CYTOLOGICAL AND GENETIC OBSERVATIONS OF MEIOTIC DRIVE IN DROSOPHILA MELANOGASTER

> Thesis for the Dagree of Ph. D. MICHIGAN STATE UNIVERSITY Mary Fae Rengo 1969





This is to certify that the

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presented by

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has been accepted towards fulfillment of the requirements for

\_degree in \_\_\_\_\_ Ph.D.

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#### ABSTRACT

#### CYTOLOGICAL AND GENETIC OBSERVATIONS OF MEIOTIC DRIVE IN DROSOPHILA MELANOGASTER

Ву

#### Mary Fae Rengo

Many exceptions to Mendel's law of independent segregation have been noted. If the inequality of recovery of two homologous chromosomes is due to a meiotic event, these homologues are said to exhibit meiotic drive.

This study was designed 1) to ascertain the effect of irradiation at the onset of meiosis on the meiotic mutant <u>Segregation-distorter</u> in order to determine the meiotic stage during which the mechanism of <u>SD</u> is sensitive, and 2) to observe the meiotic drive systems of <u>SD</u> and the abbreviated  $sc^4sc^8$  X together ("Double-Drive") both cytologically and genetically at 18° and 28°C. Analysis of the interaction of these systems and correlation of the nonrandom polar segregation of  $sc^4sc^8$  with its genetic recovery tested the hypothesis that the mechanism of both drive systems involves a predetermined functional-nonfunctional polarity which is independent of the drive systems which respond to it.

Irradiation treatments were administered to very late third instar male SD larvae. To observe the effects of sperm storage, part of the experimental and control groups were aged after eclosion as virgins. The first four daily broods from unstored irradiated males exhibited depressed  $\underline{k}$  values. The percentage of aberrations and the depression of the  $\underline{k}$  values per brood appear to be positively correlated. Using the proportion of aberration products as an index, stored males appear to transfer few sperm which were in meiotic or premeiotic stages at the time of treatment.

Depressed <u>k</u> values were exhibited by both irradiated and control groups which were stored. Lack of mating activity and possibly resultant prolongation of Meiosis I appear to "unstabilize" and depress the drive of SD.

The sex ratios (44/total) of <u>cn</u> <u>bw</u> progeny in experimental and control groups were in excess of 60%. A modification of Hiraizumi's <u>SD-X</u> chromosome homology hypothesis is presented to account for the lack of a reciprocal decrease in the sex ratios of <u>SD</u> progeny.

In the study of "Double-Drive," the nonrandom segregation of  $\underline{sc}^4 \underline{sc}^8$  or the nondisjunctional nullo product away from the bacterial pole correlated with genetic recovery, supporting the hypothesis that a predetermined functionalnonfunctional polarity exists in the primary spermatocyte. At  $18^{\circ}C$ ,  $\underline{sc}^4 \underline{sc}^8$  expresses increased drive when in the presence of <u>SD</u>. Both drives are slightly reduced when together at  $28^{\circ}C$ . Thus, the data indicate that the drive systems of <u>SD</u> and  $\underline{sc}^4 \underline{sc}^8$  are not competitive in their interaction, and that the mechanism of <u>SD</u> is not independent of that to which  $\underline{sc}^4 \underline{sc}^8$  responds.

# CYTOLOGICAL AND GENETIC OBSERVATIONS OF MEIOTIC DRIVE IN <u>DROSOPHILA MELANOGASTER</u>

By

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### A THESIS

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To my mother and father

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#### I. INTRODUCTION

Many exceptions to Mendel's law of independent segregation have been noted. These abnormal progeny ratios may be due to 1) aberrant meiotic segregation, 2) gametic selection, or 3) post-fertilization selection. If the inequality of recovery of two homologous chromosomes is due to a meiotic event, these homologues are said to exhibit meiotic drive (Novitski and Sandler, 1956).

One of the meiotic drive phenomena occurring in Drosophila melanogaster, Segregation-distorter (SD), has been intensively studied. SD is known to exhibit the following properties: 1) A male heterozygous for SD and a structurally normal SD chromosome produces almost all SD progeny, with rare exceptions. 2) Females heterozygous for SD produce normal segregation ratios; their heterozygous sons exhibit abnormal k values (with certain exceptions). (The parameter k is defined as the proportion of functional SDbearing sperm.) 3) When the <u>SD</u> chromosome is heterozygous with a structurally rearranged homologue having one breakpoint near the centromere, such as In(2LR)Cy, segregationdistortion is inhibited. 4) An SD chromosome appears to be insensitive to the action of another SD chromosome. 5) The complex SD locus lies in the centromeric region of chromosome II, between purple and cinnabar. To the right of SD is

Activator of <u>SD,Ac(SD</u>), which is necessary for <u>SD</u> to exhibit distortion. 6) There is a stabilizing modifier of <u>SD,St(SD</u>), at, or near, the tip of IIR. (Sandler, Hiraizumi, and Sandler, 1959; Sandler and Hiraizumi, 1960a, b, 1961a; Dennell and Judd, 1968).

Sandler, Hiraizumi, and Sandler (1959) presented a model for <u>SD</u> action which assumed that <u>SD</u> caused a break at  $\underline{SD}^+$ . The lethality of the union of  $\underline{SD}^+$  sister chromatids would cause an excessive recovery of <u>SD</u>-bearing chromosomes. Their model was supported by preliminary cytological observations of dicentrics at Metaphase II and bridges at Anaphase II in about half of the division figures examined (Crow, Thomas, and Sandler, 1962).

Peacock and Erickson (1965) did an extensive cytological investigation of meiosis in the <u>SD</u> male. Completely normal spermatogenesis was found. They found that all products of meiosis in the male are cytologically evident and are presumably transferred to the female. A <u>Drosophila</u> <u>melanogaster</u> male can transfer 3,000 to 4,000 sperm to the vagina during a single mating (Kaufmann and Demerec, 1942). The capacity of the female storage organs is about 700 sperm (Kaplan, Tinderholt, and Gugler 1962; Lefevere and Jonsson, 1962). Peacock and Erickson reported that, when the number of sperm transferred by <u>SD</u> males was less than the storage capacity of the female the ratio of stored sperm to progeny was 2:1. This ratio approached 1:1 when the number of sperm transferred exceeded the storage capacity of the

female or when the insemination was of the order of 60 sperm or less. These same relationships were exhibited by control, wild-type males.

In order to explain abnormal ratios recovered from certain translocation males  $\underline{T}$  (<u>1:4</u>)  $\underline{B}^{S}$ , Novitski and I. Sandler (1956a, b) proposed that some of the sperm produced by a <u>Drosophila melanogaster</u> male were regularly nonfunctional, and that particular chromosomes had certain propensities for being included in functional sperm.

The cytological observations and the sperm count versus progeny count ratios reported by Peacock and Erickson support this functionality hypothesis, and indicate that functional and nonfunctional sperm are a regular aspect of spermiogenesis in Drosophila. Peacock and Erickson postulated that the mechanism of <u>Segregation-distorter</u> was nonrandom orientation of the <u>SD</u> chromosome to the functional pole at Meiosis I.

The propensity of <u>SD</u> to orient towards the proposed functional pole of Meiosis I appears to vary. <u>SD</u>-bearing chromosomes with different strengths (meiotic drives) have been found in many natural populations, and exhibit <u>k</u> values from about .7 - 1.0 (Mange, 1961; Greenberg, 1962; Hiraizumi and Nakazima, 1965).

At least some <u>SD</u> chromosomes are temperature sensitive. From treatment of four different <u>SD</u> chromosomes with heat and cold, Mange (1968) found that temperature sensitivity was dependent on the stage of meiosis which a

sperm is undergoing and independent of the age of the male being treated. The most sensitive period occurred around early meiosis. The <u>SD</u> chromosomes which she reported to be greatly sensitive to heat treatment (30<sup>o</sup>C.) were recombinants. Both recombinants and natural <u>SD</u> chromosomes exhibited varying sensitivity to cold (190 C.).

Other meiotic drive systems in <u>Drosophila melanogaster</u> have been shown to be temperature sensitive at early meiosis, with the effect of temperature treatment being a more normal segregation ratio. The <u>RD</u> effect (high recovery rate of the X chromosome) is greatly depressed at  $18^{\circ}$ C, with the sex ratio ( $\frac{99}{7}$ /total) being most reduced 7-8 days after treatment (Erickson and Hanks, 1961). Segregation ratios of both  $X^{D}$ : IV and  $X^{P}$ : Y from A-type <u>Bar of Stone</u> translocation males were very much altered by  $18^{\circ}$ C treatment of prepupae (Zimmering and Perlman, 1962).

Gershenson (1933) suggested that the unequal recovery of XO and XXY progeny resulting from primary non-disjunction in the <u>Drosophila</u> male was due to chromosomal loss because of lack of synapsis. In XY and XYY males with chromosomes which differed in amount and distribution of heterochromatin, chromosome loss appeared to occur only where homology was drastically reduced, supporting Gershenson's hypothesis that lack of pairing is related to apparent loss of chromosomes (Sandler and Braver, 1954).

Yet, males with sex chromosomes with greatly reduced homology exhibit more normal recovery of the Y chromosome

at  $18^{\circ}$ C than at  $26^{\circ}$ C (Zimmering, 1963). Since a modified univalent Y shows a similar increase in recovery at  $18^{\circ}$ C, the increase in recovery of the Y from XY males observed at cold temperatures does not appear to be due to increased pairing between X and Y. Using  $sc^4sc^8/Y/Y$  males, Zimmering and Green (1965) found that the frequency of XY sperm recovered increased from 23% at  $26^{\circ}$ C to 43% at  $18^{\circ}$ C. Since, in such males, the  $sc^4sc^8X$  is usually a univalent while the Y chromosomes form a bivalent, the normalizing effect on segregation again does not appear to be due to increased pairing. However, this was not demonstrated cytologically.

