are also labeled in this manner, since they are modified pinacula. But since many verrucae are fused pinacula, the labelling is simplified. If 2 dorsal verrucae are present, they are D1 and D2, but if only 1 dorsal verruca is present, it is verruca D. But if 2 verrucae of different primary groups are fused (for example the fused D and SD verrucae on TII & TIII in the Arctiidae, Fig. 100) it is the D+SD verruca.

Thoracic Legs. Caterpillars typically have 3 pairs of well sclerotized legs. In many families with specialized habits (leaf miners) legs are rudimentary or lost. Otherwise they are remarkably consistant in structure. The leg consists of a coxa, trochanter, femur, tibia, tarsus, and a distal claw. In the Micropterygidae the coxa, trochanter, and femur are fused and the leg appears 3-segmented. In the other families, the trochanter is little more than a narrow band fused with the femur (Gerasimov 1952) and the leg appears 4-segmented.

<u>Prolegs</u>. Prolegs are paired ventral muscled outgrowths of the body wall. In all suborders except the Zeugloptera, prolegs are primitively present on A3-6 (ventral prolegs) and on A10 (caudal prolegs). A different compliment of prolegs (usually reductions) is thought to be a derived condition (Hinton 1955a). In the Zeugloptera true prolegs occur on A1-8.

Prolegs show a wide range of development. In many leaf-mining and internal-feeding groups they are absent or rudimentary. In the Nepticulidae prolegs are lost, but "ambulatory warts" have evolved on A2-7 (Hinton 1955a). In mature gracillariid larvae, prolegs are lost on A6. The Megalopygidae have pairs of lobes on A2 and A7 which resemble prolegs but lack crochets. The Nolidae lack prolegs on A3. The loss of prolegs in most Geometridae (usually the first 3 pairs) and in some Noctuidae (the first 1 or 2 pairs in the Acontiinae, Lithacodiinae, and Plusiinae) is associated with a specialized means of locomotion called "looping". In the Drepanidae the caudal prolegs are lost, and in many Notodontidae they are lost, rudimentary, or modified to serve as defensive structures.

The proleg consists of 2 parts: a proximal <u>base</u> which bears the SV setae, and a distal <u>planta</u> from which the crochets arise (Figs. 118-123). Contraction of retractor muscles inserted on the planta pulls its center inwards, disengaging the crochets. Turgor pressure pushes the planta out once the muscles are relaxed (Hinton 1955a).

Two kinds of prolegs are found in caterpillars. In the first, found in the Tineoidea, Yponomeutoidea, Cossoidea, Tortricoidea, Gelechioidea, Copromorphoidea, and Pyraloidea the proleg base is little more than a ring encircling the base of the planta (Figs. 118-120). The planta is

usually cylindrical and may be almost indistinguishable from the proleg base (Fig. 119), or elongate (many Plutellinae, Glyphipterygidae, Pterophoridae; Fig. 120). In this form of proleg, crochets are usually in transverse bands, a complete or incomplete circle, or a mesopenellipse. The second kind, found in the Zygaenoidea, Papilionoidea, Geometroidea, Saturnioidea, Sphingoidea, and Noctuoidea, the proleg base is elongated and forms the greater part of the proleg (Figs. 121-123). Often it is partially sclerotized and bearing numerous secondary setae (Fig. 122). The planta is reduced to a lobe at the distal end of the proleg base, with the crochets usually in a mesoseries. These 2 kinds of prolegs parallel feeding habits. The first kind is typical of internal-feeding caterpillars (Microlepidoptera), and the second kind is typical of exposed feeders (Macrolepidoptera).

Crochets. Crochets are small hooks arranged in rows or circles around the periphery of the planta. Their development probably began from the enlargment of cuticular granules which gradually assumed the shape of hooks (Gerasimov 1952). The primitive arrangement would thus be large spines arranged in circular multiple rows. This multiserial arrangement is found in the Gracillariidae, Acrolophinae, Yponomeutidae, Hepialidae, Adelinae, Tischeriidae, and Castniidae (Figs. 20, 30). Further specialization led to a reduction in the number of rows to a uniserial circle and differentiation of the size of crochets. If all the crochets in a circle are the same length they are uniordinal (Fig. 6); if 2 alternating lengths occur, they are biordinal (Fig. 7); while 3 alternating lengths are triordinal (Fig. 8).

Circles are often broken into rows, semi-circles, bands, etc. An incomplete circle has a gap in an otherwise continuous row. If the gap

occurs on the mesal side, the arrangement is a <u>lateropenellipse</u>. If the gap is on the lateral side, it is a <u>mesopenellipse</u> (Fig. 12). If a circle has a gap on the lateral side and another on the mesal side, then the crochets are in 2 <u>transverse bands</u> (Fig. 5). If a single row of crochets is present, then the arrangement is a <u>single transverse band</u> (Fig. 4). Most arboreal feeders have a single longitudinal row of crochets on the mesal side of the planta, parallel to the meson. If it is a single longitudinal row with crochets of uniform length, it is a <u>homoideous mesoseries</u>. If the crochets in the center of a mesoseries are longer than those at either end (Arctiidae, Ctenuchidae, Pericopidae) then it is a <u>heteroideous mesoseries</u> (Fig. 56). If two longitudinal parallel rows occur, the outer is a <u>lateroseries</u> and the inner is a mesoseries. (Fig. 14).

C. KEY TO FAMILIES

How to Use the Synoptical Key

A synoptical key based on a design by Leenhouts (1966) has been incorporated into the analytical key (couplets 75-88). Included in the synoptical key are 10 families of Gelechioidea and Tortricoidea for which reliable key characters are lacking, making identification difficult. Diagnostically useful characters are arranged as numbered couplets. A couplet begins with the name of the character followed by 2 or more states marked with letters. After each state, abbreviations (example, C=Cosmopterygidae) denote all the taxa for which the state of the character applies. Taxa listed after more than 1 state are those which are variable for the character.

To use the synoptical key, list the taxa (C M W B O E G S T P).

Begin with any couplet 75-88 and proceed in any order. Compare the specimen with a character, and select the state which fits best, and cross off the families which are NOT listed. For example, if the specimen has the fronto-clypeus extending to the epicranial notch, or nearly (couplet 75, state A) cross off Blastobasidae, Ethmiidae, Stenomidae, Tortricidae, and Phaloniidae (C M W B O E G S T F). Next, proceed to any other couplet and repeat. For example, try couplet 85 (anal comb present or absent). If the specimen has an anal comb, Cosmopterygidae, Momphidae, Walshiidae and Oecophoridae are eliminated, leaving only Gelechiidae, which is most likely the correct family.

If this does not lead to a satisfactory identification (either because all taxa have been crossed off, or more than one remain), a second more tedious approach is possible. This involves tabulation of similarities between the specimen and all the taxa, rather than elimination of taxa. Again, list the taxa. Assume the specimen has the fronto-clypeus extending to the epicranial notch, or nearly (state A). Tally (/) after C M W O G. Assume state A for couplet 76, and tally (/) after C S. Continue in this fashion until the specimen has been compared with all the couplets, then total the number of similarities for each taxon on the list. The taxon with the greatest number of similarities is most likely the correct one.

The couplets leading to the synoptical key (55, 70, 71, 72, 73, 74) use reliable characters which help narrow down the list of possibilities. If directed to the synoptical key, begin with the list of taxa at the couplet leading to it. For example, if at couplet 70, your specimen has an anal comb you can begin the synoptical key with all but Tortricidae

and Gelechiidae crossed off your list. The list of possibilities will have been narrowed to 4 or fewer by all couplets except 74, before entering the synoptical key.

Due to a lack of good characters and to unstable family definitions (especially in the Gelechioidea), even the synoptical key will not guarantee a correct identification. The greatest difficulty will be in separating tortricids and phaloniids; gelechiids, ethmiids and oecophorids; and cosmopterygids, momphids, and walshiids.

FAMILY KEY TO LEPIDOPTERA LARVAE

* = Kare	group, not likely to be collected.
1.	Antennae nearly as long as width of the head, and inserted above the ocelli (Fig. 23); setae modified as thick bulbous scales (Fig. 21); prolegs present on A1-8 (Fig. 22; mature larvae less than 5 mm.; sluglike, found in mosses, liverworts, and lichens, or in litter and duff)MICROPTERYGIDAE* Antennae much shorter than width of the head, and inserted between the ocelli (if present) and the base of the mandibles (Fig. 1); setae not modified as above; complement of prolegs never as above
2(1).	External parasitoids on Fulgoroidea and other Homoptera; primary setae absent; body stout, cyphosomatic, with head retracted into prothorax; thoracic legs present but rudimentary; prolegs absent, but uniordinal crochets arranged in a complete circle are present on A3-6, 10 (Fig. 42) (mature larvae 5 mm. or less)
3(2).	Crochets present on at least 1 segment of Al-94 Crochets absent on all abdominal segments, or present only on AlO (irregularly spaced spines may be present on prolegs)
4(3).	Crochets absent on AlO
5(3).	Segmented sclerotized thoracic legs absent
6(5).	Fronto-clypeus rectangular, circular, or trapezoidal in shape; apex rounded or open (Figs. 59, 65, 67, 69, 74)7 Fronto-clypeus more triangular or pentagonal in shape; apex ending in a more acute angle, never open (Figs. 61, 63, 68)

- 8(7). Fronto-clypeus much less than 2X as high as wide, never wider behind than in front (Figs. 67, 69) body cylindrical; prolegs, if well developed, present on A2-7; TII & TIII usually with pairs of ventral fleshy lobes (Fig. 27; miners in leaves, bark, or fruits, or forming galls in twigs or petioles)......NEPTICULIDAE* Fronto-clypeus at least 2X as high as wide, may be wider behind than in front (Figs. 59, 71, 72, 73); compliment of prolegs not as above; lobes on thoracic segments present or absent...............9

11(6).	Body swollen, cyphosomatic; head with 3 ocelli on each side; prolegs absent (borers in flowers of Yucca)
	Body not swollen, more cylindrical or dorso-ventrally flattened; head with fewer than 3 ocelli on each side; prolegs present or absent
12(11).	Ecdysial lines joining anterior margin of head well laterad of adfrontal suture and including the antennae within the adfrontal areas (Fig. 68); head with a single ocellus on each side (Fig. 70; leaf miners)ERIOCRANIIDAE* Ecdysial lines inconspicuous or joining anterior margin of head between antennae and adrontal sutures and excluding the antennae from the adfrontal areas (Fig. 61); head with 2 ocelli on each side (Fig. 62; leaf miners with pupation in an oval case cut from the leaf at the end of the mine)
13(5).	Numerous secondary setae, prominent urticating scoli, or long hairy protuberances usually present; sucker—like structures usually present on ventral surface of abdominal segments (external feeders on decidu—ous trees)LIMACODIDAE Secondary setae and scoli absent; sucker—like structures absent on ventral surface of abdominal segments
14(13).	Prolegs present on A3-6 (rudimentary on A10); ventral part of prolegs covered with spines (not true crochets); head not retracted into prothorax; mature larvae 50 mm. or more (borers in stems of banana; tropical & subtropical)
15(14).	Body with prominent fleshy lobes; all setae minute or absent (tropical & subtropical external feeders on deciduous trees)

16(4).	All prolegs rudimentary or absent; crochets on A3-6 in multiserial transverse rows; secondary setae absent (very small larvae, internal feeders in leaves, needles, flowers, seeds)(Nematois sp.) ADELINAE* Only caudal prolegs rudimentary or absent, ventral prolegs well developed; crochets on ventral prolegs in a mesoseries; secondary setae present at least on prolegs (large larvae, external feeders on deciduous trees and shrubs)
17(16).	Crochets in a uniordinal or biordinal mesoseries plus a uniordinal lateroseries (Fig. 14); anal shield usually modified into a spinose caudo-projecting elongate process
18(4).	Segmented sclerotized legs absent on all thoracic segments (ventral fleshy lobes may be present); head prognathous and extremely dorso-ventrally flattened (Fig. 59)(part) TISCHERIIDAE* Segmented sclerotized legs present on at least 1 thoracic segment; head prognathous or not
19(18).	Well developed crochet-bearing prolegs present only on A3-5 & 10 (absent on A6); ocelli 1 & 2 close together and separated from 3, 4 & 5; distance between ocelli 2 & 3 at least 3x the distance between ocelli 1 & 2 (Fig. 85; external feeders on foliage, may construct folded or rolled leaf shelter)

abdominal segments; arrangement of ocelli variable.....20

20(19).	A6 with 7 or fewer setae on each side below the top edge of the spiracle (Fig. 3; L1, L2, L3, SV1, SV2, SV3, V1, (MV3 usually too minute to see easily)) (spinules may be present on cuticle, if they are large and resemble secondary setae, try also couplet 111)
21(20).	At least some secondary setae grouped on verrucae or verricules, or scoli present; if verrucae or verricules resemble pinacula, then with 7 or more setae on each (Figs. 55, 57, 58, 99-102)
22(21).	A3 with no more than 5 setae on each side above the spiracle; if more than 5 are present (a few Pterophoridae, Cossidae, Scythridae, Notodontidae, Dioptidae), the extra setae are usually on primary pinacula (D1, D2, or SD1) and are usually nearly as large as the primary setae

23(20).	penellipse, mesopenellipse, transverse bands, or reduced in number, never in a mesoseries; if arranged as a mesopenellipse the gap free of crochets is usually distinctly less than 1/3 the circumference of projected circle; also, the entire series of crochets on each caudal proleg is more nearly perpendicular than parallel to the meson (Figs. 5, 6, 8, 12)24 Crochets on ventral prolegs in a mesoseries or pseudocircle, never in a lateropenellipse; if a mesoseries resembles a mesopenellipse (some Noctuidae, Epiplemidae, Ethmidae, and Pterophoridae), then the gap free of crochets is equal to or greater than 1/3 the circumference of the projected circle; also the entire series of crochets on each caudal proleg is more nearly parallel than perpendicular to the meson (Figs. 11, 56)
24(23).	Crochets uniordinal (Fig. 6)
25(24).	TI with L (prespiracular) group unisetose or bisetose26 TI with L (prespiracular) group trisetose32
26(25).	TII & TIII with the SV group bisetose, and with L3 absent; head with 01 usually between ocelli 2 & 3 or more cephalad than 2 & 3 (Fig. 87; leaf rollers and stem borers of cruciferous and other plants) THYRIDIDAE* TII & TIII with the SV group unisetose, and with L3 present; head with 01 usually caudad of ocelli 2 & 3 (Fig. 84)
27(26).	A9 with D1 closer to SD1 than to D2 (D1 and SD1 often on the same pinaculum), distance between D1 and D2 at least 3x distance between D1 and SD1 (Fig. 117)28 A9 with D1 more equidistant between SD1 and D2 or closer to D2 (D1 and SD1 never on the same pinaculum) (Fig. 113)30

28(27).	A8 with SD1 cephalad or cephalo-dorsad of the spiracle (Fig. 111); TI with SD2 almost directly dorsad of SD1 (Fig. 96; fruits, flowers, seeds of Umbelliferae)
	A8 with SD1 more nearly dorsad of the spiracle (Fig. 110); TI with SD2 caudad of SD1 (Fig. 95)
29(28).	Spiracles on A8 closer to the dorso-meson than spiracles on preceding abdominal segments, and caudo-projecting (Fig. 108)(Machlotica) GLYPHIPTERYGIDAE* Spiracles on A8 on the same level as spiracles on preceding abdominal segments and never caudo-projecting (Glaphyrinae, inquilines in nests of social Hymenoptera; Schoenobiinae, borers in stems and roots of grasses; Chrysauginae, feeders in pods of trumpet-creeper; and Crambinae, feeders in roots and stems of grasses)(part) PYRALIDAE
30(27).	Distance between ocelli 2 & 3 at least 3x the distance between ocelli 1 & 2 (Fig. 85); Al-7 with L1 caudad of the spiracle and as close or closer to the spiracle than it is to L2 (Fig. 105; fungus feeders)
31(30).	TI with SD2 almost dorsad of SD1 (Fig. 96); L2 & L3 on TII & TIII, and L2 on A1-8 may be absent (needle miners on pine)(Ocnerostoma) YPONOMEUTIDAE* TI with SD2 more nearly caudad of SD1 (Fig. 95); all L setae on TII, TIII, and A1-8 present (in stems, fruits, flowers of Compositae and honeysuckle, often pro- ducing galls)
32(25).	Crochets biserial or multiserial, or with a single row of well developed crochets plus many rows of smaller crochets or spines encircling the row of large crochets (Figs. 20, 30)

33(32).	(which may be continuous with the prothoracic shield; Fig. 26) (on clover and in roots of grasses, may construct silken galleries and webs)(Acrolophinae, a few Tineinae) TINEIDAE TI with the spiracle excluded from the L pinaculum or all pinacula indistinct
34(33).	A3 with L1 and L2 equally distant from the spiracle (Fig. 103); head with 01 closer to ocellus 3 than to ocellus 2 (Fig. 90); crochets biserial, with the inner row in an incomplete circle, sometimes resembling a mesopenellipse, and the outer row in a complete circle (Fig. 76)(part) PLUTELLINAE A3 with L1 closer to the spiracle than L2 (Fig. 105); head with 01 equidistant between ocelli 1 & 2, or closer to ocellus 2; crochets biserial or multiserial, usually in complete circles
35(34).	TII & TIII with the SV group bisetose; prothoracic shield indistinct; body irregularly patterned with brown spots (leaf folders and tiers)(Atteva) YPONOMEUTINAE TII & TIII with the SV group unisetose; prothoracic shield variable; body without distinct patterns36
36(35).	A8 with the SV group bisetose; antenna with 3rd segment no longer than 2nd segment (leaf folders)(Zelleria) ARGYRESTHIINAE A8 with the SV group unisetose, antenna with 3rd segment conspicuously longer than 2nd segment (borers in stems, twigs, flowers, fruits; and leaf folders)(part) YPONOMEUTINAE
37(32).	Crochets in transverse bands (Figs. 4,5) or reduced in number (less than 5 on a proleg), if arranged as a semicircle around the perifery of the proleg, then the gap free of crochets is greater than 1/2 the circumference of the projected circle

38(37).	TI with legs absent or conspicuously smaller than legs on TII & TIII; head prognathous, dorso-ventrally flattened, elongate; A3-6 with D1 or both D setae minute or missing; crochets on caudal half of proleg only (a "caudoseries"), 1 or 2 crochets on cephalic part may be present (leaf miners on grasses and sedges(Cosmiotes, Dicranactetes) ELACHISTIDAE: TI with legs never absent or reduced in size; head semi-prognathous or hypognathous, not extremely dorso-ventrally flattened; A3-6 with both D setae present and conspicuous; crochet arrangement variable
39(38).	Ocelli 3, 4, & 5 not equally spaced, distance between 4 & 5 at least 2x the diameter of 4, and distance between 3 & 4 usually equal to or less than the diameter of 4 (Fig. 88); TII & TIII with L1 equi- distant between L2 & L3, or L1 closer to L3; crochets in 2 transverse bands or a complete ellipse40 Ocelli 3, 4 & 5 more equally spaced, distance between all 3 ocelli less than 2x the diameter of 4 (Fig. 85); TII & TIII with L1 closer to L2 than to L3; crochet arrangement variable
40(39).	TI with SD2 almost dorsad of SD1 and usually as close to XD2 as to SD1 (Fig. 96); A9 with D1 closer to SD1 than to D2; D1 often on the same pinaculum with SD1 (Fig. 117; borers in roots, trunks, branches of trees and shrubs, or in stems and roots of herbaceous plants)
41(39).	A3 with L1 & L2 as far as from each other or farther than either is from the spiracle; L1 usually caudad of the spiracle (Fig. 104)

42(41).	Prolegs absent or rudimentary; crochets in a single transverse band with up to 12 crochets in a band (Fig. 4); ocellus 6 present; setae at distal end of tarsi not spatulate (construct flat oval cases from maple leaves)(Paraclemensia) INCURVARIIDAE Prolegs well developed; crochets in 2 transverse bands, with never more than 8 crochets in a band; ocellus 6 usually missing; setae at distal end of tarsi usually spatulate (early instars form serpentine mines, later instars are external skeletonizers)
43(41).	Abdominal spiracles and SD2 encircled by dark sclerotized spots; from 1 to 14 crochets arranged in a lateroseries (borers in stems of wheat and other grasses)
44(43).	AlO with crochets divided by a gap into 2 groups (Fig. 13); prolegs well developed on A3-6 & 10 (varied habits)(part) GELECHIIDAE AlO with crochets never divided by a gap into 2 groups; all prolegs rudimentary (construct portable cases and mine or feed externally on leaves, flowers, fruits, seeds(part) COLEOPHORIDAE
45(37).	TII & TIII with SV group bisetose

46(45).	Prothoracic shield including the L (prespiracular) group and the spiracle (Fig. 26); all 6 ocelli present and easily visible, and with ocelli 1-5 evenly spaced; thoracic legs large and heavily sclerotized (polyphagous feeders constructing portable bags)PSYCHIDAE Prothoracic shield excluding the L group and the spiracle which are often on their own well-sclerotized pinaculum; usually with at least 3, sometimes all 6 ocelli missing; if more than 3 are present, then 1 & 2 are conspicuously separated from 3, 4 & 5 (Fig. 85); thoracic legs not conspicuously large (feeders on dried animal and vegetable matter, sometimes constructing cases)
47(46).	A3 with L1 or L2 closer to the spiracle than L1 and L2 are to each other (Fig. 105), or distances equal (Fig. 103), or L2 absent
48(47)	With fewer than 5 ocelli present on each side of the head
49(48).	A3 with SD2 almost as large as SD1 and with L1 much closer to the spiracle than L2; body elongate and covered with minute spinules arranged in vertical rows; (scavengers, may construct silk galleries in fungus)(1 species in U.S., Oinophila v-flava) OINOPHILIDAE; A3 with SD2 much smaller than SD1 and with L1 & L2 more nearly equidistant from the spiracle (Fig. 103); body not fitting above description (mostly feeders in fungus or decaying wood, occasionally found in dried food products)(part, Nemapogoninae) TINEIDAE
50(48).	Ocelli 2 & 3 farther apart than ocelli 1 & 2, distance between 2 & 3 at least 3x distance between 1 & 2 (Fig. 85); A3 with L1 more caudad than ventrad of the spiracle (Fig. 105)(part, Nemapogoninae) TINEIDAE Ocelli 1,2, 3 more evenly spaced, distance between 2 & 3 less than 3x distance between 1 & 2; A3 with L1 ventrad or caudo-ventrad of the spiracle (Fig. 103)

	1
	1

51(50).	A3 with L2 absent; thoracic legs with the claw elongate, narrow, not recurved at tip; prolegs on A3 smaller and farther apart than those on A4-6 (leaf miners)(Bedellia) LYONETIIDAE A3 with L2 present; thoracic legs with the claw short and conspicuously recurved at tip; prolegs not as above (varied habits, may form tube-like shelters on foliage)(Plutellinae, part) YPONOMEUTIDAE
52(47).	Distance between coxae on TIII greater than 1.5x the width of the coxa base
53(52).	Fronto-clypeus open behind (Fig. 65); ocelli difficult to count but head appearing to have 2 dark "eyespots" on each side of the head (Fig. 66; leaf miners)
54(53).	A3 with D1 & D2 adjacent, distance between the two D1 setae at least 4x distance between D1 & D2 (Fig. 109; leaf miners)(Lampronia) INCURVARIIDAE* A3 with D1 & D2 farther apart, distance between the two D1 setae less than 4x distance between D1 & D2
55(54).	Ventral side of TI without darkened sclerotized spots (leaf miners of Cosmopterygidae and Momphidae, Cosmopteryx, Aeaea, etc.; and cosmopterygid borers in stems and seeds of cattail); may be impossible to key further)(synoptical key) 75-88 Ventral side of TI with darkened sclerotized spots(Coelopoeta) ELACHISTIDAE*
56(52).	Ocelli 2 & 3 farther apart than ocelli 1 & 2, distance between 2 & 3 at least 3x distance between 1 & 2 (Fig. 85)

