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DEVELOPMENT OF IMPROVED DENSITY ESTIMATORS FOR LARVAE OF THE CEREAL LEAF BEETLE, <u>OULEMA MELANOPUS</u> (L.)

Ву

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A DISSERTATION

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ABSTRACT

DEVELOPMENT OF IMPROVED DENSITY ESTIMATORS FOR LARVAE OF THE CEREAL LEAF BEETLE, OULEMA MELANOPUS (L.)

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Good pest management decisions begin with an accurate estimate of pest density and rely on close predictions of future densities. Sweepnet sampling was evaluated as a low cost method of arriving at an index of density. Attempts at determining instar specific multiplication factors to convert catch per sweep to catch per square foot lacked precision. Incorporation of degree days, windspeed, rainfall and crop height into the conversion factors did little to improve the model. Use of logarithmic transforms (i.e., a multiplicative model) improved the characteristics of the statistical residuals slightly. An additional data set was also evaluated, but this served only to cloud the picture, casting doubt on the generality of the conversion factors.

"Two linear feet" or square foot quadrat samples were also evaluated. Instability of the variance of egg and larval counts was characterized by a double log second order relationship with the mean. Taylor's "power law" was brought to question as a result and was found to be subject to density dependent mortality effects. Negative binomial statistics were calculated for all samples and a common k was derived and found to be independent of mean density and population age structure. The search for variance stabilizing transforms led to selection of a simple log transform over inverse hyperbolic sine, power or more complex log transforms, although the differences were small.

Of several environmental factors, crop height and degree days were found to be of most use in fitting density data. Instar, rainfall, field moisture and sprouting date by themselves were less significant, though some of this may have been due to measurement error. A simple regression of degree days (2nd order), crop height and an interaction term was used to develop a filter for conserving previous measurement information, allowing its incorporation into present measurement estimates. Extension of the density rate change equation to prediction of seasonal incidence curves was also attempted and may have general utility in extension work.

Extension of sweepnet, quadrat and filtering from CLB larvae to the problem of sampling internal larval parasitoids was impaired by inaccuracies in the dissection data. to Ron Wilson, Bert Murray, and Jack Mullins for getting it all started

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Kingston, R.I.
November 15, 1977

"...always bear two points in mind: firstly, that to the biologist computation is always a means to an end, the real objective being some greater insight into a biological problem; secondly, that the ease with which computation is now possible places on the biologist an even greater responsibility than before to understand something of the principles on which his numerical or statistical analyses are based."

(Davies, 1971)

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INTRODUCTION

1.1 Statement of the thesis problem

The absolute density of a phytophagous insect population is assumed to relate predictably to future densities and thus to the potential for insect induced crop damage. Decisions on whether or not to suppress the insect are based on these predictions. The value of a pest management decision depends on information that is only as good as the accuracy of the estimate of absolute density and the closeness of the pest's predicted density to its actual future density.

Many pest ecosystems include beneficial predators or parasitoids. Optimization of management decisions over more than one year requires a prediction of interactions between pests and beneficials. The limitations of sampling and prediction are confounded when more than one population is involved.

This thesis is an attempt to improve the ability of pest management specialists to estimate and thence to predict larval densities of a small grain pest, <u>Oulema melanopus</u> (L.) (Coleoptera: Chrysomelidae), the cereal leaf beetle (CLB). Since the density of the larval parasitoid <u>Tetrastichus julis</u> (Walker) (Hymenoptera: Eulophidae), may also be obtained from CLB larval sampling, this problem will also be discussed.

There are at least two reasons for wanting to improve density estimates and predictions. First, a sample taken early in the growing season could be used as a basis for implementing appropriate management

strategies during the same season. Second, management strategies must also include consideration of the between generation population dynamics of the beetle and its parasitoids. For example, a slightly increased loss during the current growing season resulting from refraining from insecticide use may be more than offset by the beneficial effects of parasitoids in subsequent years. Prediction of between generation dynamics rests on the accuracy of initial density estimates and on the corresponding projection of the within generation dynamics of both populations.

This thesis, accordingly, deals with two problems. The first is the problem of estimating absolute density of the larval cereal leaf beetle. The second is predicting within generation dynamics of CLB larvae. Before evaluation of the accuracy of the density estimate and prior to analysis of environmental or biological factors affecting the dynamics, fundamental statistical problems must be addressed. Most of the concern of this thesis is thus directed toward solution of these underlying statistical questions.

Data for the following studies was gathered under a joint pilot research program involving the Agricultural Research Service (USDA) and Purdue and Michigan State Universities, titled "The Effect of Pubescent Wheat on the Population Dynamics of the Cereal Leaf Beetle". This program is being conducted (1975-1978) in a 16 square mile area near Galien, Michigan, where the CLB was first discovered in North America. In 1975, the area had a high density of beetles and well established populations of egg and larval parasitoids.

Although this thesis is meant to emphasize the goals stated above, additional information from the Pubescent Wheat Study (PWS) will be

included in appendices for future reference. The results of 3 years of MSU CLB population dynamics survey work are also relegated to the appendix. Finally, a laboratory and field study of some aspects of noncrop populations of the CLB and $\underline{\mathbf{T}}$. $\underline{\mathbf{julis}}$ are also included in the appendix. In this way I hope to attain a meaningful balance between conciseness in reaching thesis goals and completeness in presenting reference data for future use.

1.2 Literature review

The cereal leaf beetle is an economically important pest on small grains. Originating in Europe, it was found in southwest Michigan in the 1950's but was not identified until 1962 (Castro, Ruppel, and Gomulinski 1965). Information for constructing life table responses to several environmental parameters was obtained by Yun (1967), Castro et al. (1965) and by Shade, Hansen, and Wilson (1970). Helgeson and Haynes (1972) have presented a model for within generation population dynamics. Haynes (1973) has reviewed features of CLB life history of most importance in developing pest management models. Several aspects of the ecology of the CLB and its principle larval parasitoid, <u>T. julis</u>, have been summarized in a simulation of the beetle ecosystem by Tummala, Ruesink, and Haynes (1975).

T. julis was first released at the MSU W. K. Kellogg Biological Station Experimental Farm in 1966 and was established by 1969 (Stehr 1970). This parasitoid is bivoltine and has a facultative diapause. Some of the offspring of the first generation emerge in midseason while the rest enter diapause. Diapausing first generation parasitoids and all of the second generation winter in soil in the pupal cells of the host. There are usually 4 to 6 parasitoids per host larva.

North American releases of other CLB parasitoids are documented by Dysart, Maltby, and Brunson (1973). These include the egg parasitoid Anaphes flavipes (Foerster) and 3 larval parasitoids, Lemophagus curtus Townes, Diaparsis carinifer (Thompson), and Diaparsis new species. It is uncertain whether D. carinifer has become established to date (Miller 1977). Further subcolonizations and recoveries made by the CLB Parasitoid Rearing Laboratory, Niles, Michigan, are updated annually by the Animal and Plant Health Inspection Services (USDA-APHIS 1972a,b; 1974).

<u>T. julis</u> was the subject of an extensive subcolonization program throughout the lower peninsula in 1971-1974. Agents from the Michigan Cooperative Extension Service collected parasitized larvae from the Kellogg Farm area and released them at preselected nursery sites in their home counties. A follow-up Extension Service recovery program in 1972-1975 revealed widespread establishment and dispersal (Fig. 1; Appendix 1).

1.3 Biomonitoring and cereal leaf beetle management

In an optimal pest management program the tactics chosen should supplement the effects of natural control agents if possible (Rabb 1970). Following its successful subcolonization, \underline{T} . \underline{julis} may be regarded as a natural control agent for the cereal leaf beetle. Pest control recommendations should be adjusted accordingly. This requires identification and quantification of the relationships between pest density, a critical set of biotic and abiotic factors (eg. parasitoid density and accumulated heat units) and crop damage.

To a certain extent, many biotic states can be related to abiotic

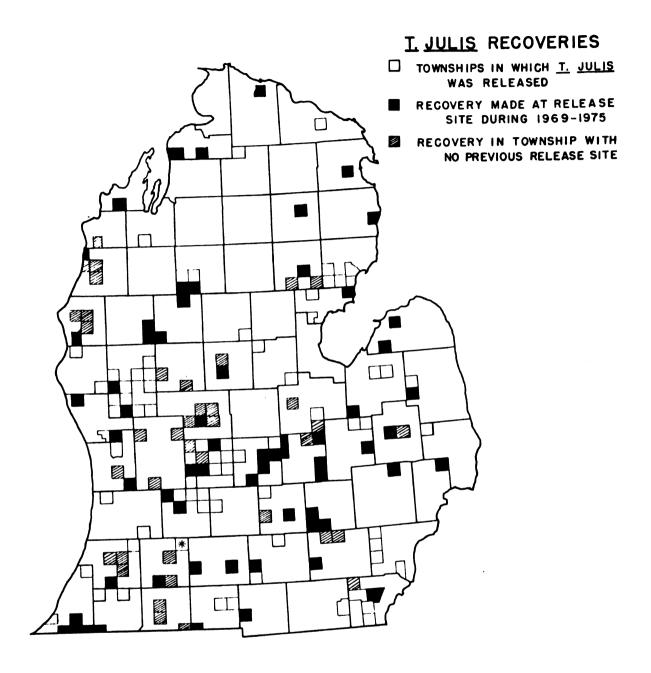


Fig. 1. Michigan townships in which <u>T. julis</u> has been released and/or recovered, 1966-1975.

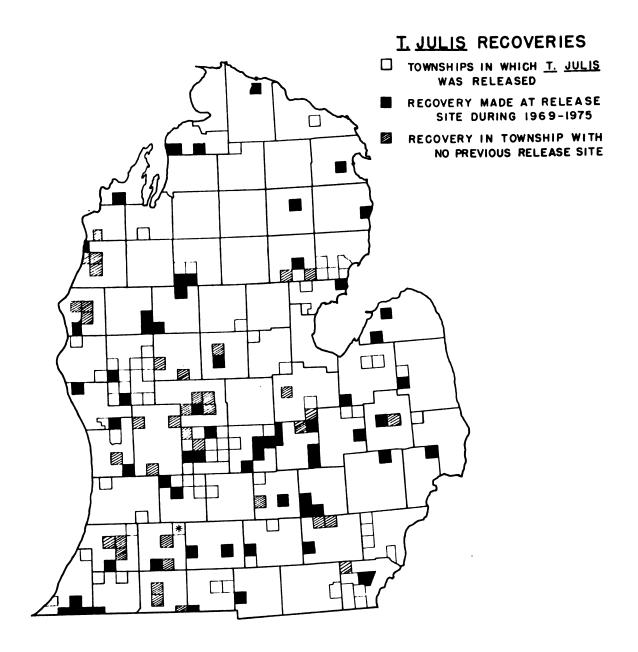
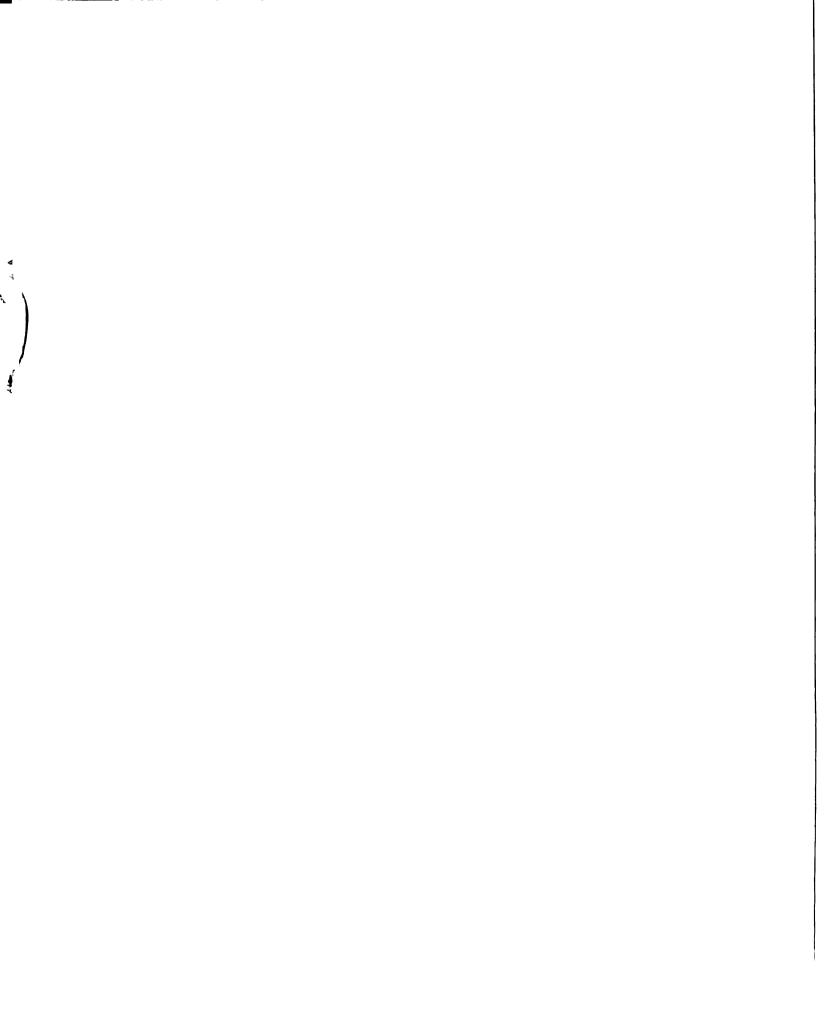


Fig. 1. Michigan townships in which \underline{T} . $\underline{\text{julis}}$ has been released and/or recovered, 1966-1975.



factors. Plant growth, for example, may be predicted from plant age, rainfall, temperature, and soil nutrients. Since monitoring abiotic factors can often be done cheaply and in an automated fashion (Haynes, Brandenburg and Fisher 1973), there have been attempts to use abiotic measurements to make regional projections of biotic states (eg. population maturity) (Fulton and Haynes 1977). The reliability of population projections is restricted by the fidelity of the predictive models and the accuracy of the initial measurements on the population.

Since projection errors will arise from even the highest resolution models, periodic field measurement are needed for recalibration. The nature of these measurements, ie. the design of the sampling program, depends on the type of projection being made. An individual farmer will tend to be most interested in the within generation dynamics of a pest. His sampling program would be intensive, marked by somewhat detailed and frequent counts of pest density, measurements of average larval size, feeding damage, etc. If a population management approach is being used, between generation dynamics of the pest and major biological control agents should also be of interest. Here, the sampling would be over a larger region and would be limited in detail and frequency by the logistics and economics involved.

A general requirement of pest management schemes is that the cost of information must be lower than the cost of proceeding without it (Headley 1975). For several years, the sweepnet has been used to sample CLB adult and larval populations as part of a continual monitoring of several major small grain producing regions in Michigan, Indiana, Ohio, Pennsylvania and Canada (Fulton, Haynes, unpublished reports). The sweepnet has the advantages of low cost per sample,

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minimal time and labor requirements, and minimal damage to the crop sampled. It is also a simple tool, making it possible for relatively untrained personnel to serve as biomonitors.

The sweepnet's value as a sampling tool rests on the assumption that the number of insects per sweep can be converted to an estimate of absolute density. Ruesink and Haynes (1973) developed a multiplicative conversion factor for adult CLB's that was affected by crop height, wind, temperature, and solar radiation. For larvae, they could find no relationship between the conversion factor and the environmental factors. They settled on a coefficient of 1.02 to convert sweep catch to a square foot density estimate. Recent questions (Gage 1974, Fulton 1976) have prompted a reevaluation of this relationship. This is presented in section 2.

In addition to possible biases in sweep catch, the sweepnet is not useful when the crop is short. For the intensive early season sampling needed for the within generation forecasts of most concern to farmers, a more direct population estimate is needed. Square foot quadrats (or equivalently 2 linear feet of crop row) and individual stem counts have been used as sample units. Square foot units were used in this study because past experience indicated this provided the best tradeoff between variance and sample cost (Southwood 1966, Helgeson and Haynes 1972).

A statistical artifact of such counts is the dependency of their variance on the mean. There are two problems caused by this. Since counts which have a higher mean also tend to have a higher variance, a standard error on the estimate is not usable (Sokal and Rohlf 1969, p 381f). This must be replaced by a skewed confidence interval with width dependent on the mean. The second problem caused by the insta-

bility of the variance (ie. dependency of the variance on the mean) is the violation of normality assumptions necessary for performing hypothesis tests or analyses of variance. These difficulties are discussed in section 3.

Density is a function not only of time but also of accumulated heat, rainfall, windspeed, and probably other environmental factors. Many of the relationships between environmental factors and fecundity, mortality, and development rates have been studied previously (Castro et al. 1965, Yun 1967, Helgeson and Haynes 1972, Gage 1974). The role of these factors is reexamined in section 4 using new data and a transformation developed in section 3.

Estimates of changes in populations of \underline{I} . \underline{julis} are most directly made by sampling adults. However, the parasitism rates of CLB larvae and pupae also contain usable information on parasitoid density. Larval samples are much easier and less costly to obtain than pupal samples. For purposes of between generation regional projections of parasitoid densities, larval samples may be optimal, despite the difficulties of estimating seasonal density. These problems and estimation procedures are discussed in section 5.

A CEREAL LEAF BEETLE LARVAL SWEEPNET MODEL

2.1 The sweepnet as a sampling tool for cereal leaf beetles

Accuracy in biomonitoring is the result of the mechanical efficiency of the sampling method in detecting (capturing) the creature being monitored. Although direct counts of such torpid insects as cereal leaf beetle larvae are generally accurate, the cost of making enough counts to obtain a minimal acceptable sampling variance (ie. variance within 10% of the mean) may be prohibitive for most survey programs. A less accurate index such as sweepnet count is frequently the only acceptable alternative.

The sweepcatch index is related to other factors in addition to density (Morris 1960). Delong (1932), in one of the first reports on factors affecting sweep efficiency, reported that capture of active insects such as leafhopper adults varied with temperature, plant height, and type of stroke or backstroke. Hughes (1955) found wind affected catches of a chloropid fly in wheat. Larval size and time of season affected sweepnet efficiency with green cloverworm in soybeans (Shepard, Carver and Turnipseed 1974). Time of day and changes in vertical distribution also affected sweep catch of several small grain insects (Vickerman and Sunderland 1975).

Ruesink and Haynes (1973) developed 2 models for sweepnet sampling of the cereal leaf beetle. A conceptual model related the "multiplication factor", M, required to convert sweep catch to density, to net diameter, d, length of the sweep stroke, l, the proportion of the



population in the net's path, b, and the probability of getting caught, p, once in the path. That is,

$$M = 1 / (dlbp).$$
 (1)

A regression model for each instar suggested that wind, crop height, temperature or cloud cover did not affect M. There were no conclusive differences in M values between the instars (for the ranges; crop height, 38 to 112 cm; temperature, 24 to 29°C; wind, 6.4 to 16 kph; and solar radiation, .7 to 1.1 cal/cm²/min). Relating this to the conceptual model, there were no detectable differences in instar specific probabilities of getting into the path of the sweepnet or of getting caught once there.

This conclusion was in conflict with an earlier Ruesink data set (Ruesink 1970) which found higher M values for younger larvae (averaging 1.41, 0.84, 0.72, and 0.40 for 1st through 4th instars). However, the inability to accurately determine instar and small sample size (5 fields in the original data set, 10 in the later) made both sets of results inconclusive. The instar determination problem has since been solved (Hoxie and Wellso 1974). Head capsule widths have 4 discrete ranges and are now used regularly to determine the 4 instars. The ranges established by Hoxie have been found to apply to Michigan CLB survey data from 1971-72 (Fulton 1975) and 1973-74 (Logan, unpublished data).

Gage (1974) found that catch per sweep underestimated the total larvae per square foot by at least half, noting that very few 1st instar larvae were being caught in the net. Fulton (1975) reported serious discrepancies between instar proportions in sweepnet samples and the proportions found in square foot samples taken at the same

time and place. Fulton concluded that sweepnets are unreliable as a method to assess average age of the population.

Reasons for the discrepancies in instar specific catch efficiency were not given by any of the above. Ruesink (1970) found more younger larvae on the lower 2/3rds of 38 inch wheat but was unable to establish this as a general rule. Jackman (1976) found the majority of all instars were on the upper 3 blades of oats throughout the season. Wellso and Cress (1973) observed a significant difference in oviposition and feeding between top and next lower blades with most eggs laid on the upper, more succulent leaves. Thus it is still indefinite what factors affect the probability of a CLB larva getting into the path of the sweepnet. Also, there have been no studies of the factors influencing the probability of getting knocked off the plant and into the net.

Because the many advantages of the sweepnet were placed in jeopardy by questions of its accuracy, a large set of sweep and square foot samples were gathered for comparison as part of the PWS (section 1.1) program. Methods used are described in section 2.2. Correlations and simple regressions of sweep and square foot catches are given in section 2.3. Section 2.4 presents an outline of a statistical technique appropriate to analysis of subsample data. Section 2.5 further examines the data to determine whether more detailed statistical models would improve sweepnet conversion factors. Section 2.6 presents an additional data set and its implication (and, alas, complications) for the sweepnet model.

2.2.1 Sweepnet and quadrat sampling methods

Wheat and oats were sampled in the PWS area near Galien. Sweeps were made after the host field was at least 8 to 10 inches tall.

A sweep is one level pass of a 15 inch diameter net. A sweep covers about a 5 foot swath through the top 8 to 15 inches of the crop.

The sweepnet was lined with a new 30 gallon plastic bag for each field to prevent small larvae from working through the mesh and to keep counts from each field distinct. The sweeper took 150 sweeps, beginning 20 to 30 feet from the edge of the field. The choice of 150 sweeps was arbitrary. Previous survey work had used a standard of 100 sweeps. The sweeper walked at a vigorous gait 120 to 180 feet into the field, made a wide u-turn, and exited sweeping until the 150 sweeps were complete. If there appeared to be less than 10 larvae in the bag the sweeper would take 100 more sweeps with the same bag. In practice, workers soon recognized certain fields were sparcely populated and took the 250 sweeps virtually automatically. Having completed the sweeps, the worker washed down the larvae on the sides of the bag with FAA, a preservative, and labeled the bag with field number and date before sealing. We used a formalin:acetic acid:alcohol:water mixture (FAA) in a 2:1:50:47 ratio. This preserved larvae suitably for dissections.

While the sweeper worked, a second sampler made square foot counts. The sampler walked 20 to 30 feet into the field carrying a lightweight 2 foot wooden stake. He tossed this, letting it fall where it would. This was done to help improve the randomness of the quadrat location. Workers took samples from points far apart in the field. The result was a semi-uniform grid of sample areas with a single count taken

randomly in each area. The stick was moved 5 feet down the same row to reduce the disrupting effect of the toss. Care was taken to avoid sampling where the field had just been swept. The sampler picked over the plants in 2 feet of 1 row, recording the number of eggs on a hand counter while mentally counting larvae.

The first 50 larvae found were put in a vial for later study of field age distribution. Vials were labeled with date and field number. They were half filled with water and were returned to the lab and frozen.

After collecting the larvae and counting the eggs in a 2 foot section, the sampler moved to another site and again tossed the stick. A minimum of 10 such sample points per field was about as much as could be taken with PWS resources. However, if 50 larvae had not been collected within this minimum, an extra 5 points were covered or enough to get 50 larvae, which ever came first. When time permitted, separate records of each 2 foot egg and larval count were made to provide a measure of within field variance.

The ruled sweepnet handle was used to estimate average standing crop height. Time of day, number of sweeps and square feet, windspeed (taken with a styrofoam ball anemometer), and field wetness were also recorded. Field wetness was classified as either "dry", "wet" (full of dew or fresh rain), or "damp" (not glistening wet but able to dampen trousers of someone walking through it). When possible, sampling was limited to times when fields were dry.

Sweepnet sample bags were taken to the lab and were stored in 9^{0}C refrigerators until processing. Three weeks' samples had accumulated before processing began but there was no apparent damage to

larvae as a result of the delay. Larvae were removed from the bag by floatation in alcohol.

A sample cleanup unit was constucted from an inverted 5 gallon plastic carboy with the bottom removed (Fig. 2). A large nylon funnel was fit into the carboy to eliminate the shoulder. A 3/4 inch ID clear tygon tube was attached to the bottom of the funnel. A 2 cork system. one on the bottom of the tube and a second attached to a stick and inserted from above into the funnel, allowed rapid isolation of the larvae. The carboy was partially filled with 95% alcohol. The plastic sweep bag was inverted and washed in the alcohol. Insects quickly sank to the bottom, accumulated in the tube, and were drawn off into a 2 oz. plastic cup. Plant debris was skimmed from the surface of the alcohol. Since the volume of alcohol was relatively small, replacement of all the alcohol was infrequent. All that was necessary was a periodic topping up and a daily flushing of the whole system. Two carboys were used and kept one man busy. Labels were transferred to the new cups and date and field number were repeated on the cup lid. After separation alcohol was poured off and fresh FAA was added to the cups.

Larvae and adults were then counted. The contents of a cup were poured onto a tray and sorted. Adults were first removed and their number recorded. Larvae were separated from remaining insect and plant debris as they were counted.

In very large samples (greater than 1000 larvae) a quartering system was used to speed counts. All adults were first counted and removed. A slurry of larvae was made so that larvae were distributed approximately uniformly on the tray. Larvae in 2 opposite quarters were removed. The remaining larvae were again slurried. Counts were

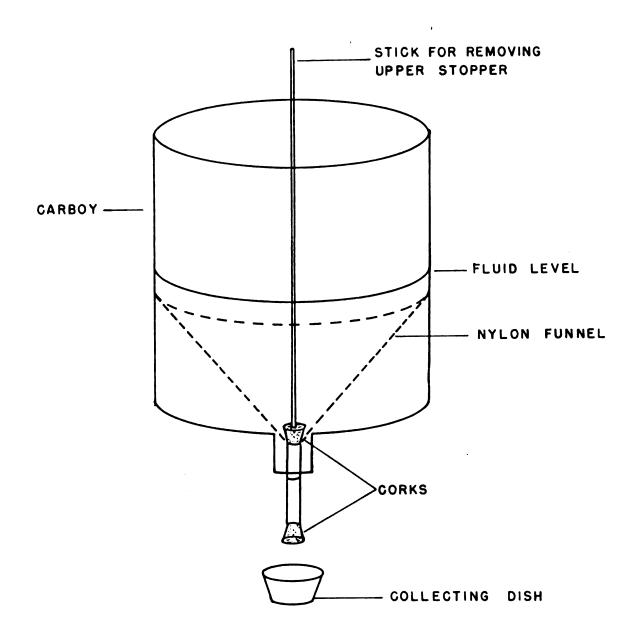


Fig. 2. Carboy unit used to extract cereal leaf beetles from sweepnet sample bags.

made of 2 opposite corners and the result was multiplied by 4. About 5% of all samples had to be treated this way.

Larval instar was determined by measuring head capsules with an ocular micrometer. An aliquot subsample was prepared from the contents of a sample cup. Up to 50 larvae from each sweep collection were measured. All larvae retained from the 2 foot counts were measured. At the same time, the larvae were dissected to determine species, life stage and number of parasites.

2.2.2 Sampling technique errors

Several counting errors can arise during the above processes.

Briefly, these are as listed:

A. Precount errors

- 1) When hand picking, not seeing or losing a larva that is present in the sample quadrat.
- 2) Sweepnet bias, as discussed in section 2.3.
- 3) Transfer losses from sweepnet to count dish.

B. Count errors

- 1) Miscounting.
- 2) Age group misclassification.
- 3) Failure to count any members of an age class.
- 4) Aliquot biases.
- C. Analysis errors, as discussed in section 2.4.

With the PWS data set I did not measure the efficiency of finding larvae in the field. I assume 100% of the larvae in a sample unit are found and counted or returned to the lab, as over optimistic as this may be. The washing down of larvae on the sides of sweepnet bags and later removal from the bag is highly efficient. Once in a particular bag, there is little chance that a larva will not make it into a sample

cup, ready for counting.

Repetition of counts on a few samples of 50 to 200 larvae indicated precision to be within about 3% of the actual number present. Failure to properly classify the development stage of an insect is unlikely because the overlap of head capsule widths between instars is quite small (Hoxie and Wellso 1974) and ocular micrometer measurement is relatively precise.

By limiting the number of measurements taken from sweep samples to 50, the probability of failing to count any individuals in a particular age class is increased. If there are 4 lst instar larvae in a sweep catch of 100, there is a probability of .34 that the lsts will be underestimated and .06 that they will not be counted at all in a subsample of 50, all other biases ignored (Larson 1969). Fulton (1975) suggested that the average age of a sample would be estimated with reasonable accuracy (5% of the mean) when 50 larvae were measured. Logistics restricted the measurements done during the PWS to 50 per subsample.

There was a major failing with the aliquot method. Too many large (4th instar) larvae were sampled. This is reflected in Fig. 3 in which 2 groups of 50 larvae were measured from each of 48 samples. The age estimate from the first group varied by roughly .5 to .2 from the second estimate. Because of this, a second batch of up to 50 larvae were measured for all of the samples used in the following analysis. This meant measurement and dissecting an additional 2271 larvae from the oat samples (for a total of 5677 measurements in 85 cases) and an extra 1100 larvae from the wheat samples (total 4136 in 62 cases). In the oat samples, 45 cases had all the larvae in the net measured. The 40

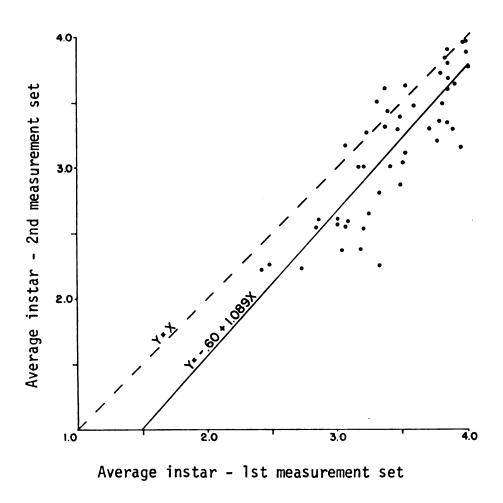


Fig. 3. Error in estimating average age class from sweepnet data.

cases with an incomplete measurement set averaged 89 measurements per case. In the wheat samples, 31 cases had all the larvae measured. The average subsample size for the remaining 31 cases was 70. Table 1 compares the measurement sets further. This includes the slope and intercept coefficients of the square foot and sweep catch regressions.

The problem with interpreting the differences of Table 1 is that there is no way to tell whether the higher proportions of 3rd and 4th instars in the partial measurement samples are the result of measurement bias or are only reflective of the older age structure of the higher density samples. In oats, comparison with associated square foot samples suggests the latter is true. In wheat, there is less of a distinction. The decision to either disgard the incomplete measurement sets or to combine the 2 was resolved in favor of combination (see Table 3), with the caveat that the result may be an overstatement of the relative proportion of older larvae in the sweepnet samples and a corresponding bias in the sweepnet conversion factors.

2.3 Absolute density versus sweep catch: simple correlations and regressions

For a sweepnet index of CLB larval population density to be of use, the catch per sweep of each instar must be correlated to absolute density. Table 2 gives the correlation matrix for the 4 instars in the paired sweep and square foot samples, separated into cases for wheat and oats. Correlation coefficinets significantly different from 0 at the .05, .1, and .2 levels are indicated (Sokal and Rohlf 1969, p. 516). Note the diagonals of the lower left 4 x 4 submatrices, the correlations between sweep and square foot catch for each instar. In oats, the proportion of 4ths in the 2 sample methods is highly

Table 1. The relationship between catch per square foot and catch per sweep for CLB larvae. y_i = average catch per square foot of instar i, i = 1...4. x_i = average catch per sweep of instar i. s.d. = standard deviation of catch. a_i = intercept. b_i = slope. s.e. = standard error of coefficients. Model is y = a + bx + error.

			01	OATS								퓔	WHEAT				
		(EN	(ENTIRE SAMPLE MEASURED)	PLE MEA	SURED)						(ENT	(ENTIRE SAMPLE MEASURED)	LE MEAS	URED)			
			Squa	Square foot	4.1		Sweep					Squar	Square foot			Sweep	а
Cases				45			45		Cases				31			33	
Total measurements	ements			1359			2112		Total measurements	ements			1031			1949	
Instar (i)	٧,	s.d.	.j.	s.e.	اً م	s.e.	× ×	s. d.	Instar (i)	۲,	s.d.	.j.	S. e.	ام	S.e.	, , , , , , , , , , , , , , , , , , ,	s.d.
-	.169	.720	.01	10.	31.57 9.651	159.6	.005 .010	010	-	.629 1.179	1.179	.298	.253	18.195 8.533	.533	. 810.	.024
2	.553	2.211	.199	404	10.797 7.320	7.320	.033 .045	345	2	1.048 1.591	1.591	.426	.475	5.808 3.595	. 595	. 107	.079
e	.672	.672 2.246	767.	.435	843 2.700	2.700	. 411.	.115	m	.899 1.052	1.052	.574	.400	1.435 1.558	.558	. 722.	.124
4	961.	.199	.133	.041	. 393	.393 .185	. 931.	.156	4	.122	.195	600.	.029	1.715 .268	.268	. 990.	.087
Mean Instar 3.113	3.113	965.							Mean Instar 2.332	2.332	.405						
		(PA	RTIAL SA	MPLE M	(PARTIAL SAMPLE MEASURED)						(PAR	(PARTIAL SAMPLE MEASURED)	MPLE ME/	ASURED)			
Cases				6			40		Cases				33			33	
Total measurements	ements			1757			3565		Total measurements	ements		_	1536			2187	
Instar (i)	۲,	s.d.	a i,		اً م	S.e.	x .	s.d.	Instar (i)	y,	s.d.	 e	s. e.	م	s.e.	, ×	s.d.
	.07	.167	.078	.028	075	075 1.680	.004 .016	910	-	1.409 1.734	1.734	.980	.313	8.060 2.731	.731	.053 .103	103
2	.330	. 584	.273	.122	.424	. 583	.136 .161	191	2	1.725	1.704	1.017	.334	.947 .277	773.	.748 .966	996
ю	. 798	.894	.364	.179	.365	.108	1.187 1.180	180	٣	2.375	2.902	1.553	.545	.390	.134	2.109 3.545	545
4	1.043	1.043 1.187	.150	771.	.533		.075 1.676 1.679	679	4	772.	.405	.243	.870	.031 .421	.421	1.111 1.770	770
Mean Instar 3.271	3.271	. 504							Mean Instar 2.335	2.335	.527						

Table 2. Correlation coefficients for count data from oats and wheat. SW1 = estimated catch of instar 1 per sweep; MISQ = mean instar of square foot catches, etc.