Peacock (1965) found that, while failure of the  $sc^4sc^8x$  and Y chromosomes to synapse was common, these subsequent univalents usually travelled to the same pole at Anaphase I. The nondisjunction frequency was directly related to the frequency of synaptic failure. The occurrence of meiotic loss of unpaired chromosomes was negligible. Peacock postulated that the  $sc^4sc^8$  meiotic drive system was another case of nonrandom segregation of both disjunctional and nondisjunctional reciprocal products into functional and nonfunctional sperm.

The experiments in this study were undertaken to 1) ascertain the effect of irradiation at the onset of meiosis on <u>Segregation-distorter</u> and 2) observe two meiotic drive systems together, the  $sc^4sc^8X$  and <u>SD</u> ("Double-Drive"), both cytologically and genetically, at 18° and 28°C.

### Effect of Irradiation

The effects of heat and cold shock on <u>SD</u> and other chromosomes which exhibit abnormal segregation implicate early meiosis as the time of operation of these drive systems. Temperature treatments, however, are administered over relatively long periods of time. As a result, it is impossible to determine precisely the meiotic stage (or stages) which is critical to the drive systems. The functionality hypothesis would require the sensitivity to treatment to be exhibited prior to Anaphase I.

Irradiation treatments can be administered over periods of very brief duration. By irradiation of very late third instar larvae, primary spermatocytes will be the most advanced meiotic stage undergoing treatment (Cooper, 1950). Sensitivity of <u>SD</u> at this stage would support the hypothesis that functional and nonfunctional sperm are determined a Meiosis I and that the operation of the <u>Segregation-distorter</u> chromosome involves nonrandom orientation to the functional pole.

#### "Double-Drive"

The proposed functional and nonfunctional poles at Meiosis I provide a good working hypothesis to explain the mechanism of all meiotic drive phenomena. The operation of the meiotic drive systems of <u>SD</u> and  $sc^4sc^8$  has been attributed to the nonrandom segregation of the chromosome or chromosomes involved to the functional or nonfunctional pole at Anaphase I

(Peacock, 1965; Peacock and Erickson, 1965). Analysis of the interaction of these systems together would test the hypothesis that the operation of both systems involves a predetermined functional-nonfunctional polarity which is independent of the drive system or systems being studied, or, if functionality is not predetermined, would provide information concerning the time of determination of the functional pole.

#### Genetic Tests

If the polarity of the primary spermatocyte is determined prior to Anaphase I, the two systems would not be competitive. If both phenomena utilize functional-nonfunctional polarity, the drives exhibited by <u>SD</u> and the  $sc^4sc^8$  systems in "Double-Drive" should be similar to those exhibited by these chromosomes when alone. In addition to the factors mentioned above, there are possible segregational interactions between the chromosomes involved. This would be reflected in further deviation from randomness in the association of chromosomes in recovered sperm.

If the driving chromosomes themselves determine the functional pole,  $\underline{sc}^4 \underline{sc}^8$  and <u>SD</u> would be in a competitive situation if the mechanisms of both systems involved functionality. When these systems are present in the same role, there are eight possible gametic products:

Disjunctional products	Nondi <b>s</b> ju <b>n</b> ctional products
4 8	4 8

SD sc sc	<u>SD</u> <u>sc sc</u> Y
SD Y	<u>SD</u> nullo
$sd^+ sc^4 sc^8$	$\underline{SD}^+$ $\underline{sc}^4 \underline{sc}^8$ y
<u>sd</u> y	<u>SD<sup>+</sup></u> nullo

If <u>SD</u> is stronger than  $\underline{sc}^4 \underline{sc}^8$  in polar determination, the drive of <u>SD</u> in the "Double-Drive" situation would approximate the drive it exhibits alone, whereas the drive of the disjunctional and nondisjunctional products of the  $\underline{sc}^4 \underline{sc}^8$ system would be reduced due to the loss of  $\underline{sc}^4 \underline{sc}^8$  products which resulted in nonfunctional gametes. The reverse would be true if <u>SD</u> is the weaker competitor. If both systems are equal in polar determination strength, a high proportion of <u>SD  $\underline{sc}^4 \underline{sc}^8$  and <u>SD</u> nullo products would be recovered, due to the competition between  $\underline{SD}^+$  Y and <u>SD  $\underline{sc}^4 \underline{sc}^8$  and between</u> <u>SD</u> nullo and  $\underline{SD}^+ \underline{sc}^4 \underline{sc}^8$  Y.</u>

### Cytological Tests

The abbreviated  $\underline{sc}^4 \underline{sc}^8$  chromosome can be distinguished from a normal X and from its Y homologue, thus allowing cytological observation of its segregational behavior at Anaphase I. This behavior was scored in the  $\underline{sc}^4 \underline{sc}^8$  X system, and with the  $\underline{sc}^4 \underline{sc}^8$  X system in the presence of <u>SD</u> ("Double-Drive") at 18<sup>o</sup> and 28<sup>o</sup>C. The granular inclusions reported by Peacock and Erickson (1964) were present in the stocks used in this study. These granules have been shown to be Bacteria or Rickettsiae, and their polar association was concluded to be an independent response to the cytoplasmic gradient in meiotic cells to which "driving" chromosomes respond (Yanders, Brewen, Peacock, and Goodchild, 1968).

The presence and position of these inclusions in relation to the segregation of the chromosomes were noted. A positive correlation between non-random segregation to the pole lacking granular inclusions and subsequent genetic recovery would support the conclusion of Yanders <u>et al</u> (1968) that the polar association of these inclusions reflects a cytoplasmic gradient in meiotic cells associated with the determination of functionality of the gamete.

With the polar association of the granules, the segregational behavior of  $\underline{sc}^{4}\underline{sc}^{8}$  alone and in the presence of <u>SD</u> can be examined. If the granules are indicators of polarity, a difference in the behavior of  $\underline{sc}^{4}\underline{sc}^{8}$  in the two systems should be supported by genetic recovery ratios of the reciprocal products.

Evidence will be presented which supports the hypothesis that the mechanisms of the <u>SD</u> and  $sc^4sc^8$  meiotic drive systems involve nonrandom segregation to pre-determined functional and nonfunctional poles at Anaphase I. The data also indicate an interaction between the two systems and apparent repulsion between the <u>SD</u>-bearing chromosome and the sex chromosomes which is influenced by temperature.

#### II. MATERIALS AND METHODS

The stocks of <u>Drosophila melanogaster</u> used are listed below. Nutrient medium, modified after Carpenter (1950), was used for all experiments. All virgin females used were aged four to five days before mating.

#### Irradiation of SD

Very late third instar <u>SD-72</u> male larvae were irradiated with 0 (Control) or 450r of X-rays with a General Electric Maximar-250-III, operating at 250kv, 15ma, with a .5mm copper, 1.0mm aluminum filter, giving an average dose of 110r/minute. The larvae used were from half-hour egg collections. For treatment, control and experimental larvae were suspended in 0.7% saline solution (Beadle and Ephrussi, 1936), .5mm in depth, in an open plastic petri dish.

In experiment I, 20 control and 50 experimental males were transferred without etherization to two virgin <u>cn bw</u> females per day for seven days from the day of eclosion. In experiment II, 10 control and 25 experimental males were transferred as described above for seven days, from the day of eclosion. Ten control and 15 irradiated males were stored for seven days as virgins, then mated as above for seven days. All progeny resulting from these matings were collected and scored for sex and eye color. Male progeny

with aberrant colors were tested for meiotic drive by matings with cn bw virgin females.

### Cytological and Genetic Studies of Meiotic Drive

The  $\underline{sc}^4 \underline{sc}^8 \times chromosome$  used was a crossover product derived from an  $\underline{In(1)sc}^4$ ,  $\underline{In(1)sc}^8$  female. The distal breakpoints of these two inversions are similar. The proximal breakpoints, in the basal heterochromatin, are such that the crossover chromosome  $\underline{Ins(1)sc}^4 \underline{sc}^8$ , having the distal region of  $\underline{In(1)sc}^4$  and the proximal region of  $\underline{In(1)sc}^8$ , is deficient for a large portion of basal heterochromatin, (Cooper, 1959).

For the study of the  $Ins(1)sc^4sc^8, y, y^+ Y$  meiotic drive system, alone and in the presence of <u>SD</u>, the  $sc^4sc^8$ stock, stock A, was infected with granules from <u>cn bw</u> females known to exhibit cytoplasmic granules. The granules are maternally inherited. Crosses between  $sc^4sc^8, y/y^+Y$  of and  $sc^4sc^8$ , y/Basc PP (with granules) or between  $\pm \pm y + Y$ : <u>SD</u>, <u>cn bw/SD<sup>+</sup>++</u> of (Stock B) and  $sc^4sc^8$ , y/Basc PP (with granules) yield two types of F<sub>1</sub> males which are distinguishable cytologically:  $sc^4sc^8$ ,  $y/y^+Y$  and  $Basc/y^+Y$ .