37(30).	6 about as far from ocelli 3, 4, & 5 as it is from ocelli 1 & 2 (Fig. 75); A9 with D1 closer to SD1 than to D2 or equidistant between SD1 and D2; D1 and D2 never on the same pinaculum (leaf miners)
58(56).	Prolegs elongate, peglike, at least 3x longer than wide (Fig. 9)
59 (58)	Tarsi conspicuously slender and elongate (Fig. 94); head with setae Al, A2 & A3 forming an obtuse angle (Fig. 89); A9 with D1 usually missing, and with SD1 without the usual seta base or pinaculum, but with a sclerotized ring around the seta base (Fig. 92); SD2 absent on A9; (skeletonizers in rolled, folded, or webbed leaves)(part) GLYPHIPTERYGIDAE Tarsi not conspicuously slender and elongate; head with setae A1, A2 & A3 forming an acute angle (Fig. 86); A9 with D1 always present, and SD1 not modified as above; SD2 sometimes present on A9 (found on Persea)
60(24).	TI with L (prespiracular) group unisetose or bisetose61 TI with L (prespiracular) group trisetose63
61(60).	TII & TIII with SV group bisetose; A1-8 with deep folds below the spiracles; most setae on chalazae(Lactura) YPONOMEUTIDAE* TII & TIII with SV group unisetose; not fitting combination of other characters

62(61).	A8 with SDl cephalad of the spiracle (Fig. 111); A9 with Dl nearly equidistant between SDl & D2 (Fig. 115) (1 species, southeastern U.S.)
63(60).	Prothoracic shield including the L (prespiracular) group(Paraprays) GLYPHIPTERYGIDAE Prothoracic shield excluding L (prespiracular) group, which is often on its own well sclerotized pinaculum64
64(63).	Al-7 with L1 & L2 distinctly cephalad of the spiracle, usually farther cephalad than D1; body elongate
65(64).	AlO with 9 or fewer setae on each side below the transverse anal slit (including the proleg)(Symmoca) SYMMOCIDAE* AlO with 10 or more setae on each side below the transverse anal slit (including the proleg)(a few) GELECHIIDAE
66(64).	TII & TIII with L1 equidistant between L2 & L3, or closer to L3, or L3 absent; prothoracic shield usually elevated, bearing a roughened caudal area (cornicula); A8 with spiracle often distinctly near the caudal margin of the segment (mature larvae may exceed 50 mm.; borers in wood, roots, stems)(part) COSSIDAE TII & TIII with L1 closer to L2 than to L3; prothoracic shield never elevated or bearing cornicula; A8 with spiracle usually located near the center of the segment, or only slightly caudad of center
67(66).	AlO prolegs with crochets divided by a gap into two groups, forming a broken row (Fig. 13)

68(67).	A3-6 with crochets interrupted on mesal side by a lobe, forming a lateropenellipse; A8 with SD1 cephalad of the spiracle (Fig. 111)(Setiostoma) STENOMIDAE
	A3-6 with crochets usually in a complete circle, A8 with SDI more nearly dorsad of the spiracle (Fig. 110)
	(leaf rollers, leaf tiers, leaf miners, gall makers, or in stored products)(part) GELECHIIDAE
69(67).	A9 with L group trisetose, arranged in a nearly hori- zontal line and with SD1 closer to L group than to D1 (Fig. 114; primarily foliage feeders on oak)
	A9 with L group variable in number, but if more than 1 is present, they are never arranged in a horizontal line, SD1 variable in position
70(69)	A9 with D1 closer to SD1 than to D2 (Fig. 117), and with the D2 setae on a common mid-dorsal pinaculum or closer to each other than each is to its associated D1 (Fig. 112); (leaf rollers, leaf folders, borers in fruits, seeds, stems, a few webworms; most TORTRICIDAE, PHALONIIDAE, may be impossible to key further(synoptical key) 75-88
	A9 with D1 more nearly equidistant between SD1 and D2, or closer to D2 (Fig. 113), and with the D2 setae rarely on a common mid-dorsal pinaculum, or closer to D1 than they are to each other
71(70).	Anal comb present (many TORTRICIDAE, GELECHIIDAE)(synoptical key) 75-88 Anal comb absent
72(71).	A9 with 3-20 setae in the SV group (many OECOPHORIDAE, ETHMIIDAE, may be impossible to key further(synoptical key) 75-88
	A9 with not more than 2 setae in SV group

73(72).	AlO with more than 9 setae on each side below the transverse anal slit (including the proleg) many COSMOPTERYGIDAE, OECOPHORIDAE, ETHMIIDAE, GELECHIIDAE, may be impossible to key further
74(73).	Seta SDl on Al-8 with a sclerotized semi-circular ring encircling its base (Fig. 93); ocelli 3 & 4 almost touching, closer to each other than either is to nearby ocelli (Fig. 35)(most) BLASTOBASIDAE Without the above combination of characters: (many) COSMOPTERYGIDAE (C) (many) MOMPHIDAE (M) (many) WALSHIIDAE (W) (few) BLASTOBASIDAE (B) (many) OECOPHORIDAE (O) (very few) ETHMIIDAE (E) (many) GELECHIIDAE (G) (few) STENOMIDAE (S) (very few) TORTRICIDAE (T) (very few) PHALONIIDAE (P)* (May be impossible to key further)(synoptical key) 75-88
75.	(55, 70, 71, 72, 73, 74). Fronto-clypeus: a- extends all the way to the epicranial notch, or nearly (Fig. 79)
76.	Ocelli 4 & 5: a- farther apart than ocelli 3 & 4, distance between 4 & 5 at least 2x distance between 3 & 4 (Fig. 88)C

77.	Seta A2 on head: a- closer to A1 than to A3 (Fig. 89) b- equidistant, or nearly, between A1 & A3 (Fig. 91)	.C.M.W.B.O.E.G.S.T.P.
78.	Seta A3 on head: a- closer to A2 than to L1 (Fig. 84) b- equidistant between A2 & L1 (Fig. 91)	MGC.M.WO.E.GT.PC.MB.O.E.G.S.T.P.
79.	Distance between TIII coxae: a- greater than 1.5x the width of a coxa b- less than or equal to 1.5x the width of a coxa	.CWG
80.	A8 with seta SD1: a- more nearly dorsad than cephalad of spiracle (Fig. 110) b- more nearly cephalad than dorsad of the spiracle (Fig. 111)	.C.M.W.B.O.E.G.S.TCOG.S.T.P.
81.	A9 with seta D2: a- more nearly dorsad that caudal of D1 (Fig. 110) b- dorso-caudad of D1 (Fig. 114) c- more nearly caudad than dorsad of D1 (Fig. 115)	.C.M.WOG.S.T.P. .C.MB.O.ES.T
82.	A9 with L group: a- unisetose b- bisetose	.C.M.W

83.	A9 with L1 & L2: a- very close together, distance between L1 and next seta below (either L3 or SV1) at least 3x distance between L1 & L2 (Fig. 117) b- farther apart, distance between L1 and next seta below (L3 or SV1) equal to or greater than distance between L1 & L2 (Fig. 115)	.C.MO.E.GT.P.
84.	A9 with L setae (if less than 3, go to 85): a- all 3 close together, distance between any 2 L not more than 1/3 distance between any L and SD or any L and SV (Fig. 113) b- all 3 farther apart, distance between any 2 L greater than 1/3 distance between any L and SD	
	or any L and SV (Fig. 115)	.C.M.W.B.O.E.G.S.T
85.	Anal comb: a- presentb- absent	
86.	Crochets: a- uniordinal b- biordinal or triordinal	.C.M.W.B.OGT.PC.MB.O.E.G.S.T.P.
87.	Crochets on A3-6 arranged: a- in transverse bands b- in a lateropenellipse c- in a mesopenellipse d- in a complete circle	.C
88.	Spiracles: a- circular b- elliptical	

89(23).	TI with L group unisetose or bisetose90 TI with L group trisetose92
90(89).	A5 with prolegs absent or conspicuously smaller than A6 prolegs (loopers; exposed feeders on foliage of deciduous plants)(part) GEOMETRIDAE A5 prolegs as large as A6 prolegs91
91(90).	Transversely striped and spotted black by large contrasting pinacula or chalazae; A8 gibbose (humped); no conspicuous granules or spinules present on cuticle (external feeders on grape, virginia creeper, etc.)
92(89).	A3 with L1 or L2 closer to the spiracle than to each other, or distances equal (Figs. 103, 105); crochets uniordinal(many Plutellinae) YPOMONEUTIDAE A3 with L1 & L2 closer to each other than either is to the spiracle; crochets uniordinal, biordinal, or triordinal
93(92).	TII & TIII with SV group unisetose; prolegs usually not conspicuously elongate; cuticle without spinules resembling secondary setae; (feeders in a web in flowers and on leaves of Hydrophyllaceae and Boraginaceae)

94(21).	A2 and A7 with pairs of ventral lobes resembling prolegs but lacking crochets; head retracted into prothorax; setae long, hairlike, in dense tufts (foliage feeders on shrubs and trees)MEGALOPYGIDAE A2 and A7 without conspicuous ventral lobes; head retracted or not; secondary setae variable95
95(94).	Prolegs and crochets absent on A3 (mature larvae not more than 20 mm., feeders on foliage of various hosts, constructing a weak shelter formed from a folded leaf or webbed terminals)
96(95).	Crochets uniordinal
97(96).	Crochets in a heteroideous mesoseries (Fig. 56)98 Crochets in a homoideous mesoseries (Fig. 11)100
98(97).	TII & TIII with at least 4 verrucae between coxa and mid-dorsal line (SD and D verrucae separated, Fig. 99), (foliage feeders on wide variety of plants, favoring herbaceous plants)
99(98).	A7 with verruca L1 the same distance or only slightly farther from spiracle as the same verruca on A1-6; distance from spiracle to verruca on A1-7 usually not more than the diameter of the verruca (Fig. 101), (mostly feeders on grasses, lichens, oleander)CTENUCHIDAE A7 with verruca L1 distinctly farther from spiracle than L1 on A1-6; distance between spiracle and L1 on A7 usually greater than the diameter of the verruca (Fig. 102); known hosts include Beaucarnea, bluebell)
100(97).	Spiracles small, more nearly circular than elliptical101 Spiracles large, elliptical102

101(100).	A3 with verricules, 4 per side (D, SD, L, SV), and usually with more than 12 setae on a verricule; crochets in a mesoseries on a lobate planta (Fig. 122); skeletonizers on deciduous plants including viriginia-creeper and grape)
102(100).	Dorsal meson of A7 (and sometimes A6) with an eversible gland; secondary setae usually in tufts of different lengths and density on verrucae; often with middorsal pencils on A1-4, (feeders on deciduous trees, many are forest defoliators)LYMANTRIIDAE Dorsal meson never with an eversible gland; secondary setae variable, if on verrucae, then tufts usually of similar length; mid-dorsal pencils never present on A1-4
103(102).	Labral notch shallow, extending less than 1/2 the distance to the base of the labrum (Fig. 80); spinules never conspicuous on cuticle; (feeders on plantain, dandelion)(Holomelina) ARCTIIDAE Labral notch usually deeper, extending at least 1/2 the distance to the hase of the labrum (Fig. 81); spinules usually conspicuous on cuticle; (general feeders on deciduous and evergreen foliage)
104(96).	TI with a mid-dorsal osmeterium (osmeteria may be inverted into prothorax: look for elongated transverse mid-dorsal slit on anterior part of thorax)
105(104).	A8 with at least 1 mid-dorsal horn, scolus, chalaza, tubercle, or verruca

106(105).	A7 (and usually A1-6) with a mid-dorsal scolus; a ventral prothoracic gland usually present; crochets usually triordinal (feeders on wide variety of herbaceous and woody plants)(most) NYMPHALINAE A7 (and A1-6) without a mid-dorsal scolus; ventral prothoracic gland absent; crochets biordinal
107(106).	A9 with a mid-dorsal scolus or tubercle(Hemileucinae) SATURNIIDAE A9 without a mid-dorsal scolus or tubercle(Saturniinae) SATURNIIDAE
108(105).	Scoli present; verrucae, verricules, and pinacula usually absent
109(108).	A9 with a mid-dorsal scolus; dorsal scoli on TII (Anisota, Dryocampa) or on TII & TIII (other genera) much longer than scoli on abdominal segments (feeders on deciduous trees and shrubs)(Citheroniinae) SATURNIIDAE A9 without a mid-dorsal scolus; dorsal scoli on TII & TIII if present, not conspicuously longer than abdominal scoli (feeders on herbaceous & deciduous plants)
110(108).	Row of crochets interrupted or reduced in size near center by a lobe (Fig. 10); A3 with 2 distinct verrucae(Riodininae) LYCAENIDAE* Row of crochets not interrupted, no lobe present on prolegs; A3 with more than 2 verrucae which may resemble pinacula or may be obscured by numerous long secondary setae

111(22).	Planta cylindrical and elongate (Figs. 37, 120); mature larvae small, rarely over 20 mm
112(111).	Thoracic segments with SV setae always conspicuous and larger than surrounding secondary setae, and bisetose on TII & TIII; (stem borers or foliage feeders within a webbed mass)(part) PTEROPHORIDAE Thoracic segments either with SV setae obscured by numerous secondary setae of same length, or if SV setae are conspicuous then they are unisetose on TII & TIII (Fig. 36); (fruits, stems, berries of apple, hawthorne, Aronia, or seed pods of Croton)
113(111).	Crochets in a complete circle, or with 1 or 2 small gaps in an otherwise continuous row (gaps always less than 1/3 the circumference of the projected circle)114 Crochets in a mesoseries or a mesoseries plus a lateroseries; if a mesoseries resembles a mesopenellipse, then the gap free of crochets is at least 1/3 the circumference of the projected circle (a few widely spaced isolated crochets may be present on the lateral side of the proleg)
114(113).	Head distinctly larger than prothorax in profile (Fig. 49); anal comb usually present (Fig. 48) (mature larvae usually less than 40 mm., external feeders on grasses, herbacious plants, usually constructing shelters)

113(113).	2x larger than the others, and with 4, 5 & 6 spaced farther apart (Fig. 51); anal plate usually bifurcate at tip (Fig. 50); mandibles usually thick, quadrate, lacking teeth, but with ventral sharp cutting edges (external feeders on grasses)(Satyrinae) NYMPHALIDAE Ocelli not arranged as above, and ocellus 3 not conspicuously larger than others; anal plate not bifurcate; mandibles not modified as above
116(115).	Crochets uniordinal
117(116).	Labral notch deep, v-shaped, and usually continued as a groove to the base of the labrum (Fig. 82); caudal prolegs usually smaller than ventral prolegs, and may be without crochets(Melalopha, Ichthyura, Datana) NOTODONTIDAE Labral notch u-shaped with parallel sides, and never continued as a groove to the base of the labrum (Fig. 81); caudal prolegs never conspicuously smaller than ventral prolegs, and always with crochets(many Acronyctinae, Hadeniinae) NOCTUIDAE
118(116).	Long fleshy filaments or osmeteria present (osmeteria may be inverted into prothorax: look for elongated transverse mid-dorsal slit on anterior part of prothorax)
119(118).	TII with at least one pair of long fleshy filaments; osmeterium present or absent; a lateroseries of crochets never present

120(119).	Larva with transverse black and green stripes on all segments and head; osmeterium absent; lateral and subventral lobes or filaments on abdominal segments absent; all pairs of filaments dorsal or subdorsal in position (feeders on milkweed, oleander, fig, and other plants)(Danainae) NYMPHALIDAE Larvae black with dorsal rows of white spots; osmeterium present; with long lateral filaments on thoracic segments and smaller lateral and subventral filaments on abdominal segments (feeders on pipe-vine)(Battus) PAPILIONIDAE
121(119).	Thoracic and abdominal segments with pinacula in the positions of primary setae with each pinaculum bearing 4 to 6 short setae; larva black with dorsal and subdorsal rows of white spots (feeders on violet, stonecrop, Sempervivum, occurs in Alaska and mountainous regions of western North America)(Parnassius) PAPILIONIDAE All segments lacking pinacula; coloration variable, may be aposematic or cryptic (wide variety of hosts, wide distribution(Papilio) PAPILIONIDAE
122(118).	A8 with at least one mid-dorsal horn, chalaza, tubercle, or scar
123(122).	Abdominal segments divided into 6 to 8 annulets (folds) (Figs. 49, 54); distances between bases of prolegs usually no greater than length of each meso- series of crochets (hornworms, variety of hosts)(part) SPHINGIDAE Segments not divided into 6 to 8 annulets; distance between bases of prolegs greater than length of each mesoseries (domestic silkworms)BOMBYCIDAE*
124(122).	Segments divided into 6 to 8 annulets (Figs. 49, 54)125 Segments not divided into 6 to 8 annulets126

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125(124).	Head equal to or larger than prothorax in profile; anal comb usually present; most setae on chalazae (Fig. 53; many hosts but favor Cruciferae and Leguminoseae)
126(124).	Secondary setae short, stiff, bristle-like; if primaries are present, then secondaries always shorter than primaries
127(126).	Row of crochets reduced in size or interrupted near center by a conspicuous fleshy pad (Fig. 10); head smaller than prothorax and retractible (external feeders on foliage, myrmecophilous, or predaceous on Homoptera)LYCAENIDAE Row of crochets not interrupted by a fleshy pad; head not retractible (1 widespread species, Libythea bachmani Kirtland, hosts are hackberry, Symphoricarpos occidentalis)LIBYTHEIDAE*
128 (126, 110).	A2-8 and sometimes TII & TIII with mid-dorsal tufts of dark setae (feeders on deciduous trees and shrubs; only 2 genera in U.S., Ocleclostera, Apatelodes)

129(22).	the SV group) extensively developed, forming the greater part of the proleg; planta usually modified as a lobe at the distal end of the basal part, never cylindrical or rudimentary (Figs. 121, 122, 123); crochets usually in a mesoseries (external-feeding and cutworm-type Macrolepidoptera)
130(129).	Ll (seta or pinaculum) farther from the spiracle on A7 than on A1-6, or with Ll on A1-6 caudad of the spiracle and Ll on A7 caudo-ventrad of the spiracle (Fig. 106); Tl with SD1 usually thinner and shorter than SD2
131(130).	TI with SD1 & SD2 very close together and excluded from the prothoracic shield (Fig. 97) (SD setae may be on their own pinaculum which may also have a few extra setae)
132(131).	AlO with 1 to 4 extra setae near the anterior margin of the segment and located approximately as high as as SD1 on A9 (Fig. 116); labral notch usually acute, v-shaped, and continued as a groove to the base of the labrum (Fig. 82)

133(132).	Caudal prolegs often modified into caudo-projecting
	processes, or smaller than ventral prolegs
	(reliable charcters separating dioptids and noto-
	dontids are lacking, check descriptions); family of
	widespread distribution and many common species,
	some are forest defoliators(part) NOTODONTIDAE
	Caudal prolegs never modified as above, and rarely
	smaller than ventral prolegs; distribution confined
	to California and southwestern U.S., 1 common
	species, the California oakworm (Phryganidia
	californica), and a few (?) rare speciesDIOPTIDAE*

- 134(131). Crochets in a homoideous mesoseries; TII & TIII with L1,
 L2, & L3 on a single large pinaculum (Fig. 98)
 (Lithosiinae, Hypoprepia) ARCTIIDAE
 Crochets in a heteroideous mesoseries; TII & TIII with
 L1 & L2 on a single pinaculum, and L3 on a separate
 weak pinaculum(Utetheisa) ARCTIIDAE

- 137(136). TI with SD1 & SD2 excluded from the prothoracic shield; A3 with more than 4 setae between spiracle and proleg (feeders on deciduous foliage, may construct loose folded leaf shelter)......THYATIRIDAE*

 TI with SD & SD2 included on the prothoracic shield;
 A3 with only 4 setae between spiracle and proleg
 (L1, L2, L3, L4) (feeders on Vibernum, honeysuckle; construct webs in early instars).....EPIPLEMIDAE

138(129).	circle (Fig. 20; borers in roots, stalks, stems)
139(138).	Ocelli 4 & 5 farther apart than ocelli 3 & 4 (Fig. 88), distance between 4 & 5 usually much greater than the diameter of ocellus 4
140(139).	TI with L group bisetose; TII & TIII with L1 closer to L2 than to L3 (4 species in southern U.S., late instars construct portable cases of leaves and silk)
141(139).	Crochets uniordinal
142(141).	TI with L group bisetose; A8 with the spiracle closer to the dorsal midline than the spiracles on A1-7 (Fig. 108); submentum with paired flaps or protuberances on caudal half (may be difficult to see) (within fruits of hawthorn, barberry, wintergreen)
143(142).	TII & TIII with SV group bisetose; prolegs often longer than wide (Fig. 120)(part) PTEROPHORIDAE TII & TIII with SV group unisetose; proleg usually shorter than wide; (miners and webbers on Sphacele, Lantana)(Anoncia) COSMOPTERYGIDAE*

144(141).	SD1 on A1-7 with its base set in a conspicuous pale
	area, ringed with dark brown (Fig. 92); SD1 on A8
	usually thinner than SD1 on A1-7; prothoracic shield,
	D, SD, and L pinacula often with secondary setae
	(miners in grasses and webspinners in flower heads of
	thistle)SCYTHRIDAE
	SD1 on Al-7 with its base not modified as above, but
	sometimes on a well sclerotized pinaculum; SD1 on A8
	not thinner than SD1 on Al-7; prothoracic shield, D,
	SD, and L pinacula never with secondary setae
	(nort) FTHMIIDAE

D. DIAGNOSTICS OF THE SUPERFAMILIES AND TAXONOMIC OBSERVATIONS

Most of the problems with older suborder groupings of the Lepidoptera (Comstock 1892, Tillyard 1926, Meyrick 1895) have been reviewed by Hinton (1946) and Common (1970, 1975). The new system recognizes 5 suborders, based on all stages of development: the Zeugloptera (Micropterygoidea), Dacnonypha (Eriocranioidea), Monotrysia (Nepticuloidea, Incurvarioidea), Exoporia (Hepialoidea), and Ditrysia (the rest of the Lepidoptera, 16 superfamilies). At the family and subfamily levels, arrangements differing from those found in Forbes (1923), McDunnough (1938, 1939), and Peterson (1956) are based on more recent literature. These changes are discussed briefly.

Suborder Zeugloptera

Based on larval characters some workers believe the Micropterygidae should be excluded from the Lepidoptera. Hinton (1958) listed 12 attributes of micropterygid larvae which separate them from all other Lepidoptera. The most important are: a) the head lacks ecdysial lines, b) a distinct spinneret is absent, c) thoracic legs with fused coxa, trochanter, and femur, d) abdominal prolegs lack muscles, e) functional metathoracic spiracles are present, f) the chaetotaxy and structure of the setae are quite different from the rest of Lepidoptera. The Zeugloptera are thought to be more archaic than the Lepidoptera and the Trichoptera, and if they are to be included in the Lepidoptera, the Trichoptera must also be included (Hinton, 1946). However, on the basis of pupal and

adult morphology, there are no grounds for separating the Zeugloptera from the Lepidoptera (Common 1975).