ABS (r) = absolute value of the correlation coefficient.

```
(cases = 85)
                                       Oats
SW1
        1.
                                       r significant at .05 if ABS (r) > .214
SW2
         .072 1.
                                       r* significant at .10 if ABS (r) > .180
SW3
        -.026
                .698 1.
                                       r+ significant at .20 if ABS (r) > .140
SW4
         .018
                .350
                       <u>.735</u> 1.
SQ1
         .245
                .028 -.055 -.082
                                        1.
SQ2
         .262
                .052 -.045 -.076
                                        .962 1.
                                        <u>.815</u> <u>.8</u>80 1.
                        .170+ .150+
SQ3
         .221
                .108
         .057
                        .433 _.805
SQ4
                .120
                                        -.095 -.068
                                                       .140 1.
MISW
       -.259
              -.218
                        .047
                              .313
                                        -.435 -.461 -.326 .292
                                                                        1.
MISQ
       -.226
              -.056
                        .085
                              .261
                                        -.327 -.331 -.269
                                                              .338
                                                                        .637 1.
                                                         SQ3
                                                 SQ2
                                                                SQ4
                                                                        MISW MISQ
          SW1
                 SW2
                        SW3
                                SW4
                                          SQ1
                                       Wheat (cases = 62)
SW1
       1.
SW2
        .409 1.
                                       r significant at .05 if ABS (r) > .250
                                       r* significant at .10 if ABS (r) > .211
SW3
        .337
               <u>.636</u> 1.
                                       r+ significant at .20 if ABS (r) > .163
SW4
        .302
                .629
                       .923 1.
        .467
SQ1
                .566
                       .139
                               .156
                                       1.
        .176
               .447
SQ2
                       .376
                               .297
                                        .551 1.
SQ3
        .059
                .408
                       .513
                               .464
                                        .213*
                                               .692 1.
                                                .197+ .342 1.
SQ4
       -.186+
                .107
                       .170+
                               .217*
                                       -.018
       -<u>.387</u> -.025
                       .194+
                              .332
                                       -.203<sup>+</sup> -.006
                                                       .253
                                                               .429
MISW
                                                       .200+
                                                                        .601 1.
       -.445 -.210<sup>+</sup>
                       .180
                                                               .436
                               .146
                                       -.667 -.270
MISQ
                                                                         MISW MISQ
                                                                SQ4
         SW1
                 SW2
                        SW3
                                SW4
                                         SQ1
                                                 SQ2
                                                        SQ3
```

correlated. Firsts and 3rds are also significantly different from 0 correlation at the .05 and .2 levels, respectively. In wheat, the 4 correlations of interest are all different from 0 at a .1 level. The average age as determined by sweepnet has about a 60% correlation with square foot mean instar in both crops.

Table 3 combines the 2 data sets from Table 1 and describes the data in more detail. For oats, 95% confidence limits on the 1st instar square foot over sweep catch regression overlap the limits for 2nd instar. The confidence limits for 3rd instar surround those for 4ths and in turn fall within those for 2nds. For wheat, distinction between the instars is easier as only 2nd and 3rd instars have overlapping confidence limits. Note that only 2nd instar larvae come close to the slope of approximately 1 suggested by Ruesink and Haynes (1973). The general trends tend to support the findings of Gage (1974) and Fulton (1975), reflecting a sweepnet bias against picking up smaller larvae.

It should be clear from the high coefficients of variability and the low coefficients of determination (R² values) (Table 3) that these simple models lack the precision desired for accurate predictions. One difficulty with the regressions is that they all provide nonzero estimates for square foot density even when no larvae are caught in the sweepnet. While this may be useful when the number of sweeps is small (100 or less, say), it may be an overestimation. Table 4 provides the slopes and corresponding confidence intervals for zero-intercept models fit to the same data. These estimators may be more useful when many sweeps are taken at low densities.

An attempt to further improve these regressions by taking into account various environmental factors is made in section 2.5, following

Table 3. Statistics of combined data set, used in simple (y = a + bx) larval sweepnet models.y = catch per square foot; x = catch per sweep.

	s.d.	.077	.752	2.662	1.350	! ! !	s.e	2.272	.256	.095	.030		C.V. R2	133.2 .22		108.7 .20		121.0.26		160.4.05	
	Xí	.036	.428	1.168	. 588	! !	Ď,	9.303	.993	.441	.052	ı	SIG		٠.00	15.0	٠.001	21.4	٠.011	2.9	60.
WHEAT	•					! ! !					1		¥.	30.87	1.84	34.05	2.27	84.05	3.93	.30	.10
	s.d.	1.522	1.670	2.289	. 325	! !	S.e.	.190	.220	.275	.044		SS	30.87	110.49	34.05	136.14	84.05	235.56	. 30	6.14
	ži	1.019	1.387	1.637	.199	! ! !	91	.687	.962	1.122	.169		d.f.	-	09	_	09	-	09	-	09
	_					! ! !	_				•		Source	Regression (R)	Residual Error (E)	œ	ш	œ	w	œ	ш
	Instar (i)	_	2	٣	4	1 1 1	Instar (i)	_	2	e	4	إبيا	Instar (i)	Regres	Residual						
	,					ANALYSIS					1	F VARIANC	Insta	-		2		က		4	
						REGRESSION ANALYSIS					(ANALYSIS OF VARIANCE	R ²	90		00		.03		.65	
	s.d.	.013	.126	.972	1.379	<u> </u>	s.e.	4.308	1.440	.178	.044	4.1	,	413.5		370.5		217.3		92.8	
	xi	.005	180.	.619	.873	! ! !	b;	9.920	.682	.280	•	i i i	S16 C		.024	.224 37	.637	2.47 21	.120		٠.001
	1	•	-•	•	•	! !		0				! !	•	<u>ب</u>				٠		8 152.9	
OATS	s.d.	.535	1.653	1.602	.925	, ,	. s	090.	.215	. 205			MS.	1.45	.27	.62	2.76	6.22	2.52	46.58	.30
	S.	•:	7.6	7.6	Ο;	! ! !		ų.	`*	``.			SS	1.45	22.62	.62	228.76	6.22	209.39	46.58	25.29
	ίż	. 126	.446	.731	.595	' ' '	a.	.081	.393	.558	.123		đ.f.	_	83	_	83	_	83	-	83
	(±)					1 1 1	Ξ				•			Regression (R)	Residual Error (E)	œ	w	œ	w	œ	w
	Instar (1)	~	2	က	4		Instar (1)	-	2	m	₹ .		Instar (1) Source	1 Regres	Residual	2		æ		4	

Table 4. Zero-intercept (y = bx) larval sweepnet models.

		OATS	WHEAT
Instar (i)	b _i	95% C.L.	b _i 95% C.L.
1	11.838	3.708,19.967	12.792 8.294,17.289
2	2.116	0. , 4.552	1.549 1.043, 2.055
3	.542	.233, .852	.598 .403, .793
4	.581	.507, .655	.099 .037, .160

a brief statistical argument, presented for future reference in section 2.4.

2.4 Weighted regression analysis

It is important to note that the instar specific variables "catch per sweep" and "catch per square foot" are themselves estimates. They are based on subsamples of larvae. For example, catch per sweep of 2nd instar larvae in sample j is

$$x_{2j} = (I_{2j}/n_j)(L_j/S_j)$$
 (2)

where I_{2j} is the number of 2nd instar larvae in the subsample n_j ; L_j is the total number of larvae from which the subsample was drawn; and S_j is the number of sweeps taken. Sampling errors for these estimates are related to subsample size. Since the standard error of the sample is

s.e. = SQRT (
$$\Sigma(x_{ij} - \mu_{ij})/n_j^2$$
) (3)

where $\mu_{i,j}$ is the unknown population mean (estimable only in repeated data sets) and SQRT() is the square root operator, sampling error is inversely proportional to square root of subsample size. In this sence larger samples may be said to be more reliable than smaller ones.

There are no standard criteria for evaluating the relative reliability of a sample estimate. By comparing values of 1/n for different n's, Searle (1971, p. 365-73) suggested ratios of such values were possibly useful. Ratios of 4: l or higher were indicative that sample error should be taken into account. It seems clear that there is little loss in reliability in going from a sample size of 100 to 50. This is equivalent to a ratio of 1.41: l, which would rule out an adjustment under Searle's criteria. However, comparing samples of 6 and 50 would result in greater than a 4: l ratio. As Searle noted, the 4: l ratio is an arbitrary norm. If one were to use a more conservative 2: l ratio, adjustment of sampling error would be considered appropriate when less than 12 larvae were present in a sample. This assumes that 50 is the optimal sample size (Fulton 1975). (If 30 were considered optimal, based on some other analysis, 8 larvae would be the lower limit, using the 2: l ratio.)

Comparing plots of sample size for the sweepnet samples with their paired square foot samples in Fig. 4, 21 of the cases in oats (24.7%) have at least 1 of the samples smaller than 12. Only 3 (4.9%) of the samples in wheat are less than 12. This suggests that the age specific density estimates in oats ought to be subjected to a weighted regression analysis to adjust for the large percentage of small, "less reliable", subsamples. Here again, the criteria are arbitrary. As a suggestion for future use, perhaps a maximum of 10% "small" samples should be considered as a basis for deciding the appropriateness of such analysis.

To perform a weighted regression analysis, the total sum of squares and the reductions in sums of squares used in the regression must be weighted. This is done by weighting each term in the sum of squares in inverse proportion to the variance (Searle 1971; Steel and Torrie 1960,

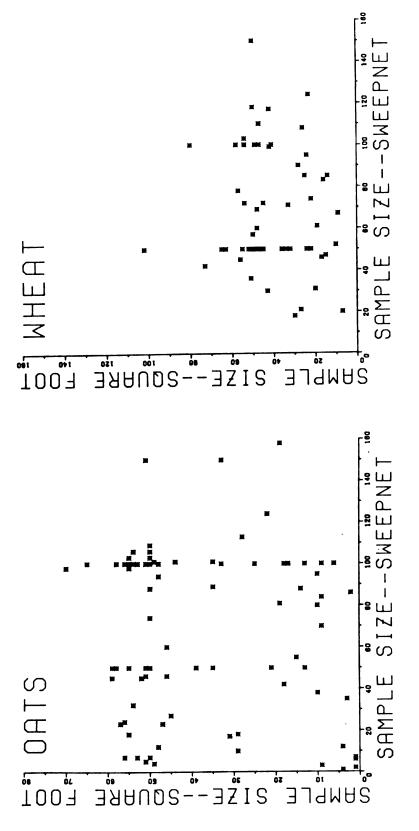


Fig. 4. Distribution of sample sizes for paired data used in sweepnet regressions.

p. 180),

$$w_i = s_v^2 / var(\bar{x}_i)$$
 (4)

where w_i is the weighting coefficient, $s_{\overline{x}}^2$ is the variance of all the sample means for variable x, and $var(\overline{x})$ is the variance associated with the specific subsample mean. The usual simplification of (4), keeping in mind that the variance is inversely proportional to sample size, is to use subsample size over the total number in all subsamples as the weight

$$w_i = n_i/n. (5)$$

where the "n." notation indicates summation over all n (Searle 1966).

Steel and Torrie (1960, p. 181) point out that it is the relative rather than the actual weights that are important. Since samples do not really add new information to the estimates in direct proportion to their sample size, it is proposed that a modification of (5) be used,

$$w_i = SQRT (n_i/n.) .(6)$$

Using a weight proportional to subsample size, a sampling of 100 would count as twice as reliable as a sample of 50. Using weight proportional to square root of subsample size, 100 would only be judged 1.41 times as reliable. Samples less than 12 receive a rapidly decreasing relative weight. This seems a reasonable compromise between using no weights and using subsample size as a weight factor.

Regression based on weighted means uses a weighted variance covariance matrix, each element of which has the general form

$$cov(x_j,x_k) = \sum w_{ij}(x_{ij}-\bar{x}_j)w_{ik}(x_{ik}-\bar{x}_k)$$
 (7)

where summation is over all i, w_i 's are the weights from (6), x_j and x_k are any variables in the regression and \bar{x} 's are weighted means,

$$\bar{x} = \sum (w_i x_i) / \sum w_i$$

Weights are based on (6) for means (number per age class per sample unit) but are set equal to 1 for all other variables. Covariances (and variances by an analogous process) are computed from the expansion of (7),

$$cov(x_{j},x_{k}) = \sum_{i,j} w_{ik}(x_{ij}x_{ik}-x_{ik}\bar{x}_{k}-x_{ik}\bar{x}_{j}+\bar{x}_{j}\bar{x}_{k})$$
 (9)

which for parameters with weights equal 1 reduces the usual form,

$$cov(x_{j},x_{k}) = \left[\sum (x_{i,j}x_{i,k}) - \left(\sum x_{i,j}\sum x_{i,k}\right)/m\right]/(m-1)$$
(10)

where m is the number of cases. A correlation matrix based on the above with elements

$$r_{jk} = cov(x_j, x_k) / SQRT(var(x_j)var(x_k))$$
 (11)

can be used to compute least squares estimates of transformed correlation coefficients, $\mathbf{a_i}$. The normal equations are

$$\begin{bmatrix} 1 & r_{12} & \dots & r_{1n} \\ r_{21} & 1 & \dots & r_{2n} \\ \dots & & & \ddots \\ r_{n1} & r_{n2} & \dots & 1 \\ with solution \end{bmatrix} \begin{bmatrix} a_1 \\ a_2 \\ \dots \\ a_n \end{bmatrix} \begin{bmatrix} r_{1y} \\ r_{2y} \\ \dots \\ r_{ny} \end{bmatrix}$$

$$(12)$$

$$\hat{\underline{\mathbf{a}}} = \underline{\mathbf{R}}_{\mathbf{x}}^{-1}\underline{\mathbf{r}}_{\mathbf{y}} \tag{13}$$

Estimates of the original coefficients are obtained from

$$\hat{b}_{i} = \hat{a}_{i} SQRT(var(y)/var(x_{i})), \qquad (14)$$

for i = 1...n, and from

$$\hat{b}_{0} = \bar{y} - \hat{b}_{1} x_{1} - \hat{b}_{2} x_{2} - \dots - \hat{b}_{n} x_{n}, \qquad (15)$$

where a_i 's are the transformed regression coefficients, var(y) and $var(x_i)$ are weighted (by w_i or by unity) variances, and \bar{y} and \bar{x}_i are the weighted means.

It should be noted that the same logic applies to models containing classification terms. For example, if a measurement were assigned a coded (0,1) value depending on whether it was taken from a particular group, mean, variance, and correlation terms could be calculated as above. The normal equations from the transformed correlation coefficients would be

$$\begin{bmatrix}
\frac{R}{X} \cdot X & \frac{R}{X} \cdot Z \\
\frac{R}{Z} \cdot X & \frac{R}{Z} \cdot Z
\end{bmatrix}
\begin{bmatrix}
\frac{a}{b} \cdot \\
\underline{b} \cdot
\end{bmatrix} = \begin{bmatrix}
\underline{r}'_{a} \\
\underline{r}'_{bv}
\end{bmatrix}$$
(16)

where the $\underline{R}_{\chi'\chi}$ submatrix is an incidence correlation matrix, $\underline{R}_{Z'Z}$ is the regression matrix as in (12). The diagonal of $\underline{R}_{\chi'\chi}$ would have all elements equal 1 and all off diagonal elements would be negative with absolute value less than 1.

Having computed the weighted means, variances, and the correlation matrix, standard statistical packages are available to complete (13) through (15)(Nie, Hull, Jenkins, Steinbrenner and Bent 1975). Carrying out the above on the data used for the sweepnet regressions involved weighting only the sweepnet and square foot instar specific densities by subsample size, assigning a weight of 1 to all other measurements. The resulting correlations and regression statistics were not appreciably different from the results presented here, which are based on an unweighted analysis. Since the amount of programming time required to adhere to the weighted regression analysis was not thought to be justified by the refinement in results, the technique was dropped. Apparently there were sufficient cases that disparities caused by small samples were averaged out. With smaller data sets or with sets involving a large number of small sample versus large sample pairs, this may not prove to be the case. Application of the above to a correlation analysis by Fulton (1975) using data with these latter characteristics reversed an original conclusion of no significance

between instar specific square foot and sweepnet catches.

2.5 Inclusion of additional variables in the sweepnet models

The simple regression of catch per square foot on catch per sweep (section 2.3) did not provide the precise conversion factors desired for the sweepnet model. It was hypothesized that environmental factors were influencing sweep catch, although this hypothesis was rejected by Ruesink and Haynes (1973). Several environmental measurements had been made for each PWS sweep and square foot density estimate (section 2.2.1). These included crop height and windspeed, which had been taken in each field. Temperature and rainfall were estimated from a single station in the center of the study area, within 3 miles of each field. Because there was some doubt as to the accuracy of the central site thermograph, data from a New Carlisle, Indiana (5 miles south), and a South Bend, Indiana (12 miles southeast), weather station were averaged with the thermograph when computing degree day accumulations. These data are listed in appendix 8. This decision to average may not have been justifiable but was not reconsidered until later (see section 4.3).

The full regression model that was evaluated was

$$y_i = a + b_1x_i + b_2dd + b_3ch + b_4r24 + b_5W + e$$
 (17)
where $y_i = instar$ specific catch of CLB larvae per square foot;

a = a constant common to all observations;

x_i = instar specific catch of CLB larvae per sweep;

dd = accumulated degree days (base 48F) on sample date;

ch = standing crop height on sample date (inches);

r24 = rainfall during previous 24 hours;

W = windspeed (mph) at time of sample;

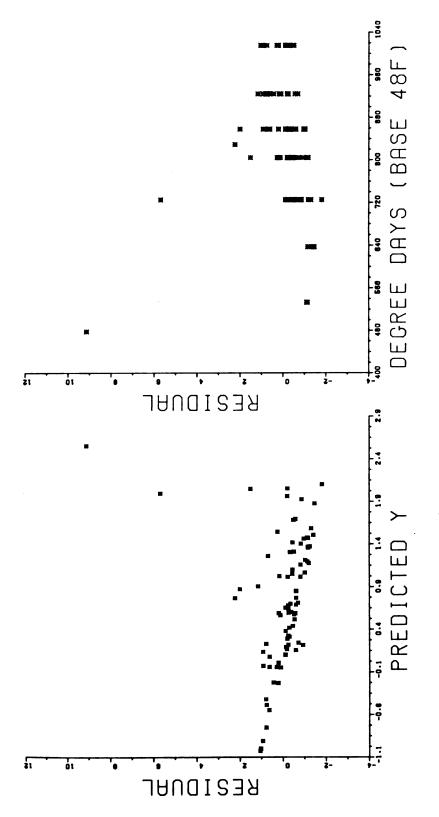
e = residual error, assumed $\sqrt{N(0, I\sigma^2)}$.

The regressions were carried out in a stepwise inclusion following the initial fitting of sweep catch (see Draper and Smith 1966, p. 171f). Results are listed in Table 5. Most of the parameters in the model (17) are included in the regressions because the regression program used was set for very loose inclusion criteria. For practical purposes, variables that add less than (arbitrarily) 5% to the R² values could have been excluded.

The differences between predicted and observed y's are called residuals. For each regression in Table 5, plots of residuals over predicted y and over the independent variables (x,dd,ch,r24, and W) were made. This technique is advocated and well discussed by Draper and Smith (1966, p. 86-99, 132) as a way to check model accuracy. Briefly, these authors recommend examining such plots for dependencies of the variance on the prediction or on the independent variables. a linear slope to the residuals, or curves in the plots of the residuals. These conditions are indicative respectively of variance instability (see section 3), incorrect fit or omission of the intercept, or model inadequacy and a need to include higher order terms or interactions in the model. The first type of abnormality (variance dependent on the predicted y) is especially crucial as a visual test of the critical assumption that the error term is normally distributed, an assumption necessary for valid testing. A few examples of such plots are given in Fig. 5. These indicate, respectively, a) overall model inadequate as there is a clear dependency of the variance on the prediction, b) adequacy over degree days and c) crop height as there are no apparent additional curvatures that would demand higher order terms.

To adjust for the variance instability, a multiplicative model of the form

		OATS					WHEAT		
		Regression	Standard				Regression	Standard	
	Order of	Coefficient	Error	•		Order of	Coefficient	Error	
ariable	inclusion	(a)	(s.e.b)	24	Variable	Inclusion	(a)	(8.8.0)	۲ _' ۵.
y ₁ (independent)	t.)				y, (independent)				
ř	-	7.48	4.02	. 060		_	6.13	2 40	, C
pp	2	0019	9000	. 203	- 5	- 2	1961	0.35	246
124	m	-1.35	11.	.226	3	, ~		562.	356
5	4	0083	.0062	.244	.24	, ,		2 -	007.
3	\$	0081	.0157	.247	. 5	, u	1,700		, oc
(constant)		1.39	.50		(constant)	,	1.5300.1) . .	•
F = 5.17***	C.V. = 379.5	ئ ة • .229				C.V. * 131.9); * 1.81	<u> </u>	
y, (independent)	Ę				v. (independent)				
, ,	-	39	1.37	.003	72 (-	, 0,	,,	200
, p	2	•.0055	7100.	.163	25.	٠ ،	59.	÷ -	
5	e	40	.02	.198	: -	, ~	- 012		325
r24	4	-3.81	2.4	.223	.	, 4	310.	<u>.</u>	225.
3	s	0066	.0497	.223	. 8	, ,			?
(constant)		5.36	1.60		(constant)		71	2 07	
F * 4.53**	C.V. • 335.2	₹ = 2.26				C.V. * 109.7	12 = 2.31	5	
y ₃ (independent)	t)				v. (independent)				
, ×	-	91.	.17	.029		-	46.7	760	~
5	2	005	.020	.136	¬ €	2	=	.05	.320
Đ	m	•.0032	9100.	.223	3	e	650.	1.	. 321
124	4	-1.84	2.31	.230	r24	4	.85	2.14	. 322
3	5	.028	.048	.233	B	s	003	600	. 323
(constant)		4.34	1.51		(constant)		.159	4.56	
F * 4.81 **	C. V. • 197.8	5.09			F = 5.35*** C.V.	V. • 120.0	it = 3.80		
y ₄ (independent)	Ç				v. (independent)	_			
×.	-	.52	.0 5	.648	***************************************	_	022	026	7
£	2	010	800.	.653	7.24	. ~	14.	59.	2.0
99		.00068	.00065	.656	. 5	. ~	900	. 100	328
3	4	.01	610.	.657	3 3	, 4	4.0	56	36.
r24	s	.32	.89	.658	. t	, <u>1</u> 0	5:00.	1,000	366
(constant)		27	.57					•	:
			;		(constant)		-1.43	19	



Plots of residuals $(y_i - \hat{y_i})$ from sweepnet model for 3rd instar in oats (Table 5) over predicted y and over degree days. 5. Fig.

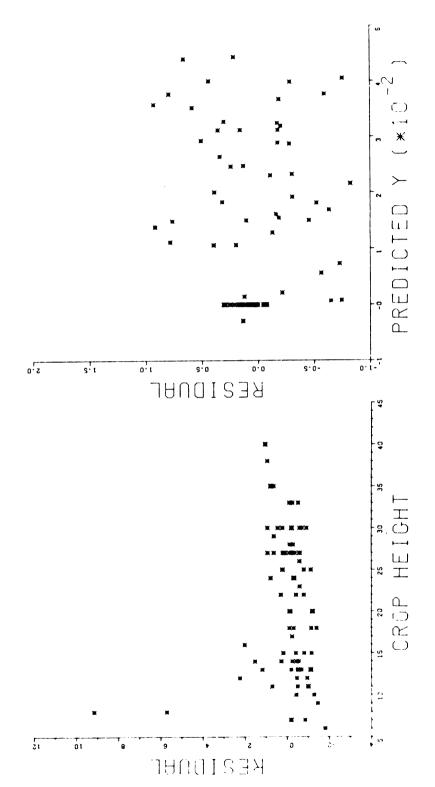


Fig. 5. (cont.) Plot of residuals $(y_i-\hat{y}_i)$ from sweepnet model for 3rd instar in oats (Table 5) over crop height (left). Residuals from transformed model for 3rd instar in oats (Table 6) over predicted (right).

$$y_1 = ax^{b_1}dd^{b_2}Ch^{b_3}r_24^{b_4}W^{b_5}e$$

was tried, where the variables are defined as on page 30 (eqn. 17). This model was evaluated by first taking logs of both sides. Some variables were recorded by adding 1 to avoid taking logs of 0's (Sokal and Rohlf 1969, p. 384; Draper and Smith 1966, p. 132). The new model was

$$log(y_i+1) = log a + b_1 log(x_i+1) + b_2 log dd + b_3 log Ch + b_4 log(r24+1) + b_5 log(W+1) + log e$$
 ...(19)

Recoding and transforming the variables did seem to produce a better fit to the data (Table 6). As expected, the effects of outliers are greatly reduced as is indicated by a general increase in R^2 values. The question of whether this makes for a better model, however, is largely a matter of personal statistical preference. Certainly the transformation did reduce the variance instability, as can be seen in the residual plot (Fig. 5d). For most practical purposes, though, the increases in R^2 values caused by adding environmental effects to the sweepnet model do not seem to justify the extra effort needed to take and use the measurements. The predictions still lack precision.

Regretfully, the simple regression models of Tables 3 and 4 are recommended as "best". Instar specific sweep catch does not appear to bear a 1:1 relationship to square foot catch. The effort to establish the expected true relationships has led to equations that may be useful yet lack precision.

2.6 An additional data set

Previous data sets comparing sweep catch with absolute density
were thought to be either too small or to have unreliable instar identification methods and thus could not serve as a check for the sweepnet

Table 6. Regression of log transforms of catch per square foot (y) on catch per sweep (x), accumulated degree days (dd), crop height (ch), rainfall (r24), and windspeed (W). Catch per sweep was forced into the regression first. C.V. is coefficient of variation. 37 is estimated residual error.

Septemble Septemble Standard Standar			OATS					MHEAT		
Coefficient From From Coefficient From Coef			Regression	Standard				Berret	brebar 42	
1,		Order of	Coefficient	Error			,	Regression	D'andard	
2.93 1.51 .065 105(y ₁ -1) (Independent)	Variable	Inclusion		(s.e.b)	n ²	4 (4) 4)	Urder of	Coerricient	בייסי	20
2.33 1.51 .062	log(y,+1) (indep	andent)			I		וועכו מאונטע	(9)	3	.
-67 - 13 - 299 193(4+1) 1 4 03 - 39 -107 - 13 - 239 193(4+1) 2 - 27 115 -107 - 104 - 397 199 cn 430 .22 -1080056 .400 .109(7-2+1) 543 .61 -1380056 .400 .109(7-2+1) .10 .20 -1.0206426 .109(7-2+1) .10 -1.0206426 .109(7-2+1) .10 -1.0209 .370 .109 cn 3 .117 .27 -1.03019 .10 .136 .109(7-2+1) .10 -1.42019 .136 .109(7-2+1) .10 -1.42019 .136 .109(7-2+1) .10 -1.43019 .10 .109 cn .10 -1.4409 .370 .109 cn .10 -1.45019 .370 .109 cn .109 cn -1.4671 .414 .109 cn .26 .318 .26 -1.47 .247 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .247 .269 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .269 .270 -1.47 .247 .247 .247 .269 -1.47 .247 .247 .247 .269 -1.47 .247 .247 .247 .269 -1.47 .247 .247 .247 .269 -1.47 .247 .247 .247 .269 -1.47 .247 .247 .247 .269 -1.47 .248 .268 .268 .268 .269 -1.47 .248 .268 .268 .268 .268 .268 -1.47 .248 .268 .268 .268 .268 .268 -1.47 .248 .268 .268 .268 .268 .268 .268 .268 -1.47 .248 .268 .	100(x.+1)	-	2 93	5	Ş	10g(y, •;) (1nde	Dendent)			
07	100 00		; ;	:	8 8	195(*191)	-	4.03	66.	.250
102	00 60:	, ,	(o :	c:	667	(1+M)501	~	.27	.15	.280
10204197 log cn	(1.474)601	-	87	Ξ.	.354	log dd	٣	.61	1.22	305
2.12 2.12 2.13 2.13 2.14 2.15 3.2 .0056 4.5 .33*** C.V. * 94.4 37044 2.13 2.13 2.14 2.13 2.14 2.15 2.15 2.15 2.16 2.16 2.17 2.18 2.18 2.18 2.18 2.18 2.19 2.19 2.19 2.19 2.19 2.19 2.19 2.19	log ch	4	102	Š	.397	100	•	۶	90	916
2.12 2.12 3.12 3.14 3.14 3.14 3.15 3.16 3.45 3.45 3.45 3.45 3.45 3.47 3.10 3.10 3.10 3.10 3.10 3.10 3.10 3.10	109(₩+1)	•	610.	.03	400	10.60	, ,	3 :	9 (9 9
1.13	(constant)		2.12	.45		100, 12, 101	n	÷ :	٠ ;	775
.13 .34 .012	F * 10.52*** C.V.	• 113.7	9500. • 56					2.05 37 • .044	3.20	
13	log(y,+1) (indepe	ndent)								
15	109(4, 1)	_	=	7.		log(y ₂ +1) (inde	pendent)			
-1.01	, e e e		: *	!	210.	log(x, 1)	-	87.	.18	.253
-1.01 .28 .395	5 7		e :	8.	.275	100.41)	~	21.	115	.273
-1.02	PP 601	m	-1.01	. 28	. 395	1 40 60	, ,	: :	? ?	6.4.
3.4506426 logic7441)	10g(r24+1)	4	-1.02	09:	.420	10 601	n .	-	77:	8/7
13.45 .81	109(14+1)	٠,	\$0:	90	426	109(1541)	•	92.	5.	. 282
(constant) 1.33 1.10 1.34 1.35 = .019 (constant) 1.35 = .014 (constant) 1.36 = .5.38*** (.V. * 69.0 5? * .044 1.42 = .09 1.37 = .012 (constant) 1.39 = .390 (constant) 1.34 = .399 1.34 = .044 1.05 1.04 1.04 1.04 1.05 1.04 1.0	(constant)		3.45	· 6	?	log dd	-			
F = 5.33*** C, V, = 69.0 5; * .044 42	F = 11.72*** C.V.	- 145 g	22 010	5		(constant)		.349	330	
.33 .10 .136 log(x ₃ +1) (Independent)42 .09 .370 log ch62 .32 .406 log ch66 .771 .414 log ch .05 .07 .419 log(x ₄ +1) n.i. 2.42 .94 (contant) 2.42 .94 (contant) 2.5026 log(x ₄ +1) (Independent)098 .066 .626 log(x ₄ +1) (Independent)098 .065 .632 log(x ₄ +1) 1 .11 .05477 .547 (constant)477 .547 (constant)477 .547 (constant)477 .547 (constant)478 .548 (constant)479 .549 (constant)470 .540 log ch471 .547 (constant)471 .547 (constant)472 .548 (constant)473 .547 (constant)473 .547 (constant)474 .055 .055			60.				V. • 69.0	•		
.33 .10 .136 log(x ₃ +1) (Independent)42 .09 .370 log(x ₃ +1) 1 .61 .1062 .32 .406 log cn 2 .38 .2666 .71 .414 log cn 2 .38 .2666 .77 .419 log(x ₄ +1) 3 .33 .44 2.42 .94 (contint)08 .06 .626 log(x ₄ +1) (Independent)09 .065 .632 log(x ₄ +1) 1 .11 .05477 .547 log ch 3 .85 .40477 .547 log ch 3 .85 .40477 .547 log ch 3 .23 .05477 .547 log ch 3 .23 .05477 .547 log ch 3 .23 .05477 .547 log ch 3 .23 .05	log(y,*1) (indepe	ndent)								
42	109(4,+1)	-		2	31.1	log(y ₃ *1) (1nde	pendent)			
-66 .71 .414	10 01	٠,		? 8	97 -	109(1,101)	-	19:	٥٢.	396
	log dd	. ~	; ;	j s	0/5.	log ch	~	96.	92.	.426
	100(-24+1)	٠.	7	¥. 1	93	109: (24+1)	e	.33	87	4.38
2.42 .94 .105(#-1) n.i1.58 2.70 .2.42 .94 .2.22 .2.42 .2.26 .2.42 .2.26 .2.22 .2.23 .2.23 .2.23 .2.23	100(441)	, ,	8 9	ξ.	7	109 63	4	94.	8	440
2.42 .94 (contait) -1.58 2.70 5.4 .026	(1000)	n	s :	.00	.419	100(#•1)	1.5			
7.* .026 1.58 .06 .626 log(y4*1) (Independent) 2.98 .06 .626 log(x4*1) 1 11 .05 2.2 .20 .637 log(x4*1) 4 .052 .052 2.47 .547 (constant) -2.33 1.03 6. 8.56*** (Constant) -2.33 1.03	(cc.) 41000 C.:			8.		(constant)		85.1-	2, 70	
.58 .06 .626 log(y ₄ *1) (independent)098 .065 .632 log(r ₄ *1) 1 .11 .05 .22 .20 .637 log(r ₄ *1) 2 .44 .20 log(r ₄ *1) 2 .44 .20 log(r ₄ *1) 4 .20 r ₄ r ₄		8.				F - 11 13*** C.	۷. • 56.9	3: . .037	•	
.58 .06 .626 log(x ₄ +1) (independent)096 .065 .632 log(r24+1) 2 .44 .20 .22 .20 .637 log dd 3 .85 .40477 .547 [constant) -2.33 l.03477 .547 [constant) -2.33 l.03477 .547 [constant) -2.33 l.03	log(y ₄ +1) (indepe	dent)								
098 .065 .532	10g(x4+1)	-	.58	8	424	log(y4+1) (inde	pendent)			
.22 .20 .637 log(w+1) 2 .44 .20 .22 .20 .637 log dd 3 .85 .40 log(w+1) 4 .052 .052 .477 .547 (constant) 2 .036 .097 .2 .012 F 8 .5 .005	log ch	2	980 -	ž	3.5	10g(x4+1)	_	Ξ.	5 0.	151.
-477 -547 (constant) -2.33 1.03 (-2.32 (-2.32 -2.33 1.03 (-2.32 -2.33 1.03 (-2.33 1.03 (-2.33 1.03 (-2.33 1.03 (-2.33 1.03 (-2.33 1.03 (-2.33 1.03 (-2.33 1.03 (-2.33 1.03 (-2.33 1.03 (-2.33 (log dd	_	33	8 8	260.	109(124+1)	~	7.	۶.	.378
log(W+1) 4 .052 .052 477 .547 log ch 5 .026 .097 ô?012 (constant) -2.33 1.03 F + 8.76*** (C.V. + 106.1 5?005	100 (14.1)		į	8.	.63/	log dd		88.	0	429
477 .547 log ch 5 .026 .097	100(000)					100(14-1)	•	640	630	7
477 .547 (constant)026 .09703012012	(1.474)501	-				()	, ,	30.	260	9
.2.33 .2.012 (constant) -2.33 .7.06.00 (constant) -2.33 .7.00 .7.0	(constant)		477	.547		40 bor	r	920	.097	.439
F = 8.76** C.V. = 106.1 32 =	F = 47.43*** C.V.	. 68.2				(constant)		-2.33	1.03	

models developed here. A new set was obtained from the APHIS (USDA) parasitoid rearing laboratory at Niles, Michigan. This included paired sweep and square foot samples from 15 wheat and 13 oats fields. Each field was divided into 14 equal size units and a single square foot was sampled in each unit. All larvae were retained for later age classification. A large portion of each sweep sample was measured and there is no reason to suspect any biases were introduced by the counting or measuring processes.