1.  $\frac{sc^4sc^8}{sc}$  without <u>SD</u>

	sc <sup>4</sup> sc Bas	$\frac{8}{y}; \frac{cn}{cn}$	bw 4	¥x	<u>sc</u> <sup>4</sup>	<u>sc<sup>8</sup>,y</u> ; + <sub>v</sub>	<u>cn</u>	bw bw	66	1		
<u>sc<sup>4</sup>sc<sup>8</sup>,y</u> ;	<u>cn bw</u>	sc <sup>4</sup> sc <sup>8</sup> ,	2 2; <u>cn</u>	bw	sc <sup>4</sup> s	sc <sup>8</sup> ,y;	<u>cn</u>	bw	Bas	<u>BC</u> ;	<u>cn</u>	bw
Basc	cn bw	Basc	cn	bw	<u>у</u> +	Y	cn	bw	<u>y</u> +	Y	cn	bw
			Cyto	Log	icall	ly dis	stin	gui	shal	ole	ma]	les

2.  $sc^4sc^8$  with <u>SD</u> ("Double-Drive")

$$\frac{sc^4sc^8, y; cn bw}{Basc cn bw} \stackrel{\text{QQ}}{+} x \stackrel{\text{+}}{+}; \frac{SD}{y^+} co bw$$

 $\frac{sc^4sc^8, y; cn bw}{+} \frac{Basc; cn bw}{cn bw} \frac{sc^4sc^8, y; cn bw}{y^+ Y} \frac{Basc; cn bw}{cn bw} \frac{Basc; cn bw}{y^+ Y} \frac{Basc; cn bw}{cn bw}$ 

 $\frac{sc^4sc^8, y; cn bw Basc; cn bw sc^4sc^8, y; cn bw Basc; cn bw}{+ SD + SD y^+ Y SD y^+ Y cn bw}$ 

Cytologically distinguishable males

The sib males with <u>Basc</u> X served as controls. These crosses were cultured at  $18^{\circ}$  and  $28^{\circ}$ C. Testes from F<sub>1</sub> prepupae and very early pupae were dissected in 0.7% saline, stained in acetoorcein, then squashed for observation under phase contrast. The prepupal and early pupal stages were selected because granule populations diminish greatly after these stages.

All primary spermatocytes with suitable Anaphase I divisions were scored. These cells were divided into three categories of granule populations: 1) polarized, where the granular mass is concentrated at one pole, 2) non-polarized, where the granular mass is distributed throughout the cell, situated between the poles at the equator, or evenly distributed at both poles, and 3) those cells with few or no granules. Disjunctional and nondisjunctional divisions were scored. In the case of polarized cells, the chromosomes travelling to the granular and non-granular poles were noted. The sibs of those males dissected were mated to  $\underline{y}$ ; <u>cn</u> <u>bw</u> females for genetic tests of the  $\underline{sc}^4 \underline{sc}^8$  drives, nondisjunction rates and <u>k</u> (SD/total) values of the F<sub>2</sub> progeny. These crosses were cultured at 22.5<sup>o</sup>C.

### <u>Genetic Tests of Male Sibs</u> of Dissected Males

1.  $\frac{sc^4sc^8}{sc}$  without <u>SD</u>

from disjunctional gametes from nondisjunctional gametes

<u>Stocks</u>

- <u>SD</u> <u>Segregation-distorter</u>, SD-72, second chromosome (Sandler, Hiraizumi, and I, Sandler, 1959).
- <u>cn bw</u> <u>cinnabar</u> and <u>brown</u>, second chromosome, exhibits granules (Madison).
- y; <u>cn bw yellow</u>, <u>cinnabar</u>, and <u>brown</u> (derived by Grant Brewen  $y^{b-32}$  (Oregon) and <u>cn bw</u>, Madison).

Stock B:  $\frac{+}{x}$ ;  $\frac{cn \ bw}{cn \ bw}$   $\stackrel{qq}{+}$   $x \frac{+}{y}$ ;  $\frac{SD-72}{cn \ bw}$  (derived by Yanders). +  $cn \ bw$   $y^+ \ Y$   $cn \ bw$ Males used for "Double-Drive" cross.

"Stock A with granules":

1. Yanders' "Stock A":

 $\frac{\mathrm{sc}^{4}\mathrm{sc}^{8},\mathrm{y};}{\mathrm{Clb}} \xrightarrow{\mathrm{cn}} \frac{\mathrm{bw}}{\mathrm{cn}} \stackrel{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}{\overset{\text{H}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{H}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}{\overset{\text{H}}}}{\overset{H}}{\overset{H}}}{\overset{H}}{\overset{H}}}{\overset{H}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}}{\overset{H}}{\overset{H}}$ 

2. <u>Basc</u> (Philadelphia).

- 3.  $\frac{+}{y^+ y}$ ; cn bw of from "Stock B".
- 4. cn bw with granules, (Madison).

Der	ivation of "Stock A with Granules"
1.	$\frac{Basc}{Basc}; \frac{+}{+} + \frac{cq}{+} x + \frac{+}{Y}; \frac{cn \ bw}{cn \ bw}$
2.	$\frac{Basc}{Y} ; \frac{cn \ bw}{+ + +} x \frac{+}{+} ; \frac{cn \ bw}{cn \ bw} $ (granules)
3.	$\frac{Basc}{+}; \frac{cn \ bw}{cn \ bw} \times \frac{sc^4sc^8, y}{Y}; \frac{cn \ bw}{cn \ bw} $ (granules)
	<u>Basc</u> ; <u>cn bw</u> 44 sc <sup>4</sup> sc <sup>8</sup> ,y; cn bw
	(for "Stock A" with granules)
4.	$sc^4sc^8$ , v; cn bw ++ x + ; cn bw

 $\overline{\mathbf{y}}_{\mathbf{y}}^{+}$ 

cn bw

(for "Stock A" with granules)

 $sc^4sc^8, y$ ; <u>cn bw</u> of

ClB

cn bw

yy+

cn bw

#### III. RESULTS

#### Effects of Irradiation on SD

Tables 1 through 6 and Figures 1 through 4 summarize the results of irradiating <u>SD</u> male larvae at or near the onset of meiosis. In experiment 1, the <u>k</u> value (<u>SD</u>/total) of the progeny of irradiated unstored males was depressed below that of the control group for the first six days of mating, with the lowest <u>k</u> values resulting from days 2 and 3. The brood of day 7 exhibited a <u>k</u> higher than any of the control group. In experiment 2, the <u>k</u> values of the unstored, irradiated males were depressed in all seven daily broods, with day 4 showing the greatest depression. Due to the large numbers of progeny, a test of significance is nct necessary.

Certain aberrations appeared in progeny of irradiated larvae which did not occur in control groups. These aberrations provide an indication of which sperm batches were most affected by irradiation. The aberrations detectable are <u>cinnabar</u> and <u>brown</u> eyed flies. These aberrant progeny are probably the result of radiation-induced chromatid breaks between the <u>cinnabar</u> and <u>brown</u> loci. Breaks in analogous regions of non-sister chromatids and subsequent chromatid exchange would produce such aberrants. The <u>SD Ac(SD) cn</u><sup>+</sup> chromatid would thereby carry the bw allele but no stabilizer

Paternal Age in Days	k(SD/total)	Percent Aberrations	Total Progeny	To Abera bw	tal tion <b>s</b> cn
1	92.96	.47	428	2	0
2	85.72	.63	1265	4	4
3	85.62	.24	3701	8	1
4	91.79	.11	5534	6	0
5	9 <b>2.</b> 31	.08	6155	4	1
6	90.32	.07	6120	3	0
7	97.25	.02	6120	1	0

Table l.	SD-72 this	rd instar	larvae	irradiated	(450 r),	
	unstored,	control 1	k = 95.8	, 50 males,	Experiment	1.

Table 2. 20 control males, unstored, Experiment 1. No aberrations observed.

Paternal Age In Days	k	Total Progeny
1	95.71	632
2	95.69	1974
3	95.83	3302
4	95.71	2571
5	95.87	2699
6	96.01	2878
7	95.81	3005

Paternal Age in Days	k(SD/total)	Percent Aberrations	Total Progeny	T Aber bw	otal rations cn
1	97.84	.37	1067	4	0
2	97.95	.42	2405	10	0
3	96.44	.22	2249	5	0
4	95.98	.49	2436	10	0
5	96.82	.09	3430	3	0
6	97.02	.00	3695	0	0
7	97.09	.025	3954	1	0

Table 3. <u>SD-72</u> third instar larvae irradiated (450 r), unstored, control k = 99.5, 25 males, Experiment 2.

Table 4. 10 control males, unstored, Experiment 2. No aberrations observed.

Paternal Age In Days	k	Total Progeny
1	99.43	1051
2	99.20	1496
3	99.44	1070
4	99.46	1303
5	99.36	776
6	99.64	822
7	99.42	517

Paternal Age In Days	k	Percent Aberrations	Total Progeny	Total Aberrations	
<u> </u>			······	Wd	<u>cn</u>
7	97.86	.089	3360	3	0
8	97.11	.000	2389	0	0
9	98.41	.000	2008	0	0
10	98.68	.000	2039	0	0
11	97.66	.000	1880	0	0
12	97.09	.000	1753	0	0
13	98.61	.000	2083	0	0

Table 5.	15 <u>SD-72</u>	third	instar larvae,	irradiated	(450 r),
	stored 7	days,	Replicate 2.		

Table 6. 10 <u>SD-72</u> third instar larvae, control group, stored 7 days, Replicate 2.

Paternal Age in Days	k	Percent Aberrations	Total Progeny	Total Aberrations
7	99.23	none	2691	none
8	98.22		2139	
9	96.77		1918	
10	96.48		1703	
11	99.33		1649	
12	97.99		1541	
13	98.45		1160	

locus, <u>St(SD)</u>. As such, it would exhibit a moderate, unstable <u>k</u>. The <u>SD<sup>+</sup>AC(SD)<sup>+</sup>cn</u> chromatid would bear the normal allele for <u>brown</u>, <u>bw<sup>+</sup></u>, and the <u>SD</u> stabilizer, <u>St(SD)</u>, and exhibit no drive.

All <u>cn</u> and <u>bw</u> male progeny were tested for drive by mating them with <u>cn bw</u> virgin females. The mean <u>k</u> for <u>brown</u> males was .772. The <u>k</u> for <u>cinnabar</u> males was .521.

Irradiated males mated from day of eclosion show the greatest number of aberrations in broods from days one through four in both replicates 1 and 2. The percentage of aberrations and the depression of  $\underline{k}$  values per brood appear to be positively correlated. Due to the small numbers of aberrations in the data concerned, a nonparametric test of correlation is not applicable.