Suborder Dacnonypha

The Dacnonypha was proposed by Hinton (1946) to include the Eriocraniidae, Neopseustidae (exotic), and the Mnesarchaeidae (exotic). Characters distinguishing them from all other Lepidoptera are: larva with ecdysial lines joining anterior margin of the head behind the antennae such that the antennae and seta Al are included in the adfrontal area, and body apodous (without legs & prolegs); pupa exarate (with free and movable appendages) and decticous (with functional mandibles); adult with homoneurous venation, aculei present, wings coupled with fibula, and females with a single genital opening (monotrysian).

Suborder Monotrysia

The Monotrysia was first proposed by Borner (1939) to include all Lepidoptera with a single genital opening: the Micropterygidae, all Homoneura, and a few Heteroneura. Hinton included the Hepialoidea, Nepticuloidea, and the Incurvarioidea in the suborder, and excluded the Zeugloptera and Dacnonypha. Common (1975) pointed out important differences between the 3 superfamilies included in the suborder by Hinton and objected to their close association. He proposed a new

suborder for the Hepialoidea based mainly on the recent discovery of 2 genital openings in the females.

Distinguishing characters of the Monotrysia are: <u>larva</u> with ecdysial lines joining anterior margins of the head excluding the antennae and Al from the adfrontal areas, and body apodous or with both legs and prolegs; <u>pupa</u> incomplete obtect, and adecticous; <u>adult</u> with heteroneurous or reduced (Nepticuloidea) venation, aculei present, wings coupled with a frenulum (Incurvarioidea) or with a frenulum & jugum (Nepticuloidea), females with a single genital opening.

Incurvarioidea

Forbes (1923) was the first to group the Prodoxinae, Adelinae, and Incurvariinae in one family, the Incurvariidae. Based on adult characters Davis (1967) raised the Adelinae to family, and left the Incurvariinae and Prodoxinae in the Incurvariidae. He thought of the Prodoxinae as a New World specialization derived from some primitive extinct incurvariine. Common (1970) raised the Prodoxinae to family rank and left the Incurvariinae and Adelinae as subfamilies of the Incurvariidae.

On the basis of larval characters, Common's arrangement is supported here. The Incurvariinae and Adelinae share many larval characters. On TI both have XD1 and D1 close together and minute, while SD1, XD2, and SD2 are close together and large. Both subfamilies have large lightly sclerotized dorsal and ventral areas on the thoracic segments. On A1-8, L1 and L2 are caudad of the spiracle with L2 closer to the spiracle than L1. Most genera have at least rudimentary prolegs with corchets in transverse bands. Both groups show a preference for a leaf mining or

flower boring habit, and many construct cases. The Prodoxinae in contrast, usually have most primary setae minute. On TI SD2 is dorsad of SD1 and both are closer together than either is to XD2. The Prodoxinae also lack large dorsal and ventral sclerotized areas on TI-III. Thoracic legs are very small or absent, and both crochets and prolegs are entirely lacking. Lastly, the Prodoxinae are restricted to feeding in Yucca, and as Davis suggests, are probably closely associated with the evolution of that group.

Nepticuloidea

The Opostegidae are specialized stem miners whose taxonomic affinities have been in dispute. Forbes (1923) placed them in the Tineoidea near the Lyonetiidae and Gracillariidae. Common (1970) placed them in the Nepticuloidea, probably based on the presence of a single genital opening in the females. Heinrich (1918) pointed out unique morphological features of Opostega larval head capsules, and argued that many trends carried to extreme in Opostega are also found in Nepticula, Ectoedemia (Nepticulidae), Tischeria (Tischeriidae), Leucopterygidae, (=Lyonetiidae) and that Brenthia, Bedellia (Lyonetiidae), Gracillariidae, and Bucculatrigidae (=Lyonetiidae in part) represent morphological trends in other directions.

Suborder Exoporia

This suborder consists only of the Hepialoidea (Common 1975).

Distinguishing characters of the suborder are: <u>larva</u> with antennae and Al excluded from the adfrontal areas with thoracic legs and prolegs on A3-6 & 10; <u>pupa</u> advanced incomplete obtect and adecticous; <u>adult</u> with homoneurous venation, aculei present, wings coupled with a jugum and female genitalia intermediate between monotrysian and ditrysian (exoporian).

In addition, the Hepialoidea can be distinguished from the Ditrysia by chaetotaxy. The XD, D, SD, and L setae on TI are on a single large prothoracic shield; microscopic setae MD1, MSD1 & MSD2 on TII & TIII and MV3 on A1-8 are nearly as large as the tactile primary setae; the SD group on A9 is bisetose, and crochets are biserial or multiserial.

Suborder Ditrysia

Distinguishing characters of the Ditrysia are: <u>larva</u> with antennae and Al excluded from the adfrontal areas, thoracic legs and prolegs (on A3-6 & 10) usually present; <u>pupa</u> usually complete obtect (with appendages glued to the body) and adecticous; <u>adult</u> with heteroneurous venation, aculei restricted or absent, wings coupled with a frenulum or a frenulum plus a reduced jugum (more primitive groups), and females with 2 genital openings (ditrysian).

Features of Ditrysian chaetotaxy which distinguish them from the Exoporia are: the L group on TI usually excluded from the prothoracic

shield, all microscopic setae minute, the SD group on A9 usually unisetose, and secondary setae often present. Crochets are usually uniserial.

Tineoidea

Important characters in recognizing a tineoid larva are: Ocelli reduced in numbers or in 2 distinct groups with 1 & 2 separated from 3, 4, & 5. TI with the L (prespiracular) pinaculum encircling the spiracle, and sometimes joined to the prothoracic shield. L1 & L2 on A1-8 far apart, SD2 on A1-8 nearly as large as SD1 (Tineidae, Oinophilidae). Crochets sometimes multiserial (Acrolophinae). The relationships among the 6 families in North America are unclear. The Tineidae seem to be the most generalized group. The Gracillariidae, Ochsenheimeriidae, Oinophilidae, and Psychidae, are relatively specialized groups which may have been derived from tineid ancestors (Brock 1971). The Lyonetiidae are thought to be a heterogeneous collection of genera (Forbes 1923, Common 1970).

MacKay (1972) put <u>Bucculatrix</u> in a separate family, the Bucculatrigidae, on the grounds that L1 & L2 on A1-8 are far apart, whereas they are closer together in other lyonetiid genera. This character she considered "of family status". Examination of other lyonetiid genera showed this character to be more variable than she implied. <u>Bucculatrix</u> larvae are admittedly unusual in appearance, but they have few consistant distinguishing characters. In <u>Lyonetia</u> L1 and L2 are as far apart as in <u>Bucculatrix</u>, and in <u>Bedellia</u> L2 is missing. Bucculatrix may merely represent an extreme development of a

specialized larval type. According to Forbes (1923) there are no important differences between adults of <u>Bucculatrix</u> and other lyonetiid genera.

Hinton (1955b, 1956) recognized four subfamilies of Tineidae, and his arrangement is followed here: Acrolophinae (mainly West Indian), Scardiinae (mainly Palearctic), Nemapogoninae (cosmopolitan), and Tineinae (cosmopolitan). Many authors, including Peterson (1956) and McDunnough (1936) gave the Acrolophinae family status.

Yponomeutoidea

That the Yponomeutoidea is a heterogeneous collection of families and genera has been admitted by nearly everybody and is supported by examination of the larvae. Of the 6 families occuring in North America (no larvae of Douglasiidae were available), only the Sesiidae and Glyphipterygidae are reasonably consistant in their characters. Too few larvae of Epermeniidae and Heliodinidae were available for assessment of their relationship to other groups in the superfamily.

Consequently, larvae of the Yponomeutoidea are difficult to characterize. In many Argyresthiinae, Sesiidae, Epermeniidae, and Heliodinidae, SD2 is dorsad of SD1 on TI. The L group on TI is trisetose in most families but bisetose in the Epermeniidae. L1 & L2 are far apart on A1-8 in the Plutellinae, Yponomeutinae, and Epermeniidae, but close together in other groups. Elongate planta and tarsi are found in some Plutellinae and most Glyphipterygidae. Crochets are usually uniserial but are multiserial in some Plutellinae and Yponomeutinae.

The taxonomic affinities among genera of the Yponomeutoidae are confusing. The treatment in the key follows that of Forbes (1923) and Hodges (1971) who recognize 3 subfamilies: the Yponomeutinae consisting of the American genera Yponomeuta, Atteva, Lactura, Swammerdamia, Podiasa, Ocnerostoma, Xyrosaris, Eucatagma, Orinympha; the Plutellinae consisting of Plutella, Prays, Acrolepia, Plinioca, Euceratia, Abebaea, Trachoma, Harpipteryx, Cerostoma, Melitinympha; and the Argyresthiinae consisting of Argyresthia and Zelleria. Examination of available larvae 'suggested a different grouping of these genera. Atteva, Swammerdamia, Yponomeuta, and Zelleria formed one group with many shared characters, and Abebaea, Cerostoma, Acrolepia, and Plutella formed another. Argyresthia and Zelleria (the only 2 genera of Argyresthiinae) shared few characters, but Argyresthia and Acrolepia shared many. Prays shared a greater number of characters with Atteva, Swammerdamia, etc., than it did with Abebaea, Cerostoma, etc., but did not fit well into either group. Lactura shared very few characters with any of the above genera, and does not resemble any other yponomeutoid.

Castnioidea

The Castnioidea is a primitive tropical & subtropical group often thought to be ancestors of the Papilionoidea (Ehrlich 1958). Others associate them with the Tortricoidea and Cossoidea based on adult characters (Brock 1971). Larvae of the Castnioidea can be distinguished from the similar Cossoidea by their possession of irregular rows of spines rather than true crochets as in the Cossoidea.

These spines may be an intermediate condition in the development of crochets.

Cossoidea

The Cossoidea is usually regarded as one of the more primitive Ditrysian superfamilies. The venation of the adults suggests a relationship to some of the more primitive Tineoidea, but larval chaetotaxy and male genitalia suggest a relationship to the Tortricoidea (Common 1970). Important characters in recognizing a cossoid are: Ocelli 1-4 evenly spaced in a semi-circle with 5 conspicuously separated from 1-4. TI with the L group trisetose. Al-8 with L1 & L2 close together below the spiracle. Crochets are uni-, bi-, or triordinal in a transverse ellipse sometimes broken at either end. A cossoid larva sometimes has features associated with the wood-boring habit, including roughened dorsal areas (cornicula) on the prothoracic shield, and large caudo-projecting spiracles on A8.

Tortricoidea

Important characters in recognizing a tortricoid larva are: Anal comb often present on AlO. A8 with SD1 usually cephalad of the spiracle. A9 with D2 usually dorsad or dorso-caudad of D1 and with the D2 setae often on a single pinaculum closer to each other than either is to its associated D1. Prespiracular group on T1 trisetose. L1 & L2 on Al-8 close together below the spiracle. Crochets uniordinal, biordinal, or triordinal, and arranged in a complete circle.

Although larvae of the Tortricoidea are usually easy to recognize, groups within the superfamily are barely separable. Neither MacKay (1959, 1962) nor Powell (1964) could find larval characters that would separate the tortricines, olethreutines, sparganothines, and phaloniids. Larval characters given in Forbes (1923) as points of distinction between the Tortricidae and Phaloniidae were: setae L1 & L2 (iv & v) on Al-8 obliquely or vertically placed in the Tortricidae but forming a horizontal line in the Phaloniidae; the SV group on A7 bisetose or trisetose in the Tortricidae, but unisetose in the Phaloniidae; and crochets multiordinal in Tortricidae, but uniordinal in the Phaloniidae. None of these differences were found to hold in the specimens examined. MacKay (1959) included two phaloniid genera, Phalonia and Hysterosia in her key to olethreutine genera because of their similarity. Possibly the Phaloniidae are not deserving of family rank.

Gelechioidea

Important characters in recognizing a gelechioid larva are: Head usually semi-prognathous (prognathous in some Elachistidae). TI with prespiracular group usually trisetose. Al-8 with L1 & L2 close together. A8 with SD1 usually dorsad of the spiracle. Crochets uniordinal, biordinal or triordinal, usually arranged in a complete circle, but may be in a mesopenellipse (some Ethmiidae, Gelechiidae, Blastodacnidae), a lateropenellipse (some Stenomidae), or in transverse bands (most Coleophoridae, some Gelechiidae). Anal comb present only in some Gelechiidae. Secondary setae sometimes present on the

prothoracic shield and other pinacula (Scythridae), as extra setae in the SV group on abdominal segments (many Oecophoridae, Gelechiidae, most Ethmiidae), or as numerous tiny setae evenly distributed over the cuticle (Blastodacnidae).

Definitions and limits for many gelechioid families are far from settled. Works relied on for clarification were those of Hodges (1962, 1964, 1966, 1969, 1971, 1974), Duckworth (1964, 1973), and Powell (1973).

The Cosmopterygidae, Momphidae, and Walshiidae cannot be separated on the basis of larval characters. Specialized larval types were found only among a few leaf mining genera (Cosmopteryx, Aeaea, Perimede).

Among other genera, few meaningful statements can be made about possible affinities. In the Momphidae, Batrachedra, Mompha,

Chrysoclista, and Cystioecetes are similar larval types and share some characters, while Homaledra is very different and superficially resembles some Yponomeutidae.

The Coleophoridae and Elachistidae appear to be specialized groups and are among the easiest gelechioids to recognize. The Coleophoridae construct portable cases and feed either as leaf miners or externally on foliage. Morphological specializations were consistant in the species examined. The Elachistidae are miners, mostly in grasses and sedges and share consistant morphological specializations associated with the leaf mining habit. Of the adults, Braun (1948) says they have a "common aspect which assists in placing them in the family."

Although the erection of the Blastodacnidae is relatively recent,

I have been unable to find its original author or definition. Hodges

(1962) referred to "Blastodacna and its allies" as a well marked group within the Momphidae but failed to mention which other genera are "allies". Clarke (1962, 1964, 1965) also failed to explain the source of the name. No earlier references to the family were found.

Blastodacnid larvae are unique gelechioids in their possession of numerous small secondary setae scattered over the cuticle. This single character separates blastodacnid larvae from other closely related groups (Walshiidae, Momphidae).

Five blastobasid genera were available, (Blastobasis, Eubolepia, Valentinia, Holocera, Zenodochum), and they shared the following combination of characters: presence of a sclerotized semi-circular ring around SD1 on Al-7, (Fig. 93) SD1 on A8 thin and hairlike, ocelli 3 and 4 nearly touching and closer together than adjacent ocelli (Fig. 35), the L group on A9 trisetose with L1 & L2 on a common pinaculum (Fig. 34), A9 with SD1 pinaculum smaller than the other pinacula. The similarities among these genera suggest that Blastobasidae is a valid family.

MacKay's (1972) contention that the Scythridae are nothing more than a specialized group in the Blastobasidae is supported by larval characters. The presence of the sclerotized ring around SDI on Al-7, the thin, hair-like SDI on A8, and the relative positions of the L setae on A9 are all characters which most scythrids share with blastobasids. Powell(1976) agreed with MacKay, but stressed the inconsistancy of the characters.

Many Oecophoridae, Ethmiidae and Stenomidae cannot be separated by larval characters. Most of the characters by which ethmiid larvae have

been distinguished from oecophorids by past authors are unreliable.

Powell (1973) retained the family level for ethmiids "merely for convenience" and suggested they will be permanently placed in the Oecophoridae when the world fauna is better known. Some stenomid genera have unique sets of larval characters (Antaeotrichia, Setiostoma) but other genera may be impossible to distinguish from some Oecophoridae.

Perhaps the Stenomidae should also be included in the Oecophoridae.

A single symmocid, Symmoca signatella H.-S., was examined.

Although Symmoca is unusual in its appearance, several gelechiids

(Stoeberhinus, Autosticha) were very similar and shared with Symmoca a modified seta base around SDl on Al-8 (Fig. 92). Common (1970) is possibly correct in placing Symmoca in the Gelechiidae.

Copromorphoidea

Meyrick (1928) was the first to place the Carposinidae, Alucitidae and Copromorphidae (palearctic) in a single superfamily, the Copromorphoidea. Although this arrangement has been followed by most recent authors (Common 1970, Munroe 1972, Hodges 1971), the systematic position of the 3 families is in doubt. On larval characters there is little to support the close association of at least the Carposinidae and Alucitidae. Other than a bisetose L group on TI the families bear little resemblence to each other in terms of separation from other superfamilies.

Pyraloidea

Important characters in recognizing a pyraloid larva are: Head never prognathous or highly modified for leaf-mining. Prespiracular group bisetose (trisetose in Pterophoridae). The SV groups unisetose on TII & TIII (bisetose in Thyrididae). L1 & L2 on A1-8 close together and below the spiracle. SD1 on A8 above the spiracle (cephalad of the spiracle in Hyblaeidae). Biordinal or triordinal crochets arranged in a complete circle (usually a mesopenellipse in Pterophoridae). Secondary setae absent (present in Pterophoridae).

Both Common (1970) and Munroe (1972) stressed important differences among the families of the Pyraloidea: "The presence of tympanic organs (in the adults) sharply divides the Pyralidae from the other families, and neither the Pterophoridae nor the Thyrididae regularly possess any of the main characters that distinguish the Pyralidae from other lepidopterous families" (Munroe, 1972). Brock (1971) excluded the Hyblaeidae (a small mostly tropical group) from the Pyraloidea. Other authors have placed them in the Noctuidae. The Pterophoridae have certain characters in common with the pyralidae but due to their unusual adult morphology, have been placed in a separate superfamily (Brock 1971, Common 1970). Only the Thyrididae seem to be reasonably closely related to the Pyralidae.

A superfamily rank for the Pterophoridae is partially supported by larval characters. Their possession of numerous secondary setae on primary pinacula separates them from all other Pyraloidea. Most Pterophoridae have elongate prolegs with uniordinal crochets arranged in a mesopenellipse. Also, in pterophorids where primary setae are not

obscured by the secondaries the L groups on TI and A9 are trisetose. In contrast, the Hyblaeidae, Pyralidae, and Thyrididae have the prespiracular group on TI bisetose, have biordinal or triordinal crochets arranged in a complete circle, and commonly have 1 or 2 L setae missing on A9.

Zygaenoidea

The Zygaenoidea is a collection of specialized larval types. There is a superficial resemblance between the Zygaenidae, Megalopygidae, and the Limacodidae, but each family has its unique set of larval characters. The Megalopygidae and Zygaenidae both have tufts of setae on verricules, a similar arrangement of ocelli, retractile heads, and tubercles near the abdominal spiracles. But they differ in arrangement and number of verricules. The Zygaenidae have lobate plantae resembling the type found in Noctuoidea, whereas the Megalopygidae have plantae and crochet rows divided by a deep fold. The Megalopygidae have pairs of ventral lobes on A2 and A7 which resemble extra pairs of prolegs. The Limacodidae have retractile heads, but otherwise do not resemble the other families. In place of verricules, they often have urticating scoli. In place of prolegs and crochets, they have suckerlike discs. Epipyropid larvae are specialized ectoparasitoids of planthoppers (Fulgoroidea), and do not resemble any other Zygaenoidea.

The Zygaenoidea may be nothing more than a heterogeneous collection of primitive ditrysian groups which developed the external feeding habit early (Forbes 1923). Brock (1971) pointed out the primitive nature of

the adult zygaenoid morphology and placed the Limacodidae, Megalopygidae, and Dalceridae (a tropical group) in the Cossoidea, and the Epipyropidae in the Tineoidea, leaving only the Zygaenidae in the superfamily. His splitting of the superfamily is supported by larval characters.

Hesperioidea, Papilionoidea

The butterflies provide a good example of a progression towards an external feeding habit. Megathymid larvae bore in stems and roots of Yucca. Hesperiid larvae live in rolled or folded leaves or in a nest of several leaves. All the Papilionoidea are exposed feeders in the larval stage. Many Lycaenidae, the most specialized group of butterflies, have adopted unusual feeding habits including predation and symbiotic relations with ants. Structural changes have paralleled this progression in feeding behavior. Megathymid larvae are pale in color and although secondary setae are present, the primaries are not obscured. Hesperiid larvae are cryptically colored, and secondary setae usually obscure the primaries. Crochets in the Hesperioidea are arranged in a complete or nearly complete circle as in most internal feeding Microlepidoptera. All the Papilionoidea share with the Hesperioidea a similar pattern of secondary setation. But radiation in adaptive strategies for external feeding has occured. Papilionid larvae have developed defensive glands (osmeteria) and striking color patterns, some cryptic, others aposematic. Pierid larvae, mostly herbaceous feeders, have adopted cryptic coloration. Nymphalid larvae have adopted a variety of defense mechanisms: aposematic coloration in the Danainae, cryptic coloration in the Satyrinae, defensive spines,

barbs, etc. in the Nymphalinae. Lycaenid larvae show many specialized characters including a retractible head, lobate planta, and a fusiform body shape. Crochets in the Papilionoidea are arranged in a mesoseries (sometimes a mesopenellipse in the Papilionidae), a clear adaptation to crawling about on twigs and leaves.

The classification of the Papilionoidea used in the key is that of Ehrlich (1958) who recognized five families of butterflies: Pieridae, Papilionidae, Nymphalidae, Libytheidae, and Lycaenidae. The Parnasiidae and Riodinidae were reduced to subfamilies, under Papilionidae and Lycaenidae respectively. The Danaidae and Satyridae were similarly treated under Nymphalidae.

Geometroidea

Important characters in recognizing a geometroid larva are: Head hypognathous. Secondary setae usually present either on the prolegs (Geometridae, Drepanidae), or as extra setae in fixed positions (Geometridae, Epiplemidae, Thyatiridae), or as numerous small setae evenly distributed over the cuticle (Drepanidae). SD1 & SD2 included on the prothoracic shield (excluded in the Thyatiridae). Prespiracular group bisetose on TI (trisetose in Thyatiridae). SV group bisetose on TII & TIII (unisetose in Geometridae). Al-8 with L1 & L2 far apart, with L4 present, and usually with extra SV setae. A9 with L group unisetose. Prolegs usually present on A3-6 & 10 but absent on A10 in Drepanidae and absent on A3-5 in most Geometridae. Crochets biordinal or triordinal, and arranged in a mesoseries.

Mimallonoidea

Franclemont (1973) excluded the tropical American Mimallonidae from the Bombycoidea and erected a new superfamily. The combination of a bisetose prespiracular group, a complete circle of crochets, and secondary setae confined to the SV group on Al-8 will distinguish the few mimallonids occurring in our area from the Bombycoidea.

Bombycoidea

Important characters in recognizing a bombycoid are: Head hypognathous. Secondary setae always present and profusely covering the body. In the Lasiocampidae and Apateloididae setae are as long or longer than primaries, but of irregular length. In the Saturniidae and Bombycidae, setae are usually very small and short. Primary setae are almost always obscured, and secondaries are never grouped in dense tufts on verrucae, or in pencils (except Apatelodidae). Chalazae and scoli, often profusely branched and bearing urticating spines are sometimes present (Saturniidae). Crochets are biordinal or triordinal and arranged in a mesoseries.

The Saturniidae have often been given superfamily rank, as the Saturnioidea, but the arrangement used in the key is that of Ferguson (1971). The three subfamilies of Saturniidae that occur in North America, the Saturniinae, Hemileucinae, and Citheroniinae, are very distinct and have sometimes been treated as families.

Larvae of the Lasiocampidae and Apatelodidae (only 2 genera of the latter occur in the United States; Apatelodes and Ocleclostera) are very similar. The only reliable character separating them is the

presence of mid-dorsal tufts of secondary setae on abdominal segments in the Apatelodidae.

Sphingoidea

A single family, the Sphingidae, comprise the Sphingoidea. Larvae can be recognized by the following: Head hypognathous. Secondary setae numerous but small, often on tiny chalazae. Scoli, pinacula, and verrucae absent. Primary setae are usually missing, but often SD1 and L1 on A1-8 are present. If L2 is also present on A1-8, then L1 & L2 are close together below the spiracle. All but 1 genus (Lapara) have a mid-dorsal horn or tubercle on A8. Biordinal crochets are arranged in a mesoseries.