Table 7 describes the APHIS data set. In comparison with the PWS data (Table 3), all slopes from the square foot over sweep regressions are steeper and all intercepts but for 3rds in wheat are not significantly different from 0. The density per square foot had a higher average for all instars in both crops than the densities found in the fields described in Table 3.

To a certain extent, the high R² values for 1st and 2nd instars were aided by the presence of many 0's. There were 53 and 42 pairs of 0's for 1sts in oats and wheat, respectively (51% and 45% of the samples) and 16% of both crops had paired 0 counts for 2nd instars. However, the good fits and high slopes of the regression coefficients in general cannot be explained away. These results were not altered by including windspeed, crop height, or accumulated degree days in the regressions.

It must be concluded that the relationship between sweep and square foot catches of CLB larvae is not well understood. Although differences between sweepers has been thought to be negligible (Ruesink and Haynes 1973), perhaps this should be reinvestigated. Until then, use of sweepnets for intensive population studies of the CLB has been generally curtailed (A. Sawyer, E. Lampert, pers. comm.). When used in

Table 7. The APHIS (USDA) data set, describing the relationship between catch per square foot and catch per sweep for CLB larvae. y_i = average catch per sweep of instar i. s.d. = standard deviation of catch.

a_i = intercept. b_i = slope. s.e. = standard error of coefficients. Model is y = a + bx + error.

Sweep 93 5718 5718 5718 6961 14 1.48 .087 .241 08 1.52 .131 .267 33 3.06 .317 .404 90 1.89 .214 .382 105.7 .55 101 101 102 103 103 104 105 105 105 105 105 105 105	WHEAT Square foot 93 7950 20933 4. a i s.e. b i 57 .329 .379 13.14 74 .244 .449 16.08 36 -5.163 1.564 41.33 67 -1.619 .824 27.80 923.6 78.4 1071.6 11.77 <.001	Cases Total measurements Total larvae in sample Instar (i) y ₁ s.c 2 2.345 5.77 3 7.935 20.4 4 4.323 12.66 Instar Source d.f. I R (a,b) 1 E 91 2 R (a,b) 1 E 91 3 R (a,b) 1 E 91 3 R (a,b) 1	Sweep 104 5022 5105 .029 .066 .073 .105 .220 .220 .164 .211 Analysis of Variance R2 .46	b ₁ 20.12 20.12 15.93 34.67 17.71 17.71 17.71 12.45 4.001 18.03	9415 Square foot 104 5857 1857 18788 1874 -2.762 2.532 06 .658 1.777 289.45 2370.97 23.24 5988.7 33879.6 332.2	sample sample s.d. s.d. s.d. s.d. s.d. s.d. s.d. s.d
		A 2000 ((A c) 0 A	9	7.03 402.4	4.55.9	(a,b) x
01	140.3				M)	102
149.3	:	R (a,b) 1	.15			-
10	15.5	16		.001	_	102
165.7	:	R (a.b) 1	E.			-
10	11.77	16		001		
233.2	:	-	.46			_
ر. د.	æ	Source	R ² -			Source
		e) 	Analysis of Variar	u		
1.89 .214	.824					3.552 14.706
3.06 .317	1.564				-2.762	4.857 19.674
1.52 .131	.449			15.93		1.279 5.082
1.48 .087	.379	1 1.472 4.657				.457 1.952
s.e. x _i	s.e.	y,		}	٠,-	بأ
6961	20933	Total larvae in sample	5105		14788	larvae in sample
5718	7950	Total measurements	5022		5857	measurements
93	93	Cases	104		104	
Sweep	Square foot		Sweep		Square foot	
	WHEAT				OATS	



extensive surveys, sweepnet samples should be compared to a few sets of paired square foot or other absolute, instar specific density estimates.

ANALYSIS OF QUADRAT SAMPLES OF CEREAL LEAF BEETLE EGGS AND LARVAE

3.1 Introduction

The goal in this section is to improve understanding of the error term that is associated with estimates of CLB larval density. Some characteristics of CLB larval count data are discussed and the relationship between variance and mean for count data is described. Based on this relationship, a choice of transformations is made so that analysis of causes of change in beetle density (section 4) can proceed on valid statistical grounds.

When crops are short (8-10 inches) or when accurate density estimates are desired, sweepnet use is not recommended. Other sample units are selected following criteria like those of Morris (1955). That is, all units in the sampling universe should have an equal chance of selection, the unit should not change size in time, the unit should be easily converted to other unit areas, and the unit should be of such a size as to provide a reasonable balance between variance and cost. For the CLB, selection of square foot units over square yards or individual stems was made by Helgeson and Haynes (1972) on the basis of relative efficiency and expense. Examination of about 3 times as much plant material was needed when using square yards to obtain the same coefficient of variation as from square foot units. In practice, 2 linear feet of row are nearly the same as 1 square foot. The terms are used synonomously here.

A final sample unit selection criteria is that the proportion of

the population using the sample unit must not change during the season. Although the expected count per sample will always be the mean density, the variance between counts may change disproportionately, along with the confidence limits on the estimate. The use of Normal Theory statistical techniques will be invalid until an appropriate choice of transform can be made to approximately normalize the frequency distribution of counts. The characteristic statistical distributions of CLB count data are therefore discussed in section 3.2. Using these, several transforms are derived and evaluated in section 3.3.

3.2.1 Pubescent Wheat Study adult sampling methods

During the PWS, simultaneous sets of sweep and square foot counts were taken in each field for adult, egg and larval life stages. Egg and larval sampling was discussed in section 2. Similar procedures were used for adults. In each field, 6 sets of 25 sweeps were taken from different parts of the field. At higher densities, set size was reduced to 10 sweeps. All of the adult beetles were counted and the 6 counts and corresponding number of sweeps were recorded. At the same time, in areas between where the sweeps were made, visual counts were made. The stick toss of section 2 was used to locate a semi-random starting point. A sample set then consisted of either 1) the total count of all adult beetles in 10 linear feet of row (= 5 square feet) or 2) a count of up to at least 10 adults, whichever came first. If no insects were counted in the first 6 sets, 4 more sets were counted. Each of the 6 sets of sweep counts and 6 to 10 sets of square foot counts were converted to either catch per sweep or catch per square foot before calculating mean and variance. Data for adult counts is recorded in appendix 7a and for larval and egg counts in appendix 7b.

Although these procedures introduce biases in the adult data that tend to overestimate the mean and may bias the variance in an unknown direction, the amount of the basis should be low, bias should be restricted to lower densities, and the relationship between mean and variance should be unaffected at higher densities. For egg and larval data, the amount of bias is believed to be negligible as virtually all samples were the same size.

3.2.2. The relationship between variance and mean for CLB counts

Aggregation is a tendency to be found in groups. It is 1 of 3 ways in which insect count data commonly violate the assumptions of the Normal distribution underlying such statistical procedures as the analysis of variance. These procedures were devised for continuous variables, normally distributed with equal variances (Hayman and Lowe 1961). Cereal Leaf Beetle counts from square foot samples differ from these assumptions in 3 ways: They are discrete, they have a skew distribution, and their variance increases with the mean. The problem of discreteness can usually be alleviated by increasing sample size, working in higher density populations, or by using a weighted analysis when a small proportion of the samples are too small (section 2.4).

The essential question when dealing with the other departures from normality is whether ignoring these will produce seriously misleading results. Another way to say this is to ask whether the rejection region of a test (or, alternatively, the test size, α , itself) is sensitive to changes in the underlying distribution. Tests that are not sensitive are called "robust" (Box 1953). Those that are sensitive are "frail" (Kendall 1973). Kendall (1973, chapter 31), summarizing earlier investigations, concluded "whereas tests on population means (ie.

Student's t test for the mean of a normal population and for the difference between means of 2 normal populations with the same variance) are rather insensitive to departures from normality, tests on variances (eg. the χ^2 test for the ratio of 2 normal population variances) are very sensitive to such departures.

If there were doubt about the validity of results, Kendall (1973) suggested 2 alternatives: to transform the data with a simple normalizing function or to use distribution-free procedures. Rather than abandon the more powerful parametric tests, it is usual to seek a transform first. The difficulty with this approach is "that we must have knowledge of the underlying distribution before we know which transformation is best applied, information which is likely to be obtainable in theoretical contexts like the investigation of the sampling distribution of a statistic but is harder to come by when the distribution of interest is arising in experimental work" (Kendall 1973).

The variance x mean relationship of aggregated populations may be characterized as

$$s^2 = Km, (20)$$

or
$$s^2 = m + m^2 / k$$
 .(21)

For low density randomly distributed populations (eg. Poisson, or K = 1),

(20) may prove useful. At higher densities or in non-uniform habitats,

(21) is more often used. More recently, a logarithmic alternative,

$$\log s^2 = a + b \log m \tag{22}$$

has been fit to many examples (Wayman 1959, Hayman and Low 1961, Taylor 1961).

The amount of variance among sample counts from non-uniform populations depends on the size of sample units (Kershaw 1964).

Sweepnet data should be converted to catch per sweep before use. This provides an unbiased estimate of the mean but leaves a bias in computing variance as those sample sets with higher numbers of sweeps hypothetically have a smaller between sample variance, depending on the area covered by the sweeper and the cluster sizes of the population. An example of this comes from the regression for adults by Ruesink (1970) who used sweep samples of from 1 to 50 sweeps per sample with 6 to 63 samples used to compute each mean and variance. The regression equation for within field variance was

$$\log s^2 = .04+1.34\log m$$
, (R² = .99, cases = 17, p < .001) (23) for catch per sample. Conversion to catch per sweep gave

$$\log s^2 = -.42 + 1.26 \log m$$
 ($R^2 = .91$, cases = 17, p < .001) .(24) Although establishment of standard numbers of sweeps or square feet for sample size would aid in making comparisons of data sets, this is seldom feasible. Insect numbers fluctuate and logistically demand adjustments in sample size. In making comparisons, one should be careful to note that sample size differences will likely affect variance.

Other studies of the CLB variance x mean relationship include that of Gage (1974) who found a good fit for the standard error of pupal cells per half square yard with the mean (s.e. = .52+.11m, $R^2 = .97$, range of means: 1 to about 110). Sawyer (1976) reported a good fit for the regression of both egg and larval log variance on log mean for square foot counts. These relationships are summed up in Table 8.

The PWS adults per sweep regression (Table 8) had the same intercept but significantly greater slope than the Ruesink regression.

Comparing adults per sweep to adults per square foot for the PWS data

Table 8. The relationship between log within field variance, y and log mean, x. Model is y ≈ a + bx + error.

! به			12.3***	41.5***	52.7***			.5n.s.		26.3***	36.6***			22.4***	28.3***	24.5***	26.6***	
R ²			.910	.884	.887			.072		.885	.987			.848	.956	996.	.970	
s.d.			.700	.565	.562			.288		.567	.763			.263	.630	.995	1.020	
×			072	395	-1.013			178		.127	220			.092	.037	096.	916.	
s.e.			.108	.039	.026			.494		.053	.035			.118	.445	.055	.053	
ام			1.261	1.660	1.408			.754		1.388	1.254			1.331	1.245	1.280	1.363	
s.e.			.074	.022	.015			.643		.030	.026			.030	.027	.053	.053	
R 0			422	441	382			829		.179	.256			.197	.239	.511	.518	
s.d.			.925	.997	.840			.812		.837	.965			.760	.803	1.315	1.411	
>			512	-1.098	-1.808			-1.419		.356	019			.319	.285	1.758	1.768	
- I		1	11	227	335		1	2	oot	35	56		00 t	35	36	23	24	
Source	ADULTS	Sweepnet	Ruesink ¹ 17	PWS ²	Square foot PWS ² 335	LARVAE	Sweepnet	Ruesink 5	Square foot	PWS ²	Sawyer	EGGS	Square f	PWS ² 92	Sawyer ³	Gage w.	Gage o.	

^{1968,} Gull Lake. Wheat, oats, barley, 1 to 50 sweeps/sample, 6 to 63 samples/set, converted to catch/sweep. (Ruesink 1970)

^{1975,} Pubescent Wheat Study. 2

^{3 1975,} Gull Lake. Oats + alfalfa, 30 x l square foot/set. (Sawyer, pers. comm.) 4 1970, Gull Lake. w = wheat, o = oats, 30 x l square foot/set. (Gage 1972)

also gave non-different intercepts but a significantly greater slope for the catch per square foot regression. This latter result is especially curious since it seems more logical to expect the relatively greater area covered in the sweepnet samples (50 to 125 square feet per sample) to produce less between sample variance than the approximately 5 square feet covered for each of the absolute counts. The ratio of catch per square foot to catch per sweep, a crude "M" value (as per section 2), for the PWS adult data was 1.96, compared to an average ratio of .66 for 15 sample sets by Ruesink and Haynes (1973). These 2 discrepancies suggest that the adult sweepnet model (Ruesink and Haynes 1973) bears reexamination.

There are no significant differences between the regressions of larvae per square foot from the PWS and the 1975 Gull Lake data (Sawyer 1976)(Table 8). There is also no significance to the difference between PWS and Sawyer data for the egg regressions. It is interesting that the regressions for eggs per square foot fit most closely to the larvae per square foot regressions for the same locations, but, again these differences are not statistically significant.

It is unlikely that there is any difference between the variance x mean relationships for eggs and larvae for wheat, oats, and barley. Ruesink's adult results (Table 8) were from 3 crops and there were no detectable differences. Gage's (1970) data likewise had no difference between wheat and oats (Table 8).

3.2.3 Statistical distributions of CLB egg and larval counts

Although the PWS regressions of Table 8 adequately fit the data $(R^2 = .848 \text{ and } .885 \text{ for eggs are larvae})$, the fitting of 2nd order regressions leads to interesting results (Fig. 6). The 2nd order

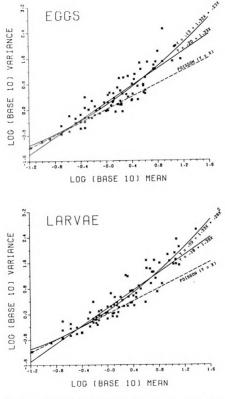


Fig. 6. The relationship between variance and mean for CLB egg and larval count data.

regressions provide only a slight increase in R^2 (.035 and .028), but they do suggest that the variance x mean relationship is not a simple log-linear one. It is useful to establish the significance of this in terms of the underlying distributions.

From Fig. 6 it appears that samples from low densities (less than .5 per square foot) have a poisson variance x mean relation, which is closely approximated by the 2nd order regression. This had been suggested by Ruesink and Haynes (1973). Regressions through those points with mean less than .5 confirms that the variance:mean relationship is approximately 1: 1. Tests for goodness of fit of the pooled samples' frequency distribution also suggest a poisson (Table 9).

The Poisson distribution is a limiting case of the Negative Binomial. As the negative binomial statistic,

$$k = m^2/(s^2-m)$$
 (25)

((21) rewritten) goes to infinity, the distribution approaches the Poisson. The variance x mean relationship at higher densities is that of (25) but, from Fig. 6, may be different from that of lower densities. It is therefore reasonable to ask if the distributions of count data are in fact negative binomial and whether they share a common k.

Although several computational schemes exist for estimating k (Anscombe 1949, Beall 1942, Bliss and Fisher 1953, Bliss and Owen 1958) most estimates have biases. An unbiased, not necessarily minimum variance, estimate of k can be obtained from the maximum likelihood estimates (MLE's) of the k's for individual samples, as in Table 10. In Table 10, column 1 refers to the samples listed in appendix 7b. Columns 2 and 3 are the individual sample's mean and variance. The k for the sample, column 4, is computed using Fisher's MLE (Bliss and Fisher 1953,

Regressions of $\log s^2$ on $\log m$, using $\log s^2 = a+b \log m$, and tests for goodness of fit of joint count frequency distributions to the Poisson for egg and larval samples (given sample mean less Table 9.

	•	+ 1	7.02***	6.11***							$x^2 = 1.67$; $x^2.05(1) = 3.84$			$K/S = .011$; $K/S .05(3) = .708 x^2 = 1.44$; $x^2.05(1) = 3.84$
	c	الار	.701	.700			$\frac{\chi^2}{\chi^2}$				$\chi^2 = 1.67$			$x^2 = 1.44$
		s.d.	.260	.265										= .708
	S	×	602	697							K/S = .025; K/S.05(4) = .624			/S .05(3)
	TATISTIC	s.e.	.045	.169	OF FIT	gorov	۸				.025; K			.011; K
	REGRESSION STATISTICS	٩	1.207	.975	GOODNESS OF FIT	Kolmogorov	Smirnov				K/S =			K/S =
	REGR	s.e.	.179	.044	G	þ	χ	•	-	10	01	0.1	0	m
foot).		Ф	.184	.005		Expected	Frequency	241.9	71.4	10.5	1.2	198.2	46.0	5.8
		y s.d.	.375	. 309		/ed	sucy	0	21	6	e +	5	41	8
than .5 per square		X	542	665 .309		Observed	Frequency	250	?9	0.	•	201	•	
than .		Cases	23	18			Count	0	-	2	3+<	0	_	2+<
			Eggs	Larvae				Eggs				Larvae		

Table 10. Estimated negative binomical k's for egg and larval sample counts and test for goodness of fit using sample k and common $\mathbf{k_c}$. $\underline{\text{EGGS}}$

NC.	MEAN	VAR	ĸ	(S.E. K)	CHI-SQUAR	SAMFLE K	USING C	FIT OMMON K ARE (DF)
12745678901275678961273456	0733300 66533300 673300 67300 67300 673300 67	7-10-2-11-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1	17 123 170 344.1997.67 17 123 170 344.1997.67 1 170 344.1997.67 1 170 344.1997.67	(6.61) (3.58) (1.28) (1.34) (1.34	3.91665 9.1665 9.1665 9.1665 1.0282 1.0282 1.0282 1.0282 1.0283 1.028	(1) (2) (1) (2) (2) (2) (2) (8)	\$225029637676767496403. 2653276686376765376496403. 26531766837663376403. 265283 265283 265283	(8) (8) (5) (6) (6) (2) (1) (1) (1) (2) (1) (2) (2) (2) (2) (2) (2) (2) (2
25 25 27	3.867 3.800 5.467	3.27 5.31 17.41	-16.25 10.90 2.22	(.09) (17.33) (13.95) (1.12)	3.335 2.007 3.926	(4) [5) [7)	44.349 7.290 4.739 4.170	(10)*** (5) (6) (7)
27 28 20 32 33	.267 3.100 1.300 1.000	5 · 21 3 · 12 · 89 3 · 99	.13 5.89 1.66 -2.20 -20.85	(.14) (6.03) (.96) (1.33) (25.09)	.010 (1.274 (2.493 (3.221 (1.334 ((3) (2) (1) •	.733 1.901 1.385 5.133	(J) NA (4) (2) (1) ••
377373749023	10007 1.50667 1.66667 1.0067 1.0067 1.0067 1.0067 1.0067 1.0067	1. 77 1. 55 1. 55 38 9. 62 2. 97 2. 78	.858 -4.918 2.78 2.78 2.79 3.54 5.55	(.58) (2.66) (.85) (8.37) (11.06) (15.57) (7.35) (1.53)	.336 .429 .142 .511 .003 2.163 2.772 4.324	2) 1) 1) 1) 0) NA (4)	. 598	(2) (2) (1) (1) (0) NA (6) (6) (8)
34567134578	1.067 1.6607 1.6607 1.0000 1.66007 1.0000 1.66007 2.66007	1.34 17.64 1.69 1.89 1.83 1.12 10.60	-6697 -6697 -6417 -26379 -120 -120 -13795	(.42) (3.42) (5.67) (1.72) (1.33) (.20) (.74) (.34) (.32)	95 (954 (954 (954 (954 (954 (954 (954 (9	1) (2) (8) (2) (1) (1) (1) (1)	2.811 11.251 .957 5.133 1.304	(2) (1) (8) (2) (1) (1) (1) (1) (1)
56666668777777777888	175333300007373733377737000 175333300007373733377737000 110000000000000	2555777 25557777 363-6-4355345526217 27222 31 1 292392	1	(.35) .79) .179) (.189) (.189)	2.094 (1.098 (5.602 (5.307 (1.377 (1.352 (1.251	5) 71) 4) 4) NA 0) NA 1) NA 1) NA 1) NA 1) NA 1) NA 1) 1) 1) 1) 1) 1) 1) 1) 1) 1) 1) 1) 1)	1.8556 9556 9556 95567 19.7759	(2) (8) (6) (6) (7)
# 4 6 7 C 1	.ACO .400 .533 3.133 1.C67 1.900 1.400 MATED CO	1.98 1.98 .92 3.03	-6.61 -37 -7.61 -4.28	(7.69) (.23) (.27) (2.66) (2.10) (.89)	.486 (4.811 (1.488 (.245 (.42) (.973 (.739 (32.59445)	6) 1) 3) 1) 3) 2)	1.708 18.233 3.866 6.557 1.398 1.257	(i) (6)*** (1) ** (4) (2) (3) (2) * PLES: 75.

Table 10 (continued)

LARVAE

MC. MEAN VAR K (S.E.K) US CHI-	-TEST FOR GOODNESS OF FIT ING SAMFLE KUSING COMMON K SQUARE(OF) CMI-SQLARE(OF)
2 10.667 35.81 4.76 (2.44) 8.6 3 2.067 14.92 33 (.19) 6. 4 24.030 430.57 1.81 (.57) 13. 5 3.200 44.74 2.64 (1.20) 8.6 6 4.067 25.78 .45 (.22) 3.7 7 1.933 16.21 .16 (.09) 2.7 10 3.667 71.81 .83 -6.42 (2.07) .10 .11 1 .300 3.34 1.11 (.80) .11 1 1.300 3.34 1.11 (.80) .11 1 1.700 2.68 1.99 (2.36) 4.6 12 1.700 2.68 1.99 (2.36) 4.6 14 2.400 2.27 -13.35 (13.25) .15 2.830 4.40 5.00 (7.01) 2.17 5.933 4.92 266.59 (21.40) 3.61 1.70 2.800 3.31 3.31 3.31 3.31 3.31 3.31 3.31 3.	435 (8)
20 2.533 3.70 5.40 (5.53) 1.7 21 2.400 5.26 1.27 (.86) 8.1 22 11.867 216.12 .81 (.27) 4.7 23 6.467 38.64 .44 (.15) 8.0 24 3.067 7.50 1.54 (.87) 6.4 25 2.467 5.41 2.07 (1.54) 1.3 26 667 .81 2.39 (4.25) 2.5	765 (4)
31	221 (1)
37 11.133 79.84 2.91 (.91) 7.1 38 11.667 36.52 5.48 (3.55) 4.1 40 12.933 124.07 2.43 (.73) 8.0 41 7.067 8.74 30.62 (79.71) 4.1 42 5.333 47.38 31 (.14) 7.3 43 4.733 8.92 7.51 (4.76) 4.3 44 4.600 25.69 4.6 (.21) 9.5 45 1.267 1.07 4.51 (2.20)	125 (8)
47	181 (1) 2.395 (1) 176 (0) NA .564 (0) NA 179 (2) 1.432 (3) 179 (2) 1.581 (2) 192 (1) .236 (2) 174 (1) 1.417 (1) 126 (3) 4.971 (3) 129 (8) 6.755 (7)
59 4.030 47.14 .44 (.17) 9.2 60 11.533 50.27 3.94 (1.72) 4.9 61 5.200 20.45 1.46 (.73) 6.9 62 1.133 4.41 .50 (.27) .6 63 .600 .54 -2.69 (3.42) .3 64 .200 .60 .04 (.03) .0	?72 (5) * 15.415 (7) ** 957 (8)
67	93 (1) 3.785 (1) 6.104 (1) 6.218 (1)
79 1.200 1.03 -3.73 (2.73) 3.2 80 +933 20.21 1.16 (.51) 5.7 81 1.400 .83 -2.87 (.39) .7 84 .533 .41 -1.56 (.68) .0 85 .333 .34 3.72 (11.06) .0 86 .267 .35 .77 (1.40) .0 89 .200 .31 .25 (.34) .0	110 (0) NA

p. 182). For some of the samples, a k was not computable. This was so whenever 1) the mean and variance were equal, giving an infinite k,

2) the mean was close enough to the variance to cause a failure to converge on an estimate of k within a reasonable series of calculations, or 3) the ratio of mean: 1st trial estimate of k was less than -1 (see Davies 1971, p. 368-374). Although the negative binomial parameter k is usually considered to be a positive number, negative estimates were also calculated (cf. Bliss and Owen 1958).

As a check on whether the individual samples were in fact distributed negative binomially, the theoretical sample distributions were generated from sample mean and MLE k. Observed and theoretical distributions were compared by a χ^2 test for goodness of it (Table 10, columns 6 and 7). Of the 68 testable samples of eggs and larvae, only 2 egg and 3 larval count distributions were significantly different from expected at a .1 α level. Due to the small sample size, 10 to 15 counts per sample, the minimum expected cell size for the χ^2 test was set at 1, rather than the usual 5 (see Steel and Torrie 1960, p. 350; Sokal and Rohlf 1969, p. 565). Such test results should be evaluated cautiously, but it is safe to conclude that the samples can be represented by the negative binomial distribution.

From the individual estimates of k and their corresponding variances, a weighted average was computed as the estimated common k,

$$k_{c} = \sum w_{i}k_{i}/n \tag{26}$$

where w_i , the weight, is the inverse of the variance of k_i and n is the number of sample sets (Steel and Torrie 1960, p. 180). The pooled variance estimate is

$$var(k_c) = \frac{(n_1-1)var(k_1)+(n_2-1)var(k_2)+...+(n_m-1)var(k_m)}{n_1+n_2+...+n_m-m}$$
(27)

These estimates are listed in Table 10 (cf. Bliss and Fisher 1953, p. 194).

Two tests were used to determine the commonness of the estimated k_C 's. The individual sample distributions were compared to theoretical distributions generated from k_C and the respective sample mean. Goodness of fit was tested by a χ^2 statistic (Table 10, columns 8 and 9). For eggs, 17 of 70, and for larvae, 13 of 69 testable cases (24 and 19%) were significantly different at α = .1, causing some question as to the commonness of the k's. It should be remembered, however, that setting minimum expected cell size to 1 makes the χ^2 test more susceptable to fluctuations in smaller cells.

A 2nd test, illustrated in Fig. 7, was suggested by Bliss and Owen (1958). The individual sample k's are plotted against the sample mean (here transformed to logs to give clearer separation). For neither eggs nor larvae are the points substantially different from the mean common k, and this is confirmed by regression through the points (Table 11). Although there is considerable variance about the k_c 's, the estimates are not correlated with the mean.

A check was made to determine if the aggregation of larvae was related to age. Although the individual larval square foot counts were not subdivided into counts of each instar, an overall estimate was made for each field. The individual field MLE k and the variance: mean ratio from Table 10 were each compared to the mean instar for that field (Table 12). There were no correlations and plots of the data suggest no higher order relationships. It is assumed, therefore, that there is no significant change in the distribution of larvae attributable to instar. The common k may be accepted as common for all age

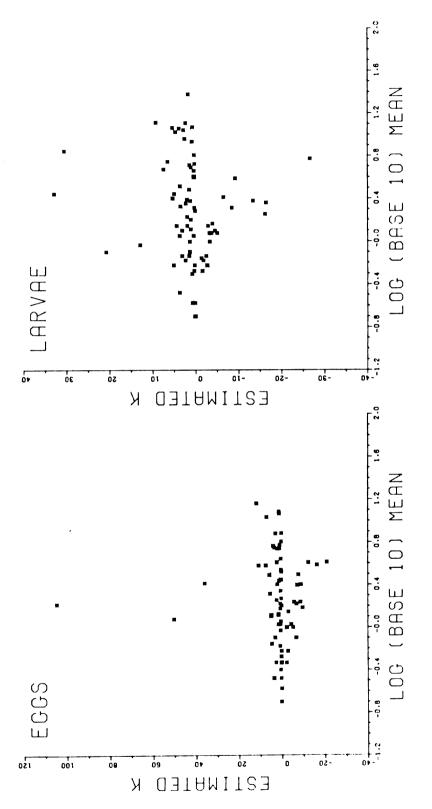


Fig. 7. The relationship between MLE sample k and log of the sample mean for CLB egg and larval count data.

The relationship between sample negative binomial k's and log mean for egg and larval counts. Model is $k_i = a+b \log m_i$. Table 11.

اب	.144n.s.	.900n.s.
$\frac{R^2}{R^2}$	000.	
s.d.	.43	.48
log m	.236 .43	.275
۵	.59 4.45	1.74
s.e.	1.73	.91
Ø	2.22 1.73	.55
s.d.	14.93	7.99
ואין	2.35	1.02
Cases	75	9/
	Eggs	Larvae

The relationship between sample negative binomial k and the variance : mean ratio over mean instar. Model is y = a + bx, where y is k_i or Var : m, and x is mean instar. Table 12.

اب	.717n.s.	-1.231n.s.
R ²	.007	.021
x s.d.	.54	.54
×	3.21	3.21
b s.e.	1.26 1.75	.78
٩	1.26	96
a s.e.	.94	.42
Ø	-2.82 .94	6.05
s.d.	8.05	3.62
χ	1.21	2.96
Cases	73	73

groups.

There seems to be no reason for not taking the estimate of $k_{\rm C}$ for eggs to be the same as that for larvae, using the above tests. A single estimate of common k, $k_{\rm C}$ = 2.75, is the arithmetic mean of the egg and larval $k_{\rm C}$'s. It is adopted for the development of the related transforms, below.

3.3 Variance stabilizing transforms

The negative binomial is a skewed distribution, especially at lower densities. Transforms that reduce variance instability (dependency on the mean) may also reduce the skewness of the frequency distributions. Results from different transforms are often similar and choice of transform may be up to user preference, with little apparent difference in the variance: mean relationships after transformation.

The choice of transform depends on the perceived relationship between variance and mean. For contagious negative binomial type distributions, Beall (1942) used an inverse hyperbolic sine transform,

$$z = \sinh^{-1}(SQRT(kx))/SQRT(k)$$
 (29)

with k, the negative binomial statistic, z, the transformed count, and x, the original count. Kleczkowski (1949) used the simpler

$$z = \log(x+c) \tag{30}$$

with c, a constant. Noting that the transform was less effective at low densities, Kleczkowski (1955) proposed the curvilinear modification

$$z = \log ((x+c+SQRT(x^2+2cx))/2)$$
 (31)

Choice of c was dependent on the intercept of a standard error x mean regression for small, untransformed counts.

To compensate for the apparently common difficulty of transform failure at low density, Hayman and Lowe (1961) used the log variance x

log mean relationship (22) of Wayman (1959). This is equivalent to

$$s^2 = 10^a m^b$$
 (32)

which was simultaneously used by Taylor (1961a) to describe the variance x mean relationship of count data from 24 biological surveys on various animals, from viral lesions to ocean fish. From this relationship, these authors used Kendall's (1973 (1st ed. 1948)) transform,

$$z = \int^{X} (1/SQRT(\bar{x})) dx, \tag{33}$$

which leads to

$$z = x^{(1-b/2)}$$
 .(34)

The "power law", equation 32 (Taylor 1965), and its transform (34) minimized the range of variances of several aphid count sets (Taylor 1970) and was found to be slightly better at this than the commonly used $\log (x+1)$. Note that (34) is not useful when b=2, in which case Taylor used the $\log t$ transform (Taylor 1961a).