Aberrations appeared only in the first brood of the stored, irradiated males. These stored males exhibited  $\underline{k}$  values similar to that of the stored control group. However, lower  $\underline{k}$  values were exhibited by the stored control group than those of the unstored males. Lack of mating activity appears to increase the percentage of  $\underline{SD}^+$  sperm transferred to the female. If aberrations are an indication of which sperm batches were in meiotic or premeiotic stages at the time of treatment, the stored males do not appear to transfer most of these sperm.

The over-all sex ratio (44/total) of <u>cn</u> <u>bw</u> progeny from experiment 1 was .619. This represents a total of 2484 <u>Cn</u> <u>bw</u> progeny. Experiment 2 <u>cn</u> <u>bw</u> progeny (682) had an

over-all sex ratio of .625. This supports the hypothesis of Hiraizumi and Nakajimi (1967) that <u>SD</u> has some homology with the X chromosome which reduces the probability of both travelling to the same pole.

Where the  $\underline{SD}^+$  <u>cn</u> <u>bw</u> chromosome reached the functional pole, it was accompanied by the X chromosome over 60 percent of the time. Thus, when  $\underline{SD}$  <u>cn</u><sup>+</sup> <u>bw</u><sup>+</sup> travelled to the hypothesized non-functional pole at Anaphase I, it was accompanied by the Y chromosome more than 60 percent of the time. This implies a degree of repulsion between the <u>SD</u>-bearing chromosome II and the X chromosome comparable to the repulsion exhibited between homologues.

# Cytological Observations of sc<sup>4</sup>sc<sup>8</sup> and "Double-Drive"

The results of the cytological studies are given in Tables 7 and 8. The presence and position of the granular inclusions varied. In about half of the primary spermatocytes of all classes observed, the granules were polarized, the great majority of granules being situated in a mass or near one pole of the cell. The <u>Basc</u> control males exhibited polarization to about the same extent as the experimental groups. The granular masses of nonpolarized cells in both types of males were usually distributed on one side of the cell along the equator.

The position of the granular inclusions was undoubtedly disturbed by the squash procedure. In most cases where the granules were scored as polarized, the granular mass was

Figure 1. <u>k</u> values per daily brood. Irradiated and control males, Experiment 1.



Figure 2. <u>k</u> values per daily brood. Irradiated and control males. Experiment 2.




Figure 3. Percent aberrations per daily brood. Irradiated males, Experiments 1 and 2.



Figure 3. Percent aberrations per daily brood. Irradiated males, Experiments 1 and 2.

			Polar	ized C	ells				Ъe	Non Pol w or no	larize o Grar	ed Nules		Totals
		Dis	junctio	nal		N	.D.*	Di	sj.**	N.D.*	Dis	**。[	N.D.*	
Temp.	لر o	<b>4-8</b> :o <b>gran-</b> ules	4-8 away	Basc to	Basc away	XY to	XY away	4-8	Basc	XX	4-8	Basc	ХХ	
28 <sup>0</sup>	D.D.***	33	51			69	37	43		33	19		25	310
	Basc			61	66				61			34		222
28 <sup>0</sup>	4-8	19	34			25	14	38		17	31		25	203
	Basc			47	66				56			43		212
18 <sup>0</sup>	D.D. ***	31	43			23	15	29		6	20		12	182
	Basc			41	48				51			13		153
18 <sup>0</sup>	4-8	34	42			6	ß	21		19	17		5	149
	Basc			44	39				43			22		158
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Table 7. Cytological Observations.

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Percent M-5 to non-gran. pole	52.0	48.7	53.9	45.8
Percent sc4sc8 to non-gran. pole	60.7	64.2	58.1	55 • 3
Percent XY to gran.	65.1	64.1	60.5	64.3
Polar- ized n.d.	.558	.424	.339	.156
Observ. n.d. rate	.529	.339	.324 .000	.235
Percent Polar- ity	61.3 57.2	45.3 53.3	61.5 58.2	60.4 52.5
	Double-Drive <u>Basc</u> sib	<u>sc sc sib</u> Basc sib	Double-Drive <u>Basc</u> sib	<u>sc4sc</u> <u>Basc</u> sib
	28 <sup>0</sup> c.		18 <sup>0</sup> C.	

Table 8. Summary of cytological observations.

not located at the pole indicated by the spindle axis, but was positioned to one side of that pole. Usually, this difference between spindle axis and granular axis was slight. However, once the granular mass approached the equatorial region, it had to be considered to be unpolarized. Thus, these nonpolarized primary spermatocytes may have actually exhibited polarization prior to disturbance of the rigid spindle and granular position by squashing.

The observed nondisjunction rate was higher when  $\underline{sc}^{4}\underline{sc}^{8}$  was in the presence of <u>SD</u> at both 28° and 18°C. No nondisjunctional events were observed in the <u>Basc</u> class. At 18°C, the rates of nondisjunction in both  $\underline{sc}^{4}\underline{sc}^{8}$  and "Double-Drive" primary spermatocytes was lower than those observed at 28°C.

In <u>Basc-y</u> Y cells, the sex chromosomes tended to undergo anaphase movement at the same time as the autosomes. In  $\underline{sc}^4\underline{sc}^8$  and "Double-Drive", however, the movement of the X and Y to the poles was generally precocious with respect to that of the autosomes. This undoubtedly is due to lower frequency of synaptic association because of the reduced X-Y homology with the  $\underline{sc}^4\underline{sc}^8$  gross heterochramatic deficiency.

The anaphase behavior with respect to the position of the granules in disjunctional or nondisjunctional polarized cells did not appear to be random. In the  $28^{\circ}$ C "Double-Drive" disjunctional class,  $sc^4sc^8$  travelled to the nongranular pole with a frequency of 61%. At  $18^{\circ}$ C, this frequency was reduced to 58%. At  $28^{\circ}$ C, the  $sc^4sc^8$  chromosome

oriented to the non-granular pole with a frequency of 64% of the time. At  $18^{\circ}$ C, this frequency was reduced to 55%. The nullo pole of nondisjunctional cells was usually that pole without the granular mass. In  $28^{\circ}$ C "Double-Drive" nondisjunctional cells, the pole to which  $sc^4sc^8$  X and Y travelled was the granular pole in 65% of such events. At  $18^{\circ}$ C, this frequency was 61%. The corresponding frequencies for  $sc^4sc^8$  were 64% at  $28^{\circ}$ C, and 64% at  $18^{\circ}$ C. The behavior of the <u>Basc</u> X appeared to be random with respect to its orientation to the granular pole.

# <u>Genetic Tests of sc<sup>4</sup>sc<sup>8</sup></u> and "Double-Drive"

Tables 9, 10 and 11 summarize the results of the progeny tests of the sibs of males sacrificed for cytological study. The <u>k</u> values exhibited by "Double-Drive" males increased from 91.3% at  $28^{\circ}$ C to 98.9% at  $18^{\circ}$ C. The corresponding over-all rates of nondisjunction decreased from .512 at  $28^{\circ}$ C to .305 at  $18^{\circ}$ C.

When the  $\underline{sc}^4 \underline{sc}^8$  drive system operated in the absence of <u>SD</u> the over-all rates of nondisjunction were lower than with <u>SD</u>. At 28°C and 18°C, the nondisjunction frequencies were .323 and .149 respectively. These results are in accord with those of Zimmering (1963), with the frequency of nondisjunction being reduced by almost half at 18°C (Zimmering cultured his  $\underline{sc}^4 \underline{sc}^8$  males at 18° and 26°C.)

The presence of <u>SD</u> ostensibly increases the efficiency of the  $\underline{sc}^4\underline{sc}^8$  meiotic drive system at both high and low

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"Double-Drive.
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tests
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Summary
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Table

lotal	4327	4194
over- all n.d.	.512	. 305
cnbw n.d.	.452	.467
SD n.d.	.518	. 303
×	91.3	98.9
cnbw Y + Q	166	20
cnbw Y đ	4	m
аз + <sup>у</sup> 6 + <sup>у</sup>	19	117
sD Y o	2028	1141
cnbw Y f	61	m
cnbw Yy <sup>+</sup> o	145	17
us ot ot	1453	1792
sD Yy <sup>+</sup>	451	1011
Temp.	28 <sup>0</sup>	18 <sup>0</sup>

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Segregat	ion		Frequency
I	II	<u>28<sup>0</sup>C.</u>	<u>18<sup>0</sup>C.</u>
$\frac{\text{sc}^4 \text{sc}^8}{\text{Y}}$	<u>SD</u>	.427	.336
	cnbw	.004	.034
$\frac{\mathrm{sc}^{4}\mathrm{sc}^{8}}{\mathrm{Y}}$	<u>cnbw</u>	.001	.014
	<u>SD</u>	.263	.104
nullo	<u>SD</u>	.272	.469
<u>sc<sup>4</sup>sc<sup>8</sup>-Y</u>	cnbw	.005	.038
nullo	<u>cnbw</u>	.001	.001
<u>sc<sup>4</sup>sc<sup>8</sup>-Y</u>	<u>SD</u>	.028	.004

Table 10. "Double-Drive" segregation frequencies.

Table 11. Summary of progeny tests  $\underline{sc}^{4}\underline{sc}^{8},\underline{y}$ .

Temp.	cnbw Yy <sup>+</sup> o	cnbw y 4	cnbw y d	cnbw y <sup>+</sup> 4	rate of n.d.*	x/x+y	o/o+xy	Total
28 <sup>0</sup>	490	1722	1020	36	.323	.778	.966	<b>3</b> 268
18 <sup>0</sup>	399	487	99	54	.149	.550	.647	1039

temperatures. Yet, in the "Double-Drive" situation, the meiotic drive of <u>SD</u> increases at  $18^{\circ}$  whereas that of  $\underline{sc}^{4}\underline{sc}^{8}$  decreases. Without  $\underline{sc}^{4}\underline{sc}^{8}$ , this  $\underline{SD}(+/y+Y; \underline{SD-72} + +)$  has a  $\underline{SD}^{+}$  cnbw

<u>k</u> value of 99.6 at 28°C and 99.1 at  $18^{\circ}$ C. Therefore, the presence of the <u>sc<sup>4</sup>sc<sup>8</sup></u> system at 28°C appears to effectively reduce the drive of <u>SD</u> while increasing the drive of <u>sc<sup>4</sup>sc<sup>8</sup></u>. At  $18^{\circ}$ C, the interaction of the systems increases <u>sc<sup>4</sup>sc<sup>8</sup></u> drive while the k of SD is apparently unaffected.