Noctuoidea

Important characters in recognizing a noctuoid larva are: Head hypognathous. Secondary setae absent in cutworm-type noctuids. In the Lymantriidae, Arctiidae, Ctenuchidae, Pericopidae, and Nolidae, secondaries are present on verrucae in primary positions: D, SD, (or D+SD fused), L, and SV on the thoracic segments; and Dl, D2, SD, L1, L2, L3, SV & V on the abdominal segments. Verruca L1 is caudad of the spiracle on A1-6 & 8, but usually caudo-ventrad of the spiracle or fused with L2 on A7. In the Notodontidae and Dioptidae, secondary are always present on the prolegs and usually in fixed positions elsewhere. Many Notodontidae and Lymantriidae and a few Noctuidae may be profusely hairy and lacking verrucae. Horns and scoli are never

present, although dorsal humps or gibbosites are present in some
Notodontidae.

In groups where the primaries are conspicuous, the following arrangements are diagnostic: SD1 & SD2 on TI close together and excluded from the prothoracic shield, with SD1 often thinner and smaller than SD2. Prespiracular group bisetose on TI. SV group unisetose on TII & TIII. L1 & L2 far apart on A1-8 with L4 present in most Notodontidae, and Dioptidae. On A7, L1 is farther from the spiracle and closer to L2 than on the preceding abdominal segments. The L group is unisetose on A9. Crochets usually uniordinal (a few biordinal) and arranged in a mesoseries. In the Ctenuchidae, Pericopidae and most Arctiidae crochets are in a heteroideous mesoseries. Prolegs are usually present on A3-6 & 10, but may be reduced or absent on A10 in some Notodontidae, reduced or absent on A3 & A4 in some Noctuidae, and absent on A3 in a11 Nolidae.

The Ctenuchidae, Pericopidae, and Arctiidae share many important characters including the presence of secondary setae on verrucae and heteroideous crochets on a lobate planta (Fig. 56). The Ctenuchidae and Pericopidae are possibly nothing more than specialized groups within the Arctiidae. A number of arctiid genera (most notably <u>Utetheisa</u>, <u>Hypoprepia</u>, <u>Holomelina</u> differ more from the rest of the Arctiidae than do the Pericopidae and Ctenuchidae.

Based on available larvae, the validity of the Dioptidae as a family is questioned here. A single species, Phryganidia californica (Packard) occurs in the United States (Borror & DeLong 1971), and it has many characters in common with the Notodontidae, including a v-shaped

labral notch with a groove extending to the base of the labrum (Fig. 82). Another specimen from the USNM collected in Arizona, and identified as a dioptid by H. W. Capps has all these features plus reduced AlO prolegs, a common feature of many notodontids. Clearly the Dioptidae and Notodontidae are very close, if indeed they are separate families.

Larvae of <u>Doa ampla</u> Grote are very unusual and difficult to associate with any family. Although they undoubtedly belong in the Noctuoidea, they have been placed in the Lymantriidae (Holland 1903), the Pericopidae (McDunnough 1938), and the Dioptidae (Forbes 1948).

A COMPUTER GENERATED KEY

A. HISTORICAL DEVELOPMENT OF KEYS

As with many biological ideas, the germs of the development of keys can be found in the thinking of Aristotle (Voss, 1962). His attempts at classification used dichotomous characters, and exemplified the type of analysis found in modern keys. In the works of the 17th century naturalists Morison, Ray, and Rivinius, there are diagrams which might be considered the first analytical keys, but an identification key of a modern type did not appear until Lamarck (1778) in his <u>Flore Francaise</u>. Lamarck differed from his predecessors in that he recognized the artificial nature of keys. As Bather (1927) stated:

"A key is not a classification, but a method of analysis. This idea was first explicitly brought forth by Lamarck at the very beginning of his career. Having asserted that every species of French plant could be more readily determined by a purely arbitrary analytic key than by the Linnaean system with its mixture of supposed reality and ordered arbitrariness, he was challenged to produce such a key and he did this within twelve months."

There have been few innovations in the mechanics of key writing since Lamarck. Apart from the bracketed (parallel) style of zoologists, and the indented style of botanists, taxonomists have been reluctant to explore new schemes of identification.

Leenhouts (1966) identified two major types of keys, and discussed their advantages and disadvantages. In the "analytical" key the first couplet divides all the taxa into two or more groups. The user begins with the first couplet and chooses between two or more states, directing him to further couplets, until an endpoint is reached. Although analytical keys are commonly used, they have major disadvantages. They

inadequately handle variable or incompletely known taxa, and are difficult to construct, revise, and edit. They force the user to follow a predetermined pathway with pre-chosen characters, making mutual comparison between taxa difficult. Advantages of analytical keys include ease of use and publication.

In the second kind, "synoptical" keys, all taxa are divided over the different states under every couplet. The user can begin with any character and continue choosing characters in any order, reducing with each step the number of taxa until only one remains. The basic synoptical key is a table or data matrix, with enumeration of taxa on the left and of characters on the top. Each taxon has its own line, each character its own column. Advantages of synoptical keys are:

1) one can make use of every character making each identification more certain; 2) they are easy to construct; and 3) mutual comparison between taxa using the same character is easier. Disadvantages are: 1) their use is limited by large numbers of taxa; 2) they are difficult to publish.

Leenhouts concluded that synoptical keys are more reliable than analytical keys. He proposed a new style for synoptical keys, eliminating their major disadvantage, that of publication difficulty (see synoptical key, p. 34).

Other alternatives to analytical keys are those by Ogden (1943), Nobles (1965), Archbald (1967), Duke (1969), Hansen & Rahn (1969), and Morse (1971, 1974). Ogden developed a synoptical key in which taxa and characters are given in coded form. His system is cumbersome, requiring 9 pages to sort 8 taxa. Nobles provided a key in which characters are

codified per taxon as a formula. If several taxa share the same formula each is given the same set of additional characters. Archbald, Hansen & Rahn and Duke provided characters and taxa on punched cards which can be sorted in any fashion by the user.

Morse (1971) distinguished between "polyclaves" and "algorithms". A polyclave is similar to a synoptical key. An algorithm is an exact identification procedure such as a classical style key. Morse (1974) developed a computerized "polythetic" polyclave interactive program in which a "variability limit" sets a threshold in the accumulation of differences between specimen and taxon. A taxon is not eliminated from the list of possible taxa until the threshold is exceeded. Polyclave identification programs with on-line interaction are powerful identification tools, provided that such equipment is available.

Important theoretical discussions of specimen identification are those of Metcalf (1954) and Osborne (1963). Metcalf discussed principles of style that should be followed in key construction.

Couplets should be stated in simple, direct, mutually exclusive pairs. They should also be dichotomous, with the first member stated in the positive, and the second in the parallel negative of the first. Keys should be constructed so they can be used backwards as well as forwards, by inserting before each couplet the number of the immediately preceeding couplet in the pathway. Osborne discussed statistical aspects of dichotomous keys and proved that the probability of error almost always increases if a key is made longer, and that the shortest key is the most symmetrical (achieved by each couplet dividing taxa into equal sub-groups). He concluded that the probability of correct identification in a key is directly affected by its shape and length.

Early attempts to use computers in the construction of keys are those of Moller (1962) and Niemala, et. al. (1968). Moller developed a statistical theory for writing a key to polythetic taxa for maximum probability of correct identification. Niemala et. al. described a system for choosing a key based on a "matrix-reduction and monothetic-divisive algorithm" (Morse 1971). These programs require complete sets of binary characters, and produce keys in numerical form only.

More recent key construction programs (Morse 1968, 1971, 1974; Hall 1970; Pankhurst 1970b; Dallwitz 1974) employ "concise recursive algorithms" (Morse 1971) for key construction. They work on a simple principle: A group of taxa is divided by an appropriate character into two or more subgroups, each of which is further divided by other characters, until every taxon is distinguished from all others. A separate key is constructed for each subgroup as it is created. program sets thresholds which must be exceeded before a "test" (=couplet) is allowed to participate in the key. Thresholds are set by desirable features of a key which the program seeks to optimize. First, only characters with high "ease of observation" values are allowed to participate in a couplet. Next, cycles for finding a suitable couplet are run first for single characters, then for combinations of two, followed by combinations of three. If a simple couplet is found (one which uses just one character), the program does not search for character combinations. Simple couplets are favored, but more complex ones are used where needed. The program makes the most symmetrical key possible by favoring even numbers of taxa in subdivisions (Osborne 1963). It also provides rapid routes to the more common taxa by

allowing the user to duplicate data sets for these taxa. Drawbacks of Hall's program are that it uses quantitative data only and prints the key in a numeric form which must be rewritten before use. Morse's program is similar, but prints a key in a form which needs additional characters, rephrasing, and other editorial changes before it is ready to use. Morse's algorithm is based on theoretical considerations of Gyllenberg (1963) as well as those of Osborne and Moller. It uses dichotomous characters only, but also allows characters to be coded as variable or inapplicable.

Pankhurst (1970b) developed the first program that printed keys in a ready-to-use form. His algorithm differed from those of Hall and Morse in the use of "character-convenience blocks" and his employment of the "attribute-value" (character - character state) rather than a hierarchical couplet concept (Morse 1971). Pankhurst's program has many desirable features. It allows for multi-state as well as binary characters. Characters can be weighed with respect to reliability. The program prints both a conventional key (bracketed or indented) and the data matrix. The maximum number of characters to be used in a couplet can be specified by the user. Unknown or missing characters are allowed. All remaining distinctive characters are automatically printed at the end of a pathway. A variety of botanical keys have been constructed using Pankhurst's program (Pankhurst 1970a, Watson & Milne 1972). These keys used over 50% fewer characters and were 10% to 20% shorter than hand made versions.

A recent program (Dallwitz 1974) has further advantages over those of Hall, Morse, and Pankhurst. A number of parameters controlling

different aspects of key generation are included in his algorithm. These allow variation of strategy producing different kinds of keys with the same data. The parameters control the weight given to reliability of characters, abundance of taxa, how the program handles intra-taxon variability, how often characters are re-used, and the maximum number of characters allowed in a couplet. Provisions are provided for masking both taxa and characters, making production of special-purpose keys possible from subsets of the data. A user can also pre-set the position of any character in the key. The output includes a list of characters and their states (App. C, p. 99), a data matrix (App. D, p. 102), a tabular key resembling a tree diagram (a diagrammatic representation of the key which helps the user assess the structure of the key) (App. E, p. 108), and a conventional bracketed key (App. F, p. 113).

A major advantage of Dallwitz' program is its low cost and high speed. A tabular key to 145 Australian grass genera was produced from 433 items (data sets) and 120 characters in 80 seconds of computer time at a cost of \$5.00. Printing a conventional key took 25 seconds more and cost \$2.00 more.

B. DESCRIPTION OF PROGRAM KEY

The purpose of an identification is to assign any individual to its taxon. KEY (Dallwitz, 1974) groups individuals into two types of class; a taxon (class of individuals with the same name), and item (class of individuals with identically coded descriptions). A taxon may include more than one item because of variation within the taxon.

The program user provides a list of characters and characters states, and a table giving the taxa corresponding to the items (App. C, p. 99). Missing values may be interpreted as not applicable or variable. The user may also provide estimates of the costs of using the characters and of the relative abundances of the items. The "cost" of a character is a combination of its probability of error and the effort in its use. Errors may arise because of misinterpretation or misjudgment by the user of the key, or because intra-taxon variability has not been completely accounted for.

KEY minimizes the average cost of an identification by assuming that character costs are additive, and that the frequency with which an item will need to be identified is proportional to its abundance. The character selection algorithm used in KEY is an estimate of the average cost of an identification. It selects a character with low cost and divides the taxa into two or more subgroups. Each subgroup is then subdivided by other characters. This continues until each subgroup contains only one taxon, or until no suitable character is found.

Options which allow control over key generation are the presetting of characters to be used in any position in the key (desirable since KEY may not necessarily use characters in a preferred order), masking of both characters and taxa allowing production of special-purpose keys from subsets of the data, and manipulation of 5 parameters (RBASE, ABASE, REUSE, VARYWT, NCONF) which optimize different aspects of key generation.

RBASE controls the weight given to cost indices. If RBASE=1, all characters have equal cost, and reliability indices do not influence

formation of the key. If RBASE=2, a more reliable character will be used over a less reliable character. The effect of RBASE on the length of a key is such that if RBASE=1, a short key using less reliable characters will result, and if RBASE>1, a longer key using more reliable characters will result.

ABASE controls the effect of the abundance indices on the structure of the key. If ABASE is large, abundant taxa will tend to key out early.

REUSE controls how often a character is used. If REUSE=1, a character is not likely to be used again, but if REUSE>1, the character will more likely be used later in the key.

VARYWT determines the treatment of intra-taxon variability. A variable character can be coded as "V" or as a dash (-). If the absolute value of VARYWT is 1, there is no special penalty for characters with intra-taxon variability. If VARYWT=0, such characters will not be used for decisions involving the variable taxa, and each taxon name will appear only once in the key. Intermediate values of VARYWT have effects between these extremes. Unknown values (dash, blank, or dot on the taxon cards) are treated as not applicable if VARYWT is positive and variable if VARYWT is negative.

NCONF is the maximum number of confirmatory characters to be sought for each main character. If NCONF=0, couplets will use only a single character. If NCONF=1, 2, or 3, multiple character couplets will appear.

The printout consists of the list of characters and their states and the reliability index (App. C, p. 99), the list of taxa and

abundance indices and the data matrix (App. D, p. 102), the key in tabular form with the sequence in which characters were used in consecutive vertical columns (App. E, p. 108), a list of characters in the order they were used in the key, and the bracketed key (App. F, p. 113).

Seven taxa arranged in a vertical column, and 5 characters arranged in a horizontal column are used as an example of how the program might run (Table 2, p. 79). The state each taxon manifests for each character is given in its adjacent horizontal column. Character 1 (C1) has 4 states: A, B, C, & D; C2 has 4 states; C3 has 3 states; C4 has 2 states; C5 has 4 states. In a tabular key, taxa are arranged on the left in the order they are sorted, and the characters in vertical columns in the order they are used (Table 3, p. 80). The same key is shown in the bracketed style (Table 4, p. 80). The characters are given reliability indices, with C4 the most reliable (9), and C3 the least (1). For simplicity, taxa were not given abundance indices.

Character 4 has high reliability, and best divides the 7 taxa into equal subgroups; and is chosen for the first couplet. Since VARYWT=1, Taxon 4 appears in each subgroup (Table 3,column 1) and keys out twice (Table 4, couplets 4 & 5). If VARYWT were set at 0 another character would be chosen for the first couplet. Both Cl and C2 divide taxa 1 and 4 into equal sub-groups, but Cl is chosen for its higher reliability (Table 3, column 2; Table 4 couplet 2). Only C3 can separate taxa 1 & 2, so it is chosen despite its low reliability (Table 3, column 3; Table 4,couplet 3). The program allowes for multi-state couplets, as shown in the choice of C1 to separate taxa 4 to 7 (Table 3, column 2; Table 4, couplet 5).

TABLE 1.

Five characters with states in coded form and reliability indices to illustrate operation of KEY.

1. (8) Character 1 (=C1) A. В. C. 4. (9). Character 4 (=C4) D. Α. В. 2. (5) Character 2 (=C2) 5. (4). Character 5 Α. В. Α. C. В. C. 3. (1). Character 3 (=C3) D. A. В. C.

TABLE 2.

Imaginary data with 7 taxa (=T1-T7) and the 5 characters from Table 1, to illustrate operation of KEY. Characters listed in row along the top, and taxa listed in the left vertical column.

	C1	C2	C3	C4	C5
T1	A	A	В	A	A
T2	A	A	С	A	A .
Т3	В	В	С	A	В
T 4	В	В	A	V	С
Т5	В	С	A	В	В
Т6	С	С	В	В	С
Т7	D	A	С	В	D

TABLE 3.

Tabular key using data from Table 2 illustrating operation of KEY. Seven taxa (T1-T7) listed in the left vertical column with characters (C1-C5) and states arranged in the order they are used in 3 vertical columns from left to right. RBASE = 2, REUSE 1, VARYWT = 1, NCONF = 0.

	1	2	3
T1	C4-A	C1-A	С3-В
Т2	C4-A	C1-A	C3-C
Т3	C4-A	C1-B	С5-В
Т4	C4-A	C1-B	C5-C
Т4	C4-B	C1-B	С2-В
T 5	C4-B	C1-B	C2-C
Т6	C4-B	C1-C	
Т7	С4-В	C1-D	

TABLE 4.

Same key as in Table 3, in the bracketed style, with characters, states, (C1-C5), and taxa (T1-T7) in coded form.

1(0).		• • • • • • • • • • • • • • • • • • • •	
2(1).		•••••	
3(2).		• • • • • • • • • • • • • • • • • • • •	
4(2).			
5(1).	C1-C	• • • • • • • • • • • • • • • • • • • •	Т6

C. IMPLEMENTATION OF KEY

KEY is written in Fortran and contains 1300 statements and 34 subroutines. The last program run (App. E & F, pp. 108, 113) took about 20 seconds of computer time at a cost of less than \$10.000.

A list of characters and character states (App. C, p. 99) and a matrix giving the taxa corresponding to the items and the states of the characters for the items (App. D, p. 102) were punched on IBM cards. Missing values were recorded as a dot (.), and inapplicable states as a dash (-). Abundance indices were provided (App. C, p. 99) based on the number of genera in each family in North America. Nine is an abundant taxon (Noctuidae), 1 is a rare taxon (Micropterygidae). Reliability indices for the characters were based partly on the data collected and partly on experience in using the characters in previous keys.

Three hundred-fifty individuals were initially examined and the states of 90 characters determined for each. Initially all characters that varied (had different states within a family) were recorded as inapplicable for that family. During first runs of the program, the error diagnostic "key incomplete, more information needed" appeared (App. F, p. 113). This was an expected result, and additional characters and items were added for the unseparated taxa.

D. RESULTS

Based on data collected for the chosen characters, KEY did not produce satisfactory results. Over a dozen keys were generated with the program, the last of which (App. E & F, pp. 103, 113) was used as a structure for developing the hand-written key in the thesis.

Allocating the correct amount of storage space for key generation was frustrating and time consuming. Each new key was tested with correctly identified specimens. When inadequacies were discovered, new taxa, new items, more characters and more data were added. This necessitated revising the parameters every time the program was run. In later runs the data set was large enough to exceed the storage capacity of the program. The families were then divided into four groups, based on character 71 (App. C, p. 99). Each group was then run separately producing four different keys (Part A, pp. 108. 113: Part B, pp. 110, 117; Part C, pp. 111, 118; Part D, pp. 112, 119).

Even in the last run of the program the results were poor. For example, many taxa keyed out twice in the same couplet (couplets 5, 6, 25, p. 113; 60, p. 114). The Notodontidae key out in 29 different places, the Noctuidae in 36. Much editing and rewriting would be necessary to eliminate these worthless couplets.

For large taxa such as Noctuidae attempts were made to account for all possible combinations of variable characters even if certain combinations were unknown to actually exist. This resulted in an exponential increase in the number of places a taxon would key out, as the program generated new items. For example, some noctuids may have scoli others may not, some lack secondary setae, some have setae on verrucae, others have setae scattered over the cuticle, and others a few secondary setae in fixed positions. Some noctuids have uniordinal crochets, others have biordinal. If one were to take into account all possible combinations of the different states of these characters, noctuids would have to key out in at least 2x4x2=16 different places.

When more specimens of variable taxa were examined, many endpoints in the key were found to represent non-existant combinations of characters for the taxon. These were eliminated from the hand-written key. The combined 4 parts of the computer key has 181 couplets, whereas the hand-written key has only 144 couplets.

In some couplets as many as 3 confirmatory characters were printed in addition to the first character (couplet 38, p. 114). Since these couplets would be a cumbersome task to any key user, their value was questioned. When tested with specimens, few were found to fit all four characters. These couplets would therefore be misleading, and would have to be deleted or shortened.

First runs of the program had absurdly large clumps of unseparated taxa. Most of these were eliminated or reduced in size, but in last runs, no manipulation of parameters or addition of new information would eliminate them (couplets 25 & 28, p. 113; 79, p. 115; 23, p. 117). These problems also would be solved only by editing and re-writing.

IV. CONCLUSIONS

There is probably no shortcut for producing a reliable key. The method requires time consuming examination and re-examination of specimens to find appropriate characters and testing the key as it is being written. Making corrections or adding new taxa and characters may necessitate the complete redesigning of a large and complicated key.

A critical problem with classical keys is that they rely too heavily on a few diagnostic characters for recognition of taxa; that is, they are monothetic in concept. If taxa are polythetic, on the other

hand, no single character or small combination of characters will be sufficient for assigning a specimen to its right taxon (Sneath & Sokal, 1973). This difference in concept between keys and taxa makes keys as identification tools extremely sensitive to single errors on the part of users.

For all its sophistication, KEY produces a classical type key and does not solve their basic problems. The theoretical aspects of key construction (Osborne 1963) implemented in KEY are probably valid but are overidden by other considerations when writing a key. The overall probability of correct identification, for example, depends less on the structure of the key, than on the stability of the taxa, careful wording of the couplets, and careful choice of characters. Even the editing advantages of KEY were exaggerated in the literature. Addition of geographical and biological information, and rewording of awkward couplets would have necessitated rewriting, even if a satisfactory key had been obtained. Lastly, time wasted in getting the program to work properly would have been used more profitably in testing the key against additional specimens.

When using computer methods in identification, it seems too easy to forget that one is dealing with biological organisms, and that both taxa (at least above species) and characters are artificial constructs. For example, KEY requires rigid pre-determination of characters and their states. Although this seems desirable as an unbiased test of characters, it was a constant source of trouble. Of 100 characters given to KEY (App. C, p. 99), most were found to be useless or misleading when new specimens were tested with the key. Fewer than 40

characters were used in their original form in the hand-written key.

Many others had to be rewritten or delineated into states differently before they would work. Continued use of the program would have necessitated tedious changes in the data matrix and character list to keep up with changing ideas of character reliability.

KEY requires a large amount of taxonomic data before the program is run. It behaves the user to supply useful data, but there is no way of knowing this until a key produced with the program is tested with specimens. When constructing a hand-written key, on the other hand, one gathers data only as they are needed.

Leenhouts' (1966) design for a synoptical key is probably the best solution to the problems of key construction short of on-line interactive programs (Morse 1974). Its polyclave nature makes identification quick, easy, and reliable. Its flexibility allows one to add or delete characters, data, or taxa without difficult rewriting or restructuring. Extensive use of Leenhouts' design would greatly improve any identification scheme.