Table 8 suggests that a common regression of

$$\log s^2 = .25 + 1.33 \log m$$
 (35)

is a reasonable expression of the within field log variance x log mean relationship for both the egg and larval square foot counts under normal farming conditions. Using Bliss's (1967, p. 128) correction for underestimation of the arithmetic mean from log data,

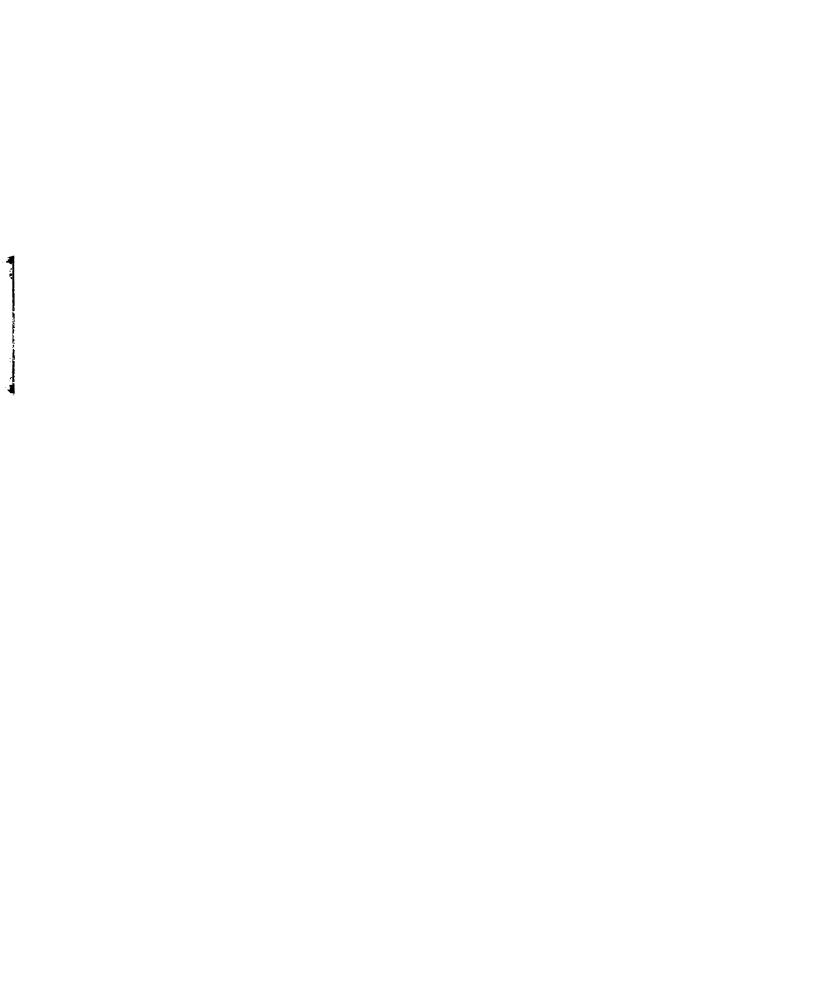
$$m_a = m_g + 1.1513s_g^2$$
 (36)

with m_a the arithmetic mean, m_g the geometric mean of the log data, and s_g^2 the error mean square (here the squared average s.e. for eggs and larvae in Table 8), leads to a variance x mean relationship of

$$s^2 = 2.11m^{1.33}$$
 (37)

for eggs and larvae. Using (34), an appropriate transform is

$$z = x^{.33}$$
 .(38)



Note that this transform depends on a 1st order relationship between log variance and log mean. Fig. 6 suggests that this may not be valid. Ruesink and Haynes (1973) observed that "b" was closer to unity when the sample means were less than .5 per sample. Perhaps this is why Taylor (1970) found the transform less effective at stabilizing variance at low density. Development of second order regressions, however, leads to expressions which are beyond the scope of this thesis to integrate.

The above transforms are compared in Table 13. Two criteria are used here to evaluate transform effectiveness. The simplest test is to compare the ratio of maximum to minimum variance with a ratio established under normality assumptions (Hayman and Lowe 1961, Taylor 1970). The test statistic is

$$\zeta = (\ln \rho)/2, \tag{39}$$

a normally distributed variable with variance $\hat{\sigma}^2$ = 1/df, where df is the degrees of freedom of the sample variance, and $\ln \rho$ is the natural logarithm of the ratio of maximum to minimum variance. Here, the average df is 12.1. The expectation of a range in a normal distribution is approximately $\sigma SQRT(n)$, where n is the size of the sample supplying the range (Masayuma 1957). Thus, under normality assumptions, for both eggs and larvae the expected minimum $\zeta = SQRT(92/12.1)/2 = 1.38$.

Initially, all transforms were evaluated using a search technique to minimize ρ , the ratio of maximum to minimum variance. That is, the transforms were optimized for ζ . Constants of transformation were selected and evaluated iteratively until ζ changed by less than .1%. Subsequently, the correlations of the variances and means of the transformed sample counts were computed (Table 13).

= ratio of maximum to minimum variance. Table 13.

Table 13.	Table 13. Comparison of transforms optimized for minimal ç.	ms optimized for		ρ = ratio of maximum to minimum varianc	minimum varianc
	<pre>r = correlation coeffi</pre>	coefficient, significant at α		= .05 if greater than .22. 🛚	$\zeta = (\ln \rho)/2$.
	Transform	Constant, c	al	N	니
Eggs	Observed		4696.43	4.23	.921
	log (x+c)	.271	19.54	1.49	.153
	log ((x+c+SQRT(x ² +2cx))/2) .740)/2) .740	18.03	1.45	.128
	×.33		22.02	1.55	.148
	٥×	.335	21.86	1.54	.216
	$sinh^{-1}(SQRT(cx))/SQRT(c)$	c) .672	17.99	1.44	.129
	sinh ⁻¹ (SQRT(cx))/SQRT(c) $2.75 (k_c)$	31.14	1.72	.093
	Normal		15.76	1.38	
Larvae	Observed		6458.57	4.39	.941
	log (x+c)	.324	24.11	1.59	860.
	log ((x+c+SQRT(x ² +2cx))/2) 1.29)/2) 1.29	23.82	1.59	.178
	x.33		30.48	1.71	.154
	٥×	.389	25.14	1.61	.350
	$sinh^{-1}(SQRT(cx))/SQRT(c)$	c) .388	23.82	1.58	.178
	sinh ⁻¹ (SQRT(cx))/SQRT(c)	(c) $2.75 (k_g)$	41.83	1.87	.134
	Normal	•	15.76	1.38	

Another criteria for optimizing transforms is to select constants so that the correlation between variances and means of the transformed counts is minimal (Table 14). This also tends to reduce the ζ statistic. A graphic check was made to be sure the lack of correlation was not due to an introduced curvilinearity. There is evidence of residual correlation at low density with all transforms (Fig. 8, for example), but only 1/3 of the overall variance after transform could be accounted for by fitting up to 4th order polynomials (Table 15).

The effect of transforms on the sums of squares and the analysis of variance has been discussed by Beall (1941). In general, gross changes in significance of test statistics will not occur as a result of transformation, but some differences about the relative importance of less correlated variables may occur. The log transform of Table 6, for example, changed few of the conclusions from Table 5, except for the cases of lsts and 2nds in wheat, in which the order of inclusion in the regression was altered.

For all of the transforms evaluated, chosing constants for minimal correlation between variance and mean helped reduce the skewness and kurtosis of the counts, although all remained leptokurtic and somewhat positively skewed. From Tables 13 and 14, all transforms are better than none for reducing variance instability and approximately normalizing the distributions. The simple log transform, with constants .132 and .170 for eggs and larvae, will be used here as it is the simplest and has marginally better ζ 's.

Table 14. Comparison of transforms optimized for minimal correlation between transformed variance and mean.

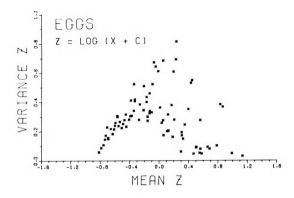
	<u>Transform</u>	Constant, c	ζ
Eggs	Observed		4.23
	log (x+c)	.132	1.64
	$\log ((x+c+SQRT(x^2+2xc))$)/2) .337	1.60
	x ²	.280	1.60
	<pre>sinh⁻¹(SQRT(cx)/SQRT(c</pre>) 1.485	1.60
Larvae	Observed		4.39
	log (x+c)	.170	1.72
	$\log ((x+c+SQRT(x^2+2xc))$)/2) .450	1.71
	x ^c	.283	1.81
	sinh ⁻¹ (SQRT(cx)/SQRT(c) 1.112	1.71

Table 15. An example of higher order variance dependencies following transform. Model is $y = \underline{b'z}$, y is transformed variance, \underline{z} a vector of orders of the transformed mean, using $z = \log(x+c)$.

	<u>Variable</u> *	<u>b.</u>	s.e.	$\frac{R^2}{R}$	ΔR^2	Overall F**
Eggs	z ²	525	.172	.290	.290	36.83
	z ⁴	.111	.222	.322	.032	21.12
	z	133	.071	.327	.005	14.23
	z ³	.256	.153	.348	.021	11.60
	constant	.386	.024		•	
Larvae	z^2	559	.134	.240	.240	28.39
	z ³	043	.193	.331	.091	22.00
	z ⁴	.247	.198	.334	.003	14.68
	z	.095	.077	.345	.011	11.45
	constant	.340	.021			

^{*} Listed in order of inclusion (Nie et al. 1975)

^{**} All F ratios significant at P <.0001



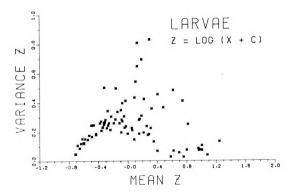


Fig. 8. The relationship between variance and mean for CLB egg and larval counts following transformation.

LINEAR PREDICTORS OF CEREAL LEAF BEETLE DENSITY

4.1 Introduction

The goals of this section are to find a way to use previous measurement information to reduce the error term of estimates of CLB density and to predict seasonal density using early season measurements. This requires a search for relevant environmental factors and linear coefficients relating these to historical changes in density. Rates of population change based on these regressions are then used to estimate present population levels, according to recorded weather and / or biological information. The most recent measurement, if any, may be averaged with the projection for an improved estimate. Models of expected future weather or biological events may be used for a density forecast over an entire season.

The PWS environmental factors measured were temperature (described as degree days, base 48F), windspeed, wetness, rainfall, and crop height (see section 2). Average larval age was estimated from subsample head-capsule measurements. Egg parasitism by <u>Anaphes flavipes</u> was determined from laboratory rearing of weekly collected field eggs.

The applicability of statistical description of change in CLB larval numbers in response to the environmental factors may depend on the relationships among the independent variables. Several of these bivariate relationships are discussed in section 4.3. The development of improved estimators for larval density follows in section 4.3. A multivariate regression is fit to the PWS data, using transformed

counts as the dependent variable. Partial derivatives of this estimator are then used as "filters", in the sense of Hildebrand (1976) and Sakawa (1971).

4.2 Parameters of the temporal distribution of CLB eggs and larvae.

Multiple regressions tend to mask relationships among variables. For the CLB, these relationships have intrinsic properties of interest which have been studied in an ongoing research program for several years. The PWS data set will be used here as a vehicle for review and corroboration of several of these underlying bivariate relationships.

The distribution of error terms on the independent variables will not be discussed here because most of the field measurements (ie. crop height, windspeed, etc.) were made only once per count period. In future studies, these error terms ought to be given the same attention given to the error term of the dependent variable (section 3)(see Box and Tidwell 1962). Out of necessity, these errors will be considered normally distributed with stable variance.

4.2.1 Adult influx

The most important variable initially affecting egg (and hence larval) density is the spring influx of adults. An estimate of the number of adults available for oviposition may come from regional survey data taken early in the season or from samples near the field of interest. Casagrande (1976) found the rates of adult emergence from wintering sites was quite similar for 4 years of Kellogg Farm data. Regional incidence curves for 3 years of sweepnet data also showed a consistant pattern with peak regional density at roughly 200 dd's and a gradual decline over the next 600 to 800 dd's. This is essentially

the pattern for PWS adults (Fig. 9). The PWS peak adult density in wheat was at 206 and 238 dd's for the square foot and sweep sample units. Buildup to this point is quite rapid as the bulk of adult emergence takes place over a relatively short span.

The adult population then decreases in an exponential fashion, $N_t = N_p e^{-\lambda(t-200)},$ (40)

where N_p is the peak population and t is "time" in dd's. The mortality rate, λ , has been estimated to be .0071, .0071, .0037, and .0045 per dd for the 4 years of Kellogg Farm survey. The factors that determine this rate are unknown. Casagrande (1976, p. 29) reported λ was independent of host crop type. The seasonal total of "adult degree days" (the area under the adult incidence curve) is sensitive to this parameter. Mortality rates of .007, .005, and .003 applied to the PWS regional density in wheat at 200 dd's resulted in 55.9, 70.9, and 100.4 adult dd's.

The number of adult dd's may be used to estimate total egg production. Table 16 reports results of several studies. Disregarding the lowest Teofilovic estimate, mean eggs per dd from 6 studies is .35 (s.d. = .044). Wellso (1976) reports average total oviposition per female in laboratory studies is similar to that of field beetles.

Multiplying .35 times 1/2 the total adult dd's gives an estimate of total eggs produced.

The rate of adult movement into oats depends on factors which have not yet been analyzed. Oats is a preferred host for CLB's and acts as a sink, with beetles leaving oats more slowly than they arrive (Casagrande 1976, p. 48). Although this should eventually establish an equilibrium with other hosts, the level at which this occurs and the factors

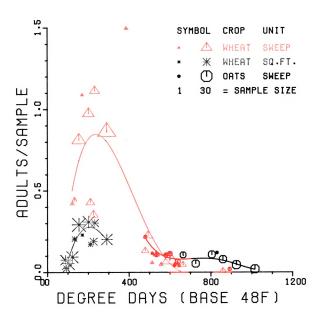


Fig. 9. Changes in adult density over accumulated degree days. Symbol size indicates number of field means included in sample point.

Table 16. Mean eggs per female CLB per degree day (48F).

Source	Eggs/dd	Conditions
Teofilovic (1969; ref. in	.325	30 ⁰ C (86 ⁰ F)
Wellso, Connin and Hoxie 1973)	.055	16 ⁰ C (61 ⁰ F)
Wellso, Connin and Hoxie (1973)	.272	26.7°C (81°F) ^a
	.362	26.7 ⁰ C (81 ⁰ F) ^b
	.381	26.7°C (81°F) ^C
Wellso, Ruesink and Gage (1975)	.388	26.7 ^o C (81 ^o F) ^d
Tummala, Ruesink and Haynes (1972)	.370	unspecified

a laboratory cages, equal numbers of both sexes

determining this level are unknown. In the 1st simulation of the CLB ecosystem, Ruesink (1972) assumed 90% of the beetles in wheat went to oats. Casagrande (1976, p. 144) plotted adult densities in strips of oats. Density increased until it was 10 to 20 times as great as in wheat. The proportions of land area in small grains and in spring grains are important in determining relative densities (Ruesink 1972, p. 28). The spatial arrangement of the host fields is also undoubtedly important in this respect. In the 1975 PWS area, oats were about 12.7% of the small grains and were about evenly interspersed with wheat (map, appendix 4).

The adult incidence curves vary considerably from field to field.

Between plus within field variance is more than within field variance

b laboratory cages, sexes separated before oviposition trial

c field collected, oviposition measured in laboratory

d mated once weekly

for all life stages (Table 17). The Ruesink regressions for adults and larvae in Table 17 are the same because they came from pooled data. There was no visible difference between the regressions for the 2 life stages (Ruesink, pers. comm.). For the PWS data, fields were grouped according to sample date (appendix 7c).

Analysis of variance for the 3 incidence curves in Fig. 9 reveals that much unexplained variance remains after inclusion of many environmental variables and after adjusting for between field differences. Table 18 includes dd's, crop height, windspeed and wetness. Between field differences were handled as classification effects, as were the field wetness ratings. Each field or wetness classification was given either a "0" or a "1", depending on whether a sample fit that classification. Singularities in the $\underline{X'X}$ (or the correlation) matrix were avoided by eliminating 1 wetness classification and 1 field, as per Draper and Smith (1966, p. 137-46). Less than 5% of the cases had missing crop height, windspeed, or wetness variables. All cases were used and the correlation matrix was constructed with pairwise deletion of missing variables (Nie et al. 1975).

Apparently, either sample technique or selection of variables were inappropriate for the adult density model. Of the environmental factors, dd's accounted for most of the variance. Between field differences accounted for about 1/3 of the variance (Table 18).

Two final observations on adult densities are appropriate here.

The amount of host crop planted is highly susceptable to economic factors that are not necessarily related to pest density. For example, Table 19 shows the change in total acreage and numbers of fields in the PWS area from 1973 to 1975, when the relative value of wheat was

Model is The relationship between log between plus within field variance, y, on log mean, x. y = a + bx + error.Table 17.

ا ب	2:	2.6**	13.6***	- 5	23.9***	7.7**
R ²	176.	.889	. 949	176.	.993	.937
s.d.	;	.204	.426	;	.287	.264
×	;	157	936	;	.382	. 335
s.e.	980.	.503	.153	980.	.095	. 279
م	1.934	2.045	1.876	1.934	1.641	1.547
s.e.	.083	.084	.063	.083	.250	.067
ro	.093	.319	.663	.093	.687	. 607
s.d.	!	.443	.820	;	.472	. 422
>		.319	-1.092	;	1.315	1.125
c I	79	9	00 t	62	00t 6	00t 6
Source	ADULTS Sweepnet Ruesink 79	PWS ³	Square f PWS ³	LARVAE Sweepnet Ruesink 79	Square f PWS	EGGS <u>Square foot</u> PWS 6 1

1 1967, 1968, Cooperative CLB survey. Unit is catch per sample. Up to 196 fields of wheat, oats, corn and barley were sample in each of 16 townships. Statistics from Ruesink and Haynes (1973).

² F tests significant at p <.001.

³ Pubescent Wheat Study, 1975.

Table 18. Analyses of variance for adult CLB's per sweep in wheat and oats and for adults per square foot in wheat. $^{\$}$ Model is $y = \mu + \underline{a'x} + \text{error}$, where $\underline{b'} = (\text{dd } \text{dd}^2 \text{ ch } \text{ch}^2 \text{ W } \underline{m'})$, dd = degree days, ch = crop height, W = windspeed, $\underline{m'} = \text{wetness}$ classifications, and f' = field effects (0,1).

<u>a</u>	SSR (d.f.)	SSE (d.f.)	<u>F</u>	$\underline{R^2}$		
Wheat: catch per	sweep					
<u>b</u>	44.9 (7)	204.1 (394)	12.38***	.180		
<u>f</u>	75.7 (60)	167.3 (364)	2.75***	.379		
(<u>b</u> <u>f</u>)'	127.3 (67)	121.7 (334)	5.22***	.511		
Wheat: catch per square foot						
<u>b</u>	4.1 (7)	27.1 (396)	8.53***	.131		
<u>f</u>	10.0 (63)	21.2 (341)	2.55***	.412		
(<u>b</u> <u>f</u>)'	17.2 (70)	14.0 (333)	5.85***	.552		
Oats: catch per	sweep					
<u>b</u>	.24 (7)	1.45 (125)	2.97**	.143		
<u>f</u>	.59 (23)	1.10 (122)	1.33n.s.	.293		
(<u>b</u> <u>f</u>)'	.67 (30)	1.02 (102)	2.24***	.397		

\$ = Reduced models are not to be confused as being parts of compound model. Differences in degrees of freedom are attributable to missing data elements.

Table 19. Wheat and oat planting in the PWS area. Figures in parentheses are for Michigan only. Michigan data, 1973-74, from M. Holmes (APHIS-USDA, pers. comm.). 1974 PWS figures from R. Gallun (USDA, Purdue University, pers. comm.).

	19	73	1	1974		1975	
	<u>Fields</u>	Acres	<u>Fields</u>	Acres	<u>Fields</u>	Acres	
Oats	(12)	(115)	14 (11)	n.a.(94)	17 (15)	181 (162))
Wheat	(11)	(111)	46 (45)	n.a.(519)	73 (58)	1234 (933))

responding to high international export volume. Oats acreage was comparatively stable during this time. The presence of a constant CLB population in the area from 1973 to 1975 would have resulted in an apparent decrease in average field density in wheat.

The distribution pattern of agricultural fields may remain stable from year to year. Fig. 10, for example, maps the distribution of 1/4 sections containing oats in the Jackson County, Pulaski township, Michigan, CLB survey site for 1973 to 1975. Although the number of oat fields dropped slightly in this area, there were few sections in 1974 or 1975 which did not contain or were not adjacent to a section with an oat field the previous year. Similar maps of all the CLB survey sites listed in Appendix 2 suggest that this continuity of pattern was general. If the degree of aggregation of a population is intrinsic to a species (D. Kendall 1948, Taylor 1965, 1970), it must nevertheless be susceptable to the spacial dynamics of crop planting pattern. Assuming that the adult spatial wintering pattern is closely related to crop pattern in the previous summer and that the spring host selection flights of adults can cover at least a mile diameter area, present crop planting habits in the Michigan CLB survey areas will do little to alter the between field CLB adult count variance x mean relationship from year to year.

The remaining error in the adult density models is large, but is understandable. Sweepnet and stick toss techniques have long been suspect as adult sampling methods, but have been retained for lack of suitable alternatives. Larger sample size would reduce error in density estimates. Further discussion of the nature and causes of between field variance of adults will be deferred to a later study (A. Sawyer,

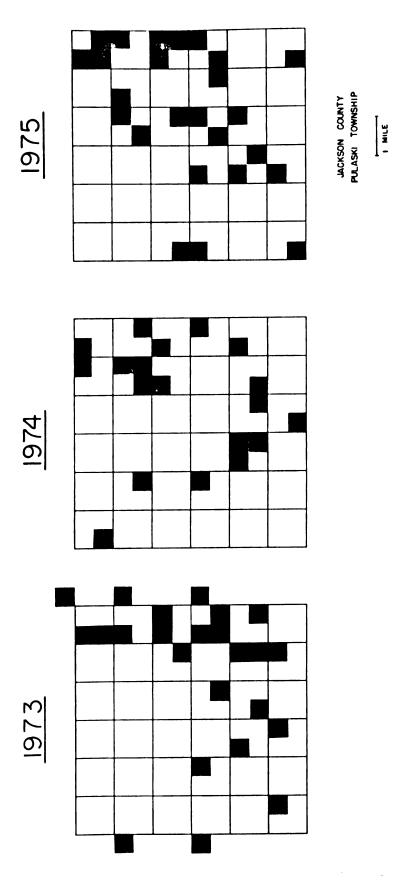


Fig. 10. Locations of quarter sections containing oat fields, Pulaski township, Jackson County, Michigan, 1973-75.

PhD thesis, Michigan State University, ca. 1978).

4.2.2 Crop height

Crop height (ch) is an indicator of several "states" of the plant, including its age. It is an indirect summary of all the factors that have been experienced by the plant, such as fertilizers, water and warmth. Each state in turn may be important for its influence on CLB density.

Those spring crops which are planted late will be exposed to a decreasing regional adult population and will receive fewer eggs, even though younger crops are preferred for oviposition (Shade and Wilson 1964a). There is a tendency for spring grains to have higher densities than the fall planted winter wheat for the same reason.

The relationship between wheat and oat ch and accumulated dd's has been graphically illustrated by Gage (1972) and Sawyer (1976). The relationship is variety specific and consistant, and, like crop maturity, may normally be subject to little modification by other environmental effects (Wiggins 1956). It is essentially that of the PWS data (Fig. 11, but note that a base of 48°F was used, not the more common base 42°F). Further analysis of this relationship suggests that growth of wheat may be much more uniform across the region than the growth of oats (Table 20). Degree days account for over 80% of the seasonal variance for wheat, but less than 40% for oats.

A hypothetical explanation for this lies in the difference in planting times for the 2 crops. Although the pattern will vary from year to year, there was a 12 day period of cool, dry weather in October 1974 (59 dd accumulated) when it is assumed most wheat was planted. Wet

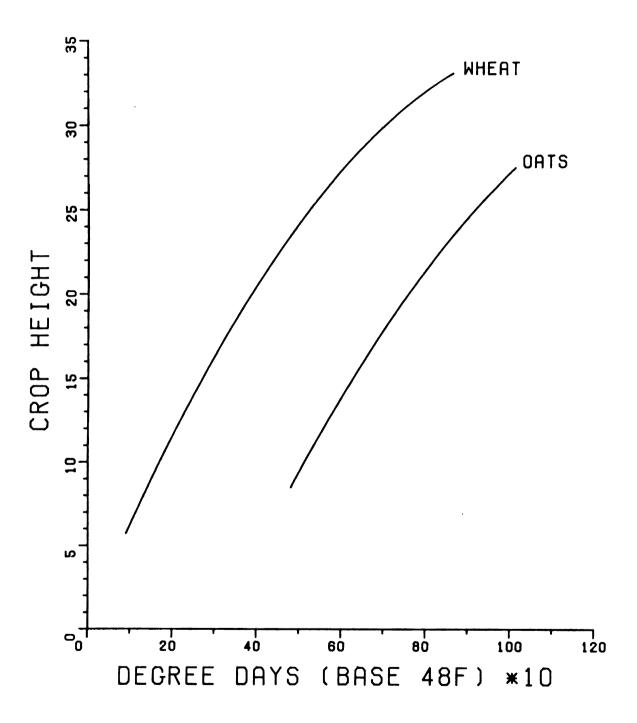


Fig. 11. The relationship between crop height and accumulated degree days for the PWS data. Regressions are: oats... $y = -19.5 + .069x - .000022x^2$, $R^2 = .388$; wheat... $y = .47 + .061x - .000026x^2$, $R^2 = .837$.

Table 20. Analyses of variance for wheat and oat crop height. dd = degree days, \underline{f} = field effects (0,1). Model is $y = \mu + \underline{a}'\underline{x}$ + error.

<u>a</u>	SSR (d.f.)	SSE (d.f.)	<u>F</u>	<u>R</u> 2	
Oats					
dd	5412.5 (1)	8652 .6 (172)	107.6***	.385	
dd, dd ²	5462.4 (2)	8603.1 (171)	54.3***	.388	
dd, dd ² , <u>f</u>	12666.0 (25)	1399.4 (148)	53.6***	.901	
Wheat					
dd	46709.0 (1)	10057.0 (840)	3901.0***	.823	
dd, dd ²	47515.0 (2)	9251.0 (839)	2154.0***	.837	
dd, dd ² , <u>f</u>	52200.0 (65)	4566.0 (776)	136.5***	.919	

Table 21. Percentage of eggs parasitized by <u>Anaphes flavipes</u>, PWS 1975 (data in Appendix 6).

Date	Degree Day (48°F)	Number of Fields	Number of Eggs Sampled	Mean Percent Parasitized (+s.d.)	
Oats					
6/2	565	18	413	11.0 (10.5)	
6/9	663	16	435	12.6 (9.4)	
6/16	804	16	449	86.6 (14.9)	
6/20	924	19	444	94.6 (8.2)	
6/25	1067	13	193	98.8 (3.2)	
Wheat					
5/14	220	17	960	0.1 (.46)	
5/21	345	17	1002	0.0 (0)	
5/28	492	17	390	7.3 (6.4)	
6/2(3)	565	17	360	21.0 (20.3)	

spring weather drew out oat planting times over roughly a 3 week period of 260 dd's in May. The result is a greater spread of oat planting times on a physiological scale than was experienced by wheat, hence the larger between field difference. Inclusion of between field differences in the oat crop height regression thus brings the R² value to .90.

4.2.3 Egg parasitism by Anaphes flavipes

Anaphes flavipes is an established CLB egg parasitoid in the Galien area (T. Burger, pers. comm.). In the PWS survey, parasitism was monitored weekly from May 14 to June 2 in wheat and from June 2 to June 25 in oats. Leaves bearing CLB eggs were clipped, returned to the lab, and transferred to a plastic petri dish. Emerged larvae, or eggs with clearly identifiable contents were noted and removed daily. All egg development was usually completed in 7 to 9 days. These data are included in the individual field summaries of Appendix 6 and are summed up in Table 21.

Table 22 lists the mean and standard deviation of total egg counts made in wheat and oats. In addition to the separate egg counts discussed above (section 3), many fields were simply given a total count per 10 or 15 square feet. All egg samples are thus included in Table 22 and the statistics given are the statistics of field means.

The individual field density means were used to determine the least squares fit of density over polynomials in degree days (up to 4th order). The percentages of parasitism in Table 21 were fit with 2nd order curves and the estimated number of parasitized eggs per square foot was calculated (Fig. 12). The areas under the total egg and parasitized egg incidence curves were compared for an estimate of seasonal egg parasi-

Table 22. Statistics of average egg density by date and degree day.

	<u>Da te</u>	Degree Day (48°F)	Number of Fields	Density (per Mean	square foot) S.D.
0ats					
	May 27 30	478 533	8 7	21.133 4.343	12.687 5.111
	June 2 3 5 6 7 9 12 16 17 18 20 25	565 579 621 637 643 663 725 804 829 858 924	13 1 4 11 17 19 17 1 21 21	3.195 .200 10.500 3.636 .500 2.388 3.002 2.543 4.067 1.248 1.457 1.111	4.276 0.000 5.696 3.261 0.000 2.202 3.048 2.823 0.000 1.679 1.577
Wheat					
	May 7 8 9 13 14 16 19 20 21 23 27 28 29 30	142 154 170 210 220 231 290 320 345 386 478 492 512 533	27 8 18 33 16 42 4 39 17 5 19 27 6 38	8.788 17.600 2.878 20.582 6.850 22.674 16.025 15.105 18.508 22.471 8.684 3.332 5.394 3.266	13.077 12.094 1.783 21.033 5.698 23.985 13.805 13.920 20.854 16.064 6.004 2.587 4.715 3.072
	June 1 2 5 6 7	544 565 621 637 643	5 5 4 12 5	2.840 2.220 1.250 1.281 .347	2.970 3.190 .994 .729 .345

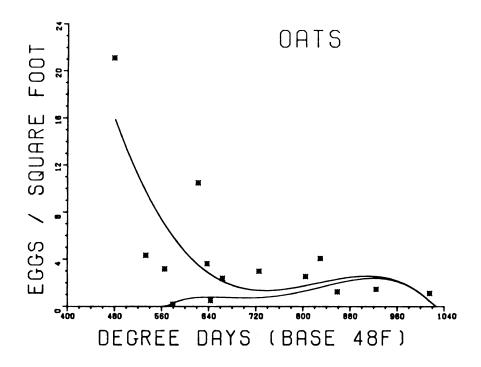
tism. For wheat, this estimate was 2% and for oats 29%, although the early part of the season was apparently missed in many oat fields (Fig. 12).

The effect of egg parasitism was to prevent the later part of the larval incidence curve from developing. <u>A. flavipes</u> thus competes with larval parasitoids, as has been discussed by Haynes and Gage (1972).

The seasonal rate of egg parasitism depends on many as yet unknown factors and varies by region. The extent of egg parasitism has been higher and has occurred sooner in several eastern U.S. survey areas (T. Burger, pers. comm.). Although the biology of <u>A. flavipes</u> has been studied (Anderson and Paschke 1968, 1969, 1970), little is known about native alternate hosts and there is no knowledge of the wintering habits. It is unknown how the <u>A. flavipes</u> density pattern of the Galien area in 1975 related to other areas or times.

4.2.4 Age of the larvae

The average age of larval insects is of interest if it relates to changes in the efficiency of a sampling technique. This problem was discussed above for the CLB. The average age also has value if it is relatable to changes in insect density. If differences in crop planting time, for example, make it impossible for insects to lay eggs in field B until much later than A, the subsequent average age of larvae in B would be expected to be less than that in A at any single time. The incidence curve in B will likewise be shifted to a later part of the time axis. The question then is: If mean instar is substituted for degree days (or another such "time axis") can the predictability of the incidence curve be enhanced?



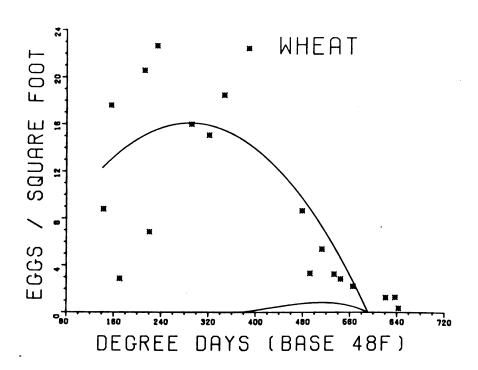


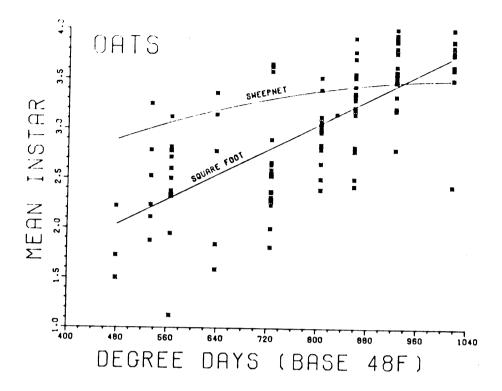
Fig. 12. Mean egg density (upper curve) and estimated incidence curve for eggs parasitized by <u>Anaphes flavipes</u>.

Following the notation of Lee, Barr, Gage and Kharkar (1976, pg. 40-43), the physiological age of an individual CLB can be represented by z, an indicator with a value of z=0 for a newly laid egg and $z=\ell$ for the oldest possible age of an individual. Then at a time, t, the number of individuals in a particular age group between $(z_i, z_i + dz)$ is $x(z_i, t)dz$. For CLB larvae, z takes on the 4 discrete integer values for the 4 instars, as no continuous indicator of age is available. The average for many individuals, however, is a continuous variable, with values from 1 to 4. Lee et al. developed a partial differential equation to describe the maturity distribution of the population as a function of time. For now, the change of the average age of larvae over time, Σ x $(z_i,t)/i$, is all that will be considered.

Fig. 13 shows the relationship between average instar and dd's for wheat and oats. The regression based upon the corrected sweepnet samples is included for comparison. The individual field instar means, as determined by square foot sampling, are plotted but not the means based on sweepnet.

The second order dd equation accounts for approximately 1/3 and 1/2 of the instar variance for square foot counts in oats and wheat (Table 23). Crop height has relatively less relation to instar and inclusion of ch and dd's in one model does not improve fit (Table 23).