The relative frequencies of the reciprocal products of disjunctional and nondisjunctional events at 28<sup>o</sup>C were:

	X/X+Y	0/0 <b>+X</b> Y
"Double-Drive"	.718	.917
$sc^4sc^8$	.778	.966

Thus, the drive of the disjunctional  $\underline{sc^4 sc^8}$  actually decreased in the presence of <u>SD</u>, as did the "drive" of the nondisjunctional nullo product. Therefore, at  $28^{\circ}$ C, the drive of both  $\underline{sc^4 sc^8}$  and <u>SD</u> are lowered. At  $18^{\circ}$ C, the results differ from those at  $28^{\circ}$ C.

	X/X+Y	0/0+X3
"Double-Drive"	<b>.6</b> 16	.893
sc <sup>4</sup> sc <sup>8</sup>	.550	.647

The drives of both disjunctional and nondisjunctional products are increased when the  $\underline{sc}^{4}\underline{sc}^{8}$  system is in the presence of <u>SD</u> at 18°C. Therefore, at 18°C, the drive of <u>SD</u> is apparently unaffected by the presence of  $\underline{sc}^{4}\underline{sc}^{8}$ , while that of  $\underline{sc}^{4}\underline{sc}^{8}$ is heightened. Table 10 shows the frequency of segregation of disjunctional and nondisjunctional  $\underline{sc}^4 \underline{sc}^8$  chromosomes with <u>SD</u> or <u>cnbw</u>. Considering disjunctional products, the association of  $\underline{sc}^4 \underline{sc}^8$  and <u>SD</u> decreases from .427 at  $28^{\circ}$ C to .336 at  $18^{\circ}$ C, while the reciprocal association of  $\underline{y}^+ \underline{y}$  with <u>SD</u><sup>+</sup> <u>cnbw</u> increased from .004 to .034. Accordingly, the segregation of  $\underline{sc}^4 \underline{sc}^8$  with <u>SD</u><sup>+</sup> <u>cnbw</u> increased from .001 at  $28^{\circ}$ C to .014 at  $18^{\circ}$ C. From genetic evidence, it appears that the  $\underline{sc}^4 \underline{sc}^8 \underline{x}$ and <u>SD</u> arrive at the functional pole (F pole) together with greater frequency at  $28^{\circ}$ C than at  $18^{\circ}$ C.

In the case of nondisjunctional products, the frequency with which the <u>SD</u> segregates to the nullo pole increases at  $18^{\circ}C$  (.272 at  $28^{\circ}C$  vs. .469 at  $18^{\circ}C$ ), as does the frequency of  $\underline{sc}^{4}\underline{sc}^{8}$ , Y - <u>cnbw</u> association. The segregation frequency of <u>cnbw</u> to the nullo pole remains the same (.001) at  $18^{\circ}C$  and  $28^{\circ}C$ . That of  $\underline{sc}^{4}\underline{sc}^{8}$ , <u>Y</u> and <u>SD</u> decreases from .028 to .004.

The segregation of the nondisjunctional product and <u>SD</u> at high and low temperatures present a different picture than <u>SD</u> segregation with disjunctional products. Both nullo-<u>SD</u> and  $\underline{sc}^4 \underline{sc}^8 x, \underline{y}$  <u>cnbw</u> segregation frequencies increase at  $18^{\circ}C$ . Apparently, the probability of <u>SD</u> reaching the nullo F pole and of <u>cnbw</u> arriving with XY at the F pole increases at  $18^{\circ}C$ . The repulsion between <u>SD</u> and the  $\underline{sc}^4 \underline{sc}^8 \underline{x} - \underline{y}^+ \underline{y}$ chromosomes increases as the temperature is lowered. The postulated homology between the <u>SD</u> complex and X (Hiraizumi, 1967) may be supported by these data. The abbreviated  $\underline{sc}^4 \underline{sc}^8$  X has less homology with the Y than a normal X (Cooper, 1959). However, the  $\underline{sc}^4 \underline{sc}^8$  X and  $\underline{y}^+$ Y together may bear enough homology to the <u>SD</u> complex to cause such repulsion.

## Correlation of Cytological Studies and Genetic Evidence

Tables 12 and 13 compare cytological observations with resultant progeny data. The frequency of nondisjunction in all primary spermatocytes and that exhibited in the  $F_2$ are in agreement. A 3 x 2  $X^2$  test (Table 14) of rate of disjunction versus the presence and location of the granules, gives a  $X^2$  value of 2.72. Therefore, the probability is about 30% that this distribution would occur in populations with homogeneous means.

These data support Yanders <u>et al</u> (1968) in their conclusion that meiotically driven chromosomes and the bacterial cellular inclusions are responding to the same cellular polarity, and that this polarity is associated with the determination of functional and non-functional gametes at Meiosis I (Peacock and Erickson, 1965). These bacterial inclusions appear to serve as an index of the normal polarity of a spermatocyte.

Yanders <u>et al</u>, derived the following parameter, <u>p</u>, as an estimate of the probability that the bacteria will be located at the nonfunctional pole of the anaphase I spindle:

### s = pt + (1 - p)(1 - t)

Where <u>s</u> is the observed frequency of occurrence of the bacteria with the Y chromosome in disjunctional anaphases,

	Cytologi- cal n.d.*	Genetic n.d.*	n.d.* in Polarized cells	sc <sup>4</sup> sc <sup>8</sup> to non gran- ular pole	sc <sup>4</sup> sc <sup>8</sup> in disjunction- al gametes	XY to granules	nullo in n.d.* gametes
28 <sup>0</sup> DD**	. 559	.518	.632	.607	.718	.651	.917
4-8	• 339	.323	.424	.642	.778	.641	.966
18 <sup>0</sup> DD**	.324	.303	.309	.581	.616	.605	. 893
4-8	.235	.147	.156	.553	.550	.643	.647
*n.d. =	nondisjuncti	ion					

Correlation of cytological and genetic observations. Table 12.

= "Double-Drive" \*\*DD

	Cytological Obsei	rvations	Genetic C	bservations		
	<b>Dis</b> junctional	N.D. *	Disjunc- tional	N.D.*		
	X + X/X	0/0 + XY	$\mathbf{X} + \mathbf{X} \mathbf{X}$	0/0 + XY	Disjunc-	Nondis-
	(s)	(s)	(t)	(t)	tional	junctional
	to non-granular pole	to non-granular pole			മ	מ
28 <sup>0</sup> D.D.**	.607	.651	.718	.917	.745	.681
28 <sup>0</sup> 4-8	.642	.641	.778	.966	.755	.651
18 <sup>0</sup> D.D.**	.581	.605	.616	.893	.849	.634
18 <sup>0</sup> 4-8	.553	.643	.550	.647	1.03	.986

Values for the parameter <u>p</u> for disjunctional and nondisjunctional products. Table 13.

\*N.D. = Nondisjunctional
\*\*D.D. = "Double-Drive"

	3 x 2	$x^2$ Test, "Do $x^2 = \frac{\Sigma (T)}{\overline{Y}}$	ouble- $(n^2) - \frac{\pi}{(1 - v)}$	Drive" a <u>G<sup>2</sup> Σn</u> )	t 28 <sup>0</sup> C.	
	Polar. cells	Non-polar cells	Agr	anular cells	Total	
Disjunc. cells	84	43		19	146	(G)
Nondisj. cells	106	33		25	164	
Sample size	190	76		44	310	(Σn)
Item	Computat	ion				<u></u>
Mean	.792	.566	.432	.4709	6 <del>7</del>	∑ means = 1.790
T <sup>2</sup>	7056	1849	361	21316	$g^2$	$\Sigma T^2/n = 69.45$
$T^2/n$	37.14	24.33	7.98	68.76	G <sup>2</sup> /Σn	
	_1	- <u>y</u>		. 529	004	
	<u>۔</u>	$(1 - \overline{y})$		.253	442	
	Σ	$E(T^2/n - G^2)$	/Σn)	.69		

2.72

Table 14. Test for homogeneity of means.

<u>x</u><sup>2</sup>

or with the X and Y chromosomes in nondisjunctional anaphases (see Table 13),  $\underline{t}$  is the coefficient of meiotic drive for X or nullo gametes from disjunctional or nondisjunctional gamete classes respectively. The calculated  $\underline{p}$  values are given in Table 13.

The mean value of p for disjunctional and nondisjunctional products of  $\underline{sc}^4 \underline{sc}^8$  males at 28°C is .703. Yanders et al (1968) reported a similar  $\bar{p}$  value (.741) for  $sc^4sc^8$  at 25-26°C. At 28°C, the p values for disjunctional and nondisjunctional events in "Double-Drive" and  $sc^4sc^8$ are comparable. At  $18^{\circ}$ C,  $sc sc^{4}$  gave p values in close agreement. In "Double-Drive" at 28°C, however, the probability that the polarized bacteria are located at the nonfunctional pole is .849 in disjunctional gametes, and .634 in nondisjunctional gametes. In all cases, the probability of the bacteria being at the nonfunctional pole is greater in disjunctional gametes than in nondisjunctional gametes. The p value for  $sc^4sc^8$  at  $18^{\circ}C$  are in close agreement (difference = .017), as are the p values for "Double-Drive" at  $28^{\circ}C$  (difference = .064). The differences between nondisjunctional and disjunctional p values for "Double-Drive" at  $18^{\circ}C$  and  $\underline{sc}^{4}\underline{sc}^{8}$  at  $28^{\circ}C$  are .215 and .104 respectively.