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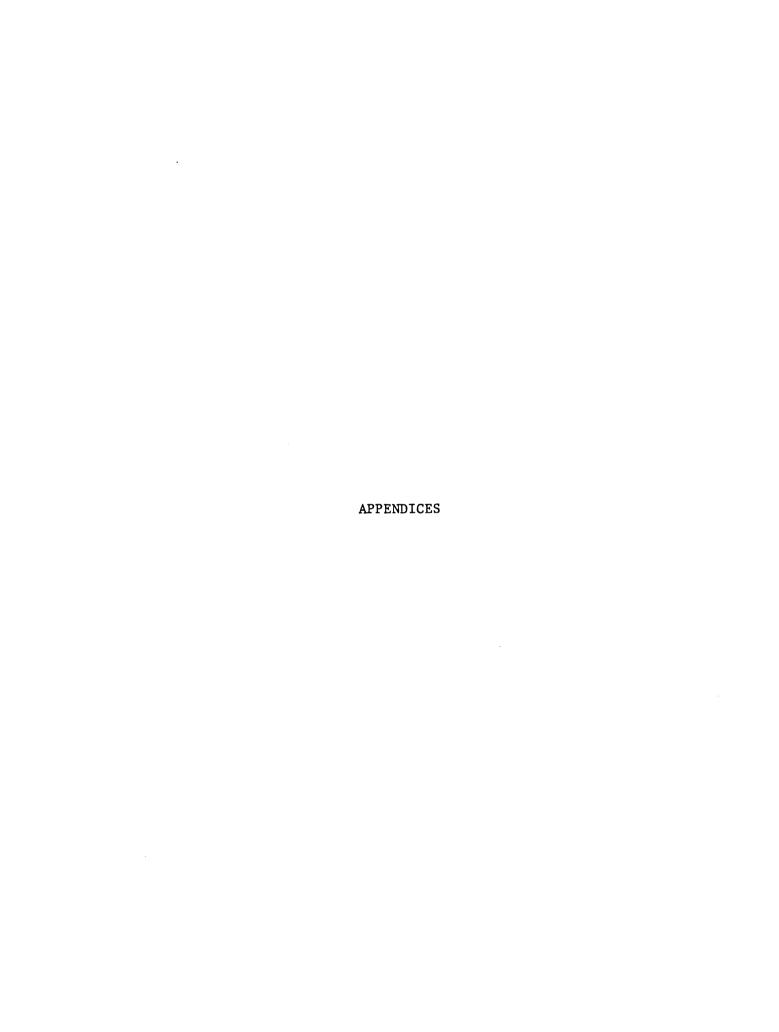
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APPENDIX A