For wheat, the regressions of Fig. 13 and Table 23 may not represent the true regional pattern as only a small part of the total number of fields were included in any day's samples (Table 24). Also, the square foot samples in wheat represent only a short part of the total incidence curve. The sweepnet samples are, of course, subject to the



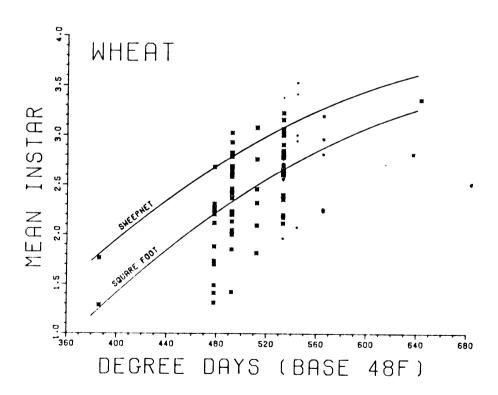


Fig. 13. Relationship between instar and degree days. Plotted points are for square foot samples only.

Table 23. Analyses of variance of larval instar. dd = degree days; ch = crop height. Model is $\hat{y} = a + \underline{bx}$.

<u>x</u>	SSR (df)	SSE(df)	\underline{F} \underline{R}^2	-
Oats-Square Foot				
dd	27.32 (1)	22.68 (116)	139.8*** .5	5
ch, ch ²	10.50 (2)	39.50 (115)	15.3*** .2	!1
dd, dd ² , ch, ch ²	30.70 (4)	19.30 (113)	44.9*** .6	51
Oats-Sweepnet				
dd, dd ²	3.20 (2)	26.80 (137)	8.2*** .1	1
ch^2	.18 (1)	29.82 (138)	.8 n.s0	11
dd, dd ² , ch, ch ²	3.93 (4)	26.08 (135)	5.1** .1	3
Wheat-Square Foot				
dd, dd ²	7.96 (2)	14.00 (92)	26.2*** .3	6 *
ch	2.22 (1)	19.74 (93)	10.4** .1	0
dd, dd ² , ch	8.37 (3)	13.59 (91)	18.7*** .3	18
Wheat-Sweepnet				
dd, dd ²	35.41 (2)	21.00 (185)	156.0*** .6	i3
ch, ch ²	10.93 (2)	45.47 (185)	22.2*** .1	9
dd, dd ² , ch, ch ²	35.97 (4)	20.44 (183)	80.5*** .6	j 4

Table 24. Summary of larval instar samples.

			Square Foot		Sweepnet			
	Date	dd	Number of	Ins	tar	Number of	Instar	
			Fields	Mean	S.D.	Fields	Mean	S.D.
0ats								
	May 27	478	3	1.816	.369	3	2.394	.345
	29	512	_	0.464	500	1	1.875	0.000
	30	533	6	2.464	.500	2	3.238	.081
	June 2 3	565 579	12	2.451	.517	6	3.118	.189
	4	599				7	3.520	.319
		621				i	3.520	0.000
	5 6 7	637	5	2.542	.789	10	3.390	.264
	9	643 663				1	2.500	0.000
	12	725	19	2.576	.532	12 18	3.396 3.109	.265 .319
	16	804	16	2.953	.288	17	3.196	.563
	17	829	1	3.153	0.000	1	3.420	0.000
	18	858	20	3.273	.391	17	3.383	.339
	19 20	891 924	20	2 (01	207	4	3.594	.081
	25	1015	20 16	3.601 3.675	.307 .372	20 20	3.617 3.467	.215 .665
			10	3.073	.572		3.407	.003
Wheat							•	
	May 23	386	2	1.526	.339	6	1.728	.094
	27	478	13	1.970	.414	19	2.705	.277
	28 29	492 512	26 6	2.377 2.421	.378 .454	27 6	2.768 2.708	.323
	30	533	36	2.706	.454	15	3.207	.279
	31	544	5	2.991	.574		0.20,	
	June 1	556	5	2.691	.428	8	3.113	.548
	2	565				_		
	3 4	579 599				2 48	3.690 3.525	.099
	5	621				40 6	3.570	.122
	5 6	637	1	2.809	0.000	13	3.575	.274
	7	643	1	3.357	0.000	4	3.423	.275
	9	663				17	3.478	.232
	18	858				17	3.663	.240

sampling biases discussed above. The samples in oats were accumulated over a longer period. A relatively complete representation of the area's fields was obtainable within one day (Table 24).

4.2.5 Rainfall

Shade, Hansen and Wilson (1970) recorded the mortality of the 4 CLB instars during 2 rainstorms in 1966. Third instar larvae experienced a significantly greater mortality than either 1st's or 2nd's. The rate of precipitation in the storms was .31 and .36 inches per hour, falling over periods of 45 and 25 minutes. Total mortality was 14.5 and 13.5 percent for the two storms. Little else has been done to look at the dynamics of populations in storm conditions. Although a rain gage kept track of the precipitation at the center of the PWS area, this is not presumed to be an accurate estimate of the precipitation in all fields. Rainfall is entered in the following regressions only as total inches in the approximate 24 hour period prior to sampling (based on the single rain gage).

4.2.6 Insecticides

One of the reasons that the goodness of fit of instar over degree days (Table 23) is not higher than .55 may be the occurrence of insecticide applications on some of the farms. These roughly coincided with "normal" peak larval density, during a period from 600 to 800 degree days. This is reflected in a decline in mean instar. The mean for the oats' square foot sample on dd 725 (Table 24) was 2.58, which is close to the 533-565 dd measurement (mean instar = 2.46) and is less than the value predicted from the overall regional regression (Fig. 13) (predicted mean instar = 2.85).

Insecticide use on CLB's is extremely common and is a major factor affecting population levels and population dynamics. Although they are usually ignored in studies on the ecology of the CLB, insecticides are in fact a part of the normal agricultural ecology of most regions. Any attempt to predict future regional beetle or parasitoid densities, or to evaluate spatial or temporal distributions, must be preceded by an understanding of the dynamic effect of insecticide usage.

For example, it should be realized that the perception of between-field variance (s^2) x mean (m) relationships may be affected by insecticides. Two cases will illustrate.

Case 1: $\log s^2 = a + b \log m$. The use of sprays at higher density will still produce a 1st order estimator with expected coefficients "a" and "b".

Case 2: $\log s^2 = a + b_1 \log m + b_2 (\log m)^2$. If a 1st order equation is fit to the data, the coefficients "a" and "b" will depend on the degree of spraying. Removing those sub-populations with higher means will result in a lower estimate of the 1st order slope. A few moments relfection on Fig. 6 will make this clear (see Pielou 1969, pg. 91-97).

Note that although use of 2nd order equations to describe variance x mean relationships may produce little increase in goodness of fit (section 3.2.4), the resulting estimator may be preferable, depending on the likelihood of insecticide usage.

Although it is not always possible to witness insecticide application directly, the incidence curves for eggs and larvae and the instar curve may contain sufficient information for inference. A precipitous drop in egg and larval density and concurrent retardation of the rate of instar

advance may be taken as indicators of insecticide use. Since actual knowledge of whether a field had been sprayed existed for only 5 of the 21 oat fields studied (maps, Appendix 4), a posteriori inference was necessary. Graphs of square foot density and instar changes in each field plus sweepnet and crop height information were all reviewed. On this basis, it was determined that 10 of the fields were not sprayed and 11 probably were. In the following derivation of a filter for reducing measurement error, only the unsprayed 1975 PWS fields were used.

4.3 Filtering

4.3.1 General considerations

Any estimate of beetle density based on count data has an error term associated with it, as discussed in section 3. This section is an evaluation of a process called filtering, which is an attempt to improve current density estimates by using previous measurement information. Extension of this same process to a complete seasonal incidence estimate is also discussed.

Filters are also called "linear minimum variance sequential state estimation algorithms" (Sage and Melsa 1971). Their basic idea is not new (Weiner 1949, Kalman 1960) but application to entomological problems is uncommon (Hildebrand 1976). The parametric approach used here is a very simple "first pass" at using the technique. This section is thus meant to be an evaluation of the potential for developing a complement to the more sophisticated discrete component simulation methods (Gutierriz, et al. 1974; Tummala, Ruesink and Haynes 1975) which will ultimately serve as the basis for an on-line pest management system for CLB (Haynes 1973). In the interim, the predictor may also provide a useful, low-cost estimator of seasonal density.

Gage (1974, p. 79) reported the initial, peak and last CLB larval populations in oats (<u>+</u> s.e.) to occur at 375 (<u>+</u>12), 723 (<u>+</u>25), and 1218 (<u>+</u>16) dd, respectively, for 6 years of Gull Lake Kellogg Farm studies. In what follows, it is assumed that the shape of the larval incidence curve is determined by the environmental factors discussed above plus the available ovipositing adult population. The incidence curve clearly cannot begin until after the crop sprouts. The last eggs will be determined by some unknown combination of adult mortality rate and plant senescence. For normally planted oats it is assumed that the curves should approximate Gage's (1974) characteristics.

The area under the curve also will determine its shape. This area is related to the potential egg input and thus to the adult influx. Either of these could be measured early in the season to be used as an indicator of the larval curve's area, but not without complicating the measuring and modeling errors.

The algorithm used here, which is meant to apply to CLB's but which is generalizable to other pest/crop systems, requires only an initial estimate of larval density and initial environmental conditions. Current environmental measurements plus the derivatives of a multiple regression of historical density and environmental variables are then used to project a current or future point. Since it is beyond the scope of this thesis to evaluate the environmental and sociological factors that govern density patterns in sprayed fields, these were not included in the analysis. The only suitable large data set available for parameter estimation was the 1975 PWS unsprayed oat fields. This provided 80 density measurements in 10 fields. The 1976 PWS data and

three fields from the 1974 Kellogg Farm work were available for evaluating the filter.

4.3.2 Density models

Table 25 lists several univariate models of density (transformed as per section 3) over environmental factors for the 10 unsprayed PWS fields. Transformation used the $z = \log(y + c)$ relationship developed in section 3 to improve variance stability prior to analysis. The relationships between log density and dd's or ch (Table 25) are clearly curvilinear for larvae. Eggs show less reliance on higher order terms probably due to failure to sample during the 1st part of the incidence curve. This is because most oat fields were not located until very close to or after peak egg density. The resulting density pattern has a negative decay rate somewhat like the adult mortality pattern (Fig. 9).

The factors, field wetness (m), rainfall in the previous 24 hours (r24), average larval age (i), seem to have little relative influence by themselves on density. This, however, may be due to the way the rainfall was measured (section 4.2.5) and to inaccuracies in instar determinations (section 2.2.2). Certainly, further work on the impact of rainfall is needed.

Sprouting date (s) was also included as a variable. These data were determined by applying the quadratic formula to the ch regression of Fig. 12, adjusting the y-intercept according to a field effect variable. That is, another regression was run using $ch = a + b_1'f + b_2dd + b_3dd^2$ as a model, with ch = height and f, a vector of field effects, coded (0,1). Combining the constant "a" and the f product into a single term produced a ch equation of the form, f ch = f a f + f and f the f complete term produced a ch equation of the form, f ch = f and f the f complete term produced a ch equation of the form, f ch = f characteristic term f characteristic term f constant f constant f characteristic term f codes.

Table 25. Univariate models of density [Y = log (y+c), section 3] over several environmental factors.

			Eggs			Larvae	
Model ^a	Cases	R^2	c.v.b	F. sig. ^C	R^2	c.v.b	F sig. ^C
dd	80	.327	84.5	.0001	.009	141.4	.393
dd, dd ²	80	.330	84.8	.0001	.323	117.6	.0001
ch	80	. 334	84.0	.0001	.012	141.2	.331
ch,ch ²	80	.358	82.5	.0001	.303	119.3	.0001
ch, ch^2, ch^3	80	. 359	83.5	.0001	.376	113.7	.0001
<u>f</u> (10)	80	.354	87.4	.0001	.347	119.5	.0001
m ₂	79	.009	102.5	.411	.006	141.6	.512
m ₃	79	.014	102.3	.305	.012	141.2	. 328
m ₂ ,m ₃	79	.031	102.0	.304	.014	141.9	.574
r24	80	.039	100.9	.079	-	-	-
i	61	.312	85.6	.0001	.021	140.8	.268
i,i ²	61	.318	86.0	.0001	.104	135.8	.041
s	80	.072	99.2	.016	.037	139.4	.089

^aModel is of form Y=a + $\frac{b}{x}$, where $\frac{x}{x}$ includes variables listed: dd=degree-days base $\frac{48}{F}$; ch=crop height (in.); $\frac{f}{f}$ =(0,1) field effect (10 fields); $\frac{m}{2}$, $\frac{m}{3}$ =damp or wet field conditions; r24= rainfall in previous 24 hours; i=instar; s=sprouting date

bPercent

^CProbability regression is N.S.

 b_2 dd. The quadratic formula was used to find the roots of this equation, one of which was easily accepted as the degree days accumulated by the sprouting date. The results (Table 25) indicate that sprouting time by itself contributes relatively little to the total variance. Of the variance in larval density explainable simply as between field effects (\underline{f}) , only about 10% could be attributable to sprouting time.

Figure 14 shows the log density regressions over dd's and ch. The predicted density was transformed $(y = 10^{Z} - c)$ before plotting. The poor fit to either variable by itself is evident, but not unexpected.

In Table 26, combinations of several of the environmental factors are evaluated. Although inclusion of additional variables will in general improve the R^2 values, up to R^2 's of about .78 for full models for both eggs and larvae (Table 26, bottom), these larger models are not very useful. The inclusion of all measured variables plus "field effects" in the full model yields an "adjusted R^2 " (Cohen et al. 1976, p. 16) of only .71. The combination of dd's (2nd order), ch, and the dd x ch interaction term provides an adjusted R^2 of .454 for larvae. This combination is simple in terms of numbers of variables to be measured but it provides significance and reasonable fit. The effect of sprouting time could be accounted for in the interaction term. Also, the fit of ch over dd's (Fig. 11) makes substitution possible, facilitating differentiation. This regression is, therefore, chosen for use in the rate calculations.

Supplying the appropriate coefficients, the larval predictor is $\log (y + .17) = b_0 + b_1 dd + b_2 dd^2 + b_3 ch + b_4 dc$ $= b_0 + b_1 dd + b_2 dd^2 + b_3 (p_1 dds + p_2 dds) + b_4 dd (p_1 dds + p_2 dds)$ (41)

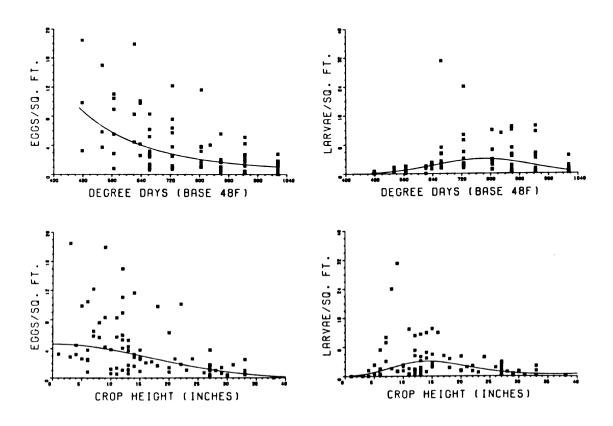


Fig. 14. Relationship between egg and larval densities and degree days and crop height.

Table 26. Representative multivariate models of density [Y = log (y+c), section 3] over several environmental factors.

		{	Eggs			-Larvae	
<u>Model^a</u>	Cases	$\underline{R^2}$	c.v.b	F sig. ^C	\mathbb{R}^2	c.v.b	F sig. C
dd,ch,dc	80	.446	77.6	.000	.417	109.9	.000
dd,ch,dd ² ,dc	80	.473	76.2	.000	.481	104.3	.000
dd,ch,ch ² ,dc	80	.474	76.1	.000	.427	109.6	.000
dd,ch,dd ² ,ch ² ,dc	80	.438	75.6	.000	.518	101.2	.000
dd,s	80	.404	80.0	.000	.045	139.7	.167
dd,s,dds	80	.459	77.3	.000	.168	128.7	.001
ch,s	80	.391	80.9	.000	.039	140.1	.217
ch,s,chs	80	.418	79.5	.000	.269	123.1	.000
dd,s,dd ²	80	.407	80.3	.000	.362	115.0	.000
ch,s,ch ²	80	.410	80.1	.000	.343	116.6	.000
dd,ch,s,dd ²	80	s	not ente	ered	.390	113.2	.000
dd,ch,s,dd ² ,ch ²	80	.501	74.6	.000	.525	100.5	.000
dd,ch,s, <u>m</u> ,r24,i,dd ² ,c	h ² ,i ² ,dds	,chs,do	² s,ch ² s				•
	79	.573	72.2	.000	.594	98.5	.000
dd,ch,s, <u>m</u> ,r24,i,dd ² ,c	h ² ,i ² ,dds	,chs,do	l ² s,ch ² s,	<u>f</u> (full n	nodel)		
	79	.578	57.0	.000	.781	75.9	.000

^aModels are of form $Y = \underline{bX}$, where elements of \underline{X} are degree-days base 48°F (dd); crop height (ch); sprouting date (s); plant wetness (m) (see text); rainfall in previous 24 hours (r24); instar (i), and field effects (\underline{f}), plus intercept (b₀).

^bCoefficient of variability, percent.

^CProbability $\underline{b} = 0$.

where:
$$b_0 = -5.516 \pm 1.071 *$$

 $b_1 = 1.345(10^{-2}) \pm 3.12(10^{-3}),$
 $b_2 = -6.814(10^{-6}) \pm 2.224(10^{-6}),$
 $b_3 = 0.105 \pm 3.302(10^{-2})$
and $b_4 = -1.515(10^{-4}) \pm 4.041(10^{-5}).$
Also, $p_1 = 6.838(10^{-2}) \pm 1.567(10^{-2}),$
and $p_2 = -2.257(10^{-5}) \pm 1.016(10^{-5}),$ from Fig. 11.
Then, since
 $\hat{y} = 10^K - .17,$ (42)

where K is the right hand side of (41),

$$\frac{dy}{dd} = (y + .17)(C_1 + C_2 dd + C_3 dd^2) \ln 10,$$
where: $C_1 = b_1 + b_3(p_1 - 2p_2 dds) + b_4(p_1 dds + p_2 dds^2),$

$$C_2 = 2(b_2 + b_3 p_2 + b_4(p_1 - 2p_2 dds)),$$
and $C_3 = 3b_4 p_2,$

$$(43)$$

with dds, the estimated dd's at time of sprouting, as discussed above.

This equation (43) describes the rate of change in the average 1975 PWS unsprayed oat field larval density in response to dd's and ch. To adjust the rate for other densities, the initial measurement in the field is compared to the predicted density from (42). The ratio (measured + .17) / (predicted + .17) is multiplied by the rate (from (43)) to produce a corrected rate. The addition of the .17 was necessary to allow the predicted density to asymptotically approach zero, instead of being restricted to a finite range.

^{* +} standard error of the coefficient

4.3.3 The weighting of the second observation with the predicted density.

An initial field measurement of larval density is needed to start the filter. The rate of change in density (43) can then be applied (section 4.3.4) for an estimate of current density. The question of interest in this section is how to best average this estimate with a corresponding current measurement. The choice is between equal weighting (a simple average) and weighting somehow based on the relative reliability of the 2 numbers. I'll evaluate the latter choice first.

The initial measurement (the one that starts the filter) has a variance associated with it which can either be measured or estimated from the equations of Fig. 6 or from (21) using the common k of Table 10. One approach to weighting is to use a rate of change in this error term similar to the rate of change of the density estimate. A simple version of this was used by Hildegrand (1976) who simply multiplied the variance of earlier measurements by an apparently arbitrary 1.05 per day, "thus weighting the past measurements less." This is intuitively appealing in that older measurements somehow seem less reliable than more current ones.

A similar result is obtainable from evaluating the rate of change of the confidence limits about the prediction equation (42). In general, the symmetric confidence interval around any estimate of E (y_0) corresponding to the set \underline{x}_0 is

$$\underline{\mathbf{x}}_{\mathbf{0}}^{'}\hat{\mathbf{b}} + \hat{\mathbf{\sigma}}\mathbf{t} \quad \underline{\mathbf{x}}_{\mathbf{0}}^{'} \left(\underline{\mathbf{X}}^{'}\underline{\mathbf{X}}\right)^{-1} \underline{\mathbf{x}}_{\mathbf{0}}$$
 (46)

(Searle 1971, p. 108, eqn. 95), where \underline{b} is the vector of estimated regression coefficients, t is a value of Student's t statistic, $\hat{\sigma}$ is an estimate of residual error variance (Searle 1971, p. 93), $(\underline{X}'\underline{X})$ is from

the original data set, and \underline{x}'_0 is a specific set of x's. For retransforming from the logarithmic relationship of (43), the confidence limits are

$$\frac{x_0'\hat{b} + \hat{\sigma}t}{10} \times \frac{x_0'}{\sigma} \times \frac{(X'X)^{-1}}{\sigma} \times \frac{x_0'}{\sigma} -.17$$
 (47)

The derivative of (47) involves the solution of a quadratic of the form d $\underline{x}'\underline{Ax}$ / dt, which at the time of this writing prohibits further progress.

Neither the divergent trend of Hildebrand's multiplier nor the rate of change of the prediction equation confidence interval is entirely satisfactory. Continuous divergence of the confidence limits is a very conservative approach, eventually negating the worth of early season measurements. Even altering the value of the constant is objectionable until an empirical basis for this change is obtained. On the other hand, the confidence limits of the prediction converge as they move toward the peak density, and diverge away from the peak, ignoring the loss in reliability due to the age of the previous measurement information.

A third approach to the variance estimate is simply to use the estimate from Fig. 6 (or (21)) applied to the predicted current density. This again ignors the age of the previous measurement information. However, it does make the larger densities, which inherently have greater variance, account for less when averaged. This is a problem since the weighted average will be used as the starting point for the next iteration of the filter. The net result will be bias toward low seasonal incidence estimates.

Simple arithmetric averaging of the predicted and the measured density produces an unbiased estimate which incorporates past information, weighting most recent measurements more than previous, though it still ignors the relative ages of the measurements. Despite the latter drawback, it is the averaging technique that will be used in the following.

4.3 Filter operation

The filter algorithm is quite simple. The filter is initiated by the first non-zero field density measurement. CH and dd accumulation are also needed. Ch is incremented in 1 dd steps according to the relationship of Fig. 11. Ch and dd are then entered into the rate equation. The calculated increase is adjusted by a factor that is the ratio of the density measurement to the predicted estimate (from (42)). This same factor is retained for each dd's rate calculation. The amount of increase is added to the density measurement and the process is repeated up until the date of the next measurement. The resulting prediction is then averaged with the observed density, as discussed above. After adjusting for ch error by using the new ch measurement (see below), a new correction factor and a new rate is calculated and the process is repeated.

In evaluating the dd and ch components of the filter, it was noticed that the regional (PWS unsprayed fields) average larval density seemed to peak later than the averages for 6 years of Gull Lake data (Gage 1974, p. 79). Re-examining the dd data used (Appendix 8), I decided that the decision to average the PWS site center data with the New Carlisle and South Bend stations (see 2.5) was a mistake. Rather than convert the dd accumulation, the rate equation was merely shifted to 30

dd's earlier. This was the difference in accumulated dd's between the PWS site center and the average, for the site center 700-750 dd period (the expected peak period).

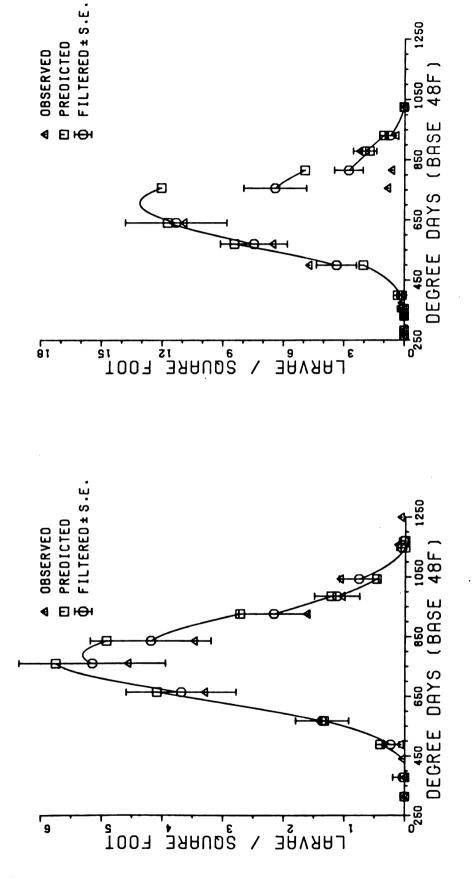
Filter operation is illustrated in Figs. 15a and b, using 1 field from the 1976 PWS data (Sawyer, pers. comm.) and 1 from the 1974 Gull Lake data (Sawyer 1976). Other fields in these studies were also used to evaluate the filter and these results are listed in Appendix 9.

The filter behaves roughly as expected, with incidence curves similar to Gage's in respect to peak and last observed incidence. Depending on ch, larval density tends to peak at around 750 dd's, and to go to zero near 1150 dd's as in Fig. 14.

The filter also noticeably conserves previous information, which is, of course, its purpose. This makes initial measurement accuracy quite important. A low measurement initially contributes to a low estimate at the next and subsequent observation periods.

The filter is sensitive to the accuracy of the dd day accumulation. A shift of the rate equation a few dd's either way can be quite important. Table 27 shows the effect of the 30 degree day shift mentioned above. The filtered peak density in this example increased 19% following the shift.

There is also a problem with the plant growth equation as used in the filter. Measurement of standing crop height is subject to errors. To allow for changes in crop height (and thus the larval density rate change equation) over time, the crop height over dd equation of Fig. 11 was used to predict the path of crop height between measurement periods. The next new measurement and the prediction were averaged arithmetically and the filtered crop height term was used as the beginning for the next crop height path prediction.



Examples of filter operation. a) (left). Application to 1976 PWS data (Sawyer, unpublished), each observation based on 15 to 20 2-linear foot samples. b) (right). 1974 Gull Lake data (Sawyer 1976), each observation based on 30 square foot samples. Fig. 15.

Table 27. An example of sensitivity of the filter to accumulated degree-days, Gull Lake field 916 (see Fig. 18B). O(Y)= observed larval density; E(Y)=predicted from rate equation; F(Y)=filtered estimate.

	0r	iginal		Shifted	by 30 dd
Degree-Day	0(Y)	E(Y)	F(Y)	E(Y)	F(Y)
313	0.00	-	-	-	-
378	0.00	0.00	0.00	0.00	0-00
440	.03	0.00	.03	0.00	.03
488	.06	.35	.20	.39	.22
567	1.40	1.02	1.21	1.28	1.34
663	3.29	2.99	3.14	3.98	3.63
760	4.56	3.99	4.27	5.64	5.10
835	3.47	3.46	3.47	4.87	4.17
926	1.61	1.80	1.71	2.71	2.16
985	1.03	.81	.92	1.21	1.12
1043	1.06	.30	.68	.47	. 76
1157	.11	0.00	.06	0.00	.06
1249	.06	0.00	.03	0.00	.03

			i

The ch equation of Fig. 11 needs to be adjusted for the sprouting time of the particular field. One way of doing this is to use the ch curve and the first measurement to project backwards to the zero height (sprouting) dd, as discussed above. Subsequent applications of the ch curve could then be based on this figure. Without knowing the accuracy of the initial measurement, however, there is no reason to accept this sprouting time as correct. This is perhaps more clearly a problem when the field is not located immediately after sprouting but instead is found when it is already, say, 8-12 inches tall, as is most commonly the case in survey work. A better estimate of sprouting time might be the filtered estimate obtained after the second measurement time. But using this procedure a logical question is, why stop here, accepting only one estimate of the sprouting time? In the filter as used here, the approach was taken that each new measurement of ch, filtered using previous measurements, provided a valid estimate of sprouting time. Crop growth rate and density rate equations were thus altered for subsequent prediction periods. There are obvious drawbacks to this approach and refinements could be made. But perhaps the most limiting is the observation that the estimated sprouting date seemed to be later as later measurement information was added. Also, estimates of crop height were most frequently higher than observed, indicating plants were growing more slowly than predicted. For ch to be used in the filter, a better model is needed. In most plant studies, however, yield is emphasized and data on height is often secondary or ignored (eg. Wiggins 1956; Holt, Bula, Miles, Schreiber and Peart 1975; Jackman 1976).

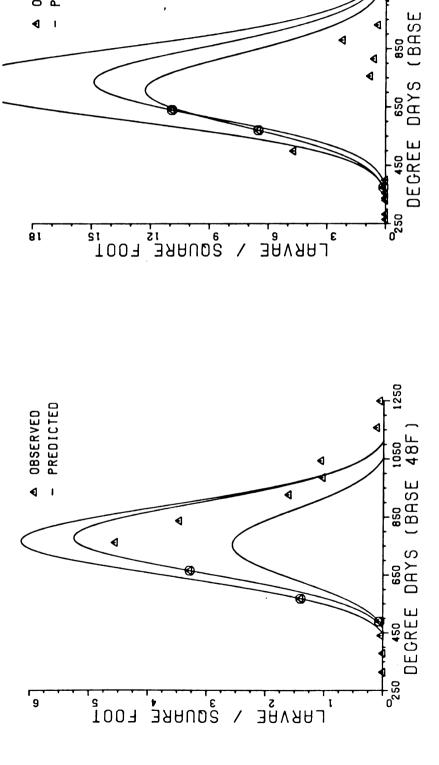
Re-estimation of sprouting time and filtering of ch changes the density rate equation slightly at each time of observation. This change is in addition to the change from the adjustment factor, section 4.3.2. Thus, for observation near peak density, for example, it is possible for a negative predicted trajectory to become positive at the time of a new observation.

An additional application of the density rate equations is to predict seasonal incidence curves. This can be done from a single measurement, projecting both forward and backward in time. Measurement error, however, is extremely critical. Fig. 16 illustrates the use of the rate equations for 3 different observation points, treating each observation as though it were unique.

4.4 Further limitations of the filter

Because of the simplistic nature of the density rate change equation, the effect of many potentially important factors is ignored. Perhaps the most critical is the effect of egg parasitism, which may vary greatly between regions. The net effect of increasing egg parasitism is to accelerate the rate of decline of the larval density in late season. On a dd basis, this may be compensated for by reducing the absolute value of the coefficients of either the 2nd order dd or the interaction term in the larval predictor equation (41). The magnitude of correction necessary for a given level of egg parasitism is unknown but experimentally determinable. The alternative, not explored here because of numerous missing elements in the data set, would be to include egg and parasitized egg densities at previous measurement points as part of the larval density regression. The spacing (separation in dd's) of

▲ OBSERVED- PREDICTED



Use of larval density equations to estimate secsonal incidence. Extrapolation from 1976 PWS representative points, Δ , is indicated by solid prediction curve. Left: data (Sawyer, unpublished). Right: 1974 Gull Lake data (Sawyer 1976). Fig. 16.

1050 48F sample periods and great difficulties in analysis made this approach virtually useless as a practical tool.

Difficulties in estimation and prediction may not be entirely the fault of the filter. Sawyer's (1976) data uniformly showed peak densities of larvae at the 640 dd observation. This is somewhat earlier than expected if the Gage characteristics (Gage 1974, p. 79) are held to be generalizable. This might be attributable to an error in measuring dd accumulation, as it was warm in late March. For several years, it has been customary to begin marking the accumulation of dd's on April 1. There is no reason for this other than the generalization that there usually are no dd's before this in many parts of Michigan. A better standard might be to begin accumulating after the end of the last full 2 week period in which no dd's were accumulated, or to use some similar standard marking the cessation of cold weather. In 1976, there were a few warm days at the end of March which could have contributed to the accumulation of dd's experienced by the population of beetles. The 1976 PWS adult population peaked near the 200 degree-day mark, however (Sawyer, pers. comm.), removing some of the doubt about dd accumulations.

Another cause could have been a more general use of insecticides than was observable. Average density on dd 640 was over 12 larvae per 2 linear feet, with some fields reaching 20 and even 43 larvae per sample. It is not known how farmers perceive these densities, nor what factors go into a decision to spray, but it should be noted that the pattern of field 1024 (Appendix 9), a field known to have been sprayed (Sawyer, pers. comm.) had a below-average density and displayed a less precipitous decline than other fields not labeled as sprayed. Investigation of egg and adult density curves and larval age structure for the

1976 PWS data may shed further light on this problem, but that work is yet to be completed (Sawyer, Ph.D. thesis, Mich. State Univ., ca. 1978).

ESTIMATION OF LARVAL PARASITISM BY TETRASTICHUS JULIS

To estimate the fraction of the CLB larval population that is parasitized by \underline{T} . \underline{julis} , or to obtain density estimates, requires taking a larval sample. The statistical difficulties in this have been discussed above. Several additional technical problems with larval dissections also require investigation before addressing the problem of improved parasitoid density estimators.

Sweepnet sampling to obtain an estimate of larval parasitism is affected by sweepnet bias. The percentage parasitism of each instar must be adjusted by the instar specific catch correction factors.

Overall apparent (i.e., at the time of sampling) parasitism rate is thus dependent on the accuracy of the correction factors. For the 85 cases of paired sweep and square foot samples used in section 2, proportions of parasitized and total CLB larvae (Table 28a and b) were corrected by use of the simple multipliers of Table 4. The averages for the corrected samples (Table 28c, column 5) generally reflect the effect of larger numbers of younger instars. Younger instars have lower rates of parasitism (Table 28d). Thus, application of sweepnet bias correction factors lowers the estimated rate of parasitism (Table 28c, column 3).

Use of the correction factor doesn't account for all the differences in parasitism rate between sweepnet and square foot, though. The remaining difference is most probably caused by preservation and/or microscope technique (section 2.2.1). Square foot sample larvae were preserved by freezing and sweepnet samples used FAA. Based on past

d = mean number dissected. Average parasitism rates of CLB larvae by T. julis, 1975 PWS. Table 28.