The probability that the nonfunctional pole will exhibit granules appears to be influenced by environmental conditions such as temperature, and the extent and direction of this influence is dependent upon the chromosomes in the

drive system (or systems). The extent of the influence of temperature may differ in disjunctional and nondisjunctional gametes.

#### VI. DISCUSSION

## The Effect of Irradiation on SD

The depression of the <u>k</u> values in the sperm batches of irradiated males indicates that the mechanism of <u>SD</u> is disturbed by irradiation. The period of sensitivity occurs prior to Anaphase I, probably during the period of synapsis at zygotene of Prophase I. The percentage of aberrations observed per sperm batch is inversely correlated with the <u>k</u> values exhibited. Since the probable origin of these aberrations is irradiation-induced chromatid breakage and subsequent rejoining of non-sister chromatids, the chromatids involved would most likely be in synaptic association. The sperm batches containing the greatest proportion of chromatids affected by these events also exhibited the lowest <u>k</u> values, suggesting that the <u>SD</u> mechanism was most sensitive to irradiation at the time of synaptic pairing.

This interpretation is supported by the work of Sandler, Hiraizumi, and Sandler (1959) who found that inhibition of pairing at the <u>SD Ac(SD</u>) region by inversions in the <u>SD</u><sup>+</sup> homologue greatly depressed the drive of <u>SD</u>. Irradiation at synapsis may reduce the effectiveness of <u>SD</u> by disturbing the intimate association of the chromatids.

Mange (1968) found the <u>k</u> values of <u>SD</u> males most reduced 7 to 11 days following treatment of temperature-

sensitive <u>SD</u> males for a 24 hour period at  $30^{\circ}$  or  $19^{\circ}$ C. The irradiation-induced reduction of <u>k</u> values in these experiments was most pronounced 7 to 10 days after treatment. Therefore, the period during which the <u>SD</u> mechanism is sensitive to temperature appears to coincide with the irradiation-sensitive stage. The <u>k</u> value depression observed after irradiation of <u>SD-72</u> is comparable to that reported by Mange after cold shock of <u>SD-72</u> for 24 hours at  $19^{\circ}$ C. (<u>SD-72</u> was reported to be insensitive to heat treatment.)

Storage of both treated and control males affected the <u>k</u> of <u>SD</u>. Only the first sperm batch of treated males carried aberration products. The sperm in later batches were most probably not yet primary spermatocytes at the time of irradiation. The erratic <u>k</u> values of both the control and experimental groups can be attributed to prolonged storage of the adult male without mating. Storage apparently "unstabilizes" <u>SD</u>. Why storage should reduce the stability of <u>SD</u> is not clear, but sexual abstinence could retard the rate of spermiogenesis, and prolongation of the primary spermatocyte stage could cause disorientation at Anaphase I.

The sex ratios ( $^{99}$ /total) of <u>cn</u> <u>bw</u> progeny of <u>SD/cn</u> <u>bw</u> males were in excess of 60% in all groups. Hiraizumi (1967) reported such an increase in the proportion of females in <u>cn</u> <u>bw</u>, with a reciprocal decrease in the proportion of females in <u>SD</u> progeny. Because of this, he postulated a homology between the X chromosome and the <u>SD</u> <u>Ac(SD)</u> <u>St(SD)</u> complex, and this type of homology would account for the current

results. This would imply that a degree of repulsion exists between the <u>SD</u> bearing II and the X resulting in their tendency to segregate at Anaphase I to opposite poles.

However, a reciprocal decrease in the proportion of <u>SD</u> females was not observed in these data. The <u>SD</u> progeny and the homozygous <u>cn bw</u> stock exhibit similar sex ratios of about 50%. Since <u>SD</u> has been maintained by back crossing it to <u>cn bw</u> every generation, the <u>SD</u> bearing chromosome II should be essentially the only difference between the two stocks.

The absence of a reciprocal depression of the sex ratio in the two classes is difficult to reconcile with Hiraizumi's original homology hypothesis, which implied an "effective pairing" before Anaphase I. To account for these data, a modification concerning the chromosomal sequence of Anaphase I segregation is proposed: 1) If <u>SD</u> precedes the sex chromosomes to the functional or nonfunctional pole at Anaphase I, the probabilities of the X or Y reaching that pole are equal. That is, the arrival of the X at that pole is effectively a random and independent event. 2) If the sex chromosomes undergo Anaphase I segregation prior to the autosomes, and the X chromosome travels to the F-pole, the probability of <u>SD</u> orienting to the F-pole is reduced. The reciprocal Y; <u>SD</u> gamete is nonfunctional.

Effectively, this is unilateral repulsion. The chromosome does not "recognize" sufficient homology with the <u>SD</u> chromosome to disorient its movement at Anaphase I. The

<u>SD</u> complex on chromosome II, however, is somewhat repelled by an X-chromosome-bearing pole. A reciprocal decrease in the <u>SD</u> sex ratio would therefore not occur. The <u>SD</u><sup>+</sup> <u>cn</u> <u>bw</u> chromosome would have an increased potential of reaching the F-pole if the X preceded it. The reciprocal Y; <u>SD</u> gamete is lost. If the X segregated precociously to the nonfunctional pole, the strong drive of the <u>SD</u> chromosome to orient towards the F-pole would be unaltered, or slightly increased. If the drive of <u>SD</u> is somewhat increased by repulsion from an X-bearing nonfunctional pole, the subsequent increase in the proportion of functional Y; <u>SD</u> gametes would compensate for the loss of Y; <u>SD</u> nonfunctional gametes resulting from the repulsion of <u>SD</u> toward an X-bearing functional pole.

## <u>Relationships of Cytological</u> and <u>Genetic Data</u>

Comparison of the rates of nondisjunction observed in primary spermatocytes and recovered in progeny of sib males indicates that the cytological and genetic data are measurements of the same meiotic event. Both methods indicated 1) an increase in the frequency of nondisjunction in  $sc^4sc^8$  when in the presence of <u>SD</u> in "Double-Drive", at  $18^{\circ}$ and  $28^{\circ}C$ , 2) a decrease in nondisjunctional events in both  $sc^4sc^8$  and "Double-Drive" at  $18^{\circ}C$  and 3) comparable rates of nondisjunction in both  $sc^4sc^8$  and "Double-Drive" at  $18^{\circ}$ and  $28^{\circ}C$ .

Yanders <u>et al</u> (1968) concluded that the polarization of the bacteria at Meiosis I was an independent response to

a cytoplasmic gradient which regularly exists in the primary spermatocyte. This gradient renders the meiotic poles unequal, supporting the cytological model for meiotic drive proposed by Peacock and Erickson (1965). Their functionality hypothesis requires a predetermined polarity of the primary spermatocyte, with only one pole yielding functional sperm. On the basis of the <u>p</u> values calculated (<u>p</u> = the probability that the bacteria will be located at the nonfunctional pole at Anaphase I, Yanders <u>et al</u>, 1968), the observed frequency of association of one or the other reciprocal class of disjunctional or nondisjunctional events in  $sc^4sc^8$  is a reasonable estimate of the genetic recovery of that class.

The probability that the pole exhibiting polarized bacteria will be nonfunctional is slightly lower in nondisjunctional cells. In the cases of  $\frac{sc^4sc^8}{sc^8}$  at  $18^{\circ}$ C and "Double-Drive" at  $28^{\circ}$ C, the <u>p</u> values for disjunctional and nondisjunctional cells differ by only .017 and .064 respectively. In  $\frac{sc^4sc^8}{sc^8}$  at  $28^{\circ}$ C and in "Double-Drive" at  $18^{\circ}$ C, the differences are somewhat greater, .104 and .215. The <u>p</u> value differences do not correlate with the characteristic differential in recovery of nondisjunctional reciprocal products reported by Zimmering (1963) and Peacock (1965), and observed in both  $\frac{sc^4sc^8}{sc^8}$  and "Double-Drive" at both temperatures. Although the propensity of the bacteria to be located at the nonfunctional pole at Anaphase I is significant in every category, there appears to be a variance with

temperature, chromosomal constitution, and segregation behavior at Anaphase I.

The 3 x 2  $X^2$  test for homogeneity of means indicated that polar, non-polar, and agranular cells did not differ significantly in frequencies of nondisjunction. Therefore, the polarized cells scored for the association of bacteria with the reciprocal products of disjunctional and nondisjunctional events can be considered to be representative of the population in general.

In addition to the cytological evidence that a polarity exists in the primary spermatocyte prior to Anaphase I movement, the genetic tests of  $sc^4sc^8$  and "Double-Drive" support the hypothesis that functionality is determined prior to chromosomal segregation. If the functional pole were determined by the driving chromosome itself,  $sc^4sc^8$ and <u>SD</u> would be in a competitive situation. Yet, at  $18^{\circ}C$ , the <u>SD k</u> value in "Double-Drive" is characteristic of the drive of this <u>SD</u> alone (.989), and the drive of  $sc^4sc^8$ is increased in the presence of <u>SD</u>. At  $28^{\circ}C$ , the <u>k</u> of <u>SD</u> in "Double-Drive" was decreased (.917), as was the drive of  $sc^4sc^8$ . These results indicate that the <u>SD</u> and  $sc^4sc^8$ systems were both responding to a gradient in the spermatocyte associated with functionality.

Hartl, Hiraizumi, and Crow (1967) proposed that the mechanism of <u>SD</u> was the production of dysfunctional  $\underline{SD}^+$  sperms. Such a mechanism would be independent of any polarity gradient in the primary spermatocyte. This model is formally

equivalent to the chromosome breakage hypothesis concerning <u>SD</u> (Crow <u>et al</u>, 1962) but does not require cytological evidence of actual breakage events.