CLASSIFICATION OF THE LEPIDOPTERA

```
Suborder ZEUGLOPTERA
  MICROPTERYGOIDEA
      Micropterygidae (=Eriocephalidae; formerly part of Jugatae, Homo-
             neura)
Suborder DACNONYPHA
  ERIOCRANIOIDEA
       Eriocraniidae (formerly part of Jugatae, Homoneura)
Suborder MONOTRYSIA
  NEPTICULOIDEA
      Nepticulidae (=Stigmellidae)
       Opostegidae (formerly part of Tineidae)
  INCURVARIOIDEA
      Incurvariidae
           Incurvariinae
          Adelinae
          Prodoxinae
       Heliozelidae
       Tischeriidae (formerly part of Tineidae)
Suborder EXOPORIA (formerly part of Monotrysia)
  HEPIALOIDEA
      Hepialidae (formerly part of Jugatae, Homoneura)
Suborder DITRYSIA
  TINEOIDEA
       Tineidae (includes Amydriidae, Setomorphidae, Acrolophidae)
           Acrolophinae (=Anaphorinae, Psychidae, in part)
           Scardiinae
          Nemapogoninae
           Tineinae
       Psychidae (includes Talaeporiidae)
       Oschenheimeriidae
       Oinophilidae (=Oenophilidae)
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TINEOIDEA (continued)

Lyonetiidae (=Tineidae in part; includes Bucculatrigidae, Leucopterygidae)

Gracillariidae (=Gracilariidae, Lithocolletidae, Tineidae in part; includes Phyllocnistidae)

YPONOMEUTOIDEA

Yponomeutidae (=Hyponomeutidae; includes Plutellidae, Argyresthiidae, Acrolepiidae; excludes Scythridae)

Yponomeutinae

Plutellinae (=Acrolepiidae)

Argyresthiinae

Heliodinidae (=Tinaegeriidae; Elachistidae in part, Lavernidae in part)

Epermeniidae

Sesiidae (=Aegeriidae, Sphingidae in part)

Glyphipterygidae (=Hemerophilidae, Yponomeutidae in part; includes Hypertrophinae, Choreutidae)

Douglasiidae (=Glyphipterygidae in part, Elachistidae in part)
(no larval specimens were available for inclusion in the key)

CASTNIOIDEA

Castniidae (includes Neocastniidae, Tascinidae)

COSSOIDEA

Cossidae (=Zeuzeridae; includes Ratardidae, Metarbelidae, Hypoptidae, Zeuzeridae)

TORTRICOIDEA

Tortricidae (includes Eucosomidae, Grapholithidae, Epiblemidae, Sparganothidae, Oecophoridae)

Phaloniidae (=Conchylidae, Tortricidae in part)

GELECHIOIDEA

Elachistidae (=Cycnodiidae in part; included in former Lavernidae) Coleophoridae

Cosmopterygidae (part of former Lavernidae)

Momphidae (part of former Lavernidae)

Walshiidae (part of former Lavernidae)

Blastodacnidae (formerly part of Momphidae)

Blastobasidae (=Gelechiidae in part)

Scythridae (=Scythrididae, Blastobasidae in part; formerly part of Yponomeutidae)

Oecophoridae (=Gelechiidae in part; includes Depressariidae, Metachandidae)

Ethmiidae (=Oecophoridae in part)

Stenomidae (=Stenomatidae, Xylorictidae, Uzuchidae)

Gelechiidae

Symmocidae (=Gelechiidae in part)

COPROMORPHOIDEA

Alucitidae (=Orneodidae, Pterophoridae in part)

Carposinidae (=Tortricidae in part)

PYRALOTDEA Hyblaeidae (formerly part of Noctuidae) Thyrididae (=Thyridae, Sphingidae in part; includes Argyrotypidae) Pyralidae (=Pyralididae; includes Pyraustidae, Phycitidae (=Phycidae), Crambidae, Crysaugidae, Nymphulidae, Epipaschiidae, Galleriidae) Pterophoridae (includes Agdistidae; excludes Alucitidae) ZYGAENOIDEA (=Psychodoidea) Zygaenidae (=Pyromorphidae, Anthroceridae in part, Sphingidae in part) Megalopygidae (=Lagoidae, Liparidae in part) Epipyropidae Limacodidae (=Eucleidae, Cochlidiidae, includes Chrysopolomidae) Dalceridae HESPERIOIDEA (formerly part of Rhopalocera) Hesperiidae (excludes Megathymidae) Megathymidae (=Hesperiidae in part) PAPILIONOIDEA (formerly part of Rhopalocera) Papilionidae (includes Parnasiidae) Pieridae (=Asciidae) Nymphalidae (=Aegyreidae; includes Danaidae, Satyridae, Heliconiidae; excludes Libytheidae) Danainae (=Lymnadidae, Euploeinae; includes Ithomiidae) Satyrinae (=Agapetidae) Nymphalinae (includes Heliconiidae) Libytheidae Lycaenidae (=Cupidinidae, Ruralidae; includes Riodinidae) Lycaeninae Riodininae (=Erycinidae, Lemoniidae, Nemerobiidae) GEOMETROIDEA (=Uranioidea in part) Geometridae Drepanidae (=Drepanulidae, Platypterygidae; formerly part of Drepanoidea: includes Auzatidae) Thyatiridae (=Cymatophoridae; formerly part of Drepanoidea) Epiplemidae (formerly part of Uranioidea) MTMALLONOIDEA Mimallonidae (=Lacosomidae, Cicinnidae; formerly part of Psychidae, Uranioidea, Bombycoidea, Drepanulidae) BOMBYCOIDEA (=Lasiocampina) Saturniidae (=Saturnioidea; includes Citheriniidae, Hemileucidae) Saturniinae Hemileucinae Citheroniinae Bombycidae Lasiocampidae (=Eupterotidae, Bombycidae, Lachneidae, Notodontidae in

Apatelodidae (=Zanolidae, Eupterodidae; formerly part of Notodontidae)

SPHINGOIDEA

Sphingidae (=Smerinthidae)

NOCTUOIDEA

Dioptidae (includes <u>Doa</u>) Notodontidae (=Ceruridae)

Lymantriidae (=Liparidae)

Arctiidae (includes Lithosiidae)

Ctenuchidae (=Amatidae, Syntomidae, Euchromoodae)

Pericopidae (includes Hypsidae)

Nolidae (=Noctuidae in part)

Agaristidae (=Noctuidae in part)

Noctuidae (=Phalaenidae, Nolidae in part; includes Agrotidae, Acronyctidae, Plusiidae; excludes Nolidae, Hyblaeidae)

APPENDIX B. NOMENCLATORIAL SYSTEMS FOR PRIMARY SETAE (TI)

Propriorreceptor MXD1 MV2 MV3	VI	SV1 SV2	L1 L2 L3	SD1 SD2	D1 D2	Tactile XD1 XD2	Hinton (1946)
PMT1 MC1 MC2	S1	C1 C2	PN3 PN1 PN2	PT4 PT1	PT3 PT2	ST2 ST1	Mutuura (1956)
VIIc VIId	VIII	VIIa VIIb	IV VI A	III II	ΙX	IX IIIa	Gerasimov (1935) Nepialidae Other
Xa VIIc VIId	VIII	VIIa VIIb	IV VI	IIIa III	III	X	(1935) Others
1 1 1	sigma	pí nu	kappa eta theta	epsilon rho	beta delta	alpha gamma	Forbes (1923)
vII	VIII	VI	III A AI	Ic IIc	IIa IIb	Ia Ib	Heinrich (1916)
- tau phi	sigma	pí nu	eta kappa theta	rho delta	alpha beta	gamma epsilon	Fracker (1915) Jugatae Frena
1 1 1	sigma	pi nu	kappa eta theta	epsilon rho	beta delta	alpha gamma	(1915) Frenatae
1 1 1	viii	via vib	iv v iii	ic iic	iia iib	i a	Dyar (1895)

APPENDIX B (CONTINUED) NOMENCLATORIAL SYSTEMS FOR PRIMARY SETAE (TII & TIII)

MSD1 MSD2 MV1 MV2 MV3	Proprio- receptor MD1 MD2	SV1 SV2 V1	SD1 SD2 L1 L2 L3	Hinton (1946) Tactile D1 D2
MT3 MT4 MS1 MC1 MC2	MT1 MT2	C1 C2 S1	ST1 ST2 PN1, SC2 PN3, SC1 PN2	Mutuura (1956) T2 T1
IXa IXb VIIc VIIc VIId	Xa Xb	VIIa - VIII	III IIIa V IV VI	Gerasimov (1935) Hepialidae Others I I I
IXA IXb VIIb VIIc VIId	Xa Xb	VIIa - VIII	III IIIa IV V VI	(1935) Others I II
- x x x	ι×	vii viii	iib iia iv v iii	Forbes (1923) ia ib
- VII VII 	1 1	viii	IIb IIa IV V III	Heinrich (1916) Ia Ib
gamma gamma nu tau omega	gamma	pi - sigma	rho delta kappa epsilon theta	Fracker Jugatae alpha beta
omega	1 1	pi - sigma	rho epsilon kappa eta theta	Fracker (1915) atae Frenatae Alpha alpha beta beta
111 11	1 1	vi - viii	iib iia iv v iii	Dyar (1895) ia ib

APPENDIX B (CONTINUED) NOMENCLATORIAL SYSTEMS FOR PRIMARY SETAE (A1-9)

Proprio- receptor MD1 MD2 MD3	VI	SV1 SV2 SV3	L1 L2 L3	SD1 SD2	Tactile D1 D2	Hinton (1946)
PMT1 - MS1	S1	SS1 SS2 SS3	PN1, SC1, SC2 SC1, SC2 C1	ST1 ST2	T2 T1	Mutuura (1956)
VIId VIId	VIII	VIIa VIIb VIIc	VI VI	III IIIa	ΙΙ	Gerasimov (1935) Hepialidae Others
Xa Xb VIId	VIII	VIIa VIIb VIIc	VI VI	IIIa	ΠΠ	(1935) Others
i i	viii	viia viib viic	v iv vi	iii iiia	íí	Forbes (1923)
1 1 1	VIII	VII VII	VI V	IIIa	II I	Heinrich (1916)
omega	sigma	pi nu tau	theta kappa eta	rho epsilon	alpha beta	Fracker Jugatae
1 1 1	sigma	pi nu tau	kappa eta mu	rho epsilon	alpha beta	Fracker (1915) atae Frenatae
1 1 1	viii	vii vii vii	iv v vi	iii iia	i i	Dyar (1895)

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FAMILY KEY TO LEPTTOPTERA LARVAE
                              1. (4) ME AD CAPSULE

A. LONGER THAN WIDE

A. LONGER THAN THE COMMAN WIDE

2. (5) MEAD CAPSULE EXTENDS EVENON THE COMMAN MOTCH

A. LONGER THAN TAILS WIDTH

3. (4) ME AD OPTEMTATION

A. POOR THAN THOSE STORM MORE SERVING THAN MIDE.

C. FURTHER THAN TAILS WIDTH

A. (3) PRONT

COMPTIONS OF STRICKY OF HORY

A. (3) PRONT

COMPTIONS OF STRICKY OF HAN MIDE.

B. S. HIDE AS LONG OF HEARLY

C. WIDER THAN THICE AS LONG AS WIDE

5. (9) PRONT

B. CLOSER BEHIND

6. (1) FRONT

C. WIDER THAN THICE AS LONG AS WIDE

C. WIDER THAN THICE AS LONG AS WIDE

6. CLOSER BEHIND

C. NITHER SERVINGED AT ANTERIOR END FRONTAL SUTURES ANGULATE OR HOT ULATE BETWEEN ANGULATE OR HOT LANGE SETTING OF HORY

C. WITTER SERVINGED AT ANTERIOR OF FRONTAL SUTURES ANGULATE. FRONTAL SUTURE

C. WITTER SERVINGED AT ANTERIOR OF FRONTAL SUTURES ANGULATE. FRONTAL SUTURE

A. EXTEND BETOING COMMAN HOTCH

A. EXTEND BETOING COMMAN HOTCH

E. STEEND BETOING COMMAN HOTCH
C. NETTHER EXPANSED AT ANTERIOR END OF FRONT

7. (S) ADPRENTITION SUPPLY CARRIER OF A MITERIOR END NOR FRONTAL SUTURES ANGULATE. FRONTAL SUTURE

2. EXTEND SECONDAL NOTCH

3. EXTEND SECONDAL NOTCH

4. EXTEND SECONDAL NOTCH

5. EXTEND SECONDAL NOTCH

6. EXTEND TO COCROMAL NOTCH

7. EXTEND TO THE CORONAL NOTCH

8. (8) FRONT EXTENDS TO THE CORONAL MOTCH

8. (8) FRONT EXTENDS TO THE CORONAL MOTCH

8. (10) COELL

10. OSCUPE OR ABSENT

9. (9) COELL

10. OSCUPE OR ABSENT

10. THREE TO FIVE PRESENT

10. THREE TO FIVE PRESENT

10. THREE TO FIVE PRESENT

11. (5) COELL

11. 12. 3. ARRANGED

12. AN ACUTE ANGLE (L.T. 88 D.)

13. (5) OCELL

14. AN ACUTE ANGLE (L.T. 88 D.)

15. IN AN ACUTE ANGLE OR NEARLY (1871-196 D.)

16. IN AN OBSIST ANGLE OR NEARLY (1871-196 D.)

17. AN ACUTE ANGLE (C. THREE NOT COLOR TO A MIT AND COLOR TO A MITH ANGLE (C. T. 88 D.)

12. (7) COELL STAN ACUTE ANGLE (C. T. 88 D.)

13. (5) AN ACUTE ANGLE (C. TRANCE OR NEARLY (1871-196 D.)

14. AN ACUTE ANGLE (C. TRANCE OR NEARLY (1871-196 D.)

15. IN AN ACUTE ANGLE (C. TRANCE OR NEARLY (1871-196 D.)

16. (7) COELL STAN ACUTE ANGLE (C. TRANCE OR NEARLY (1871-196 D.)

17. AN ACUTE ANGLE (C. TRANCE OR NEARLY (1871-196 D.)

18. (7) COELT STANCE OR NEARLY (1871-196 D.)

19. CONSTITUTIONS OF SERVICE OR NEARLY (1871-196 D.)

10. COERT TO AST THEN TO STANCE SETMEEN OCA AND OCS C. T. IV.

10. COERT TO AST THAN TO OR NEARLY OF THE THAN ALL AND AS

10. (5) READ STARS TO AT THAN TO AST THAN TO 
                     C STAP THE FOUR TO DISTANCE SETWEEN COCI A

13. (5) WE AD ST TA AZ

13. (5) WE AD ST TA AZ

14. (5) WE AD ST TA AZ

15. (6) WE AD ST TA AZ

16. (7) WE AD ST TA AZ

16. (8) WE AD ST TA AZ

16. (9) WE AD ST TA AZ

17. (9) WE AD ST TA AZ

18. (9) WE AD ST TA AZ

18. (10) WE AD ST TA AZ

19. (10) WE AD

19. (10) WE AD ST TA AZ

19. (10) WE AD

19. 
                 16. (9) MEAD SETA OI

A. INSTOR CIRCLE OF EVERY

B. OUT A THE CIRCLE OF THE WEEK OCZ AND OCS

18. (5) MEAD STATE OF MEARLY BETWEEN OCZ AND OCS

18. (5) MEAD STATE OF MEARLY BETWEEN AS AND LI

19. (5) MEAD STATE OF MEARLY BETWEEN AS AND LI

19. (5) MEAD STATE OF MEARLY BETWEEN AS AND LI

19. (5) MEAD STATE OF MEARLY BETWEEN AS AND LI

20. (3) EVERY STATE CALEND ON VENTRAL SIDE OF I STATE OF THE STATE OF
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52. (9) L JETAE ON THORAGIG SEG. I,
       53. (4) L SETAE ON MOD. SEG. 3,
     54. (5) L SETAF DW AND SEG. 9.
56. (5) 30 SE OM ALL TWEEE, 9

$7. (4) SEGRENTED SCLEMOTIZED THORACTC LEGS

$8. (4) PROLEGS ON ABD. SEGS. EXCEPT 18.

$8. (4) PROLEGS ON ABD. SEGS. EXCEPT 18.

$9. (4) PROLEGS ON ABD. SEGS. 10.

$9. (5) TOTAL POOP LEAST ON ALL ABDONIBAL SEGNETS WIDTH

$9. (1) TOTAL POOP LENGTH, F.G.L.,

$9. (1) TO
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63. (1) CERVICAL SHIELD

A. DISTINCT MEAVILY SCLEPOTIZED, LIGHTLY TO DARKLY PIGHENTED

B. DESQUEE OR REDUCED TO TRREGULA'S SCLEROTIZED SPOTS, OR ENTIRELY ABSENT

STEACLES

B. OR IT AND AB CONSPICUOUSLY LARGER THAN OTHERS

C. ONLY TI SP. LARGER THAN OTHERS

65. (7) SPIRACLES SHAPE

A. ROUND AB SP. LARGER THAN OTHERS

A. ROUND AB SP. LARGER THAN OTHERS

A. ROUND AB SP. LARGER THAN OTHERS

A. ROUND AB SP. LARGER THAN OTHERS
65. (7) SPIRACES SHAPE

A ROUND

66. (9) CROCHET ON A 3 (SERIAL)

67. (9) CROCHETS ON A 3 (SERIAL)

67. (9) CROCHETS ON A 3 (SERIAL)

68. (1) CROCHETS ON A 3 (SERIAL)

69. (1) CROCHETS ON A 3 (SERIAL)

60. (1) TRANSVERS ON CROCHETS OF COMPANY

60. (1) TRANSVERS CAND OR CROCHETS REDUCED IN NUMBER

69. (1) MAR DECEMBER OF COMPANY

69. (2) MAR DECEMBER

70. (6) MAR DECEMBER

71. (4) SECONDARY

71. (9) SECONDARY

72. (6) ANAL COMPLETE OF LONG, SCATTER DO OVER CUTTOLE, SOMET

1 FOR THANKES PRESENT, HEN SECONDARIES NO LONGER OR SECONDARY

60. (1) TRANSVERS PRESENT, HEN SECONDARIES OF SECONDARI
73. (5) HOUT OR ALL STAE CHALAZAE OR TUBERCLES
74. (5) SEGRENTS OF BOTT CHALAZAE OR TUBERCLES
75. (9) POSITION OF SETA SOL ON ABDORMAND OR CHANALATAE OR TOWN AND LETS
75. (9) POSITION OF SETA SOL ON ABDORMAND OR CHANALATAE OR THE COXA
76. (5) DISTINCT WITE FRONT OR STEAKING OR ONLY STRAFTLY ABOVE OR BELOW, IF ANY
76. (5) DISTINCT WITE FRONT OR STEAKING OR ONLY STRAFTLY ABOVE OR BELOW, IF ANY
76. (5) DISTINCT WITE FRONT OR STEAKING OR ONLY STRAFTLY ABOVE OR BELOW, IF ANY
77. (5) PROTECTION OF SETA SOL ONLY STRAFTLY ABOVE OR BELOW, IF ANY
77. (6) PROTECTION OF SETA SOL ONLY STRAFTLY ABOVE OR BELOW CAUDAL SHIELD
80. (9) HUMBER OF SETA SOL ON REAR AIS PROLEG BELOW CAUDAL SHIELD
81. (9) ELESS THAN SAL SOL ON SAME PINACULUM
80. (9) THE SETA SOL OF SETA SOL OF SETA SOL ON SAME PINACULUM
80. (9) THE SETA SOL OF SETA SO
                   96. (9) ANAL PLETE ON ARE PRESENT ON WENTRAL SIDE OF PROLEGS BETTER ON ARE PLETE ON ARE PROSESSES OF THE PROCESSES OF THE PRO
                       A. DISTINCTLY LAGGER HAND PROPHERS IN DIAMETER STANDARD IN THE STANDARD IN THE
                                                                                                                                                                                                                                  DISTINCTLY CONICAL OR ANGULATE, OF ARMED DORSALLY WITH SCOLI OR SPINES ROUNDED, MITHOUT SCOLI OR SPINES
99. (9) LABRAL NOYCH.

A. SPALLON, EXTENDING NOT MORE THAN 1/2 THE DISTANCE TO THE BASE OF THE LARRUM B. DEEP MITH PAPALLE SIDES, EXTENDING TO BASE OF LARRUM, OF MEANLY COMMENTS.

100. (9) EXPRESSION OF ABOUTE SEC. 7 (SOMETIMES B)

B. ASSENT ON DOPSOMESON OF ABO. SEGS. 6 (AND)

B. ASSENT ON DOPSOMESON OF ABO. SEGS. 6 (AND)
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FAMILY KEY TO LEPINOPTERA LARVAE (A)

	•••••	•••••	••••••	••	
TAXA, ABUNDANCE INDICES, AND CHARACTER		6.39.28. 07/2			
	12345 67898	11111 11112 12345 67890	22222 22223 12345 67891	12345 67896	4444 44445 12345 67898
1/ 1. (3) ERIOCRANIICAE	DCAAB BCAB-	A888A A	AA		-CB-A
2/ 2. (3) HEPIALIDAE	AA 888 CD8EC	AAACA ABC-9	ADCA- CCCCA	-AECB BCACC	ACCAA CC-CB
3/ % (3) NEPTICULIDAE			AA		CAB
4/ 3. (3) NEPTICULIDAE	03ABAA8- 04ABBA8-	AABAĀ	AA: :::::	::t:: :::::	######################################
5/ 4. (1) OPCSTEGIDAE	AAAMAB-	AA8AA	AA		C
	AAADAAB-	A88-AAÄ	AA		•••••
6/ 5. (3) INCURVARIINAE 7/ 5. (3) INCURVARIINAE	CARAS CDAEC	GREEK ACABE	A08 A- AC-AA A05 A: AC-AA	CABCA ACABC	ADCBA EAC
	800C848-	CBASA AACAA	AA		
8/ 6. (3) PRODOXINAE 9/ 6. (3) PRODOXINAE	C-0AB COSE-	A-B-CCB	AA	A	-0 A
	VAB-	A-B-AAA	AA	A	•••••
10/ 7. (3) ADELINAE 11/ 7. (3) ADELINAE	CAABB CGAEC	AACCC ABC-B BBADA BAOAA AACCC ABC-B	AGGAA CAAAA	AABCA ECABB	ACBBA AAACC
	-UU-G -FAB-	SBADA BADAA	AA	*****	••••
12/ 6. (3) HELIOZELIDAE 13/ 6. (3) HELIOZELIDAE	CBAA3 -44C-	AAABA A	AAA		-CBAAA-
	chim :	AAAAA ====A	*****		•••••
14/ % (1) TISCHEPITOAE	COAD CAAEB	COADA GA-AA	AA	A-EA-	-CBAAA-
15/ 10. (3) LYCHETIIDAE (LEUC)	CAAAAADA 80-60 -8488	C-B-A AAAAA	AAGGA AA-98	A-FAC	ACA
16/ 11. (3) LYCHETIIDAE	CAAMAE-	C-8-A VAAAA	AABBA AA-BB	A-BAC	ACA
17/ 11. (3) LYONETIIDAE	CAAAY AD- 80-80 -8EA8	C-B-A VAAAA	AAGBA AA-OB	THE ATENC	ACA
18/ 12. (3) LYONETIIDAE (GUCCU)	CA 8A3808 8D-6C -BE AA	COB-A AADAA	AABAA GA-68	A-8CA	ACA
19/ 13. (3) LYONETIIDAE (CED)	SAAA - EAS	cas-a aaaaa	AABBA BA-BB	-AG-A 4-80C	-CA
28/ 14. (3) OINOPHILIDAE	CAAAR CHRC	C-ABG AAAAR	-COB- CCAAA	BCA ABECA	BADAA CCACC
21/ 15. (5) GRACILARIIDAE (2ND INSTA	CAAM : 415			•••••	B-AA-
22/ 15. (5) GRACILARIIDAE (2NC INSTA	DAJAA -AA	A-0-AAA	AA	:: ::::::::::::::::::::::::::::::::::::	
23/ 16 (5) SPACILARIIDAE (LATE INST		A-8-AAĀ	AA		-CC-A -A
24/ 16. (5) GRACILARIIDAE (LATE INST	BA FAB -3A BC C EDA CAA AB EA	CBB-A AV-AA	***************************************	*****	-66-1 -1
25/ 16. (5) GRACILARIIDAE (LATE INST	BC2 -80 4- BBAAB -44	CBB-A AV-AA	AA		-cc-1 -1
26/ 16. (5) GRACILARIIDAE (LATE INST	COAAB -BA BC	CBB-A AV-AA	AA: ::::		166-1 13-1- 166-1 13-1-