'nS	Instar:		į	[! !	;	! ! !	2	 		3		1 1 1	4	1 1 1 1 1
	DD48F	Cases	اه	%T	(s.d	d.)	미	%T)	(s.d.)	٦I	LT%	(s.d.)	ы	%T7	(s.d.)
A.	Sweepnet	net sample	a												
	478	1	5.0	٥.	_	(0:	15.0	۳.	(0.)	7.0	ъ.	(0.)	0.	0.	(0.)
	533	13 2	0.	0.	_	(0:	3.0	0.	(0.)	45.5	42.9	(33.5)	20.5	69.4	(43.3)
	637	17 4	1.25	0.	_	(0:	13.25	1.8	(3.6)	37.25	21.1	(23.1)	45.5	33.5	(23.8)
	725	5 18	.44	0.	_	(0:	8.06	.5	(1.6)	22.56	4.1	(7.1)	19.28	13.2	(15.8)
	804)4 16	1.13	7.3	(25.1	.1	6.56	2.7	(8.9)	38.81	7.0	(10.3)	29.94	11.6	(11.4)
	829	1	0.	0.	_	(0:	2.0	0.	(0.)	25.0	-	(0.)	23.0	- .	(0.)
	858	91 89	90.	0.	_	(0:	4.56	1.	(3.5)	29.06	3.7	(6.7)	32.88	5.3	(6.5)
	924	14	.36	0.	_	(0:	2.57	21.3	(36.5)	19.07	29.0	(29.3)	66.07	13.4	(10.7)
	1015	13	0.	0.	_	(0.	2.0	17.9	(33.4)	11.77	41.8	(41.9)	32.77	50.3	(37.0)
В.	Square	foot	sample												
	478		7.0	0.	_	(0:	21.0	0.	(0.)	17.0	0.	(0.)	0.	0.	(0.)
	533	3 2	1.0	٥.	<u> </u>	(0:	0.9	5.0	(7.1)	0.6	18.2	(25.7)	1.5	16.7	(23.6)
	637	7 4	3.25	0.	<u> </u>	(0:	5.0	5.0	(10.0)	7.5	0.	(0.)	8.5	3.2	(3.7)
	725	5 18	3.06	0.	<u> </u>	(0.	10.89	۳.	(1.1)	11.72	3.5	(7.4)	6.94	6.7	(23.8)
	804	1 16	1.94	1.3	<u>.</u>	5.0)	6.56	9.	(2.5)	16.56	1.5	(3.4)	10.19	2.6	(9.9)
	829		0.	0.	<u> </u>	(0.	0.6	0.	(0.)	32.0	0.	(0.)	18.0	0.	(0.)
	828	91 8	.63	0.	_	(0.	3.06	0.	(0.)	16.56	.7	(1.5)	23.81	1.0	(1.9)
	924	1 14	.14	0.	<u> </u>	(0:	2.43	0.	(0.)	12.50	3.7	(8.2)	29.50	2.9	(4.9)
	1015	5 13	.08	7.7	(27	(7.	.77	0.	(0.)	6 .08	6.4	(10.1)	23.85	2.5	(4.3)

Table 28 (continued)

C. Average percent parasitism (\underline{T} . \underline{julis}) for all larval samples.*

DD48F	<u>Cases</u>	Sweepnet	Square Foot	Corrected Sweepnet
478	1	22.22	.0	10.09
533	2	33.51	12.14	29.71
637	4	29.71	1.47	20.54
725	18	10.05	1.41	7.68
804	16	4.48	.69	4.13
829	1	8.0	.0	7.21
858	16	4.08	.72	3.8
924	14	9.63	1.26	9.98
1015	13	62.08	6.32	60.38

D. Parasitism rate by instar, all samples.

	SI	weepnet	S	quare Foot
Instar	No. CLB's	No. Parasitized	No. CLB's	No. Parasitized
1	42	2	121	2
2	461	35	456	4
3	2184	308	1092	28
4	2949	581	1447	42

^{*} Average percent = 100(Σ (Σ TJ / Σ CLB)) / n ... i = 1..4; j = 1..n j i

Table 28 (continued)

C. Average percent parasitism (\underline{T} . \underline{julis}) for all larval samples.*

DD48F	Cases	Sweepnet	Square Foot	Corrected Sweepnet
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637	4	29.71	1.47	20.54
725	18	10.05	1.41	7.68
804	16	4.48	.69	4.13
829	1	8.0	.0	7.21
858	16	4.08	.72	3.8
924	14	9.63	1.26	9.98
1015	13	62.08	6.32	60.38

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2	461	35	456	4
3	2184	308	1092	28
4	2949	581	1447	42

* Average percent = 100(Σ (Σ TJ / Σ CLB)) / n ... i = 1..4; j = 1..n j i

experience with FAA preserved specimens, above stage microscope lighting was used for dissections. The ivory colored T. julis eggs and larvae were teased loose from the beetle body and were easily discernable. This was not the case with the frozen samples, however. When frozen, T. julis eggs and larvae tend to rupture. Detection of the membranous chorion is possible and reportedly there is little confusion with other membranes; but this requires substage lighting. Freezing and dissection with substage lighting has been reported (T. Burger, pers. comm.) to be superior to use of FAA or alcohol preservation and above stage lighting (i.e., the parasitism rates were closer to corresponding live sample results). However, when using above stage lighting, as was the case in the PWS dissections, T. julis eggs and larvae are much more difficult to find. Parasitism rates for square foot samples (Table 28c, column 4) are thus underestimated by an unknown magnitude. These shortcomings in the PWS data set, and in all similar existing sets, make it impossible to meaningfully proceed with development of filters or incidence curve predictors for parasitoids at this time. Nevertheless, it is possible to provide some quidelines for future efforts and to suggest some of the variables that may be most useful in model building, based on previous research on T. julis biology (Gage 1974). Accumulated degree days play an important role in the emergence of adult wasps from wintering sites. The soil temperatures experienced by the wasps will rely on soil type, ground cover and moisture variables, but Gage (1974, p. 39) concluded that at least a field average relationship between soil and air temperatures justified use of air temperatures in predictive equations.

Degree days also can be used to predict the rate of larval development (Gage 1974, p. 32). Unfortunately, the relationship between mortality rate and temperature is less clearly understood. Gage (1974, p. 70) used accumulated dd base 85°F as an index of mortality in the soil but had no data on mortality rate in CLB larvae on foliage. Thus, there is substantial but incomplete evidence, sufficient to suggest the potential utility of heat units as a significant variable for deriving rate equations for filtering.

Just as the CLB larval incidence pattern is affected by the time of host plant sprouting, so \underline{T} . <u>julis</u> density patterns would be determined by the time of availability of the host. It is not unreasonable to evaluate crop height as an indirect but possibly useful predictive variable (See Gage 1974, p. 92).

The attack behavior of \underline{T} . \underline{julis} is apparently greatly affected by wind and rain (Gage 1974, p. 58). Just as with the CLB larval density estimators, a more thorough investigation of the effects of rain duration and intensity is badly needed. Also, a concept of "wind days" above a threshold may have value in predicting rate of parasitism. \underline{T} . \underline{julis} is apparently inhibited by winds, so that a higher than average accumulation of wind days would mean a lower than average attack rate.

The development of filtering equations for \underline{T} . \underline{julis} density in CLB larvae should consider at least the above set of variables. Development of the rate equations for 1st generation \underline{T} . \underline{julis} follows analogously the CLB density change rate equations in section 4. Second generation curves need to include an addition set of factors to account for the diapause rate of T. \underline{julis} . Considerable work has been done to

determine causes of diapause rates (Gage 1974; S. Koul, pers. comm.; Logan, unpublished notes), but neither temperature, soil moisture, age of female, or constant light (e.g., 16 hrP, 8 hrS) regimes definately cause changes in diapause incidence.

Predicting the path of the bimodal parasitized larval incidence curve is difficult due to the number of potentially significant variables affecting rates. Any meaningful attempt at the multiple variate analysis and regression needed to fit data requires reasonably accurate measurement of parasitism rate throughout the season. With the current problems in the PWS and all existing data sets, any effort to develop filters along the guidelines laid down above is presently premature.

SUMMARY AND CONCLUSIONS

6.1 Summary

Sweepnet sampling for cereal leaf beetles is inaccurate. Conversion to square foot density equivalents may be accomplished by use of the multipliers of Table 3 or 4. Inclusion of some additional environmental parameters provides little improvement in the estimator. Sweepnets may be used as tools of detection but statements about the degree of confidence in the detection statement (Ruesink and Haynes 1973) ought to be subject to larval bias corrections.

Analysis of factors affecting changes in low density populations, where some samples are disproportionately more "reliable" than others due to sample size (section 2.4.) may require a weighted regression analysis for correct interpretation.

The variance:mean relationships of CLB egg and larval counts (sections 2.2.1 and 3.2.2) suggests increasing aggregation at higher densities; i.e., variance is unstable. Use of 2nd order log/log transforms produces a nearly Poisson variance:mean relationship at low densities and accounts for increased variance at higher densities.

Taylor's power law, which relies on an assumption of a first order linear relation between log variance and log mean and hypothesizes that the slope for that relationship is a biological constant, is cast in doubt by the second order fitting. Density dependent mortality thus would produce a changing first order slope, according to, for example, the regional tendencies to apply insecticide.

Several transforms are more or less equivalent in reducing variance instability and at normalizing the distribution of egg and

larval square foot count data. A log transform was selected from the alternatives due to its simplicity.

Accumulated degree days and crop height are useful in filtering larval density data, accounting for half the variance in the PWS data set. Derivatives of a log transform of density over these terms is useful for predicting future (including total season) densities, and for conserving previous measurement information to improve current density estimates. Inaccuracies in previous measurements will be retained in the filtered estimate.

The use of larval maturity as an improved scale for time (a substitute for degree or calandar days) has limited utility. The numbers of larvae to be collected and measured for precise estimates of proportions is logistically limiting at the initial research stage and would be prohibitive at an implementation phase without low cost automated age group classification techniques.

The problem of estimating <u>T</u>. <u>julis</u> densities within CLB larvae is compounded with the difficulties of larval density estimation. The complexity of the bimodal parasitism curve demands accurate measurement data before parameterization is possible and current data sets are limited in this respect. Although there is sufficient understanding of <u>T</u>. <u>julis</u> biology to allow speculative guidelines for development of filtering algorithms, accurate estimation of parameters will require more precise quantification.

6.2 Concluding Remarks

The review of larval CLB sweepnet models confirmed doubts about the net's sampling accuracy. The 1976 PWS program incorporated this and

greatly reduced the use of the sweepnet for sampling.

The work on distribution of count data relative to a standardized sampling scheme has significant applicability in the development of sequential sampling algorithms. Corroboration of the negative binomial relation with a common k along with development of a suitable dynamic damage threshold model for small grains would allow farmers to minimize the expense of detecting currently threatening population densities. The procedure for doing this is laid out in "The Rationale of Sequential Sampling, with Emphasis on its Use in Pest Management" (Tech. Bul. 1526, USDA-ARS, 1976, 19 p.). The use of filtering equations could further reduce the required number of samples for determining an updated density estimate.

Finally, the use of density rate change equations, despite their simplistic assumptions, should provide growers with an improved ability to interpret current density information. In these regards, continuation of the evaluation of and further refinements of the sweepnet and quadrat sampling and the filtering models discussed in this thesis seems certainly merited.

Appendix 1 - Michigan release sites for Tetrastichus julis (Walker)

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_	ł

Section	3333	4 4	13	2 5	36 26	34	17	_	27	19	24	34	·	33	2]	33?	~	352	Fr. claims		25	150	, O	13	26	7
Township	Sherman Reno Tablas	Deerfield	Schoolcraft	Chase	Mavfield	Kasson	Blissfield	Putnam	Iosco	Richmond	Arcadia	Riverton	=	Warren	Ingersoll	Bundee	Bedford	Ida	Raisinville	Sidnev	Ravenna	Montague	Dayton	0xford	Weare	Churchill
County	Iosco "	Isabella	Vackson Kalamazoo	Lake "	Lapeer	Leelanau	Lenawee	Livingston	=	Macomb	Manistee	Mason	=	Midland	=	Monroe	=	=	=	Montcalm	Muskegon) =	Newaygo	Oakland	Oceana	Одетам
Section	7 18 19	1,26	<u>o</u> m	33 33	34	14	20	17	35	4	33	_	34	7	22	35	7	m	18	32	80	35	10	2	12	31
Township	Harrisville Otsego	Banks Carlton	Rutland	Thornapple	Watervliet	Noble	Coldwater	Eckford	Volinia	=	Silver Creek	Wilson	Benton	Sheridan	Watertown	01 i ve	0vid	Dallas	Vermontville		Mayfield	Reading	Brookfield	Ingham	Boston	Ronald
County	Alcona Allegan Alpena	Antrim Barry	-	Rerries	=	Branch	=	Calhoun	Cass	=	=	Charlevoix	Cheboygan	Clare	Clinton	=	=	=	Eaton	Genesee	Grand Traverse	Hillsdale	Huron	Ingham	Ionia	=

County	Township	Section	County	Township	Section
Osceola	Highland	16	Genesee	Thetford	28
=	Richmond	36	Huron	Chandler	7
Oscoda	Comins	56	Ingham	Ingham	2
Otse go	Corwith	36	Ionia	Berlin	2
Presque Isle	Belknap	2	=	=	5
Saginaw	Albee	13	=	Campbell]5
St. Clair	Lynn	32	=	-=	14
Sanilac	Lamotte	12	Kalamazoo	Schoolcraft	=
Shiawassee	Vernon	7	Livingston	Unadilla	2
=	Fairfield	01	Manistee	Arcadia	24
Tuscola	Arbela	17	Monroe	Ida	<u></u> 6
Van Buren	Hamilton	13	=	=	15
=	Pine Grove	20	=	Raisinville	Fr. claims
Wayne	Plymouth		Muskegon	Sullivan	
			Oakland	0x ford	13
1972			Ottawa	Polkton	24
,			St. Clair	Lynn	32
Arenac	Whitney		Sanilac	Marlette	35
=	=		Shiawassee	Hazelton	18
Barry	Thornapple		=	Middlebury	35
= :	Castleton	10	=	Burns	9
= :	Woodland	15	Tuscola	Arbela	37
=		32	=	Almer	വ
Berrien	Lake	2]	Wayne	Northville	6
Branch	Quincy	23	=	Van Buren	23
Calhoun	Leroy	18	=	Canton	36
Clinton	Watertown	25	=	Brownstown	7
=	Bingham	21			-
=	0vid	7			
Genesee	Forest	32			

County	Township	Section	County	Township	Section
Arenac Clinton Ottawa 1974	Lincoln Clayton Duplain Jamestown	18 33 19 24	Newaygo " " " Oceana	Norwich Sheridan Sherman (2 releases) Wilcox Greenwood	20 4,15 7,21 12
Ionia Newaygo 	Berlin Ionia Keene Orange Lyons Portland Ashland Ashland Big Prarie Bridgeton Dayton 4,10	19 12 28 5 3 30 30 18 31 28 9,21,26 17 20 4,10,20,23,24,29,30			
	Garfield Goodwell Grant	6,16 21,28 17			

Appendix 2. Parasitism rates in samples from MSU Department of Entomology CLB state census, 1973-1975.

		1973					~ 1	1974					1975	
Date	LO-0	No. of Fields	Mean CLB's ³	Mean % Parasitism ²	Date	10-0	No. of Fields	Mean CLB's ³	Mean % 2 Parasitism ²	Date	٥-0	No. of Fields	Mean CLB's 3	Mean t Parasitisr
Allega 6/12 6/20 6/26	Allegan County 6/12 755 6/20 935 6/26 1050	- Dorr Towns 12 13 13	wnship 81 85 24	18 2 37	6/10 6/20 6/27	625 750 830	20 25 26	145 232 310	27 8 7	6/8 6/16-7 6/22	560 700 840	24 26 27	49 33 33	40 50 31
Barry 6/14 6/20 6/26	County - 810 960 1080	Barry County - Thornapole 6/14 810 21 6/20 960 21 6/26 1080 21	e Township 220 116 13	12 11 62	6/12 6/20 6/27	700 810 920	30 26 29	213 207 215	28 15 13	6/8 6/16 6/22*	560 690 840	13 21 3*	114 33 89	45 20 73
Emmet	County -	Emmet County - Resort Towns	wnship		6/16 6/26 7/5	455 576 751	19 19 21	342 71 28	15 25 23	6/24	670	19	150	91
Jackso 6/11 6/18 6/25	n County 592 771 945	Jackson County - Pulaski To 6/11 592 30 6/18 771 29 6/25 945 29	Township 45 99 13	21 4 4	6/5 6/12 6/21 6/27	576 720 869 948	23 24 21	31 37 37	49 19 10	5/31 6/6 6/12	515 590 690	25 26 26	200	50 60 37
Lake C 6/12 6/22 7/2	631 631 820 987	Lake County - Chase/Pinora 6/12 631 11 6/22 820 11 7/2 987 10		ips 3	6/16 6/27 7/5	606 746 911	14	96 166 8	71 E					
0scoda 6/14 6/19 6/27 Shiawa	. County . 598 674 824 8see Cour	Oscoda County - West Comins 6/14 598 5 6/19 6/4 4 6/27 824 5 Shiawassee County - New Hav	ins Township 310 860 33 Haven Township	nip 17 38 36 sshfb	6/16 6/25 7/7	594 701 937	8 / /	8 E E	26 61 57					
6/13	731 926 1044	99 88 99	363 167 7	. 17 2 67 67	6/12 6/17** 6/25 7/3	690 770 886 1060	23 23 29 29	506 389** 708 50	38 E 6	6/4-5 6/12	575 690	88	4 0 20	58 52
Degre	e-days,	base 48°F,	accumulat	Degree-days, base 48 ⁰ F, accumulated from 1 April, at nearest NOAA weather station	1, at near	rest NOA	4 weather	station						

uegree-days, base 48°r, accumulated from 1 April, at nearest NOAA weather station.
2 [(parasitism rate for field i)/ E i, given there were any larvae found.
3 Mean per 100 sweeps.
2 Samples lost in lab accident.

**Samples lost in lab accident; figure known to underestimate mean CLB's/100 sweeps.

Appendix 2b. 1974-76 MSU Cooperative Extension Cereal Leaf Beetle Parsitoid Recovery Program

Key to footnote symbols used in total larvae column:

•	Symbol	Sample Unit
1 25 sweeps 2 50 sweeps 3 125 sweeps 4 150 sweeps 5 200 sweeps 6 250 sweeps 7 300 sweeps	h 1 2 3 4 5	hand picked, no unit 25 sweeps 50 sweeps 125 sweeps 150 sweeps 200 sweeps 250 sweeps

1974-76 MSU Cooperative Extension Service Cereal Leaf Beetle Parasitoid Recovery Program. Appendix 2b.

County/ Town (sec.)	Date	Host	Tot	Larvae Tot Dis	% Par	County/ Town (sec.)	Date	Host	Tot	Larvae ot Dis	% Par
Antrim Banks (1)	14/6/74 25/6/74	0	158 956	50	36 20	Cass Volinia (21) (26)	25/6/75 10/6/75	00	120 300	50	44
(7)	3/7/74	3	28 28	282	٥٢.٢		12/6/75	02	593	20	0
(22)	23/6/74 23/6/75 23/6/75	03	70 161 4	2022	°.00	Charlevolx South Arm (8)	24/6/75 1/7/75	0	68 240	50 50	56 56
Barry Carlton (24)	9/6/75	3	29	29	0 6	Cheboygan Benton (33)	5/7/74	0	74	74	70.3
Rutland (4) (10)	27/6/74	: O 3	132	20 22	44 20	Clinton	4///71		Ø	×	8/.5
Thornapple (28)	17/6/75 11/6/74 22/6/74	0	37 100 89	20 22	2.7 32 8	Bingham (21)	11/6/74 25/6/74	0	51	42 51	45.2
Unspecified	27/6/74 20/6/74	Z	120	200	20 4 0	0vid (7)	3/ // 4 12/6/74 25/6/74 3/7/74	0	333	23 33 5	56.5 18.2 20
Benzie Joyfield (9)	10/6/75 20/6/75	733	77 285	50	0 0	Hillsdale Reading (35)	31/5/74	000	0 6	0 6	0 6
Branch Unspecified	31/5/74 11/6/74	₹X	0 -	0 -	00	Huron Brookfield (10)14/6/74)14/6/74	0	33 23	30	33.3
Calhoun Leroy (18)	3/6/74	0	26	26	19.2	me don'	27/6/74		45	42	19
Unspeci fied	20/6/74 3/6/74 12/6/74	3 3 4	130	100	23 63.6 0	Aurelius (12)	10/6/75	0	408	20	99

County/ Town (sec.)	Date	Host	Tot	-Larvae Dis	% Par	County Town (sec.)	Da te	Host	L Tot	-Larvae Dis	% Par
Ingham						Ionia					
Aurelius (22)	13/6/74 20/6/74	0	216 1322	20	18	Ronald (31)	2/6/75 9/6/75	3	97 35	50 35	38 45.7
	27/6/74	u	919	20	14		2//9		21	51	14.3
Ingham (2)	14/6/74	0	<u></u> ∞	<u>_</u>	5.9		7/9		47	47	84.7
	28/6/74		130	20	4		//9	0	95	95	79.3
Ionia						Isabella					
Berlin (1)	13/6/74	0	202	20	22	Deerfield (14)	13/6/74	0	51	5	47.3
•	21/6/74		818	20	J6	•	20/6/74		225	20	52
			406	20	0		>		75	20	48
	/9/8		37	37	100	Nottawa (15)	5/7	0	13	13	23.1
	14/6/76		22	22	95.5		20/6/74		74	20	26
	21/9/12		56	5 6	88.5		//		33	38	55.3
(19)	12/6/74	0	35	35	46.9						
	19/6/74		703	20	22	Kalamazoo					
	24/6/74		431	20	18	Brady (22)	2/1	0	36	36	33.3
(32)	2/6/15	0	154	20	46		12/6/74		279	180	20
	9/6/15		48	48	68.8		2/9		7	7	0
	16/6/75	•	က	က	66.7	Comstock (32)	2	0	573	20	2
Eaton (2)	6/6/75	X.	14	14	78.6	<u> </u>	6/7	0	4	4	75
	11/6/75	M4	15	12	41.7	Schoolcraft (11	2/9	3	7	2	20
	20/6/75	3	4	4	22		/		ა	വ	0
Ionia (1)	14/6/76	0	20	20	75		6/7	0	2	54	14.8
	21/6/75	0	30	30	53.3		6/7		237	20	∞
Otosco (32)	15/6/75	0	370	20	J6		27/6/74		98	20	18
	19/6/75	0	125	20	∞		2/9	3	17	17	0
Poland (1)	9	0	29	29	84.7		2/9	0	58	28	0
_	જે	3	183	20	5 8	(14)	7	0	_	_	0
			83	20	50						
	જે		33	33	5.6						

County/ Town (sec.)	Date	Host	Tot	Larvae Dis %	e % Par	County Town (sec.)	Date	Host	Tot	-Larva Dis	e % Par
a ye						loolansu					
Chase (2)	19/6/74	3	_	_	100	Kasson (28)	6/7	_	1259	ŗ.	16
	25/6/74		,	_	0		//9	~	640	200	2 ~
	2/7/74		_	_	0		1/7/74	0(E)	327	20	ı o
	19/6/74	0	3	3	0		4/6/7	~	348	50	^
	25/6/74		210	20	9		6/7	_	141	500	ıc
	2/7/74		က	က	0		7/2/1	(T)0	527	20	4
Pinora (36)	9/6/15	3	13	13	69		•)	•
	19/6/15		6	თ	44.4	Livingston					
						ಭ	2/9	0	24	24	91.7
Lapeer							6/7	C	19	9	00[
Arcadia (21)	11/6/75	0	12	12	20	•	5/6/7	•	6	6	52.6
	20/6/75		_	_	100		3/6/7		52	52	48.1
(22)	27/6/75	0	7	_	100	Iosco (4)	6/7	0	18	38	94.4
(56)	11/6/75	0	∞	∞	52	(22)	7	0	18	18	100
	20/6/75		∞	∞	52		2/9		25	25	9/
,	27/6/75		16	16	001	(23)	2/9	0	45	45	22.2
Deerfield (23)	9//9//	0	∞	∞	100	,	6/7		127	20	18
	14/6/76		4	4	100		2/9		82	20	8
	21/9/12		37	37	94.6		2/9	0	4	4	20
Mayfield (27)	15/6/76	0	4	4	75		2/9/9	0	33	33	36.4
	22/6/76		7	7	100		2/9	0	45	45	40
Mayfield (31)	10/6/75	3	20	20	20	(27)	6/6/7	0	17	17	100
	20/6/75		က	က	0	Putnam (2)	6/7	0	96	20	44
•						•	2/9/6		1056	20	18
Leelanau							19/		532	20	9
Kasson (21)	14/6/74	3	=	=	0	(11)	6	050	476	20	42
	24/6/74		12	15	0		/		220	20	22
	1/7/74		വ	2	0		16/7	0	44	44	38.6

County/ Town (sec.)	Date	Host	Tot	-Larva Dis	e	County/ Town (sec.)	Date	Host	L Tot	arva Dis	e % Par
Livingston Unadilla (10)	10/6/74	0	154	50	44	Mason Riverton (3)	7/9/6	0	501 228	50	00
(16)	24/6/74 9/6/75 16/6/75	0 0 0	63 27 47	50 27 47	24 51.9 53.2	(14) Sherman (11)	9/6/7 1/6/7 9/6/7	03	185 12 10	50 12 10	0 20 0
Macomb Unspecified	23/6/75	0 0	78	50	70	Custer (16)	25/6/75 12/6/75 19/6/75 25/6/75	0	33 203 85	33 50 50	0 12.1 2 14
Manistee Arcadia (12)	21/6/74	0	2729	50	00	Mecosta Hinton (2)	3/6/7	0	35	35	51.4
Brown (18) Onekama (23) Bear Lake (18)	10/7/74 21/6/74 21/6/74 17/6/75	000 41	232 415 2149 57	20020	2 2 6 38 6	Mecosta (27)	13/6/75 18/6/75 27/6/75	0 0 0	20 20 27	20 20 27 27	55.6 55.6
	16/6/75		19	20	2	Missaukee Richland (4)	1/6/7	0	က	က	100
Victory (1)	15/6/76	0	7 22	7 22	100	Riverside (16)		0	2 212 166	20 20 20	100 28 18
(15)	29/6//6 20/6/74 28/6/74 10/6/75	0	916 454 22	2022	58 18 0 27,3		7/7 7/9 7/9		L 2 4	L 2 4	0 100
	19/6/75		58 58 58	28	3.6	Monroe Dundee (33)	4/6/75 13/6/75 20/6/75	0	33 9 11	33	24.2 0 45.5

County/ Town (sec.)	Date	Host	L Tot	Larvae Tot Dis	% Par	County Town (sec.)	Date	Host	Tot	-Larvae Dis 7	1e % Par
Munroe						Montcalm					
Milan (4)	5/6/75	0	67	50	0	Evergreen (13)	10/6/75	0	6	6	22.2
	13/6//5		22	22 23 20 20 20 20 20 20 20 20 20 20 20 20 20	/ . / .		17/6/75		86	20	26
C1/ -[[::::::::::0	70/61	c	3 5	3.5	1.00.	•	24/0/12	•	S (300	09
Kalsinville (17)	/9/01)	<u>8</u> 2	<u>∞</u>	28	Fairplains (10)	8/6/76	0	32	32	9.19
	9//9//1		? 0) °	087		16/6//6		2/	27	41./
(12)	6/6/6	C	י ע	י ע	0.77	Montesta (24)	9//9/67	c	40	φ. r	4.05
17)	14/6/	>	300	9 0	16.7	MONICCA IIII (24)	10/6//5	>	9 V 0	200	۵ و ر
	22/6/74		17	17	5.9		24/6/75		23	23	78.3
(34)	/9/6	0	4	4	75	(25)	14/6/74	0	448	20	36
	17/6/76		4	4	90	•	21/6/74		1560	20	14
•	23/6/		50	24	62.5	Pine (2)	9//9/6	0	49	49	81.6
(32)	4/6/	0	5 8	5 8	28.6		16/6/76		9/	9/	63.2
	9		24	24	0		22/6/76		80	80	15
	19/6/15		91	91	18.7						
•											
Montcalm						Montague (5)	18/6/74	0	105	20	2
Bloomer (17)	9/9/8	0	ည	വ	100		22/6/74		378	20	7
			ည	വ	∞	Ravenna (5)	11/6/76	0	112	20	95
Bushnell (18)	9/9/8	0	5 5	5 5	84.6		17/6/76		73	20	72
100/		(- (- ;	り(9//9/67		ဂ	ဂ	08 80
Day (20)	10/6//5	0	97	5	∞		11/6/76		20	20	30
	1//6/75		42	42	59.5		17/6/76		41	41	70.7
	5//9/57	•	\	/	_		29/6/76		15	12	83.3
Douglas (33)	14/6/74	0	201	20	0	(8)	11/6/76	0	38	18	100
	21/6/74		360	20	7		17/6/76		46	46	9.69
	27/6/74		303	20	9		29/6/76	•	4	4	100
	9		20	20	2 5	(16)	13/6/75	o P	2	വ	0
	/9//		742	20	12	(21)	17/6/74	0	405	20	10
	24/6//2		195	20	40		3/7/74		15	15	6.7

County/ Town (sec.)	Date	Host	Tot	Larvae ot Dis	% Par	County/ Town (sec.)	Date	Host	Tot	Larvae Tot Dis	% Par
Muskegon						Очетам					
Sullivan (13)	12/6/75	o	662	20	14	Unspecified	19/6/74	0	6	თ	66.7
							20/6/74		2	10	20
Newago							જે		5 8	5 8	85.7
Ashland (12)	13/6/74	0	96	20			9		24	24	70.8
	21/6/74		304	20	10		9		33	53	62.1
	28/6/74		126	20	4		~		∞	7	100
(41)	10/6/75	0	104	20	24						
•	18/6/75		158	20	4	Osceola					
	25/6/75		27	27	14.8	Hersey (33)	21/6/76	0	98	98	45.3
Dayton (9)	13/6/74	0	38	37	13.5						
	19/6/74		46	46	23.9	Highland (16)	14/6/74	0	207	20	6 2
	25/6/74		5	5	21.6		20/6/74		203	20	88
	14/6/75		167	20	18		3/7/74		4	4	52
	21/6/15		234	20	2		3/6/15		98	20	35
	30/6/15		_	_	0		10/6/75		20	20	78
	9/9/6		52	22	100		19/6/15		226	20	30
	16/6/76		32	32	09	(11)	10/6/76	0	37	37	9.79
	21/6/16		Ξ	Ξ	72.7		16/6/76		30	30	93.3
(22)	13/6/74	0	195	20	56		22/6/76		298	20	89
	20/6/74		518	20	4	Richmond (17)	2/6/15	0	128	20	28
	25/6/74		575	20	9[(36)	13/6/74	0	189	20	50
Grant (12)	9/9/6	0	72	72	92.8		19/6/74		887	20	56
	16/6/76		9	9	20		9/6/15		88	20	6 2
	22/6/76		<u>1</u>	9[56.3		19/6/75		181	20	42
							9//9//		37	37	89.5
Oceana		•					15/6/76		53	53	72.4
Elbridge (11)	9/6/15	<u>-</u> ر	104	20	2		22/6/76		179	20	12
	16/6/75	-0	100	20	0						

County/ Town (sec.)	Date	Host	Tot	-Larvae Dis	e % Par	County/ Town (sec.)	Date	Host	Tot	Larvae Dis %	% Par
Oscoda Comins (17)	23/6/75	0	∞ ~	ω ~	37.5	St. Joseph Florence (10)	3/6/74	0	130	50	4 5
(19)	,	0	<u> </u>	35.5	69.2 75		18/6/74		96	2002	21 8 9 2
(56)	19/6/74 26/6/74	3	2-0	2-0	000	Lockport (26)	3/6/74	3	62 4	62 4	3.2
	5/7/74 23/6/75	0	0 -	0 -	100	Sturgis (6)	18/6/74 8/6/76	0	52	52	48.1
Saginaw Freemont (22)	20/6/74 27/6/74	0	011 101	50	20 30	Sanilac Marlette (33)	14/6/74	0	29	29	34.5
Maple Grove (20)	5/7/74 20/6/74 27/6/74	3	3290	9 9 9	0000	Shiawassee Burns (6)	14/6/74	0	104	50	848
(21)		0	73	20 20 20	30 30		1/7/74		120	2021	8 8 8 8
St. Clair Lynn	7/6/74 13/6/74 20/6/74	១	0 7 0	000	000	Fairfield (10)	3///5 14/6/74 24/6/74 1/7/74 25/6/75	6660	118 118 138 138	50 50 50 50 50	80 40 26 46.5 70
St. Joseph Fawn River (7)	3/6/74	0	.ღ ა	13	38.5	Hazleton (18)	1/7/75 13/6/74 24/6/74	5 53	32 108 110	32 50 50	87.5 22 42
	10/6/74 18/6/74 3/6/74		32 35	35 35 35	70 10 0	Middlebury (26)	1/7/74 24/6/75 1/7/75	<u> </u>	158 18	50 81 72	40 72.2 83.3
	10/6/74 24/6/74	66	27 50	27 50	00	(32)	13/6/74	6 6	103	50 00	28 28 24

County/ Town (sec.)	Date	Host	Tot	Larvae Tot Dis % Pa	% Par	County/ Town (sec.)	Date	Host	Tot	Larvae Tot Dis %	% Par
Shjawassee				Ċ	Ç.	Washtenaw		Ó	Ó	C	¢
Vernon (7)	13/6/74	5	67	20	5 5 7 8 7 8	Dexter (36)	31/5/74	0	2;	2 5	0 8
	25/6/14	o	128	20	30		10/6/74		94	20	30
	1/7/74	o P	110	20	ထ္က		17/6/74		107	20	14
	27/6/15	0	115	20	9/	Webster (31)	31/5/74	3	თ	0	0
	2/1/15		34	34	83.3		10/6/74		35	35	15.6
New Haven (13)	25/6/15	0	92	20	46		17/6/74		4	4	0
	2/1/75		40	40	06		3/6/15		55	20	26
							10/6/75	ι	42	42	3
Tuscola							18/6/75	£.	7	7	0
Arbela (17)	23/6/75	0	26	20	100		3/6/75	0	181	20	99
(18)	21/6/15	0	258	20	50		10/6/75	,	15	15	09
	23/6/15		26	20	36		18/6/75	0	166	20	ω
(27)	23/6/75	0	75	20	100	Sharon (4)	12/6/75	0,	10	10	50
							20/6/75	0	27	20	46
Van buren											
Unspecified	28/5/74	0	0	0	0						
	28/5/74		0	0	0						
	28/5/74	c	0	0	0						
Arlington (23)	10/6/75	0,0	99	20	0						
•	19/6/75	05	5 8	28	14.3						
Hamilton (1)	15/6/75	05	9	9	0						
	21/6/15	70	_	_	00						
Paw Paw (20)	11/6/75	20	92	20	0						
	10/6/75	70	Ç	5	7 7						

Appendix 3: The Non-crop Larval Ecosystem

Introduction

Not all cereal leaf beetles or \underline{T} . \underline{julis} are found in fields of cereal grains. Fence rows, pastures, fallow fields, scrub woods and similar uncultivated areas are also capable of supporting beetles. This appendix examines some of the factors that affect the number of CLB's and parasitoids that can be found outside of crops. These include oviposition, survival and development rates, and incidence of parasitism.