If <u>SD</u> action is independent of the cytoplasmic gradient to which  $\underline{sc}^4 \underline{sc}^8$  responds, the drive of  $\underline{sc}^4 \underline{sc}^8$ should be unaffected by that of <u>SD</u>. The occurrence of  $\underline{sc}^4 \underline{sc}^8$  with <u>SD</u> or <u>SD</u><sup>+</sup> would be randomly determined, those sperm bearing  $\underline{sc}^4 \underline{sc}^8; \underline{SD}^+$  being rendered dysfunctional. The present data indicate that the drives of these two systems are indeed related, with  $\underline{sc}^4 \underline{sc}^8$  exhibiting an increased drive when the <u>k</u> of <u>SD</u> is higher (at  $18^{\circ}$ C) and a diminished drive when <u>SD's k</u> is lower (at  $28^{\circ}$ C). This is not compatible with the dysfunctionality hypothesis.

Sandler, Lindsley, Nicoletti, and Trippa (1968) consider <u>SD</u> to be in a class of meiotic mutants which disturb normal meiotic behavior of chromosomes. In their germinal cycle model, <u>SD</u>'s action would most likely be exerted after centromeric association and the onset of pairing. The <u>SD</u> locus must pair with the <u>SD</u><sup>+</sup> locus on its homologue in order to effect segregation-distortion (Sandler, Hiraizumi, and Sandler, 1959) and subsequent chromosomal disjunction is normal (Hiraizumi and Hartl, 1968).

The aberrant segregation of the  $\underline{sc}^4 \underline{sc}^8$  chromosome is probably due to its gross heterochromatic deletions. Deleted X chromosomes characteristically exhibit greatly reduced pairing with the Y and nondisjunction (Lindsley and Sandler, 1958). It is possible, however, that a locus

(or loci) for normal genetic control of sex chromosome disjunction and Anaphase I segregation is included in the segment deleted from  $\frac{sc^4sc^8}{sc^8}$ .

While both <u>SD</u> and  $\underline{sc}^4 \underline{sc}^8$  exhibit aberrant segregation, they do so for different reasons. In the case of  $\underline{sc}^4 \underline{sc}^8$ , the deletion of proximal heterochromatin produces a chromosome which is much smaller than the normal X, and considerably smaller than the Y. It has been shown that when the homologues differ in size, the smaller of the two is recovered more frequently (Novitski, 1956a). On the other hand, there is no difference in size between <u>SD</u> and its <u>SD</u><sup>+</sup> homologue, and the <u>SD</u> locus itself, presumably a single site in the heterochromatin, is responsible for the aberrant segregation. This is a qualitative difference in homologues, while that of  $\underline{sc}^4 \underline{sc}^8$  is quantitative. Nevertheless, both <u>SD</u> and  $\underline{sc}^4 \underline{sc}^8$  exhibit a pronounced meiotic drive, even though the mechanisms initiating the drives are dissimilar.

The model which best fits these results is the functional-nonfunctional pole hypothesis rather than the dysfunction hypothesis. If the chromosomes themselves determined polar functionality, as is required by the dysfunction model, the two driving systems would compete and one or both drives would be diminished. The present data do not indicate such competition. The cytologically observed drive of  $\frac{sc^4sc^8}{sc^8}$  away from the granular pole at Anaphase I and its comparable genetic recovery support the existence of a predetermined functional-nonfunctional polarity

in the primary spermatocyte. Although the <u>SD</u>-bearing chromosome II is not cytologically distinguishable from its  $\underline{SD}^+$  homologue, the concomitant elevation and depression of the <u>SD k</u> value and  $\underline{sc}^4 \underline{sc}^8$  evident at  $18^{\circ}$ C and  $28^{\circ}$ C indicates that the mechanism of <u>SD</u> is not independent of the polarity gradient to which  $\underline{sc}^4 \underline{sc}^8$  responds.

#### V. SUMMARY

The research was designed 1) to ascertain the effect of irradiation at the onset of meiosis on <u>Segregationdistorter</u> in order to determine the meiotic stage during which the mechanism of <u>SD</u> is sensitive, and 2) to observe the meiotic drive systems of <u>SD</u> and the  $sc^4sc^8$  X together ("Double-Drive") both cytologically and genetically at 18<sup>°</sup> and 28<sup>°</sup>C. Analysis of the interaction of these systems and correlation of the nonrandom polar segregation of  $sc^4sc^8$ with its genetic recovery tested the hypothesis that the mechanism of both drive systems involves a predetermined functional-nonfunctional polarity which is independent of the drive systems which respond to it.

In the first study, irradiation treatments were administered to very late third instar male <u>SD</u> larvae. To observe the effects of sperm storage, part of the experimental and control groups were aged after eclosion as virgins. The first four daily broods from unstored irradiated males exhibited depressed <u>k</u> values. The percentage of aberrations and the depression of the <u>k</u> values per brood appear to be positively correlated. Using the proportion of aberration products as an index, stored males appear to transfer few sperm which were in meiotic or premeiotic stages at the time of treatment.

Depressed <u>k</u> values were exhibited by both irradiated and control groups which were stored. Lack of mating activity and possibly resultant prolongation of Meiosis I appear to "unstabilize" and depress the drive of SD.

The sex ratios ( $^{99}$ /total) of <u>cn</u> <u>bw</u> progeny in experimental and control groups were in excess of 60%. A modification of Hiraizumi's <u>SD-X</u> chromosome homology hypothesis is presented to account for the lack of a reciprocal decrease in the sex ratios of <u>SD</u> progeny.

In the study of "Double-Drive", the nonrandom segregation of  $\underline{sc}^4\underline{sc}^8$  or the nondisjunctional nullo product away from the bacterial pole correlated with genetic recovery, supporting the hypothesis that a predetermined functionalnonfunctional polarity exists in the primary spermatocyte. At  $18^{\circ}C$ ,  $\underline{sc}^4\underline{sc}^8$  expresses increased drive when in the presence of <u>SD</u>. Both drives are slightly reduced when together at  $28^{\circ}C$ . Thus, the data indicate that the drive systems of <u>SD</u> and  $\underline{sc}^4\underline{sc}^8$  are not competitive in their interaction, and that the mechanism of <u>SD</u> is not independent of that to which  $\underline{sc}^4\underline{sc}^8$  responds. BIBLIOGRAPHY

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#### **BIBLIOGRAPHY**

- Beadle, G. W., and B. Ephrussi, 1936 "The Differentiation of Eye Pigments in <u>Drosophila</u> as Studied by Transplantation." <u>Genetics</u> 21: 225-247.
- Carpenter, J. M., 1950 "A new semi-synthetic food medium for Drosophila." Drosophila Information Serv. 24:96.
- Cooper, K., 1950 "Normal spermatogenesis in <u>Drosophila</u>", In: <u>Biology of Drosophila</u>, M. Demerec (ed.), John Wiley and Sons, New York, pp. 1-62.
- Cooper, K. W., 1959 "Cytogenetic analysis of major heterochromatic elements (especially Xh and Y) in <u>Drosophila</u> <u>melanogaster</u>, and the theory of 'heterochromatin'." <u>Chromosoma</u> 10: 535-588.
- Crow, J. F., Constance Thomas, and L. Sandler, 1962 "Evidence that the Segregation-Distorter phenomenon in <u>Drosophila</u> involves chromosomal breakage." <u>P.N.A.S.</u> 48, 1307-1314.
- Denell, R., B. Judd, 1968 "Segregation-Distortion in <u>Drosophila</u> <u>melanogaster</u>, the location of a stabilizer of <u>SD</u>." <u>Drosophila Information Serv</u>. 43: 119.
- Erickson, J., and G. Hanks, 1961 "Time of temperature sensitivity of meiotic drive in <u>Drosophila</u> <u>melanogaster</u>." <u>American Naturalist</u> 95: 247-250.
- Gershenson, S., 1933 "Studies on a genetically inert region of the X chromosome of <u>Drosophila</u>. I. Behavior of an X chromosome deficient for a part of its inert region." <u>Journal of Genetics</u> 28: 297-313.
- Greenberg, R., 1962 "Two new cases of <u>SD</u> found in nature." <u>Drosophila Information Service</u> 38:76.
- Hartl, D. L., Y. Hiraizumi, and J. F. Crow, 1967 "Evidence for sperm dysfunction as the mechanism of segregationdistortion in <u>Drosophila</u> <u>melanogaster</u>." <u>P.N.A.S</u>. 58, no. 6, pp. 2240-45.
- Hiraizumi, Y. and D. Hartl, 1968 "Evidence for normal chromosomal disjunction in Segregation-Distortion in males." <u>Drosophila Information Service</u> 40:72.

- Hiraisumi, Y., and K. Nakazima, 1965 "Segregation-Distortion in a natural population of <u>Drosophila</u> <u>melanogaster</u> in Japan." <u>Drosophila Information</u> Service 40:72.
- Hiraizumi, Y., and K. Nakazima, 1967 "Deviant sex ratio associated with Segregation-Distortion in <u>Drosophila melanogaster</u>." Genetics 55: 681-692.
- Kaufman, W. D. and M. Demerec, 1942 "Utilization of sperm by the female <u>Drosophila melanogaster</u>." <u>American</u> <u>Naturalist</u> 76: 445-469.
- Lefevre, G., Jr. and U.-B. Jonsson, 1962 "Sperm transfer, storage, displacement, and utilization in <u>Drosophila</u> <u>melanogaster</u>." <u>Genetics</u> 47: 1719-1736.
- Lindsley, D. L., and Grell, E. H. 1944 <u>Genetic Variations of</u> <u>Drosophila</u> <u>melanogaster</u>, Carnegie Institution of Washington Publication No. 627, Library of Congress Catalog Card Number 68-15915.
- Lindsley, D. L., and L. Sandler, 1965 "Meiotic behavior of tandem metacentric compound X chromosomes in <u>Drosophila</u> <u>melanogaster</u>." <u>Genetics</u> 51: 223-245.
- Kaplan, W. D., V. E. Tinderholt, and D. H. Gugler, 1962
  "The number of sperm present in the reproductive
  tracts of <u>Drosophila melanogaster</u> females."
  Drosophila Information Service 36:82.
- Mange, E. J., 1961 "Meiotic Drive in natural populations of <u>Drosophila melanogaster</u>. VI. A preliminary report on the presence of segregation-distortion in a Baja California population." <u>American Naturalist</u>. 95: 87-95.
- Mange, E. J., 1968 "Temperature sensitivity of Segregation-Distortion in <u>Drosophila</u> <u>melanogaster</u>." <u>Genetics</u> 58: 399-413
- Novitski, E., 1951 "Non-random disjunction in Drosophila" <u>Genetics</u>. 36: 267-280.
- Novitski, E., and I. Sandler, 1956a "Are all products of spermatogenesis regularly functional?" <u>P.N.A.S</u>. (Wash.) 43: 318-324.
- Novitski, E., and I. Sandler, 1956b "Further notes on the nature of non-random disjunction in <u>Drosophila</u> <u>melano-</u> <u>gaster</u>." <u>Genetics</u> 41: 194-206.