28/ 16. (5) GRACILARIIDAE (LATE INST	CARAS -BA	COB-A AV-AA	AA: ::::A	******	:66:4 :3:::
	8CC -80 A-	CEB-V VA-VY	AA	•••••	••••
29/ 16. (5) GRACILARIIDAE (LATE INST	86643 -EA BCEOA- CBBAS -RA	CBB-A AV-AA	AA		
30/ 16. (5) GR/CILARIIDAE (LATE INST	6CC -8DA-	CBG-A AV-AA	AA: ::::4		-66-1 -3
31/ 17. (3) ACROLOPHINAE	6000- CSEAB	ce 158 82:59	ccss- ecc-c	CCB ec eca	-BBAA BCACC
32/ 10. (4) SCAPDIINAE	900C - 6-80C	C-AB- AAAAA	AAARA RAARR	-8686 88AB	AA -CAGC
33/ 19. (4) NEMAPOGONINAE	0-8 -88EC 8700C CEEAS	C-A8- AAAA	ACAA	8888	AA -GAGC
34/ 28. (4) TIMEINAE (D)		C-AB- AAAAA	AAABA BAABB	60086 £0686	AA -CACC
35/ 21. (4) TIPEINAE (X)	50500 C8EAS	C-AB- AAAAA	BARKS ABBAA	eeeee tetes	AA -CACC
36/ 22. (4) TIREINAE (T)		C-AB- AAAAA		EBE 88 60-08	AA -CACC
87/ 23 (4) PSYCHIDAE	84 E 88 -08E-	AAA-C ACS	BBCB- A-CAA	-A8CC 8-88C	A A EACCC
30/ 23. (4) PSYGHICAE	80000 BREAB 84683 -086- 80000 BEA8	C-A AABAA C-A AABAA	BBCB - A-CAA	-Aecc e-eec	AA EICCC
39/ 23. (4) PSYCHIDAE	BA 668 - D8 E -	AAA-C ACB C-A AABAA	BRCB - A-CAA	-ABCC 0-88C	AA EACCC
40/ 24 (1) OSCHENHEIMERIIDAE	BOAA3 CPAEC	BAACB A-A-B C-B AADAA	CBAB- ACBAA	-A-CA A-EBC	AAA CGC
41/ 25 (5) PLUTELLINAE (PR)	236-2 6A8	BA AC	-BBBCAB	CAB A-EAA A-BAA BBBB	A ACC
42/ 26. (5) PLUTELLINAF (PL)		BACCA ACC-3	CPBBC aG-AB		8A A3C
	SUUGI BELAA	U AAAAA	WWG REWES	-00-A 88688	•••••

43/ 27. (5) PLLTELLINAE (C)	-ABAB -CBEC BA BOOK BUEAA C-	CCA ACA-B C888C -88A9	CABCE ACECS BA ACC
44/ 28 (5) PLUTELLINAE (AB)	-A BAB -BBEC BA BDDC: BBEAA C-	CBA ACE -88BC -B-AB	CABCB A-8CB A ACC
45/ 2% (5) PLUTELLINAE (AC)		ACB AC-B -BBBC CAB	CARRC 4-8C8 A ACC
46/ 30. (5) ARGYRESTIINAE (A)			AABCB ABBCC BA AAACC
47/ 31. (5) ARGYRESTIINAE (Z)		A-B ACB-B -BC-C -B-AB ABA AAAAA AABBA BAABB BDA ABAAACAB	CB AEECA BAA BCACC
	BODC: CBEAB CB	ABA BAAAA AAGBA BAABB	AA88A 88888
46/ 32. (5) YPCNOMEUTINAE (A)	BOCCO CEEAB C-	BCB ABA-A CCBBC -B-AB BAAAA AABBA BBAB3	CAECA -EEBA CAA BCC
49/ 33. (5) YPCNOMEUTINAE	CABA - BEAS CA	-CA BAAAA AABBA GGABB	CARC AN BY AA BCC
50/ 34 (5) YPONOMEUTIMAE (L)	BBB -AAEC BA BDDCD BEEAB C-	AC 8 ABA - 8 CDA BC - 8-AB	CABA - ACABCAAC
\$1/ 35. (3) HELIODINIDAE	DA 898 CBBEC -A	AAC B	CA A-8-CAC
52/ 36. (3) EPERMENIIDAE	DA AAB C-BED BA	BBA ABG-B BCCB- ACCAA	8C8 A-8-CAACC8
53/ 36. (3) EPERMENTIDAE	DAAAB C-BED BA CCDCC BREA- C- DAAAB C-BED BA CCDCC BBEA- C-	BBA ABA-B BCCB- ACCAA	8C8 A-8-CAACC8 A88-A A8888AACC8 A88-A A8888AA
54/ 37. (5) SESTIDAE	O-EASAE- AS-	AL-3- 3-3-4 834	B-C A-C-CVVACC
55/ 37. (5) SESTIDAE		THE ACOUNT AND A CONTROL OF THE ACOUNT OF TH	1
94/ 34. (3) GLYPHIPTERYGIDAE	ADDOC BREAK C-	A AAAA AABA BAAAA	AABA EABBAC
57/ 36. (3) GLYPHIPTERYGIDAE	BOCOC BUEAA C-		AASSA SASS
50/ 39. (3) GLYPHIPTERYGIDAE (S)	SABAR SABAR SA	er telle elle ere	AABA AABB :A :C:
99/ 48. (3) GLYPHIPTERYGIDAE (P)	BOOC BEAS C-	-C - A-AAA AABA BAABA	AAGS SAGS ACC
60/ 41. (3) ELACHISTIDAE	BAM CRAE- C-	BAA AABAA AA: :::::	A
61/ 42. (5) COSHOPTERYGIDAE	CARAR -BAEC -A	41414 1 5146	AB2-A EEEOS ACC
62/ 43. (5) COSMOPTERYGIDAE (ANON)		AGG AGAGA GRAAG TABAR	BARCH ACCCCBAA AMCC
63/ 44 (5) COSHOPTERYGIDAE (TRI)		ACC ABABB CCOBABAC ABA ABABA AABBB BABBB	PERCH ACCCC 8-88A 86GAC
64/ 45. (5) CCSHOPTERYGIDAE (LIHH)		AB C ABAAA BCBAC B-CAB -BA ABAAA AABBA BAABB	BEBCB MACCC BAA MACAC
65/ 46. (3) HOMPHIDAE (CH)		A-A-EAA	C- AC AAAC-
66/ 47. (3) NOMPHICAE (MCM)		ABA AAAAA AA bb a baabb BCA A bab e C ob ac -C-Ab Aba aaaaa a ab e baabb	A-BC- ACEBC BAA -CACE
67/ 48. (3) MCMPHIDAE (HOM)		aba aaaaa aabb a baabb abc abaab ça bbb ab	
66/ 49. (3) NOPPHIDAE (BAT)	BODDC BBEAS C-	AC- ABAAA AABBA BAABB	ABBA EECO
	BODOC BAEAG C-	ABB AAAAA AASBA SAASS	-EBCS ASCCC SAA ACACC
69/ 50. (3) MALSHIIDAE (A)		AOB AAAAA AABBA AAABB	AAACB AACAC BAA ACB-C
70/ 51. (3) WALSHIIDAE		-CO AGAAB COODAD A-A AAAA AADA BAAGD	-8-C8 ACC-A 8AA AGACC
71/ 51. (3) WALSHIIDAE			
72/ 52. (6) COLEOPHORIDAE 73/ 52. (6) COLEOPHORIDAE	C-VAS -AAE- BA BDDC; BBAAB C- C-VAS -AAE- BA	ABA AAAAA AABBA BAABB	-88C8 ACA ACC
74/ 52. (6) CCLEOPHORICAE	COVE TARE SA	ABA AABAA AABBA BAABB	-Beco AC AC
75/ 53. (6) BLASTORASIDAE		AGA AĀDAĀ ĀĀBGĀ BAAGĢ A-C ABBBA CCBAA AB- ĀVĀĀĀ ĀĀĀ BĀĀBG	-86 68688 8CC
76/ 5% (7) DECEPHERIDAE (A)		AB- AVAAA AAA BAABB ACB ABA-B CCBBCBAB	ABBS BBSS
	BOCOC BEEAB C-	ABB ASAAA AABB- SASBS	AB68- 8A886
77/ 55 (7) GECOPHORIDAE (D)		ACC ABAAB CCBBCCAB ABB ABAAA AABB- BABBB	BA-CE ABCCC BCCAA SCACC
78/ 56 (7) OE COPHERIDAE		AC- ABA-8 CCB8- CC-AA AB- ABAAA AA BAABB	ABEB- EEEBB
79/ 57. (3) ETHMIIOAE	CACAB -08 EC GA	-C- AC-88 - 8CAA	AARCB ACCCC BC-AA CCC
00/ 54. (3) ETHHIIDAE (E. SEH)	CA 888 -DC EC 84	ACB ACAAB CCBBABAS ABB ABAAA AABB- BABBB	AABCB ACCCC BCCAA BCBCC
81/ 5% (3) ETHHIIDAE ((PYR)	CAR-B -UBEC BAR BODDC BEEAB C-	ACB ACABS CCBBC CCBAA -BA ABAAA AABB- BABBB	BA-CA ABCBC AAA CCACC
82/ 68. (9) GELECHIINAE (CH)		ABBC-AA	8CA A-C-C A A ACC
83/ 60. (9) GELECHIIDAE (CH)	BOD-C BHEA- C-	-BA ĀBAAV AA BĀBĒŠ ASBC-AA -BA ABEAV AA BABGB	EEEBA EEEBE 1:-66

85/ 6	1. (9)	GELECHIIDAE GELECHIIDAE	(ER)	800-3 800-3	BGEA-	8A C8A BA	VAAV	AA	BAABB	9CA 6688A 9CA PBB6A	ERRYR	AA AA	Acc
867 6	Z. (9)	GE LECHIIDAE	(AP)		BREA-	8A CBA	AADAV	AA	-C-AA BABB3	BCA A888-		AA	
		GELECHIIDAE GELECHIIDAE		800-C	BEEA-	84	AVAAV AVAAV		-C-AA BABBB -C-AA BABBB		A-C-C	AA	}ċċ
		STENON IDAE		80000	B-CE-	-8A-C C-A38	AS AA A	C-88-	BAABB	E868-	BAVBA	ACA	
90/6		STENOMIDAE STENOMIDAE		90000 -A 688	B-CE-	-8A-C C-A88 -8A-C	A88 A A	AA88- C-88-	CC-AA BAABB CC-AA	6888- 688- CA	AA-CC	¥::3	
		STENOMIDAE		80000	8-BE-	C-A88	AAAAA	AABB- C-88-	BAABB CC-AA	8688 -	BAVBA AB-CC	ACA	
93/ 6	5. (7)	CAFPOSINIDA	E	8000C C-8-8 BCD8C	-8EA- 8E- 86EA6	-446-	8-8-A 8-8-A	AABB- 88- AAA	BAABB GAABB	BCA	ABCBC	AV	-c-cc
94/6	6. (3)	ALUCITIDAE		C88 A8	C-AEC BEAG	#4CB8 C-88 A	****	AAA	ACOS- SAAOS	8-8CA -88-A	ACCCC	AC-AA	
95/ 6	7. (3)	DALGERIDAE				C-8-A		ĀĀ				-D-AA	
96/ 6	a. (4)	LIPACODIDAE				X-0		AA	••••		••••	-0	
97/ 6	% (3)	HYELAEIDAE			-DCEC	SAACB C-ABB	AB AAB ABA	C088-		8CA	ACCCC	ACCAA	
90/ 7	4. (5)	THYRIDIDAE		ec 83	-08 E-	88 A C-A8B	-ÇA-6	B-CB-				A	C-
		THYRIDIDAE		ec ဦးဦ	-0CE-	AA A	-CA-2	AA-CB-				A	
100/ 7		THYRIDIDAE		BCD80	BOE AB	C-A88 C-A88	AAAAA	B-CB-	A-	:::::		¥====	
1017	U. 17/	THYRIDIDAE		ac dao	BBEAB	€C A C - A8 B	- CA-B	AA	••••	••••	••••	•••••	••••
		PYPALIDAE		ec ove	VB E AB	ç	AVAAĂ	AA==A	E AABB	VBBBY	FRESY	AC-AA	
		PYRALIDAE PYRALIDAE		8C076	VRE AN	C	AVBAA	AAA	0 Å A 8 8 - C - Å Å 0 Å A 8 8 - C - Å Å	VBBBV	FOCOY	14-51	
		TORTRICIDAE		5v038	-8-EC -8E AB	C	AVDAÄ	AAA	CC-AS SAAVS		6868A	AC-AV	• • • • •
		PHALONIIDAE				C- A8 A	AVAAV					AAV	
		PHALONITOAE		C-EAS	-6 E A B	C8 A	AV AAŸ	Ba-	-CCAA	-88CA -88CA	ACC-C	1::14	
100/ 7	3. (5)	PHALCHIIDAE		50000	-8E A8	C8 A	AVAAY	AA8	PAAAA	488-A -88CA E8680	ACC-C	¥==¥\$	
109/ 7	3. (5)	PHAL CHIIDAE		BODCS BOCCS	-AEAB	CBA	ŧ	11 BB-	BĂĂBS - CCAA BAABB	-88CA	ACC-C	A4 ¥	cc
		PHALONIIDAE		6 58€	-BE AB	C8 A	āŢĀĀ Ð	AAB	59911	ABB-A	466°c	A44	CC
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-		COSSIDAE		848484 848484	BE A	-C -B -	ACB	AA - A	BA -88	-66-6	4-C-C	A	
-		COSSIDAE		C-B-B	-8E A-	-0-8-	17.55	AAA	8A-85	8	A-C-C	<u> </u>	-6-66
116/ 7	5. (4)	COSSIDAE		-008	-BEA- -BEA-	C	AÇ B	8	8A-88 8A-88	-86-8 8-8	68888 A-C-C	A	-ċ-ċċ
117/ 7	6. (3)	DREPANI DAE		CAP-3	EC	g							
110/ 7	7. (4)	BREPHINAE							CECAC				
119/ 7	6 (3)	MIPALLONIDA	E						8A				
									8AV88				
124/ 7	9. (7)	NCTCOONTIOA	E	BC 0	BSEV-	V88	A-AAR AAEVA	808	AA	•••••	AA	64898	GESVB
121/ 0	6. (5)	AGARISTIDAE		aA E -8 BC DBC	-D-EC B3EA-	9A	1244X	B	BBAAA		ACECA		
		NOCTUINAE		BC C BC	-D-EC	8B	ACA-V AVEVA	VAV	CAV		A-V-A	84888	94249
		NOCTUIDAE		8C035	BUCA-	g8	AGEVA	VAV	CAV	CÀ	A-V-A	AAA BABAB AAA BABAB	8-3XE
124/ 8	1. (9)	NCC TUI DA E		A C D BC	BUEA-	çe	AGATX	VAV	CAV	CA	*-A-Y	84868	BAEB

FAMILY KEY TO LEPIDOPTERA LAFVAE (C)

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•	.18.20	11111	C-ABA	A-8	:: ;;	8A	8 0-808	 	83 C	90 CE-88	80 C-888	9	90 C0	88	C- ABB	C-ABB	MA MG	3-AE- C-ABB	A 4	3-14E-C	8C-AA C-A88			000
	• 16	67890	-8E AA	-8 A 8-	E-	-68EC -6EAA	-DCEC -6EA8	-0-E-	C-CEC	-DBEC	-0 CEC	-00E-	-VCEC	-DCEC	-DC EC	-9cE-	P)CF	- PCFFC	() (10 - 10 - 10 - 10 - 10 - 10 - 10 - 10 -	-0-FC	9	0 - 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 - 0 0 - 0 1 - 0 1 - 0 1 - 0 1 - 0
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EV 10	NOANC		(3) ZYGAE	(4) LIPAC	3	(S) P	d (6)	6	3 8	(S) A	1 (6)	(8)	(7) C	(2) N	(7)	2 (2)		33	(6)	(6) 4	3	E (6)	(6)	ž 6
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FAPILY KEY TO LEPI	TAXA, A9 UNDANCE		7	72	3/	3	2/	9	2	8	6	10/	11/	12/	13/	14/ 14	15/ 15.	16/ 15.	17/ 16.	18/	14/	201	21/	122

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AND CHARACTER												(AIGEGOMY)			
ABUNDANCE INDICES, A		SCYTHRIJAE	COSSIDAE	COSSIDAE	COSSIDAE	COSSIDAE	(4) THVATIRIDAE	GEOMETRIDAE	EPIPLEMIDAE	(7) NOTODONTIUAE	DOA	LITH CSII NAE	NOCTUIDAE	NOCTUTORE	
NOAN		(3)	(*)	3	3	3	(†)	(6)	(3)	2	3	6)	(6)	5 3	121
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TAXA.		7	72	3/	;	21	/9	2	8	6	701	11/	12/	13/	;

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FAMILY REV TO LEFICORTEMA LA-MAT (A)
PROGRAM KEY. MUP AT.18.39 CH 17/29/76. STURAGE PERUIPPO - 12379 MORDS.
CHARACTERS - 10. WEAD, BU MASKID, 6: USED IN KEY.
ITEMS - 124 READ, 124 MASKID, 142 APPEAR IN CEVI 6: TAXA,
PRASE = 6.00, ALASE = 7.01, PLUSE = 1.01, VARYME = 1.00., NUONE = 3
PRESET 2 HAPACTERS - COLUMNIGHOUP, CHAPACTER 1: 1: 61
MEY INCOMPLETE, MORE INFORMATION NO OF
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INOCTUTO AF	[+ 1C]	574	+ 73	25.0	1 3 3 1	1																
RP EPHIN AE	1016	576	4 79	126	1 1 3																	
PIPALLINIDAE	1610	570	£ 78	520	5 9																	
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EARILY REX TO FEBIDOBLESA FASAS 161
PROGRAM KEY. RUN AT. 15.47 ON 17/29/76. STORAGE REQUIRED - 3374 HOPPIS.
CHARACTERS - 10. READ. 99 MASKED. 25 USED IN KEY.
ITEMS - 22 READ. 22 MASKED. -1 APPEAR IN KEY: 19 TAXA.
RBASE = 4.GC. ABASE = 3.UL. REUSE = 1.U1, VARYNT = 1.U. . NOONE = 7
PRESET CHARACTERS - COLUMN, SPOUP, CHARACTER
CHARACTER MASK -
KEY INCOMPLETE. MORE INFORMATION NEEDED.
LASIOCAMPIDAE | 15921-661634 6781784 10019941724 204193 1253 1773
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IDANATINAE		Fie Haio Adit Tail 101 F Bai 15 d T. dai
DANAII NAE	elfac.cs:	: ‡6 44 \$6 74 \$7 5 4 \$ 1 3 C \$5 9 4 \$ 7 2 3 \$ 7 4 8 \$ 2 2 4 \$
PAPILI ONI DAE	#\$29£\$18£	E\$63A\$678\$71A\$17C\$59A\$723\$748\$2.3\$
DREPANIDAE	•\$59£3n8£	E \$6 3A \$67E \$70A \$10C \$593\$
IPIERI DAE	•มิวฮอมียฮค	FÎ63AÎ67BÎ7CBÎ
NOCTUI DAE	1545 × 85	£ \$23\$67A\$39A\$46R\$
INOTODONTIDAE	• 138± 1€ 95	: Î693 Î674 Î99A Î46C Î
NOTODONTIDAE	29 E . T d£	<u>[[693]674]993</u>]
INOTODONTIDAE	•รุ๋งฮยรุ๋ยศ	FÎE 33ÎE 74Î 390Î
INOCTUIDA:	• 54L 05L	_1633676173A1
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PAPILIONIDA -	•1:461·46	ele 311673173317231

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FAMILY K-Y TO LEPIGUPT -A LA-VUE (C)
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PROSHAM KEY. KUN AT.14.44 ON .7/29/75. STUPAGE REQUIRED - 1918 ADROS.

CHAPACTER - 12 FEM 1. FOR 12 MACKET . 12 AFPEAR IN KEY! IN TAXA.

RBASE = 4.0 . AGASE = 3. . P USE = VIRYNT = 1.1.1. NOONE = 3

PRESET CHARACTERS - COLUMN, PROCESCHARACTER 1. 25 95

CHAPACTER MASK -

KEY INCOMPLETE, MORE INFORMATION MEEDID.

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INCCTUIÚA-	544	• !						
INOCTUIDAE	550	υ 9 Α	67A	73A	•			
INOCTUIDAL	<u>ا</u> نادز <u>:</u>	094	67A	739	2.A	:		
INOCTUIDA-	536	694	674	733	208	:		
INOLIDAE	i i うりし	694	674	734	5.8			
INOCTUIDAE	ょうれい	69A	673	• ·	• • • • •	•		
INOCTUIDA:	550	. 98		•				
IPTEROPHORIJAL	\$ 54E	274	6 B	i				
IZYGAEN 1046	556	674	LIE	59A	65A			
LYMANTHILDA =	7 55.	574	635	594	65B	31 A	I G A	
ARCTIIDA: =	1 540	c 7 A	6 8 E	39A	658	9: 4	1. 31	
NOTOGONTIDA:	3 3 4 5	074	6.15	294	658	913	9.A	
INOCTUILA:	5 5 5 €	074	635	59A	658	9:9	92A	į
PERICOPIDAL =	5 5 5	.74	E DE	94	6 58	9:3	91.B	
INOCTUIDAL	į 5 g-	₹7#	6 d F	1 591	39≜	594	2 4	:
INOTOPONT:DA-	51	+74	6 3 5	593	994	59A	2.B	í
I NOC TUI DAL	551	674	68E	,93	994	59A	5.81	:
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INOTOGOTIOA_	į fai	674	685	3 9 3	998	İ		
PROTUNOCOLON	. इ.च.ह. इ.च.ह.	674	6 3 E	593	990	Ī		
IARCTIIDAE =	34	674	6 3 F	124	Ī			
ICTENUCHIJAE	Ī 554.	574	f 3F	323	914			
PERICOPIDA: =	į 50:	674	6 4F	323	918			
ROCTUIGAL	5 3 5	676	694	394	7 3A	•	, 4	
LASIOCAMPIDAE	Î ララレ	57 3	6 9 A	374	738	724	314	9231
INOCTUIDA:	i 5a_;	€ 7 €.	69A	994	738	72 A	3:9	92 A İ
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ISATURNII 'AE		58:	676	699	993	944	934	739	į	
CITHERONIIMAE			6 7 0	698	#5 9	344	93A			
INYMPHALIMA	•	546	+73	698	983	949	34.7			
NYMPHALINAL		54.	-7	643	983	948	36-3	93A	2.A	
ICITHERON119AE	•	54-	07 0	6 39	+33	34B	۶é٦	93A	219	į
RIODININAE		أعاد	37c	693	∍ •3	944	963	1:4	72A	95 A I
NOCTUICAE	•	542	57t	633	393	9 4 B	951	939	72A	95 8 į
INVMPHALINAF	•	515	6711	698	983	340	363	934	72A	958
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-EUMILY REXTIO FEBIDOBIESO FURNAE 105
PROGRAM KEY. RUN AT.18.49 ON .7/29/76. STORAGE REQUIRED - 2968 HORDS.
CHARACTERS - 10. READ, 99 MASKED, 18 USED IN KEY.
ITEMS - 14 READ, 14 MASKED, 27 APPEAR IN KEY! 9 TAXA,
RBASE = 4.00, ABASE = 3.00, REUSE = 1.61, VIRYHT = 1.000, NCONF = 3
PRESET CHARACTERS - COLUMN, SROUP, CHARACTER
CHARACTER HASK -
•••••
NOCTUIDAE
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PROGRAP KEY. RUN AT.10.39 CH [7/29/76. STOPAGE PEQUIPED - 12379 NORDS.
CHARACTIRS - 160 MEAD, OC MASKID, OJ USED IN KEV.
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64. (63)	SETA LI CN AECO CISTANCE BETH SETA C1 CLCCE AND CNLV A LI SETA LI ON AEOC FINACULUM. DI	MINAL SEG. TETAL LE AND NO TO OUR THAN THE CAUDAD HINAL SEG. 10	# FRY CLOSE FAREST LA IN TO COSE IF ANY: S NOT ASSOC	TO SW SETAL STA DE ON AE STA DE ON AE STATES HITH SW	OR ON SAME PINACU CE SET WEEK TO RECTLY CE SEG NEVER ON SA VE LESS THAN ON OT ARLY BETMEN OCS	LUM: SV: FEAD ABOVE DI ME	PLUTELLINAE (PR)
		SETA OF ON AS	O SVI HEAD S D. SEG. 9 AB	OVE AND CAL	I ISTANT OR NO UDAD OF DIE SI	ARLY BETHEEN OCZ SETAE ON ABO. SE	ANO CÇ 31 G. 9 2	ARGVRESTIINAS (2)
65. (63)	SPIRACLES SHAPE SP101-SD2 ; STRAIGHT LINE SPIRACLES SHAPE SD1 AAGLE SPIRACLES SPIRACLES SPIRACLES SHAPE	POUND: SETA J-90 D.: L S , CR MEARLY FLLIPTICAL -501-502 G.	L SDZ ON THE SETAL ON ABI (176-19: 0 OR EGG SHAI T. 36: L SI	OPACIC SEG. I D. SEG. 9, 31 PLOI SETA SOZ ETAE DA ABO.	BEHIND SDI, ANGLE OCELLI 3,4,5, ARR ON THORACIC SEG SEG. 9, 21 OCELLI	ANGEC IN A	GLYFFIPTERYGIDAE (P) HEPIALIDAE
66. (29)	SW SEIAE ON THO	RACIC SEGMEN	TS, 3 ON T	I, 1 OM TII A	NO TIII		#1
67. (66)					SHIELD HOME PRAKE		SELECHTICAE CAM
66. (67)	THORACIO SEC.	DAL PROLEGS	01 VID ED IN	TO THE GROUPS	, BROKEM SERIESI S	ETA L1 04 281251'5274	SELECHIIOAE (BR)
69. (SETA SD ON AGCOUNT CITY OF AGT	MINAL SEG. 3 STANGE RELIE VERV LONG, 3 STANGE SEG. 9 LEN SO AND O LEN	VEW CLOSE EN TO AMO AT LEAST WI FAR FROM I LLESS THAN LY LONG, LE	E TO AT LEAST TO AT DISTANCE TO GET THAN ANY L SETA, HI N AT DISTANCE ESS THAN AT A	STAFF L SETAFF CHENTER ON SAME PINAC BETWEN SO AND LEST ON LES	OR ON SAME	GLYPHIPTERYGIDAE (S)
70. (69)	FROM T FRTENDS TO	O THE COPCNA	L NOT CH AL	PORE HOOK SH.	MEARLY! CLAM ON T MEDT PROLEGS LESS OR MEARLY BETHEEM "CLAM ON THORACIO EGS ELONGATE S THAN TO OCC	ZA KA ARHT	COLEOPHORIDAT PLUTELLINAE (AB)
71. (66)	L GP. ON THOPAC CA ANG SO SET COFLET 1-8-1-1 AJ EQUIDISTAN L GP. ON THOPAC CONTINUOUS CO 3/41 OCELLI 3	IC SEGON DI AGRANGED IN T CR NEARLY IC SEGON DE IC SEG	STINGTLY STANDS TO THE STREET AZ	EPAPATED FROM ME CORONAL NO T LIME, OR NE AND LI SO SETAE LOCA EVERNOS TO THE NGLE G.T. 190	GERYICAL SHIELD A TRIV (173-196 0:) I FEO ON ONE OSSIING COMONAL NOICH FE D.I HEAD SETA AS	MO FROT XD.	TPONCHEUTINEE (L) PSYCHIOAE
72. (291	OCCLLI THREE TO	FIVE PRESEN	TI SEIA LL	ON 480. SEG.	3 CLOSER TO SP. T TO LE THAM TO SP	MAN 10 LZ	TACHELLIOVE (BACCA)
73. (72)							SESTIONE SESTIONE
74. (73)	MEAD SFTA OL CU	TSTÁL GÍÁCLE OKSPICUOUSLY	OF EVES	FROM OCELLI	L-4. DISTANCE DET	EEN CC4 AND	SESTIME
		STRATGHT LINE SOIL ANGLE S OCCULING S CONSP	THE DISTANC -S01-S02 10 ICUDUSLY SER L.T. FOUAL T	0 0 1 2 1 5 6 1 6 6 1 6 6 1 6 6 1 6 6 1 6 6 1 6 6 1 6 6 1 6		DISTANCE BETWEEN COLORS SEE TO	OCS AND OCS	75