Literature Review

The literature on the relationship between the cereal leaf beetle and various hosts is extensive. Hodson (1929) assigned a descending order of feeding preference to barley, oats and wheat, respectively. He also noted that orchard grass (<u>Dactylis glomerata</u>) became a staple food after cereal grains had been harvested. Venturi (1942) claimed that the CLB was polyphagous within the Gramineae. Balachowsky (1963) found the CLB in Europe would feed on quack, timothy, rye, orchard, oats and canary grasses, among others.

Wilson and Shade (1964a) described the feeding and oviposition preferences on several grasses and related these to the survival rates of the larvae. They found oats to be preferred over barley and wheat of the same age for oviposition, and the latter 2 were preferred over corn. The age of the plant was a critical factor, however, and wheat or barley were more preferable to oats that was 10 days older. Wilson and Shade (1964b) also studied differences in post-diapause adult weight gain and survival on different grasses. Any small grain was accepted before sorghum or corn. Foxtails (<u>Setaria</u> sp.) were antibiotic to CLB's. They hypothesized that <u>D</u>. <u>glomerata</u> was used as a spring adult food if small grains were unavailable.

Larval survival and development rate also varied according to host (Wilson and Shade 1965). The small grains plus orchard, timothy, brome, quack and rye grasses all had larval laboratory survival rates above 50%. Corn, which was an acceptable food for prediapause adults, was less suitable for larvae.

Leaf-vein spacing was one factor that affects larval survival (Wilson and Shade 1966). The mouthparts of first-instar larvae were too large for insertion between veins, and the veins themselves were too tough to chew through. Leaf pubescence was also a deterrent to oviposition and feeding (Schillinger and Gallun 1968, Wellso 1973, Webster, et al. 1973).

A general theory of host suitability was developed by Caswell, Reed, Stephenson and Werner (1973). The plants described above as unfavorable or antibiotic to the CLB were found to have a common metabolic pathway, designated C-4. Plants suitable for CLB's had a different pathway, labeled C-3. There was a good correlation between metabolic pathways and leaf-vein spacing.

There are at least 3 reasons to expect to find cereal leaf beetles in wild grasses. Adult beetles may be drawn to non-crops when crop age (Wilson and Shade 1964a). Feeding damage to crops may reduce moisture content and succulence, prematurely aging the crop and causing adults to look for new hosts. Random flight activity may carry adults out of the

field, perchance to an acceptable grass host. Surveys of adult beetles at the Kellogg Farm during 1973-1974 (Casagrande 1975) showed that during the spring and early summer grasses in fence rows, pastures, and stubble fields held a sizeable reservoir of adults.

Methods

1. Field studies: Three types of experiments were carried on in the field during the summer of 1973 using the MSU Kellogg Farm as a research area. In one set, large (18 x 6 x 6 cu. ft.) cages were set up half over grass and half over adjoining winter wheat, and others for grass and spring wheat, and grass and oats (as in Figs. 3-1). The cages on wheat were moved later, and the experiment was repeated to look for changes due to the age of the wheat crops. The cages on oats were set up for only one period. For each experimental treatment, 2 cages were used at a time. About 250 adult CLB's were introduced to each cage, and 100 were added 2 days later to make sure there would be a large number of eggs laid. After 1 week, the crop and the grasses were sampled to determine egg counts, densities of various grasses, and distribution of eggs within the cages.

A second field experiment involved taking a sweepnet catch of larvae in both an oat field and an adjoining grass field. Fifty sweeps were taken in oats and 100 sweeps were taken in the grasses, where the density was somewhat lower. This was done as a check on the distributions found in the cage studies.

Finally, beetles that had been reared in the laboratory, and which could not have been previously exposed to parasitoids, were set out on pots in the same oat field and at varying distances into the grass field.

This was to see whether parasitoids could find the larvae in the grasses or whether the parasitoids would avoid this area. After 2 days, the larvae were brought back into the lab and were dissected for parasitoids.

2. <u>Laboratory studies</u>: Four different series of experiments were conducted in the lab. All 4 series were done in the CLB rearing room in the MSU Plant Science greenhouse. The temperature throughout the experiments was a constant 24°C with 60-80% RH and a 16 hour daily photophase. Seeds for 4 grasses were collected near the Kellogg Farm. These included orchard grass (<u>Dactylus glomerata</u>), 2 types of brom (<u>Bromus inermis</u> and <u>B. tectorum</u>) and barley. Timothy (<u>Phleum pratens</u>) and quackgrass (<u>Agropyron repens</u>) seeds failed to germinate, even when held for 3 months in a refrigerator. It was suggested later that a better way to get quackgrass would be to dig up roots in the autumn. When used in these experiments, all grasses were 4-6' tall.

A test was made to check for oviposition rates on each grass by itself. A minimum of 24 pairs of adult CLB's were set up on individual caged pots for each of the 3 grasses and for barley. The cages were made of "lumite" screening, which fit snugly into a 2-inch flower pot. The cages were checked daily for eggs, the numbers were recorded, and the fresh eggs were removed. Plants were replaced every 3 days. Counts were made daily for 3 weeks.

A 2nd series of experiments was conducted on oviposition preference. For this test, five 2 x 2 x 2 cu. ft. cages were used. In one set of experiments, the 5 cages each held barley, orchard grass and \underline{B} . $\underline{tectorum}$ ("downy brome"). A second set also included \underline{B} . $\underline{inermis}$ ("smooth brome") and the timothy that would germinate. A third set added "smooth brome," but had no timothy.

In each cage, 10 pairs of sexually mature adults were established.

After 2 days, the plants were removed and counted for eggs.

A 3rd test was run to check egg-laying preference on unequal amounts of orchard grass, "red brome" and barley. Five 2 x 2 x 2 cu. ft. cages were set up with a total of 6 potted plants per cage. The eggs laid were counted at the end of a 3-day period.

A final test was undertaken to study the survival and development rates of larvae on the 4 grasses. A minimum of 30 individually caged pots (like those of the 1st series of lab experiments) were set up for each of the grasses. 2 eggs were placed on the upper side of the lower leaves of the plant. Twice daily checks were made to note the larval instar and when pupation occurred. Survivorship was also noted.

Results

1. Field studies: Cage studies to evaluate the relative oviposition in grasses as crops are extremely susceptible to artifacts. That is, the presence of the cage alters adult behavior. Fig. A3-1 shows the results in 1 of the cages. The beetles tend to pile up along the edges of the cage, especially in corners. Square foot foliage samples taken 1 foot away from the edge have 1/3 the density of eggs as samples taken against the cage wall. In analyzing egg counts, a paired-t test was used, grouping square-foot counts from opposite sides of the cage (eg. 45-20 in Fig. A3-1). Results of the cage counts are given in Table A3-1, with mean egg density for both sides of the cage. In addition to the counts taken in the 4 early season wheat/grasses cages, 3 square-foot foliage samples were taken from each cage. Grasses were divided into 3 distinguishable types: bluegrass (Poa canadense), quackgrass and a conglomerate "wild oats," which later proved to be a mixture of timothy, oats (Avena sp.) and

3	Σ + υ	u			
	Wheat			Grass	
20	12	6	12	7	13
30	_	8	7	ω	27
<u>142</u> 45	10	21	16	59	20

Figure A3-1. Square foot counts of eggs in a cage study.

Distribution of eggs per square foot in cage studies, "n" = number of square feet Table A3-1.

sampled on each side of the cage.

Eggs/Sq. Ft. Mean±s	-n- Crop Grass Difference t	9 $20.00+10.6$ $18.00+9.0$ +2.0 .30 9 $7.67+6.4$ $13.89+12.4$ -6.22 2.82**	9 $19.10+9.24$ $14.89+3.02$ $+4.2$ 1.50 9 $20.67+17.36$ $15.44+8.32$ $+5.23$.94	10 3.40±2.72 1.90±1.6 +1.5 1.50	10 3.40±3.6 1.10±1.1 +2.3 2.26*	10 14.30+7.4 2.00+1.56 +12.3 5.00**
Egg A	-n- Crop	$\begin{array}{ccc} 9 & 20.00+10.6 \\ 7.67+6.4 \end{array}$	$\begin{array}{ccc} 9 & 19.10+9.24 \\ 9 & 20.67+17.3 \end{array}$	3.40+2.72	3.40+3.6	10 14.30+7.4
	Crop	Winter Wheat	Spring Wheat	Winter Wheat	Spring Wheat	Oats
i	Time of Season	Early-10" to 15"	crop neignt	Late-wheat in "boot"	stage	

downy brome, with others possible. The number of stems in each set of 3 sq. ft. and the total eggs on those stems are given in Table A3-2. The "oats" made up only 6 percent of the stems, but contained 41 percent of the eggs. Forty-eight percent of the stems were bluegrass, but these contained only 12 percent of the eggs.

A separate count a week later of eggs on 30 stems gathered in each of 3 cages had a mean eggs per stem of .823, 1.80, and 0.07 for quack, "wild oats" and bluegrass, respectively.

A later repetition of the 30 stem egg count was made in 4 cages, just prior to the "boot" stage in the wheat. By this time timothy had been separated from the other grasses. Egg density was quite low. The average eggs per stem on wheat, quack, timothy and brome (Bromus sp.) grasses was .067, .017, .167, and 0.0, respectively.

Sweepnet catches in an oat field and an adjoining grass field (an alfalfa field over-run by quackgrass), taken at the Kellogg Farm in 1973 (June 13), reflect the same relative preference for oats over grasses. This is summarized in Table A3-3, which also lists results of dissections to estimate parasitism.

There were 7 cases of superparasitism in the 332 larvae dissected for Table A3-3, all of them in the grasses. No hypothesis to explain this relatively high level can be offered at this time. It is known that adult Disparsis n.sp. feeds frequently on a variety of common hosts, including daisy fleabane (Erigeron annus), yarrow (achillea millefolium), chickweed (Stellaria media), and mustards (Lepidium sp., Barbarea vulgaris, and others), (D.C.. Miller, pers. comm.). Although these may have been attractants to the grass field, the superparasitism in the face of so many unparasitized larvae remains unexplained.

Table A3-2. Relative frequencies of grass types and eggs laid on them in 4 cage studies (early season - paired with wheat). "Oats" include wild oats, downy brome, and timothy.

Cage	Grass	Stems in 3 Sq. Ft.	Relative Frequency(%)	Eggs in 3 Sq. Ft.	Relative Frequency(%)
1	Quack	497	91	100	77
	"0ats"	42	8	30	23
	Other	7	1	0	
2	Quack	108	23	17	41
	Blue	276	67	6	15
	"Oats"	27	7	18	44
3	Quack	48	11	11	10
	Blue	343	75	13	12
	"Oats"	47	10	86	78
	Clover	19	4	0	0
4	Quack	161	39	22	52
•	Blue	253	61	20	48

Results of sweep catches in oats and an adjoining grass (+ alfalfa) field, Kellogg Farm, June 13, 1973. Table A3-3.

L. curtus	-	5 **					
% Parasitism Diaparsis Spp.	ო	10.9	11.7*	4	10.9**	14.7**	7.9**
T. julis	36	14.5**	23.5**	28	23.9**	26.5**	34.2**
Number Dissected	100	55	34	52	46	34	38
Larvae* Sweep	3.22	.55	.34	.25	.46	.34	.38
Distance from Host Oat Field (Ft)	ı	∞	20	30	40	50	09
Host	Oats	Grass					

* 150 sweeps in oats; 100 in grass.

^{**} Results include incidences of multiple parasitism.

Dissection of larvae that were set out on pots showed essentially the same results as Table A3-3. At a distance of 10 feet into the field, there were no larvae parasitized. Perhaps \underline{T} . \underline{julis} is able to respond to the presence of oats at that distance so that it spends less time in the grass searching for hosts.

2. <u>Laboratory studies</u>: When caged on a single host species, ovipositing CLB females laid more eggs on barley than on orchard grass or brome grasses (Fig. A3-2). There was considerable variation between individual oviposition rates on any host. There was no apparent difference in survival of adults on the 4 hosts over the 2-week period (Fig. A3-3).

When given a choice of plants, barley is clearly preferred for oviposition, over orchard grass, smooth or downy brome, and timothy (Table A3-4a). This is true even when given a surplus of the less favored host (Table A3-4b).

Development time per instar is given in Table A3-5. Larval mortality is not listed in the table because it is not certain which larvae died and which were able to escape the cage. The numbers of larvae in the table reflect lost larvae as well as those found dead.

<u>Discussion</u>

Native grasses are capable of sustaining cereal leaf beetle adults and larvae in different degrees. Some are better as oviposition sites than others. Bluegrass, for example, does not seem to serve well as a host for egg laying.

The role of wild grasses in maintaining a non-crop population of the cereal leaf beetle is a real one. The grasses serve as food during times when the crop is unpalatable. During this time, eggs are laid on the grasses. The grasses are then capable of supporting larval development. The numbers

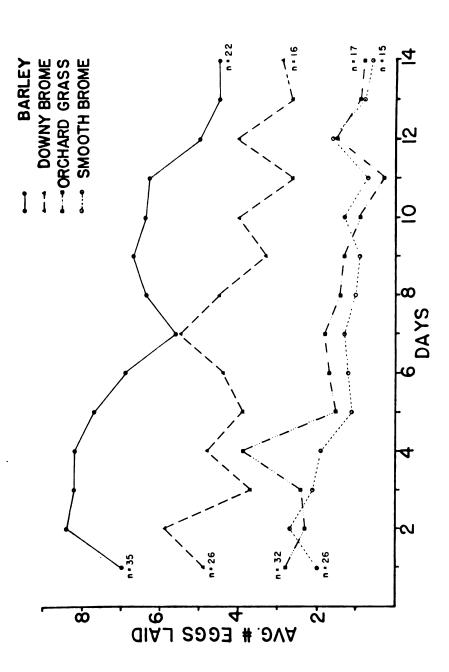
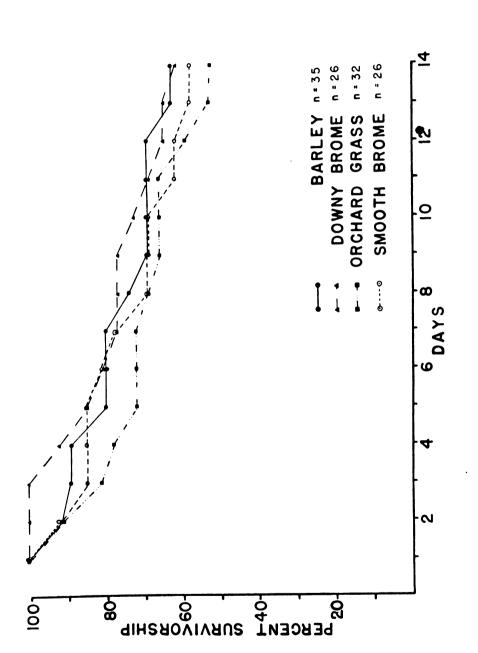


Fig. A3-2. Averal cereal leaf beetle oviposition on 4 hosts. Numbers are the number of ovipositing females at the beginning and end of the experiment. Conditions: temperature, 24-26°C; R.H., 60-80%; daily light, 16 hours.



Relative adult mortality on 4 hosts. n=number at start of experiment. Laboratory conditions: temperature, 24-26°C; R.H., 60-80%; daily light, 16 hours. Fig. A3-3.

Table A3-4a. Relative frequencies (% of total) for oviposition when presented with equal numbers of a selection of hosts.

n = total number of eggs.

Test Series	Barley	Orchard	Smooth Brome	Downy Brome	Timothy	-n-
la	44	33	n.a.	22	n.a.	306
b	71	17	n.a.	12	n.a.	579
С	80	8	n.a.	12	n.a.	696
d	57	24	n.a.	18	n.a.	148
2a	51	18	16	8	8	555
b	47	12	15	11	14	307
3	45	19	22	14	n.a.	189

Table A3-4b. Oviposition preferences when given different amounts of each host. No repetitions were made.

		Ratio of Plants				Ratio of eggs
Test	Barley	:	Orchard	:	Downy Brome	
1	5	:	1	:		6.0 : 1
2	1	:	5	:		2.8 : 1
3	3	:	3	:		1.7 : 1
4	3	:		:	3	2.8:1
5		:	3	:	3	0.3:1

Development times per instar on four hosts. Laboratory condition: temperature, 24-26°C; humidity, 60-80%; daily photophase, 16 hours. Table A3-5.

4	$-n y$ s_y $-n y$ s_y Total		15 3.0 .9 7 3.6 .6	2.3 .5 20 2.2 .3 17 3.4 .9 8 4.2 .5	2.2 .6 25 2.8 1.0 15 2.8 .6 8 4.5 .9
1	S,	<u></u>	٠.	-:	
4	>	<u>س</u>	3.6	4.2	4
! ! !		7	7	8	80
car	s ^{>}	9.	6.	6.	9.
3	>	5.0	3.0	3.4	2.8
2	Ļ	22		17	15
	s _y	9.	2.5 .7 23 2.8 .9	۴.	1.0
<u>ا</u> ح	>	2.8	2.8	2.2	2.8
	<u>-</u>	24	23	20	25
	s [×]	∞.		3.	9.
_	y s y -n- y s	2.8	2.5	2.3	2.2
	- 4	56	25	27	27
	Host	Barley	Smooth brome	Downy brome	Orchard grass

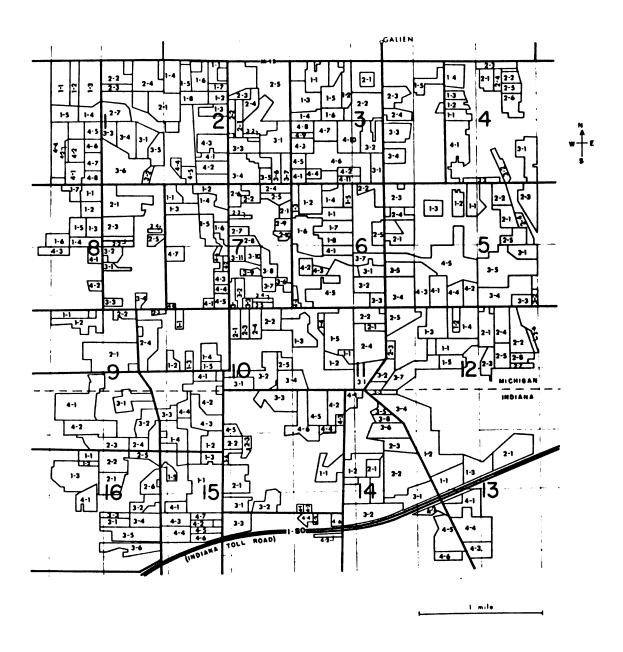
caught in sweepnet indicate a potentially large number of beetles could be found on other than crop hosts.

Non-crop populations of cereal leaf beetle larvae are in turn capable of supporting larval parasitoid populations. The level of these populations will depend on the suitability of the grasses present and the availability of parasitoid adult food sources. The field studied at the Kellogg Farm could have served well as a reservoir of beetles and parasitoids, acting as a buffer to the effects of insecticide usage in the host crop.

A more complete study of the wild grasses would require an estimate of their relative abundance over large areas. Also, the effect of age and the synchronies of grass types should be evaluated further. As least, this study indicates that significant numbers of beetles and parasitoids may reside outside of host crops.

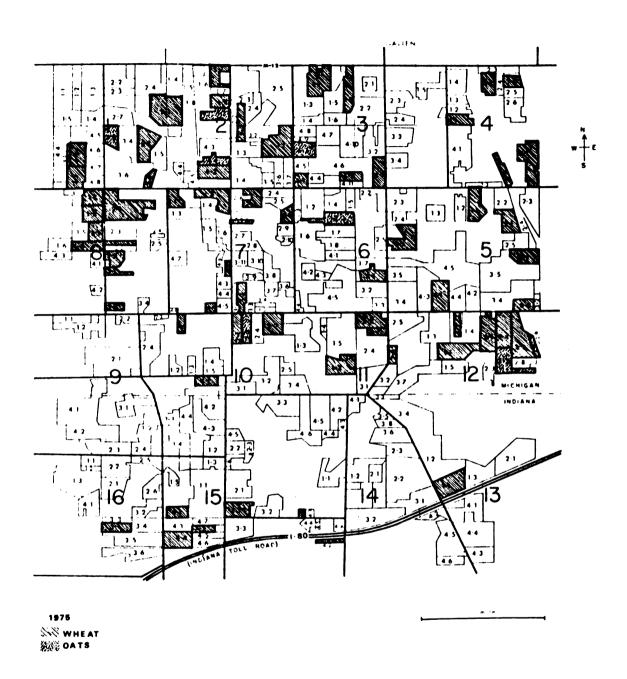
Appendix 4. 1975 PWS Field Maps

- A4-1 Field identification codes (see Appendix 5).
- A4-2 1975 PWS Oat and Wheat fields.
- A4-3 Supplementary oat fields used in 1975 PWS study (located approximately 1 mile due east of main study area).

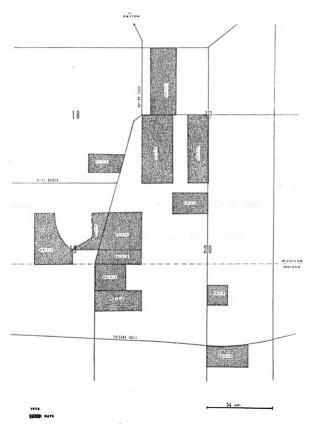


A4-1. Field identification codes (see Appendix 5).

1



A4-2. 1975 PWS Oat and Wheat fields.



A4-3. Supplementary oat fields used in 1975 PWS study.

Appendix 5. Identification of 1975 PWS Cultivated Fields (Map, Appendix 4) by Crop and Acreage.

(Survey taken 28 July through 1 August, 1975).

Field Identification

Field #	<u>Crop</u>	Acres	Field #	Crop	Acres
1-1-1	Corn	37	2-2-3	Alfalfa	9
1-1-2	Soybean	20	2-2-4	Corn	29.5
1-1-3	Corn	45	2-2-5	Hay	79
1-1-4	Corn	15.5	2-3-1	Wheat	35
1-1-5	Corn	19	2-3-2	Soybean	3.5
1-2-1	Wheat	40.5	2-3-3	Corn	24
1-2-2	Soybean	26.5	2-3-4	Corn	37.5
1-2-3	Corn	14.5	2-3-5	Hay	7.5
1-2-4	Soybean	46.5	2-3-6	Corn	16
1-3-1	Wheat	23	2-3-7	Hay	14.5
1-3-2	Wheat	6	2-4-1	Oats	
1-3-3	Oat	13.5	2-4-2	Wheat	20
1-3-4	Alfalfa	25.5	3-1-1	Wheat	25.5
1-3-5	Soybean	18.5	3-1-2	Wheat	12
1-3-6	Corn	68.5	3-1-3	Soybean	31
1-4-1	Wheat	20	3-1-4	Hay	10.5
1-4-2	Wheat	19	3-1-5	Corn	22
1-4-3	Oats	5	3-1-6	Soybean	15
1-4-4	Corn	33	3-1-7	Hay	
1-4-5	Pasture	14.5	3-2-1	Wheat	9
1-4-6	Corn	9.5	3-2-2	Corn	40.5
1-4-7	Hay	12	3-2-3	Soybean	24
1-4-8	Corn	11	3-2-4	Hay	12
2-1-1	Wheat	13	3-3-1	Wheat	21
2-1-2	Wheat	7	3-3-2	Wheat	26.5
2-1-3	Oat	9	3-3-3	Corn	16
2-1-4	Plowed	28	3-3-4	Soybean	14.5
2-1-5	Hay	11	3-4-1	Wheat	9.5
2-1-6	Soybean	16	3-4-2	Wheat	6.2
2-1-7	Hay	5.5	3-4-3	Oat	16
2-2-1	Wheat	12	3-4-4	Oat	7.5
2-2-2	Soybean	10.5	3-4-5	May	9.5

Field #	<u>Crop</u>	Acres	<u>Field #</u>	Crop	Acres
3-4-6	Corn	67	5-4-2	Corn	19
3-4-7	Soybean	20.5	5-4-3		34
3-4-8	Corn	9.5	5-4-4	Hay	17
3-4-9	Hay	6	5-4-5	Corn	90
3-4-10	Corn	37	6-1-1	Oat	21
3-4-11	Hay	4.5	6-1-2	Corn	32
4-1-1	Wheat	14	6-1-3	Hay	10
4-1-2	Corn	12	6-1-4	Pasture	19.5
4-1-3	Soybean	9	6-1-5	Alfalfa	11.5
4-1-4	Corn	21.5	6-1-6	Pasture	46
4-1-5	Soybean	17	6-1-7	Corn	17
4-2-1	Wheat	16	6-2-1	Wheat	26.5
4-2-2	Wheat	11	6-2-2	Corn	12.5
4-2-3	Hay	14	6-2-4	Corn	12
4-2-4	Soybean	8.5	6-2-5	Corn	14.5
4-2-5	Pasture	11.5	6-3-1	Wheat	17.5
4-2-6	0at	19.5	6-3-2	Soybean	21.5
4-3-1	Wheat	32	6-3-3	Hay	12
4-3-2	Wheat	10.5	6-3-4	Corn	37.5
4-3-3	Pasture	8.5	6-3-5	Hay	19.5
4-4-1	Corn	57	6-3-6	Corn	
5-1-1	Soybean	22	6-3-7	Plowed	18.5
5-1-2	Pasture	14	6-4-1	Hay.	12
5-1-3	Plowed	14	6-4-3	Hay	5.5
5-2-1	Wheat	18.5	6-4-4	Hay	7
5-2-2	Corn	18	6-4-5	Pasture	77
5-2-3	Pasture	30	7-1-1	Wheat	12
5-2-4	Pasture	10	7-1-2	Wheat	21
5-2-5	Corn	15	7-1-3	Soybean	17
5-3-1	Barley	18	7-1-4	Pasture	25
5-3-2	Wheat	9	7-1-5	Wheat	22
5-3-3	Wheat	10	7-1-6	Wheat	9.5
5-3-4	Alfalfa	49	7-2-1	Wheat	11
5-3-5	Corn	74	7-2-2	Wheat	15
5-4-1	Wheat	25	7-2-3	0at	5

Field #	Crop	Acres	<u>Field #</u>	Crop	Acres
7-2-4	Corn	13	8-2-1	Wheat	49
7-2-5	Hay	11.5	8-2-2	Wheat	5.5
7-2-6	Plowed	15	8-2-3	Corn	29.5
7-2-7	0at	10	8-2-4	Wheat	4.5
7-2-8	Hay	15	8-2-5	Soybean	7
7-2-9	Soybean	8	8-3-1	Wheat	7
7-2-10	Corn	5	8-3-2	Corn	13
7-3-1	Rye	8	8-3-3	Wheat	8
7-3-2	Wheat	9	8-3-4	Hay	10
7-3-3	Corn	7	8-4-1	Wheat	9
7-3-4	Hay	5.5	8-4-2	Plowed	8
7-3-5	Corn	6	8-4-3		11
7-3-6	Hay	7	9-1-1	Wheat	16
7-3-7	Hay	9	9-1-2	Corn	27
7-3-8	Hay	16	9-2-1	Soybean	73
7-3-9	Hay	7	9-2-2	Corn	9
7-3-10	Hay	12	9-2-3	Hay	4
7-3-11	Hay	10	9-2-4	Plowed	24.5
7-4-1	Wheat	4	9-3-1	Plowed	28
7-4-2	Wheat	5	9-3-2	Plowed	19
7-4-3	Soybean	10.5	9-3-3	Corn	17
7-4-4	Plowed	6	9-4-1	Wheat	56
7-4-5	Corn	10	9-4-2	Corn	44
7-4-6	Hay	9		•	
7-4-7	Corn	11.5	10-1-1	Wheat 	9
7-4-8	Alfalfa	2.5	10-1-2	Hay	12
8-1-1	Wheat	14	10-1-3	Plowed	6
8-1-2	Wheat	15	10-1-4	Wheat	16
8-1-3	Wheat	12.5	10-1-5	Corn	9
8-1-4	Wheat	15	10-2-1	Wheat	16
8-1-5	Corn	14.5	10-2-2	Wheat	19
8-1-6	Corn	14	10-2-3	0at	12
8-1-7	Corn	10.5	10-2-4	Corn	18
			10-2-5	Soybean	9.5

Field #	Crop	Acres	<u>Field #</u>	Crop	Acres
10-3-1	Hay	19	12-1-1	Wheat	27
10-3-2	Corn	40	12-1-2	Wheat	7
10-3-3	Corn	36	12-1-3	Hay	33
10-3-4	Hay	25	12-1-4	Corn	35
10-4-1	Wheat	9	12-1-5		18
10-4-2	Corn	58.5	12-2-1	Wheat	27
10-4-3	Hay	32	12-2-3	Corn	13
10-4-4	Plowed	11.5	12-2-4	Soybean	27
10-4-5	Hay	7	12-2-5	Oat	17
11-1-1	Wheat	7	12-2-6	Oat	6.5
11-1-2	Wheat	20	12-2-7	Oat	7
11-1-3	Hay	33	12-2-8	Soybean	10
11-1-4	Hay	6	13-1-1	Wheat	28
11-1-5	Corn	56	13-1-2	Pasture	60
11-2-1	Wheat	9	13-1-3	Hay	26
11-2-2	Plowed	23	13-2-1	Corn	44.5
11-2-3	Wheat	9	13-4-1	Wheat	33
11-2-4	Soybean	50	13-4-2	Wheat	11
11-2-5	Pasture	29	13-4-3	Wheat	20
11-2-6	Hay		13-4-4	Corn	46
11-3-1	Corn	25	13-4-5	Corn	37
11-3-2	Corn	18	13-4-6	Soybean	12
11-3-3	Hay	6	14-1-1	Rye ·	23.5
11-3-4	Pasture	58	14-1-2	Hay	30
11-3-5	Hay	8	14-2-1	Corn	14
11-3-6	Corn	25	14-2-2	Corn	70
11-3-7	Plowed	37	14-2-3	Hay	23
11-3-8		13.5	14-3-1	Pasture	29
11-4-1	Hay	12	14-3-2	Corn	47
11-4-2	Hay	28	14-4-1	Wheat	2
11-4-3	Corn	5	14-4-2	Wheat	6
11-4-4	Hay	7	14-4-3	Pasture	2.5
11-4-5		33	14-4-4	Pasture	9
11-4-6	Corn	37.5	14-4-5	Oat	4
			. , , ,		•

Field #	Crop	Acres	Field #	Crop	Acres
14-4-6	Pasture	7.5	16-3-4	Alfalfa	22
15-1-1	Corn	78	16-3-5	Alfalfa	37
15-1-2	Hay	15	16-3-6	Corn	28
15-1-3	Corn	9	16-4-1	Corn	23
15-1-4	Corn	43.5			
15-1-5	Pasture	9			
15-1-6	Plowed	9.5			
15-2-1	Corn	28			
15-2-2	Hay	12			
15-2-3		9			
15-3-1	Wheat	20			
15-3-2	Pasture	20			
15-3-3	Pasture	25			
15-4-1	Wheat	16			
15-4-2	Wheat	8			
15-4-3	Alfalfa	19			
15-4-4	Wheat	25			
15-4-5	Corn	11			
15-4-6	Soybean	9.5			
15-4-7	Soybean	12			
16-1-1	Oat	14.5			
16-1-2	Corn	7			
16-1-3	Alfalfa			•	
16-2-1	Wheat	31			
16-2-2	Hay	34			
16-2-3	Wheat	32			
16-2-4	Corn	14.5			
16-2-5	Soybean	17			
16-2-6	Corn	22			
16-3-1	Wheat	15			
16-3-2	Wheat	17			
16-3-3	Corn	7			

Appendix 6. Pubescent wheat study total egg, larval, and adult cereal leaf beetle count data.

Appendix 6a. PWS egg count data.

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DD = degree days base 48°F
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EP = number of eggs in ED parasitized by <u>Anaphes flavipes</u>

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Appendix 6b. PWS larval count data.

DD = degree days base 48°F

CH = crop height (inches)

M = field wetness (1=dry; 2=damp; 3=wet)

WI = wind speed (mph)

UNIT = either linear feet (SQFT) or sweeps

NSU = number of UNITS sampled

TOTC = total count of larvae in NSU UNITS

1...4 = numbers of larvae in a subsample grouped by instar.
First number of pair is the number of that instar in the subsample. Second number of pair is the number that were parasitized by <u>Tetrastichus julis</u>.

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Appendix 6c. PWS adult count data.

DD = degree days base 48°F

CH = crop height (inches)

M = field wetness (l=dry; 2=damp; 3=wet)

WI = wind speed (mph)

SWP = number of sweeps

ASWP = number of adults in SWP

SQ = number of <u>linear feet</u>

ASQ = number of adults in SQ

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Appendix 7. Pubescent wheat study individual egg, larval, and adult count data.

Appendix 7a. Adult count data. There are 6 sample repititions for sweepnet and 6-10 for linear foot counts. A "30-0" in a linear foot count is a combination of 3, 10 foot samples with no insects found.