- Peacock, W. J., 1965 "Nonrandom segregation of chromosomes in <u>Drosophila</u> males." <u>Genetics</u> 51: 573-583.
- Peacock, W. J., and J. Erickson, 1964 "An indication of polarity in the spermatocyte." Drosophila Information Service 39: 107-108.
- Peacock, W. J., and J. Erickson, 1965 "Segregation-Distortion and regularly nonfunctional products of spermatogenesis in Drosophila melanogaster." Genetics 51: 313-328.
- Sandler, L. and G. Braver, 1954 "The meiotic loss of unpaired chromosomes in <u>Drosophila</u> <u>melanogaster</u>." <u>Genetics</u> 39: 365-377.
- Sandler, L., and Y. Hiraizumi, 1959 "Meiotic drive in natural populations of <u>Drosophila melanogaster</u>. II. Genetic variation at the Segregation-distortion locus." <u>P.N.A.S.</u> U.S. 45: 1412-28 1960a "IV. Instability at the <u>SD</u> locus." <u>Genetics</u> 45: 1269-1287. 1961b. "VIII. Conditional Segregation-distortion: A possible nonallelic heritable aging effect on the phenomenon of Segregation-distortion." <u>Canadian</u> Journal of Genetics and Cytology 3:34-36.
- Sandler, L., Y. Hiraizumi, and I. Sandler, 1959 "Meiotic drive in natural populations of Drosophila melanogaster. I. The cytogenetic basis of segregation-distortion." <u>Genetics</u> 44: 233-50.
- Sandler, L., D. L. Lindsley, B. Nicoletti, and G. Trippa, 1968 "Mutants affecting meiosis in natural populations of Drosophila melanogaster." Genetics 60: 525-558.
- Yanders, A. F., J. G. Brewen, W. J. Peacock, and D. J. Goodchild, 1968 "Meiotic drive and visible polarity in <u>Drosophila</u> spermatocytes." <u>Genetics</u> 59: 245-253.
- Zimmering, S., 1963 "The effect of temperature on meiotic loss of Y chromosomes in the male <u>Drosophila</u>." <u>Genetics</u> 48: 133-138.
- Zimmering, S., and R. E. Green, 1965 "Temperature dependent transmission rate of a univalent X chromosome in the male <u>Drosophila melanogaster</u>." <u>Canadian Journal of</u> <u>Genetics and Cytology</u> 7: 453-456.
- Zimmering, S., and M. Perlman, 1962 "Modification of abnormal gametic ratios in Drosophila. III. Probable time of the A-type effect in Bar of Stone translocation males." <u>Canadian Journal of Genetics and Cytology</u> 4: 333-336.

APPENDIX

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### APPENDIX

Basc

constitution: In(1)sc<sup>SlL</sup>sc<sup>8R</sup>+S, sc<sup>Sl</sup>sc<sup>8wa</sup> B. synthesis: Muller synonym: M-5: Muller-5 references: Spencer and Stern, 1948, Genetics 33: 43-74 properties: Male and homozygous female viable and fertile; X/O male poorly viable, variegated for y, ac, and probably 1(1)Jl. Suppresses crossing over in X, but less so than Binsc because In(1)S = In(1)6Al-3;10Fl0-11Al less effective than In(1)dl-49 =In(1)4D7-El;11F2-4. Routinely used in detection of sex-linked recessive lethals.

#### <u>cn: cinnabar</u>

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location: 2-57.5
origin: Spontaneous
discoverer: Clausen, 2018.
references: 1924, J. Exptl. Zool. 38: 423-36
phenotype: Eye color bright red, like v or st.
    Ocelli colorless. Eye color darkens with age, but
    ocelli remain colorless. Larval Malpighian tubes
    pale yellow (Beadle, 1937, Genetics 22: 587-611).
    Nonautonomous in development of pigment of
    transplanted eye disks (Beadle and Ephrussi, 1936,
    Genetics 21: 230), cn blocks conversion of
    kynurenine to 3-hydroxyhynurenine, which has
   been identified as the cn<sup>+</sup> hormone (Butenandt,
    Weidel, and Schlossberger, 1949, Z. Naturforsch.
    4b: 242-44). RK1.
cytology: Proximal to 44C, based on its inclusion in
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Dp(2;3)P32 = Dp(2;3)41A;42D-E;44C-D;89D7-E1(E. B. Lewis).

#### <u>bw</u>: <u>brown</u>

location: 2-104.5
discoverer: Waaler, 19j15.
references: 1921, Hereditas 2: 391-94.
Sturtevant and Beadle, 1939, An Introduction to
Genetics, Saunders, p. 64 (fig.).
phenotype: Eye color light brownish wine on emergence,
darkening to garnet. Red pigments lacking; ommochromes at 87 percent normal level (Nolte, 1954,

J. Genet. 52: 111-26). Adult testes and vasa colorless. Larval Malpighian tubules pale yellow (Beadle, 1937, Genetics 22: 587-611). Produces white eyes in combination with v, cn, or st. Eye color autonomous when transplanted into wild-type host (Beadle and Ephrussi, 1936, Genetics 21: 230). RK1.

cytology: Placed between 59D4 and 59El by Bridges
[1937, Cytologia (Tokyo), Fujii Jub. Vol. 2: 74555], on the basis of its exclusion from the inner
inversion of In(2LR)bw<sup>Vl</sup> = In(2LR)2lC8D1;60Dl-2 + In(2LR)40F;59D4-El and its inclusion
in In(2R)bw<sup>V De2</sup> = In(2R)41A-B;59D6-El. Based on
the study of bw rearrangements, Slatis (1955,
Genetics 40: 5-23) tentatively places bw in 59D9,
10, or ll.

other information: Separable into at least two subunits by recombination with bw and bw<sup>75</sup> about 0.001 units to the left of bw<sup>59</sup> and bw<sup>81</sup> (Divelbiss, 1961, Genetics 46: 861).

## <u>y: yellow</u>

location: 1-0.0. origin: Spontaneous. discoverer: E. M. Wallace, lla references: Morgan and Bridges, 1916, Carnegie Inst. Wash. Publ. No. 237: 27. phenotype: Body color yellow; hairs and bristles brown with yellow tips. Wing veins and hairs yellow. Tyrosinase formed in adults (Horowitz). For the most part, y is autonomous in mosaics; tissue, however, over limited distances there is some nonautonomy [Hannah, 1953, J. Exptl. Zool. 123: 523-60 (fig.)]. Larval setae and mouth parts yellow to brown, hence distinguishable from the dark brown of wild type (Brehme, 1937, Proc. Soc. Exptl. Biol. Med. 37: 578-80; 1941, Proc. Natl. Acad. Sci. U.S. 27: 254-61). RKl cytology: Placed in region 1A5-8 on basis of its being carried by the  $X^{D}3^{P}$  element of  $T(1;3)sc^{260-20} = T(1;3)1A8-B1;61A1-2$  and by  $Dp(1;f)sc^{260-27} = Df(1;f)1A8-B1;19F$ , but not being lost from Df(1)260-5 = Df(1)1A4-5 (Sutton, 1943, Genetics 28: 210-17). In(l)sc<sup>4L</sup>sc<sup>8R</sup>: Inversion(1) scute-4 Left scute-8 Right cytology:  $In(1)1B3-4;19F-20C1^{L}1B2-3;20B-D1^{R};$ duplicated for 1B3, mitotic chromosomes deficient for the proximal third of hD, all of hC and hB, and

the distal majority of hA (Cooper, 1959, Chromosoma 10: 525-88). About 0.6 the length of a normal X

at metaphase.
origin: Recombinant containing left end of In(1)sc<sup>4</sup> and right end of  $In(1)sc^8$ . discoverer: Gershenson. 1933, J. Genet. 28: 297-313 reference**s:** 1933, Biol. Zh. (Moscow) 2: 145-59, 419-24. genetics: Duplicated for the sc locus, carrying both  $sc^4$  and  $sc^8$ ; deficient for the bb locus and the nucleolus organizer [i.e., Df(l)bb<sup>G</sup>]. Shown by Ritossa and Spiegelmann (1965, Proc. Natl. Acad. Sci. U.S. 53: 737-45) to be deficient for all the DNA that is complementary to ribosomal RNA present in a haploid chromosome set. In the male, In(1)sc<sup>4L</sup>sc<sup>8R</sup> frequently fails to pair with the Y and when it does the unpaired X and Y usually proceed to the same pole (Peacock, 1965, Genetics 51: 573-83). Furthermore, reciprocal meiotic products are not recovered with equal frequency, which Peacock interpreted as the result of non-random orientation of the first meiotic division with respect to the postulated functional pole of the primary spermatocyte. Irregularities in meiotic behavior of In(1)sc<sup>4L</sup>sc<sup>8R</sup> in the male are affected by the Y chromosome present (Peacock, 1965) and the temperature at which meiosis occurs (Zimmering, 1963, Genetics 48: 133-38).  $In(1)sc^{4L}sc^{8R}/Y/Y$  male gives quite regular segregation of the two Y's and low recovery of the X.