75. (7~1	ANGLE SPSOL	-537 G.T. 96	A18 PROLEG	RELOW CAUDAL	SHIFLD HIME OR LE	221	S ES I LOAE
. ,, ,	. •••	CROCHETS ON C	AUCAL PROLEG	\$ 4.0 0 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1	DED ENTO THO: TANT BETHEEN U "ELOW GAUDAL	SHIELD HINE OR LE GROUPS, CONTINUOUS ZANO LO MORE THÂN PS, BROKEN SERIESI	i Sérceso Alike 1	COLECPHORIDAE
		THORACIC SEG.	AUCAL PROLEG	O ES THAN	TO LI	PS, GROKEN SERIEST	SETA LI ON	SELECHIIDAE (AP)
76. (19)	LAERAL NOTCH SH	ALLOH, FXTEN	ALLE SIDE	OHE THAN 172	THE DISTANCE TO TH TO BASE OF LABRUP D BASE OF LABRUM S	OR NEARLY	NOTOCONTINA F
77. (76.)	PRESENT		F AND CAUD		• • • • • • • • • • • • • • • • • • • •	•••••	NOTCCONTIDA E
77. (76)	SETA DZ CN AGD. SETA DZ CN APD. CR OFLOM	366: 1 699Y	•••••	• • • • • • • • • • • • • • • • • • • •	. 94C4. 27 1544C4. 34.	•••••	NOTCCONTIDAE NOCTLIDAE NOCOUTON
77. (76)	SETA DE CH ARD. SETA DE CH ARD. L SETAF Sh IMES	₹{8: 1 6997 1818 ₹{8: 1:	\$:::::::	• • • • • • • • • • • • • • • • • • • •	· · · · · · · · · · · · · · · · · · ·		MOCTLIDAE
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77. (78. (79. (76)	SETA BY EN APOLICE BY LOW LINE APOLICE BY LOW LINE APOLICE BY	SEE: 9 6855 ACTO SEE: 1: ACTO	S, MEAD DE	PESSED, MORI 30071671175013 	CONTAL , ROUTH PART POUTH PARTS LOT POUTH PARTS LOT SUPS CONTINUOUS S	s otrected of Wicker	NOCTLIDAE NOTOCONTIDAE 68 GRACILARIIDAF OLITE INST NOCILLOSIONF STRALOSIONF STRALOSIONF STRALOSIONF STRALOSIONF STRALOSIONF
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PAMILY MEY TO LEPIOOPTEMA LARVAE (8)
PROGRAM KEY, RUN AT-19.47 DM . 7/29/74. STORAGE REQUIRED - 3974 HORDS.
CHARACTERS - 184 REAL MASKED, -1 APPEAR IN REVIEW IN TAXA.
RRASE . .... ABASE . B.... REUSE . 1.... VIRYMY . 1.... HONF . 3
PRESET CHARACTERS - COLUMN, SROUP, CHARACTER
CHARACTER MASK -
MEY INCOMPLETE. MORE INFORMATION NEEDED.
     1) CROCHET, ABSENT ON ALL AGOOMINAL SEGMENTS! POLEGS OF ABO, SEG. 18 49ENT OR FOR MENTALY DR AT LEAST CONSIDERING STALLER THAN POLEGS ON A 1-6. LINACODIDAE CROCHETS PLACED ON ABC. SEG. 13 FALSENT OR LEAST 1 OTHER ABOOMINAL SEGMENT FRIPTROPIBA
                                                                                                                                                                                                         HESPERIIDAE
                    S) ANAL COME ASSETT! POSITION OF SP. ON ABO. SEG. 8 IN LIME NITH SP. ON PRECEEDING AND COME PROCEEDING AND COME PROCEEDING AND COME PROCEEDING. LIBYTMEIDAE AND COME PROCEEDING AND COME PROCEEDING AND COME PROCEEDING AND COME PROCEEDING AND COME PROCEEDING. MESPERIIDAE
                    PROLEGE ON 180: 356. 1: RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RESERT 64-ADDIMENTARY 34-RY-CERSY CONSIDERAGES ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 3-E RY-CERSY CONSIDERAGE ON A 
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   12. ( 11) LABRAL NOTCH SHALLON, EXTENDING NOT NOTE THAN 1/2 THE DISTANCE TO THE BASE OF LABRAL NOTCH DISTANCE TO THE BASE OF LABRAL NOTCH DISTANCE TO BASE OF LABRAL NOTCH DISTANCE TO BASE OF LABRAN SOMETIMES NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PRESENT NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHOOSET THE PROPERTY NOTCHO
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                    9) ANAL COMB PRESENT
    ..... OREPAMIBAE
    17. 4 16) FILAMENTS OF OSMETERIA AREENT.
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    10. ( 17) EVERSIBLE CLANS ON VENTRAL SIB: 87 F1 BETWEEN LEGS RESERVE.....
                          MOST OR ALL SETAE ON ELEVATED CHALAZAE OR TUBERCLES......
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                           20 - 4 100 ABO: BEG: 8 MITHOUT LEAST BORSE DORSEL MOTHS SCOLUS CHALAZA PUBERCLE POR SCAR. SCAR.
           4 28 AMAL CLATE ON ABD. SEC. 14 BIFIRCATE AT TIP, STARING THE DISTINCT PROCESSESS

TOTAL START OF ANY TAMOR ESTABLES OF THE ARD AS CONSTITUTED THE BEST THAN
                               DIRYOLD THTO "AAN" ARMALETST SPIRICLES TON "TI AND THE CORSPICUOUSLY LIRGER THAN NAL PLANE OF THE CORONAL NOTHING OF BOOK DIVIDED THAT A TO BE ANNULTED SPIRACLES NOTHING SAME SIZE AND SPIRACLES.
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   22. ( 28) RON OF CHONETS ON VENTAL SID: OF PROLESS INTERPUTED ON REDUCED IN SIZE MEAR
RON OF CACCHETS ON VENTAL SID: OF PROLESS INTERPUTED ON REDUCED IN SIZE MEAR
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APATELOGIDAE
SPHIMEIDAE
MOCTUIDAE
LASIOCAMPIDAE
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          1 26) LABRAL HOTCH STALLOW, EXTENDING NOT MORE THAN 1/2 THE DISTANCE TO THE BASE OF LABRAL HOTCH ASSTRANCE TO THE BASE OF LABRAN THE STALL HOTCH ASSTRANCE TO SAFE OF LABRUM SORTIMES HOTCHOOMYTORE PRESENT
   20. ( 27) SETA 85 SN A38: SEE: $ 375454772 CANOS -05, 031 RAG -3409 SCIERATED -19 LAST - RAGISE - MOTUDONE MOTODONIES
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   30. 6 29) FILAMENTS OF GSMITICAL ASSISTA
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FAMILY KLY TO LEPINIPT: A LA-VAE (C)
PROGRAM K: V. WUN AT. 19.48 Ov . 7/29/79. STURAGE FFRUIRER - 2918 MORRIS.
CHARACTERS - 13 FLAD. 56 MASK D. 18 USCO IN KEY.
ITEMS - 62 MEAR. 62 MASK: . - 2 APPEAR IN KEY! IF TAKA.
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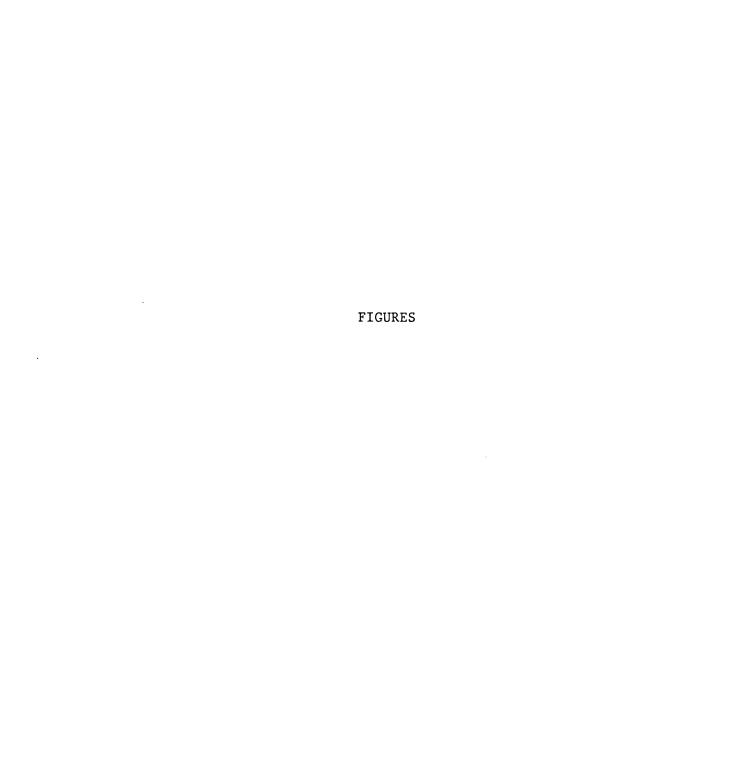
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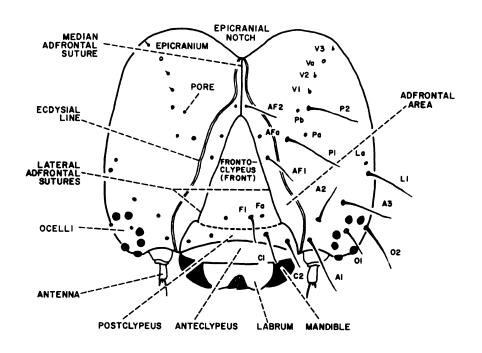
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- Fig. 1. Generalized head capsule of Lepidoptera larva, frontal aspect.
- Fig. 2. Generalized head capsule of Lepidoptera larva, lateral aspect and ocelli numbering system.



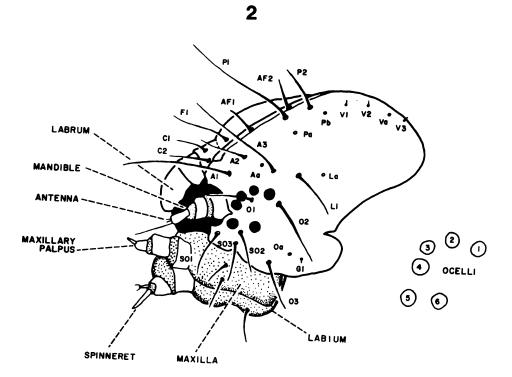
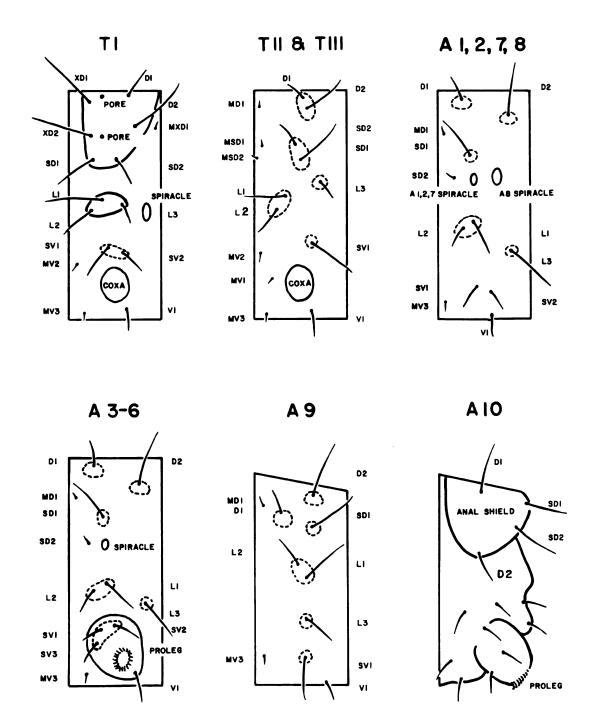
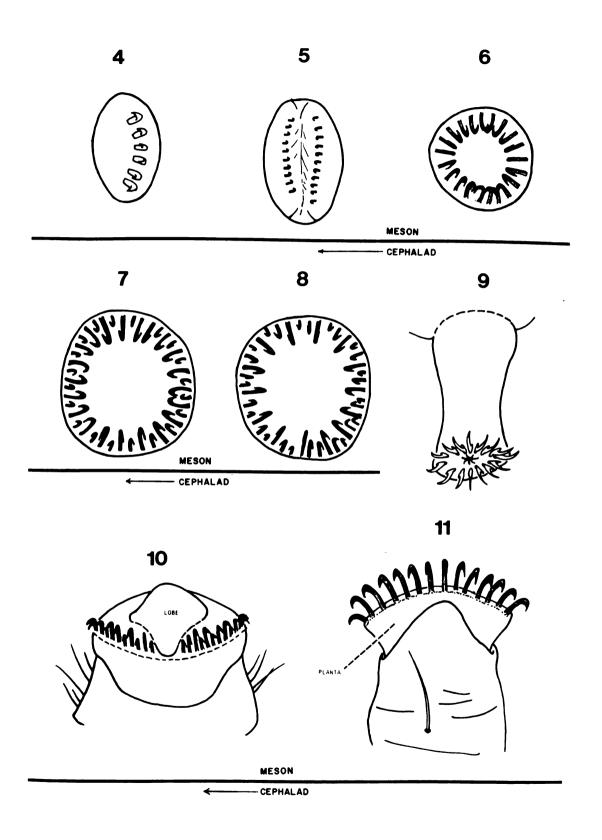


Fig. 3 Generalized setal map of Lepidoptera larva and chaetotaxy of Hinton (1946).



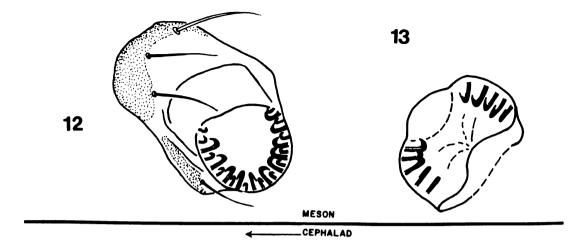
Figures 4 - 11. Prolegs and crochet arrangments found in Lepidoptera larvae.

- Fig. 4. Single transverse band.
- Fig. 5. Two transverse bands.
- Fig. 6. Uniordinal complete circle.
- Fig. 7. Biordinal complete circle.
- Fig. 8. Triordinal complete circle.
- Fig. 9. Elongate ventral proleg as in many Plutellinae, Pterophor-idae.
- Fig. 10. Lycaenidae, <u>Brephidium</u> <u>exilis</u> Boisd. (USNM). Ventral proleg.
- Fig. 11. Noctuidae, Polia sp. Ventral proleg, mesoseries arrangment.



Figures 12-14. Crochet arrangments.

- Fig. 12. Biordinal mesopenellipse on a ventral proleg.
- Fig. 13. Stenomidae, <u>Setiostoma xanthobasis</u> (coll. Habeck). Caudal proleg.
- Fig. 14. Drepanidae, <u>Drepana sp.</u> (USNM). Ventral proleg, biordinal mesoseries plus uniordinal lateroseries.
- Fig. 15. Theoretical information content of taxa at the family level.
 - A. A homogeneous family.
 - B. A heterogeneous family.



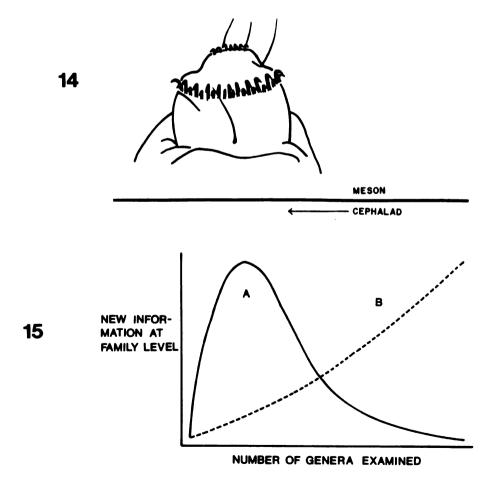
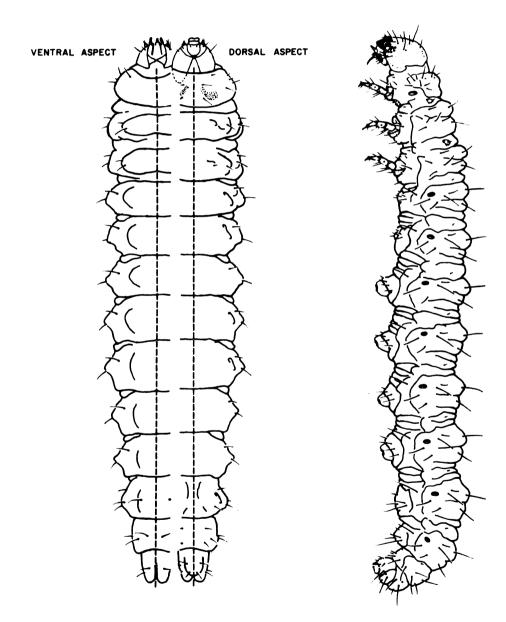


Fig. 16. Eriocraniidae, Mnemonica sp. (USNM). Dorsal and ventral aspects.

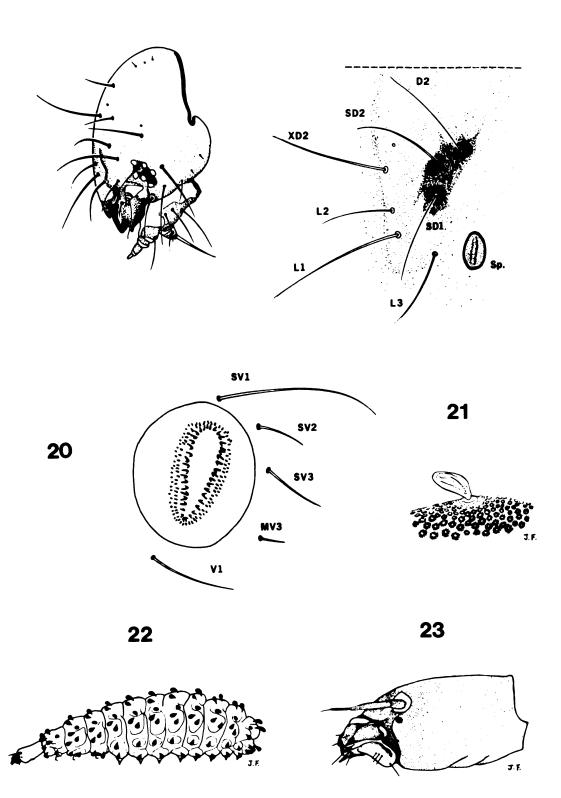
Fig. 17. Hepialidae, Dalaca sp. (USNM). Left lateral aspect.



Figures 18-23. Hepialidae and Micropterygidae.

- Fig. 18. Hepialidae, Dalaca sp. (USNM). Head lateral aspect.
- Fig. 19. Dalaca sp. Prothoracic shield, lower half of left lateral aspect.
- Fig. 20. <u>Dalaca sp.</u> Right ventral proleg showing multiserial crochet arrangment.
- Fig. 21. Micropterygidae, Micropteryx sp. (England, USNM). A body seta, detail.
- Fig. 22. Micropteryx sp. Left lateral aspect.
- Fig. 23. Micropteryx sp. Head, left lateral aspect.

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Figures 24-27. Psychidae and Nepticulidae.

- Fig. 24. Psychidae, Solenobia walshella (Clemens) (USNM). Lateral aspect.
- Fig. 25. S. walshella. Head, frontal aspect.
- Fig. 26. S. walshella. Prothoracic shield, left lateral aspect.
- Fig. 27. Nepticulidae, Ectoedemia populella Busck (USNM). Lateral aspect (see Fig. 69 for head).

Figures 28-31. Cosmopterygidae and Yponomeutidae.

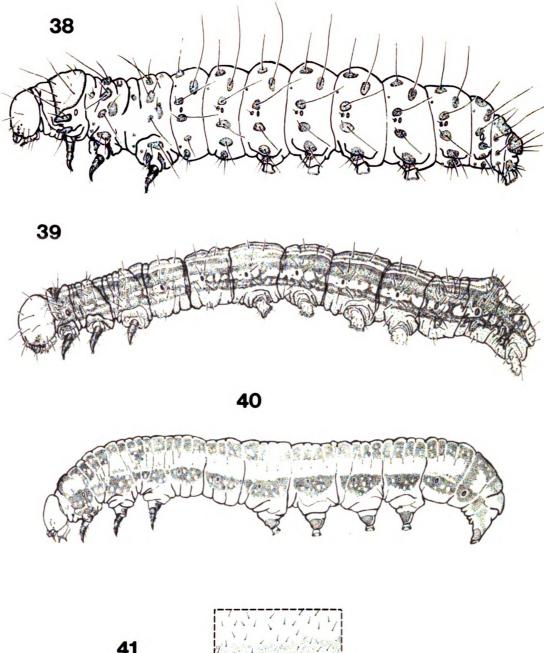
- Fig. 28. Cosmopterygidae, <u>Limnaecia</u> <u>phragmitella</u> Stainton (USNM). Head, left lateral aspect, detail; ocelli and cranial setae in immediate vicinity.
- Fig. 29. L. phragmitella. Lateral aspect.
- Fig. 30. Yponomeutidae, Zelleria pyri Clarke (USNM). Left ventral proleg, multiserial crochet arrangment.
- Fig. 31. Z. pyri. Lateral aspect.

Figures 32-37. Blastobasidae, Blastodacnidae.

- Fig. 32. Blastobasidae, Valentinia glanduella Riley (USNM). Head, TI, & TII, lateral aspect.
- Fig. 33. V. glanduella. A3, left lateral aspect.
- Fig. 34. \overline{V} . glanduella. A8, A9, & A10, left lateral aspect. Fig. 35. \overline{V} . glanduella. Head, left lateral aspect, detail; ocelli and cranial setae in immediate vicinity.
- Fig. 36. Blastodacnidae, Aetia bipunctella (Chambers) (USNM). Head, TI & TII, lateral aspect. Fig. 37. A. bipunctelia. A3 left lateral aspect.

Figures 38-41. Thyrididae, Dioptidae, Libytheidae.

- Fig. 38. Thyrididae, <u>Dysodia granulata</u> (Neumoegen) (USNM). Lateral aspect.
- Fig. 39. Dioptidae, Phryganidia californica (Packard) (USNM).
- Fig. 40. Libytheidae, Libythea bachmani (Kirtland) (USNM).
- Fig. 41. L. bachmani. Detail od spiracle and setae, A3 left lateral aspect.





Figures 42-44. Epipyropidae, Mimallonidae.

Fig. 42. Epipyropidae, Epipyrops sp. (USNM). Ventral aspect.

Fig. 43. Mimallomidae, <u>Cicinnus melsheimeri</u> (Harris) (USNM). Fig. 44. <u>C. melsheimeri</u>. Right ventral proleg.

Figures 45-49. Hesperiidae, Adopaea lineola Ochsenheimer (MSU).

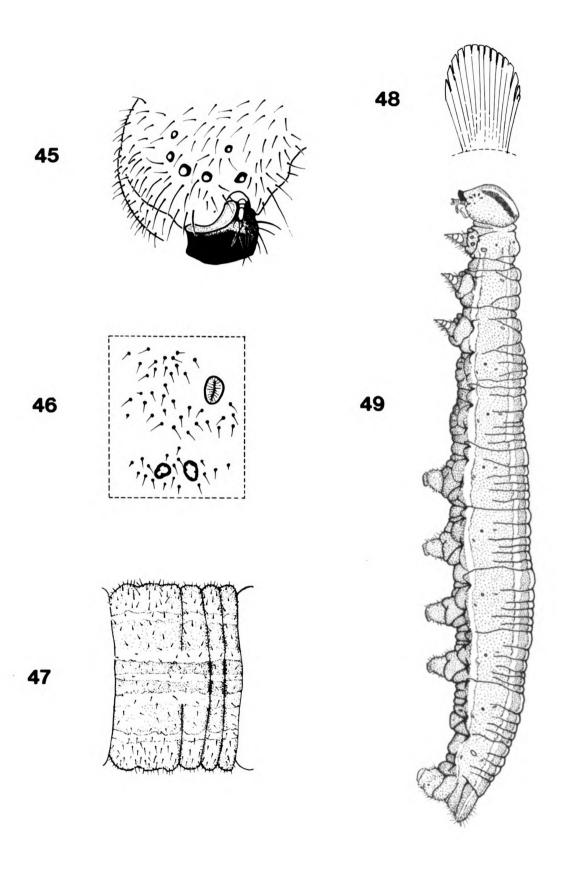
Fig. 45. Head, left lateral aspect.

Fig. 46. TI detail around spiracle, left lateral aspect.

Fig. 47. A3 dorsal aspect.

Fig. 48. Anal comb.

Fig. 49. Lateral aspect.



Figures 50-54. Satyrinae, Cercyonis pegala (=alope) (Fab.) (USNM).

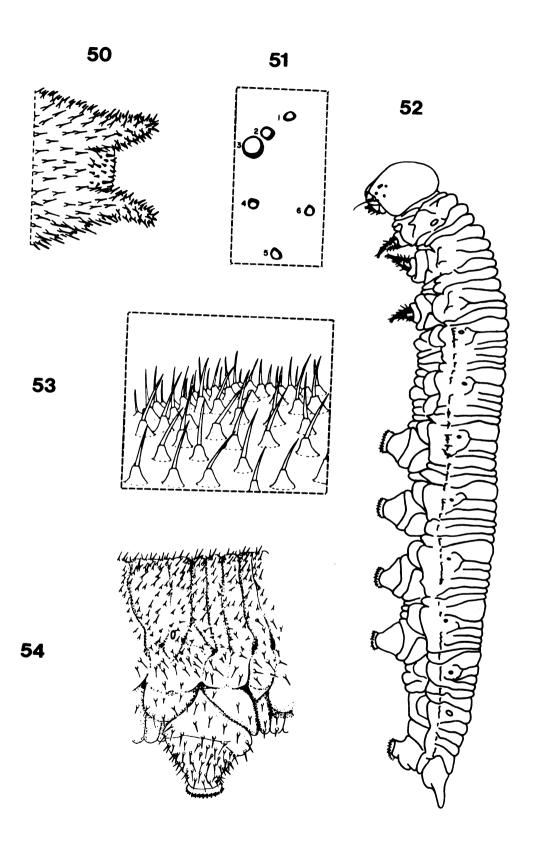
Fig. 50. AlO dorsal aspect, bifurcate anal plate.

Fig. 51. Detail of head, left lateral aspect, ocelli.

Fig. 52. Larva, lateral aspect.

Fig. 53. Detail of cuticle showing secondary setae on chalazae.

Fig. 54. A3, left lateral aspect.

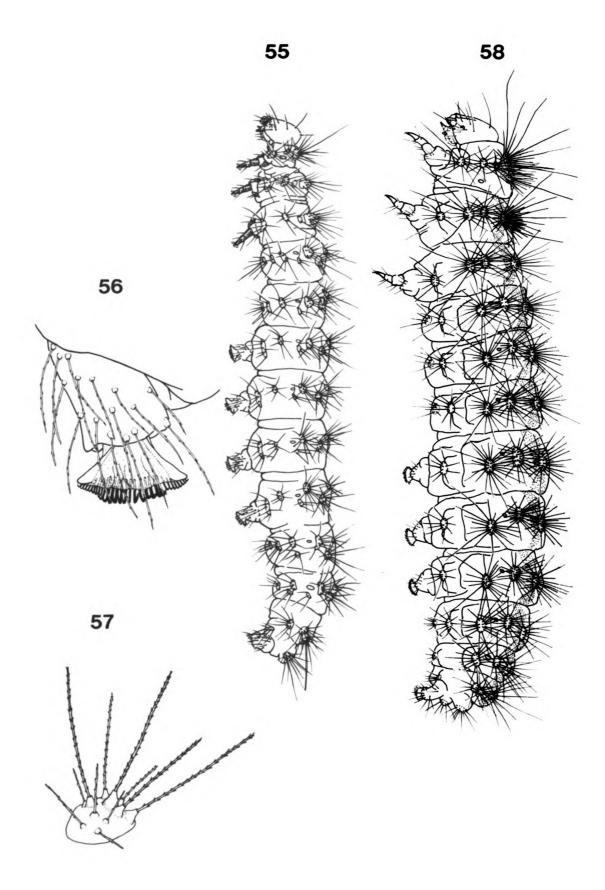


Figures 55-58. Pericopidae, Nolidae.

Fig. 55. $\underline{\text{Composia}}$ $\underline{\text{fidelissima}}$ $\underline{\text{Herrich-Schaeffer}}$. Fig. 56. $\underline{\text{C.}}$ $\underline{\text{fidelissima}}$ $\underline{\text{Ventral proleg, lateral aspect; crochets}}$ arranged in a heteroideous mesoseries.

Fig. 57. C. fidelissima. Enlarged verruca.

Fig. 58. Nolidae, Celama sorghiella Riley (USNM).



Figures 59-66. Tischeriidae, Heliozelidae, Lyonetiidae

- Fig. 59. Tischeriidae, <u>Tischeria</u> <u>malifoliella</u> (Clemens) (USNM). Head, dorsal aspect.
- Fig. 60. T. malifoliella. Ocelli, ocellar setae, and outline of anterior part of head.
- Fig. 61. Heliozelidae, <u>Coptodisca</u> <u>splendoriferella</u> <u>Clemens</u> (USNM). Head, dorsal aspect.
- Fig. 62. <u>C. splendoriferella</u>. Ocelli, ocellar setae, and outline of anterior part of head.
- Fig. 63. Lyonetiidae, <u>Proleucoptera smilaciella</u> Busck (USNM). Head, dorsal aspect.
- Fig. 64. P. smilaciella. Ocelli, ocellar setae, and outline of anterior part of head.
- Fig. 65. Lyonetiidae, <u>Paraleucoptera albella</u> (Chambers) (USNM). Head, dorsal aspect.
- Fig. 66. P. <u>albella</u>. Ocelli, ocellar setae, and outline of anterior part of head.

Figures 67-72. Nepticulidae, Eriocraniidae, Gracillariidae, Opostegidae.

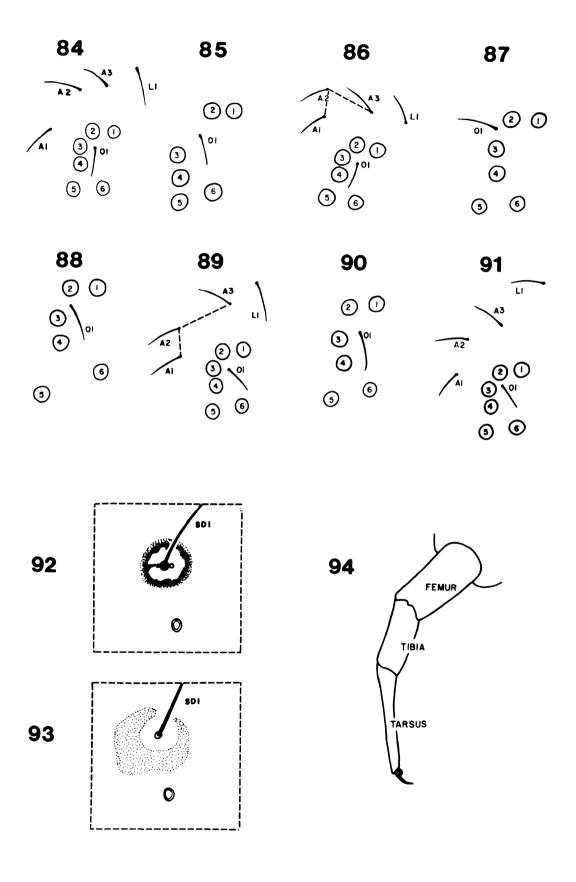
- Fig. 67. Nepticulidae, Nepticula myricafoliella Busck (USNM). Head, dorsal aspect.
- Fig. 68. Eriocraniidae, Mnemonica sp. (USNM). Head, dorsal aspect.
- Fig. 69. Nepticulidae, Ectoedemia populella Busck (USNM). Head, dorsal aspect (see also Fig. 27).
- Fig. 70. E. populella. Ocelli, ocellar setae, and outline of anterior part of head.
- Fig. 71. Gracillariidae, <u>Marmara salictella</u> Clemens (USNM). Head, dorsal aspect.
- Fig. 72. Opostegidae, Opostega sp. (USNM). Head and TI, dorsal aspect.

- Figures 73-83. Gracillariidae, Lyonetiidae, and some characters used in the key.
 - Fig. 73. Gracillariidae <u>Phyllocnistes populella</u> Chambers (USNM). Head, dorsal aspect, early instar.
 - Fig. 74. P. populella. Head, dorsal aspect, 4th instar.
 - Fig. 75. Lyonetiidae, Lyonetia speculella Clemens (USNM). Head, left lateral aspect, ocelli (cranial setae left out).
 - Fig. 76. Plutellinae, typical crochet arrangment on a ventral proleg.
 - Fig. 77. Frontal aspect of typical head, fronto-clypeus extending less than halfway to the epicranial notch.
 - Fig. 78. Fronto-clypeus extending more than halfway to epicranial notch.
 - Fig. 79. Fronto-clypeus reaching the epicranial notch.
 - Fig. 80. Typical labrum with shallow labral notch.
 - Fig. 81. Noctuidae, Acronycta americana (MSU). Labrum with a deep motch.
 - Fig. 82. Notodontidae, Heterocampa manteo Doubleday (MSU). Labrum with a v-shaped notch and a groove.
 - Fig. 83. Saturniidae, <u>Telea polyphemus</u> Cramer. Labrum with a very deep u-shaped notch.

Figures 84-94. Characters used in the key.

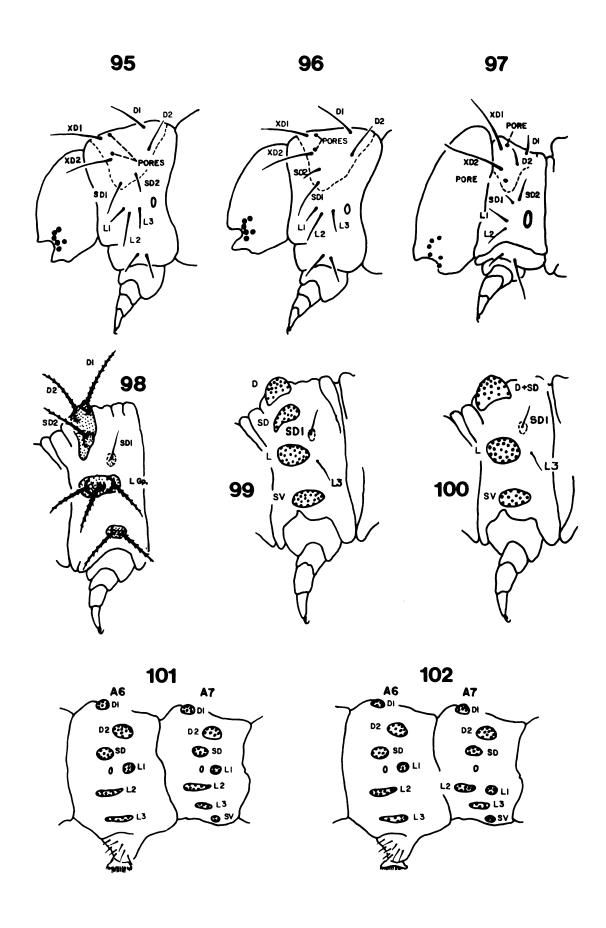
- Figs. 84-91. Various arrangments of ocelli and cranial setae Al, A2, A3, L1, and O1.
- Fig. 92. Gelechiidae, Autosticha pelodas (Meyrick) (USNM). Seta SD1 on A3 and modified pinaculum.
- Fig. 93. Blastobasidae, <u>Eubolepia gargantuella</u> Hein.(USNM). Seta SD1 on A3 and a semicircular pinaculum.
- Fig. 94. Glyphipterygidae, Rhobanda gaurisiana (Walker) (USNM).

 Thoracic leg with an elongate tarsus.



Figures 95-102. Characters used in the key.

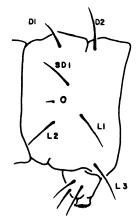
- Fig. 95. Setal arrangment on TI with SD2 caudad of SD1.
- Fig. 96. Setal arrangment on TI with SD2 dorsad of SD1.
- Fig. 97. Notodontidae, Heterocampa manteo Doubleday (MSU). TI lateral aspect. SD1 & SD2 excluded from the prothoracic shield.
- Fig. 98. Arctiidae, <u>Hypoprepia fucosa</u> (Hubner) (USNM). TII lateral aspect.
- Fig. 99. Arctiidae. Verruca arrangment on TII, lateral aspect.
- Fig. 100. Pericopidae. Verruca arrangment on TII, lateral aspect.
- Fig. 101. Ctenuchidae. Verruca arrangment on A6 & A7, lateral aspect.
- Fig. 102. Pericopidae. Verruca arrangment on A6 & A7, lateral aspect.

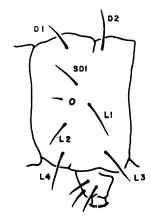


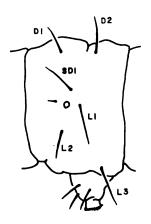
Figures 103-109. Characters used in the key.

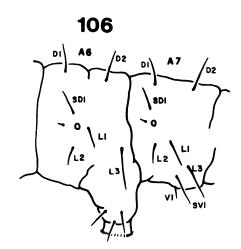
- Figs. 103-105. A3, left lateral aspect, various setal arrangments.
- Fig. 106. Noctuidae. A6 & A7, left lateral aspect.
- Fig. 107. Cossidae, <u>Dyspessa ulula</u> (Bkh.) (USNM, Italy). A3, left lateral aspect.
- Fig. 108. Sesiidae, <u>Paranthrene robiniae</u> (Hy. Edwards) (USNM). A7 & A8 dorsal aspects showing location of caudo-projecting spiracles on A8.
- Fig. 109. Incurvariidae, <u>Lampronia sp.</u> (J.A. Powell, coll.) A3 dor-sal aspect.

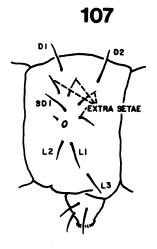


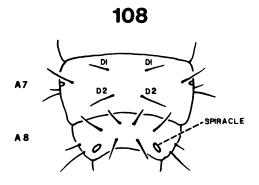


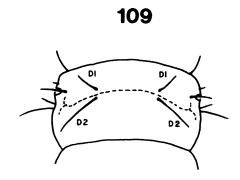












Figures 110-117. Characters used in the key.

- Fig. 110. A8 left lateral aspect, SD1 dorsad of the spiracle.
- Fig. 111. A8 left lateral aspect, SD1 cephalad of the spiracle.
- Fig. 112. Tortricidae. A9 & A10, dorsal aspect. Setae D2 on a common mid-dorsal pinaculum on A9.
- Figs. 113, 114, 115, 117. Various setal arrangments on A9, left lateral aspect.
- Fig. 116. Notodontidae, Heterocampa manteo Doubleday (MSU). A9 & A10 left lateral aspect, with extra seta on A10.

Figures 118-123. Left lateral aspects of A6 below spiracle, showing different kinds of prolegs.

- Fig. 118. Pyralidae, Ostrinia nubilalis (Hubner) (MSU).
- Fig. 119. Cossidae, Chilecomodia sp. (USNM).
- Fig. 120. Pterophoridae, <u>Platyptilia</u> antirrhina Lange (Lange, coll.)
- Fig. 121. Geometridae, Cingilia caternaria Drury (USNM).
- Fig. 122. Noctuidae, Agrotinae. Choephora fungorum (Grote) (USNM).
- Fig. 123. Noctuidae, Acronyctinae. Acronycta oblinita Smith & Abbott (USNM).

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