DD = degree days base 48°F

CH = crop height (inches)

M = field wetness (l=dry; 2=damp; 3=wet)

WI = wind speed (mph)

SQ = number of linear feet

SW = number of sweeps

KNT = count per SQ or SW

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SW-KNT S	25-039 2 25-027 2 25-036 2 25-016 2	25-004 25-019 25-078 25-088 25-088 25-037	25-023 25-027 25-098 25-098 25-073	25-007 25-041 25-002	25-024 25-027 25-007 25-021	25-018 25-025 25-007	25-012 25-004 25-006	25-013 25-003 25-032 25-014
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SW-KNT	25-159 25-072 25-083 25-032	25-037 25-029 25-143 25-146 25-098 25-098	25-005 25-010 25-210 25-217 25-092	25-043 25-023 25-074	25-044 25-011 25-076 25-042	25-015 25-031 25-011	25-011 25-028 25-025	25-025 25-035 25-027 25-105
SW-KNT	25-047 25-106 25-047 25-047	25-029 25-010 25-178 25-161 25-087	2 25-014 25-026 25-056 25-124 25-079	25-037 25-032 25-034	2 25-038 25-025 25-054 25-019	3 25-056 25-012 25-018	25-015 25-008 25-027	25-010 25-015 25-018 25-031
SW-KNT	.D: #202 25-015 25-034 25-031 25-115	25-007 25-007 25-004 25-356 25-022 25-022 25-015	25-012 25-012 25-031 25-180 25-047 25-121 25-141	LD: 5201 25-036 25-017 25-051	LD: 530 25-018 25-046 25-047 25-030	25-022 25-022 25-018 25-028	ELD: 540 17 25-006 15 25-005 19 25-005	1D: 620 25-003 25-002 25-002 25-016
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DD	210	231 290	170	290 154 225	290						
T SW-KNT	4 25-009		7 25-047 6 25-016 2 25-008	1 25-002 2 25-000 0 25-000	0 25-000 1 25-001 0 25-000 7 25-013	4 25-014 2 25-004 0 25-009 2 25-003	2 25-000 0 25-000 3 25-003	9 25-003 7 25-009 9 25-006	8 25-004 4 25-006 2 25-005	6 25-017 1 25-045 1 25-041 9 25-084	1 25-019
SW-KNT	25-004	25-02	25-027 25-026 25-002	25-001 25-002 25-000	25-000 25-001 25-000 25-000	25-004 25-002 25-000 25-002	25-002 25-000 25-003	25-009 25-027 25-009	25-028 25-014 25-002	25-006 25-031 25-031 25-099	25-041
SW-KNT	25-001		25-020 25-014 25-012	25-000 25-003 25-001	25-000 25-001 25-000 25-004	25-008 25-008 25-001 25-024	25-001 25-001 25-004	25-010 25-112 25-008	25-016 25-001 25-002	25-010 25-068 25-012 25-032	25-017
SW-KNT	25-002	25-013	25-045 25-022 25-020	25-002 25-003 25-000	25-000 25-001 25-001 25-003	25-010 25-016 25-015 25-017	25-001 25-003 25-004	25-029 25-014 10-029	25-021 25-001 25-001	25-020 25-035 25-011 25-052	25-016
SW-KNT	1 25-002	~ ?;	25-022 25-016 25-024	25-000 25-000 25-000	25-003 25-000 25-001 25-001	25-000 25-005 25-006 25-006 25-010	25-001 25-001 25-001 25-001	25-022 25-022 10-019	25-060 25-002 25-005	25-017 25-029 25-013 25-013	.D:15404 25-009 25-006
SW-KNT	IELD: 1220 12 25-001	LD:1220 25-008	25-007 25-001 25-019	TELD: 12204 04 25-000 08 25-001 09 25-002	ELD: 1310 0 25-000 0 25-001 6 25-001 2 25-010	FIELD: 14401 1 05 25-008 2 07 25-017 1 06 25-026 1 08 25-024	00 00 00 00 00	ELD: 1530 0 25-035 5 25-016 3 10-020	LD: 15401 25-031 25-001 25-003	LD: 1540 25-021 25-004 25-008 25-008	FIELD:15404 07 25-009
X	FIE 1 12	E O	3 09 1 08 1 0	FIE 2 04 3 08 1 09	FIEI 1 10 2 06 1 12	FIE 1 05 2 07 1 06 1 08	FIE 2 07 1 08 1 10	FIE 1 10 1 13	FIELD: 1 03 25 1 04 25 1 05 25	FIE 1 10 1 08 1 07	FIE 1 07
픙	20 1		127	£ 1 4	0 ± 6 ±	02 1 2 8 2 2 2 2 3 2 3 2 3 2 3 2 3 2 3 2 3 2	585	08	157	7112	12
QQ	290	154	201 225 290	154 201 225	154 201 225 290	154 210 225 290	210 225 290	154 210 290	170 210 290	154 210 231 290	154
				-			- •		- •		

Appendix 7b. Egg and larval count data. Each number is the number of insects found in 2 linear feet of row (= 1 square foot).

FIELD	/CH	-					- EG	igs /	SQ.	FT.						
DEGRE	E DA	YS:	621		DATE	։ յ	UNE	5								
7281 1303 1403 7203 7302 7401 7402	25 11 4 16 28 24 32	0 19 5 25 0 0	0 11 3 16 0 5	9 24 9 5 0 1	2 9 6 13 0 0	2 16 5 12 3 2 0	5 16 2 8 1 3 0	1 7 3 9 0 1 0	3 12 8 9 0 0	1 27 2 18 0 0	4 9 3 11 2 4	4 14 2 8 0 2 0	3 8 4 4 1 0	3 11 7 10 0 0 2	1 21 5 11 0 0	2 15 3 4 0 0 2
DEGRE	E DAY	YS:	725		DATE	: J	UNE	12								
1303 1403 2103 2401 3403 3403 6101 7203 10203 91101 91102 91103 91104 92101 94101 95101 95102 95103	7 6 8 7 6 10 20 12 30 27 33 26 33 15 22 22 25	8 9 4 8 0 7 0 0 0 1 0 1 0 2 1 2 2	5 1 11 17 5 2 0 0 0 0 0 0 0 2 3 0 1 1	10 3 11 17 13 3 0 0 2 0 0 0 0 0 3 5 0	16 3 3 21 30 2 2 0 0 1 1 0 1 2 2 1 2 2	7 0 22 19 12 2 1 0 1 1 1 0 2 1 2 0 2	10 1 10 0 9 7 1 0 0 0 0 1 3 2 1 3 1	17 1 13 0 28 5 1 3 0 3 1 0 1 4 0 3 1 3	5 0 7 1 20 3 1 5 0 0 4 1 0 2 3 0 4 2 0 4 2	4 2 6 2 2 4 1 2 6 0 1 2 0 0 2 4 1 0 0 0	2 1 11 3 5 1 2 4 0 2 3 0 3 5 2 1 0 0	0 1 5 0 16 6 0 4 3	0 3 2 0 11 7 2 5 0	0 2 5 2 2 3 4 1 0	2 5 2 0 6 1 0 10 0 4 3	3 6 2 6 2 7 2 14 2
DEGRE	E DAY	/ S:	804		DATE	: J	UNE	16								
1403 2103 2401 3402 6101 7203 10203 91101 91102 91103 91104 92101 92102	13 14 14 10 15 23 15 27 29 30 28 27 27	1 0 15 2 1 0 0 1 1 0 3 0	3 4 1 1 11 0 8 1 0 1	1 0 3 1 15 0 5 2 0 2 6 1	2 9 70 6 2 5 0 1 0 0 2 4 0	2 4 1 5 7 8 0 4 0 0 0 5 0	0 16 0 4 8 5 0 4 1 0 2 5 0	0 4 4 5 4 8 0 2 0 0 7 3	2 10 0 5 5 7 0 2 1 1 1 5 7	1 9 2 6 6 1 3 2 1 0 2 4 2	2 21 4 2 5 8 0 3 6 0 0	2 21 1 4 2 6 0	4 18 0 2 4 3 0	1 29 6 7 6 2 0	4 5 1 4 3 0 0	1 25 7 2 1 2 1

DEGRE	E DAYS	S: 8	304	ſ	DATE	: J	UNE 1	16 (CONT	.)						
94101 95101 95102 95103 1303	23 30 30 25 11	2 0 0 0 5	2 1 1 0 9	0 2 0 0 4	2 0 0 1 6	1 0 0 10	0 0 0 0 5	3 0 0 0 7	2 0 2 1 1	0 1 1 2 0	0 0 0 0	1 1 0 0 2	1 0 0 1 3	0 0 0 0 3	1 4 1 0 1	1 0 2 0 4
DEGRE	E DAYS	S: 8	358	[DATE	: J	UNE 1	8								
1303 1403 2103 2401 3403 6101 7203 10203 12202 12206 12301 91101 91103 91104 92101 92102 94101	16 11 12 13 18 19 11 24 28 27 27 27 27 27	4 1 4 1 0 0 3 1 0 0 0 2 0 0 2 0 0	2 0 3 0 1 2 4 1 0 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0	2 0 5 0 0 3 3 0 0 0 0 3 1 0	4 0 17 1 2 1 4 0 0 0 0 1 0 0 2 1 0	2 0 12 2 0 2 6 4 0 0 0 1 0 0 2 0	0 0 2 0 0 2 14 0 0 0 2 0 1 2 0	1 1 5 2 0 1 7 2 0 0 0 1 0 0 1 3 0	0 1 3 0 0 3 4 3 0 0 0 3 0 0 1 0 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	0 0 2 0 0 4 9 0 0 0 0 6 0 0 0 2 5	1 2 1 0 1 16 0 0 0 2 0 0 2 0	1 2 4 0 2 1 5 1 0 0	0 0 8 6 1 0 4 0 0 0	6 1 6 3 1 3 4 0 0 0	3 0 4 1 0 1 3 2 0 0	2 0 8 0 1 1 1 2 0 0
95101 95102 95103	30 30 27	0 0 1	0 0 1	0 0 0	1 1 0	0 0 0	0 1 0	0 0 0	0 1 0	1 0 1	0 4 0	0 0 1	0 0 0	1 0 1	0 0 1	3 0 0
DEGRE	E DAY	S: !	924	1	DATE	: J	UNE :	20								
1303 1403 2103 2401 3403 6101 7203 10203 12205 12206 91101 91102 91103 91104 92101 92102 94101 95102 95103	17 13 15 13 13 24 19 14 27 30 27 27 30 27 27 30 27 30 35 35	6 0 9 10 1 4 8 0 0 0 0 0 0 0 0 0 0 0 0	12 1 5 6 7 0 0 0 5 3 1 0 0 0 0	4 0 9 11 12 0 6 0 0 0 7 1 0 0 2 1 0 0 1 6	3 4 7 8 2 10 1 0 2 0 3 0 0 0 1 1 0 2 0 0 0	36 28 17 10 10 40 00 20 02 0	1 0 9 7 4 2 0 0 0 0 5 3 0 0 1 2 0 0	04030130006000010000	0 1 2 4 0 2 0 0 0 0 0 4 3 1 1 3 0 0	1 0 3 4 0 3 5 0 0 0 2 0 2 0 0 1 0	0 0 1 17 0 1 0 0 2 2 0 1 0 3 2 0	0 1 2 0 0 5 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0	0 0 1 0 1 0 0 0 0 0 0 0 0 0 0 0 0 2 0	2 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	1 0 2 2 0 0 0 0 0 0 4 2 0 1 1 1 0 0 0	0 0 1 2 1 0 0 0 0 0 0 0 0 1 2 0 0 1 2 0 0

DEGRE	E DAY	/ S:	1015		DAT	E:	JUNE	23								
1303	22	9	9	4	1	0	1	0	0	0	0	0	0	1	1	2
1403 2103	11 18	4	3 0	1	4	0	2	3	0	3 5	0	5	0	0 0	0	0
2401	14	1	0	0	5	11	2	0	Ó	0	Ó	0	0	0	1	4
3403	14	i	4	3	0	Ö	1	0	0	9	5	5	11	0	3	0
6101	20	ò	i	ĭ	Ŏ	3	i	ĭ	Õ	í	Ö	Ö	i	ĭ	2	Õ
7203	28	4	17	0	ì	6	i	5	1	10	14	ì	Ó	6	Ō	Ō
10203	20	0	0	3	2	1	0	2	0	0.	0	0	0	0	0	0
12301	25	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
91101	33	2	5	2	3	1	3	4	1	3	4	5	4	3	2	4
91102	33	2	3	2]	2	1	0]	0	2	0	0]	0]
91103	30	0	1	0]	0	0	0]	0	0	0	0	0	1	0
91104	33	1	0	1	1	0	0	0	1	0	0	1	1	0	0	1
92101	31	1	5	4	0	2	0	2	2	1	5	3	0	1	1	0
92102	29	1	1	3	0	2	1	2	0	1	1	3	1	1	2	2
94101	38	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
95101	40	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
95102	40	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
95103	35	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

FIELD/	CH -						- LA	RVAE	/ S	Q.FT					
DEGREE	DAYS:	621		DATE	J	UNE	5								
7201 1303 1403 7203 7302 7401 7402	8 2 0 0 2 1 0	24 1 0 0 0 1 1	12 2 0 0 5 2	6 2 0 0 1 0 2	9 3 0 0 4 0	7 0 0 0 3 0	5 1 0 0 4 5	15 2 0 0 2 0	8 4 0 0 5 2 0	29 3 0 0 6 1	3 1 0 0 1 1	12 4 0 0 3 1 2	8 4 0 0 3 1 0	14 0 0 0 1 1	3 2 0 0 2 1 2
DEGREE	DAYS:	725		DATE:	J	UNE	12		·						
1303 1403 2103 2401 3403 3403 6101 7203 10203 91101 91102 91103 91104 92101 92102 94101 95101 95102 95103	5 0 50 28 2 12 4 0 4 2 0 0 1 1 1 5 2 1 3	3 0 26 17 0 1 5 0 8 0 2 0 0 1 3 3 1 0 5	6 4 5 10 8 0 1 0 15 0 0 0 2 0 4 4 3 0 1	3 1 20 3 18 4 0 0 2 0 0 1 5 1 7 4 3 4	21 0 21 8 8 11 3 0 1 1 2 0 3 1 0 5 2 1 2	10 0 26 5 5 0 0 0 2 0 1 2 3 1 6 1 2 1	11 35 44 14 03 04 13 51 36 45	23 0 19 11 5 0 3 0 0 6 4 0 4 6 2 9 3 6 5	14 3 81 0 0 0 1 0 2 1 2 0 3 4 0 4 2 5 1	12 15 44 5 0 0 3 0 12 1 3 0 5 2 2 7 1 0 1	10 0 9 12 0 0 2 0 4	9 0 7 11 0 0 2 0 12	14 0 27 10 10 0 4 1 24	5 2 8 7 1 0 2 0 20	14 3 12 7 0 0 2 0 23
DEGREE	DAYS:	804		DATE:	J	UNE	16					•			
1403 2103 2401 3402 6101 7203 10203 91101 91102 91103 91104 92101 92102	5 53 14 9 5 0 2 3 0 1 0 4	0 33 0 6 7 0 2 0 0 1 1 3 7	6 17 0 7 4 0 2 0 0 1 1 3 2	5 5 8 3 0 0 2 1 0 2 6 5	3 2 15 4 0 0 0 0 0 1 1 0 4	2 1 14 3 1 0 0 0 2 0 8 2	0 0 10 4 0 0 2 1 1 1 3 2 3	2 0 0 1 0 0 0 0 0 4 3	4 6 0 2 2 0 0 0 1 0 1 4 4	6 2 0 4 1 0 0 0 2 0 2 3 5	1 11 14 0 7 0 0	0 7 5 1 2 0 2	0 3 12 0 2 0 0	0 23 5 3 0 0	2 13 0 0 2 0 1
94101 95101 95102 95103 1303	1 0 0 0 2	0 0 0 2 39	0 0 1 1 7	0 1 4 1 11	0 0 0 0 8	0 0 0 1 5	0 1 0 0 8	1 2 1 0 15	1 0 0 0 5	0 1 0 0 10	1 0 0 1 17	3 1 2 2 18	1 2 1 3 9	3 1 2 0 6	4 0 1 1 7

DEGREE DAY	S: 8	58	DATE	: J	UNE	18								
1303 1403 2103 2401 3403 6101 7203 10203 12202 12206 12301 91101 91103 91104 92101 92102 94101 95101 95102 95103	1	18 4 2 17 1 4 0 3 0 2 1 1 0 0 2 0 0 0 0 0	0 1 9 3 5 10	9 0 7 9 2 3 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	21 2 13 9 0 5 0 3 0 1 0 2 5 1 2 1 0 1	23 0 9 7 0 7 0 12 0 2 0 1 0 3 1 1 5 1	5 11 9 0 6 0 14 0 0 0 1 3 2 0 2 1 0	19 17 7 0 12 0 5 0 0 0 2 0 0 1 5 1 0 3 3 3 3	12 2 12 11 0 2 0 11 0 2 1 0 0 1 4 0 0 3 1 3	11 8 8 0 10 0 0 0 1 0	17 2 15 10 0 4 0 10 0 0	2 0 14 6 8 4 0 3 1 0 1	5 0 7 8 5 1 0 0 0 2 0 4	9 0 2 4 11 2 0 8 0 1 0
DEGREE DAYS	: 92	4	DATE:	JU	NE 2	0								
1303 1403 2103 2401 3403 6101 7203 10203 12202 12205 12206 91101 91102 91103 91104 92101 92101 92102 94101 95101 95102	2 5	10 4 15 15 0 3 0 0 0 0 1 1 1 0 0 1 1 1 1 1 1 1 1 1	0 0 5 13 1 25 4 8 0 0 1 2 0 0 1 1 1 1 1 1 1 1 1 3	5 1 25 0 27 5 0 1 0 3 2 2 1 0 1 0 2 0 1	2 0 10 0 13 1 0 8 0 0 0 0 0 0 2 0 0	4 1 11 0 5 0 0 0 1 1 0 0 0 1 1 3 0 0 2 0	4 0 20 0 4 5 0 0 0 0 2 0 0 0 1 1 0 0 0 2	5 0 15 2 3 8 0 0 0 0 5 0 0 0 1 0 0 1 1	3 0 12 1 6 5 0 0 1 0 1 0 2 0 1 0 1 0 3 1	7 0 14 20 15 8 0 1 1 0 0 0 0 0 1 0 0 1 0 1 0 1 0 1 0	9 6 19 21 13 2 0 0 0 0 0 1 1 3 0 0 2 0 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3	3 0 9 2 11 3 0 0 2 0 2 0 1 0 2 1 0 2 1 0 2 1 0 2 1 0 2 1 0 2 1	5 0 7 2 10 0 0 0 0 0 1 2 0 0 2 2 2 2 2 2 2 2	6 0 20 5 12 1 0 0 1 0 0 1 0 0 2 1 1 0 0 1

DEGREE	DAYS:	1015		DATE	Ξ:	JUNE	23								
1303	0	3	2	0	2	0	1	0	1	1	0	1	4	3	3
1403	0	0	0	1	0	0	1	0	0	0	0	0	0	3	3
2103	1	1	0	0	1	0	3	2	9	4	2	2	2	4	3
2401	2	0	1	0	0	2	0	0	2	1	3	1	2	2	2
3403	1	0	0	7	5	9	5	13	14	3	6	7	2	2	0
6101	1	1	1	3	1	1	1	1	0	2	3	0	2	2	2
7203	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
10203	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
12301	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
91101	1	0	0	2	1	0	0	1	0	1	0	1	0	0	1
91102	1	0	0	1	0	0	0	1	0	0	0	2	0	0	0
91103	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
91104	0	0	0	1	0	0	0	0	0	1	0	0	2	0	0
92101	0	0	0	1	0	0	0	0	0	0	1	1	0	1	0
92102	1	0	0	0	0	0	0	0	1	0	0	1	0	0	0
94101	1	0	0	2	0	0	0	0	0	0	0	0	0	0	0
95101	0	1	0	1	0	0	0	0	1	0	1	1	0	1	0
95102	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
95103	0	0	0	0	0	2	0	0	0	0	0	0	2	0	0

Appendix 7-C. Data for between + within-field log s^2 x log \bar{x} regression analysis (Table 9).

A. Adults

Degree Days		Sweepnet		S	quare Foo	t
> 48°F ^a	#Fields ^b	$\log s^2$	<u>log x</u>	#Fields ^C	log s ²	$log \bar{x}$
87				39	9927	-1.1269
91				24	-2.6076	-1.7868
101				54	-1.9663	-1.3426
110				8	-2.1918	-1.4357
121	6*	8463	4266	49	-1.4553	-1.1000
132	10*	8582	3513	15	8103	7111
154	42	.3127	0912	58	2066	5139
170	3*	5006	.0370	3*	-1.4699	6425
201	34	.3599	0110	49	3783	5068
210	23	2477	3465	12	8767	8163
225	27	7844	4688	28	-1.0590	7897
231	28	.2580	.0468	29	.0610	4028
290	57	.0947	0688	45	6174	6963

B. Larvae & Eggs - Square Foot Counts

Degree Days	l	arval		Eggs				
> 48°F ^a	#Fields ^d	log s ²	<u>log x</u>	log s ²	<u>log x</u>			
621	7	1.3227	.4021	1.5777	.6981			
725	19	1.8370	.6900	1.3733	.5099			
804	18	1.5353	.4783	1.5548	.4819			
858	20	1.4442	.4926	. 7371	.1133			
924	21	1.3169	.3910	.8290	.1616			
1015	21	.4341	1599	.6787	.0445			

^a Accumulated since 1 April.

b 6 x 10-25 sweep samples/field. See Sect. 3.3.1.

^c 6-10 Samples of up to 10 linear feet/field. See Sect. 3.3.1.

d 10-15 Square feet/sample. See Sect. 2.2.1.

^{*} Excluded in Table 9 as being two small a sample.

Appendix 8--Pubescent Wheat Study Weather Data.

Temperature records for three locations near the study area with accumulated degree days (base 48°F) for each site and mean accumulated degree days for all three sites. R24 = rainfall during the previous 24 hours (inches/hours).

April, 1975

	-New	Carl	isle	So	uth B	end	Site Center	Mean
Day	Max	Min	d-Day	Max	Min	d-Day	Max Min d-Day R24	d-Day
1	33	24		46	30			
	29	18		38	25			
2 3 4	25	18		33	25			
4	27	16		35	23			
5 6 7	29	16		38	23			
6	30	18		40	24			
7	39	18		49	23			
8	50	18		52	25	1		1
9	43	34		47	34	1		1
10	49	30		50	30	1		1
11	41	26		43	28	1		1
12	48	24		50	23	1		1
13	52	22	1	54	23	2		2 2 3 5
14	46	33	1	48	33	2 2 3 5		2
15	52	33	2	53	32	3		3
16	55	34	4	57	34			
17	68	34	11	69	35	13	Installe	
18	67	62	27	69	57	28	.5/5	28
19	56	42	30	57	43	31		31
20	47	40	30	49	36	31	Installed	31
21	47	29	30	48	30	31	.1/.5	31
22	64	42	36	65	42	38	.1/1.	
23	70	50	48	68	53	50	.1/2	49
24	58	40	51	58	42	54		53
25	54	38	53	59	43	58	Calibrated	56
26	61	33	57	62	35	63	62 32(4+est60)	61
27	49	41	57	51	42	64	48 40 64 .9/10	62
28	49	42	57	50	44	65	48 41 64 .4/2	62
29	76	40	69	75	41	76	76 40 76	74
30	68	52	81	71	55	91	68 52 88 .1/.5	87

May, 1975

Day	-New Max	Carl Min	isle d-Day	So Max	uth B Min	end d-Day	- Max	Site Min	Center- d-Day	R24	Mean d-Day
_	60	44	86	61	45	97	60	42	91		91
1 2	70	44	95	71	45	107	73	40	101		101
2	62	51	103	65	53	118	73 61	40	101	.1/.5	110
3 4	56	31 44	103	57	43	121	55	49	110	.1/.5	112
4 5	70	44	115			130	70	38	118		121
6	65		113	70 68	41				128	15/5	132
5 6 7	66	52 45	137		51	142	65	50 44	136	.15/.5	142
8	66	45 50	150	68 68	47 54	152 165	66 68	51	148		154
9	75	50 52	166	75		181	76		163		170
10	/5				53			50	175		183
11			st180 st194	76	47	195	73	48			197
	EO			77	46	209	78 50	42	188	1/ 5	
12	58 70	40	197	64	43	216	58	37	191	.1/.5	201
13		36	205	70	39	225	69	35	199		210
14	72	41	215	72	47	237	72	42	209		220
15	60	44	220	59	45	242	57	40	212		225
16	64	40	226	69	41	251	63	36	217		231
17	78	40	239	75 25	40	262	80	40	231		244
18	88	44	258	85	50	282	87	45	249	/	263
19	92	57	284	91	62	310	90	60	276	(out for	290
20	94	63	314	91	67	341	94	62	306	repairs)	320
21	82	62	338	87	63	368	85	60	330		345
22	72	60	356	73	61	387	70	56	345		363
23	87	56	380	85	57	410	88	54	368		386
24	90	66	410	88	67	440	90	64	397	.05/2.0	416
25	88	63	438	86	65	468	87	62	423	.85/1.5	443
26	77	64	460	77	65	491	79	62	445		465
27	70	50	472	72	54	506	71	46	456		478
28	81	45	487	77	47	520	79	42	470		492
29	79	57	507	80	58	541	78	55	488		512
30	80	60	529	78	62	563	80	56	508	.4/3.0	533
31	68	52	541	68	53	575	68	46	517		544

June, 1975

	-New	Carl	isle	So	uth B	end		-Site	Center		Mean
Day	Max	Min	d-Day	Max	Min	d-Day	Max	Min	d-Day	R24	d-Day
1	71	47	552	72	48	587	72	45	528		556
2	70	46	562	65	49	596	68	44	537		565
3	74	49	576	74	51	610	76	48	551	.5/3.0	579
4	83	52	596	82	55	630	82	52	570	.05/.5	599
5 6 7	80	62	619	79	61	652	81	60	592		621
6	70	56	634	71	58	668	73	59	610		637
	60	45	639	60	49	674	58	48	615		643
8	68	44	648	68	45	683	68	42	623		651
9	72	45	659	73	48	695	73	42	634		663
10	80	52	677	80	55	715	82	50	652		681
11	84	63	703	80	64	739	83		678	.35/3.5	707
12	76	56	721	76	59	759	75	55	695		725
13	88	57	745	83	58	781	86	56	718	.4/2.0	748
14	82	58	767	81	60	803	84	56	740	.6/4.0	770
15	71	60	785	71	59	820	71	57	756	.5/2.0	787
16	78	52	802	77	56	838	76	52	772		804
17	80	69	828	80	69	864	78	66	796	.4/1.5	829
18	91	64	858	88	66	893	88	62	823		858
19	90	72	891	87	73	925	92	70	856		891
20	93	70	925	90	71	957	96	68	890		924
21	92	66	956	88	60	983	92	64	920		953
22	94	68	989	91	71	1016	92	68	952		986
23	86	68	1018	84	69	1044	88	67	982		1015
24	84	67	1046	84	68	1072	86	66	1010		1043
25	79	68	1072	7 7	68	1096	79	66	1034	.2/.5	1067
26	85	65	1099	82	66	1122	89	62	1062		1094
27	90	64	1128	86	64	1149	91	62	1090		1122
28	92	66	1159	89	67	1179	93	62	1120		1153
29	90	66	1189	87	65	1207	92	63	1150		1182
30	91	62	1217	88	63	1235	91	60	1178		1210

Niles precipitation, 1975, in inches. Source: NOAA.

<u>Day</u>	<u>April</u>	<u>May</u>	<u>June</u>
1	t	.01	
2	.77		. t
3	.21	.14	.29
4			.05
5		.01	.12
6		.46	
7			
8		t	
9	t		
10			
11		.01	.50
12		.18	t
13			.36
14	t	t	.62
15		.05	.88
16	t		
17			.42
18	1.40		•
19	t		
20		.14	t
21	.21	.30	
22	.06	.05	t
23	. 38		.08
24	.05	.22	.01
25	t	.42	.04
26		t	
27	.87	t	
28	.49		
29	.05	.12	
30	.48	.35	

Appendix 9. Examples of Filter Operation.

Appendix 9a. Application of filtering technique to 1974 Gull Lake data (Sawyer 1976).

С	ROP HEI		LARVAL DENSITY					
DD	0(Y)	E(Y)	0(Y)	E(Y)	F(Y)			
FIELD: 91	FIELD: 911.							
313 378 440 488 567 663 760 835 926 985 1043 1157 1249	3.43 4.49 6.54 8.07 10.51 13.50 16.81 16.18 18.58 21.65 23.50 29.65 30.12	3.45 3.99 5.28 11.62 16.71 20.53 22.68 24.18 24.40 24.27 23.93 27.46	0.00 0.00 0.00 .06 .15 .91 .74 .60 .09 .03 0.00	0.00 0.00 0.00 .71 1.49 1.89 1.20 .48 .09 0.00 0.00	0.00 0.00 .06 .43 1.20 1.31 .90 .28 .06 0.00			
FIELD: 91	6.							
313 378 440 488 567 663 760 835 926 985 1043 1157 1249	8.10 3.98 5.71 7.32 9.21 12.56 14.25 10.83 15.47 17.40 18.94 24.13 26.06	8.11 6.08 8.96 12.97 16.74 20.09 21.28 21.06 21.47 22.51 26.00 25.73	0.00 0.00 .03 .06 1.40 3.29 4.56 3.47 1.61 1.03 1.06 .11	0.00 0.00 .39 1.28 3.98 5.64 4.87 2.71 1.21 .47 0.00 0.00	0.00 .03 .22 1.34 3.63 5.10 4.17 2.16 1.12 .76 .06			
FIELD: 91	7.							
313 378 440 488 567 663 760 835 926 985 1043 1157 1249	9.00 4.13 5.71 7.99 10.12 12.09 15.08 14.49 17.17 19.76 20.63 27.32 28.03	9.01 6.62 6.21 12.03 16.72 19.85 21.56 22.90 23.12 24.41 27.27 27.65	0.00 0.00 0.00 .15 1.72 8.26 8.02 3.16 2.64 1.29 .71 .06 0.00	0.00 0.00 0.00 1.04 4.10 9.61 8.36 3.55 1.71 .63 0.00 0.00	0.00 0.00 .15 1.38 6.18 8.81 5.76 3.10 1.50 .67 .03 0.00			

Appendix 9b. Application of filtering technique to 1976 PWS data (Sawyer, unpublished data).

DD C	CROP HEIGHT DD O(Y) E(Y)			LARVAL DENSITY O(Y) E(Y) F(Y)					
FIELD: 136.									
	4.00 6.00 5.00 8.00 10.00 16.00 22.00 25.00 24.00 28.00 34.00 33.00	4.02 5.01 6.68 8.58 15.28 19.62 24.41 28.83 29.64 29.78 31.20 36.34	0.00 0.00 .40 .30 .50 9.10 17.20 43.10 2.10 1.10 2.30 .30 0.00	0.00 0.00 1.00 1.18 7.13 21.12 33.13 47.71 20.73 7.36 2.89 .31	0.00 .40 .65 .84 8.11 19.16 38.12 24.90 10.92 4.83 1.60 .16				
FIELD: 23	6.								
264 282 330 338 354 374 399 499 570 640 756 815 879 931	3.00 3.00 4.00 4.00 6.00 7.00 12.00 22.00 20.00 22.00 18.00 29.00 32.00 30.00	3.03 3.05 3.55 3.80 3.92 6.58 12.99 16.64 23.02 27.28 27.52 25.97 29.89 34.83	0.00 0.00 0.00 0.00 0.00 .10 0.00 4.70 6.50 10.90 .80 .60 2.20 .40 0.00	0.00 0.00 0.00 0.00 0.00 .36 2.62 10.58 15.45 17.69 8.03 3.13 1.61 .16	0.00 0.00 0.00 0.00 .10 .18 3.66 8.54 13.17 9.24 4.31 2.67 1.00				
FIELD: 246.									
330 354 374 399 499 570 640 756 815 879 931 1025	5.00 4.00 4.00 12.00 16.00 18.00 15.00 14.00 24.00 28.00 28.00	5.00 4.51 4.27 10.49 15.47 19.65 24.88 23.03 21.97 25.61 31.08	0.00 0.00 0.00 .10 6.00 12.40 20.50 5.20 4.20 5.30 2.80 0.00	0.00 0.00 0.00 2.23 12.29 24.49 33.53 18.50 9.07 4.77 1.07	0.00 0.00 .10 4.12 12.34 22.49 19.36 11.35 7.19 3.78 .53				

DD	CROP HEI	GHT E(Y)		AL DENSI E(Y)	
FIELD: 3	37.				
282 330 354 374 387 487 570 640 756 815 879 931	5.00 8.00 11.00 14.00 15.00 19.00 24.00 24.00 28.00 26.00		.60 2.10 4.10 4.60 11.60 8.50 24.501 26.901 22.70 3.80 .50 0.00	16.64	2.10 4.22 5.97 10.02 38.43 69.02 74.10 56.48 25.52 8.57 2.59 .38
FIELD: 3	46.				
338 354 380 399 487 570 640 815 879 931	8.00 12.00 12.00 15.00 20.00 24.00 28.00 25.00 26.00	5.00 8.20 11.31 16.83 20.51 23.88 32.06 32.81 31.22 32.73	0.00 .30 .80 3.40 1.00 8.00 8.20 .50 0.00 0.00	0.00 .78 1.37 13.84 22.73 26.99 17.23 5.59 1.64 .13	.30 .79 2.39 7.42 15.36 17.60 8.87 2.79 .82 .06
FIELD: 5	24.				
262 282 338 354 374 387 487 570 640 756 815 879 931	6.00 6.00 6.00 11.00 13.00 12.00 18.00 22.00 26.00 27.00	2.03 2.55 4.29 5.15 6.44 12.45 16.61 18.78 21.78 22.98 25.71 28.32 31.88	0.00 0.00 0.00 0.00 .10 0.00 .50 3.80 12.10 3.30 8.90 .30 .10	0.00 0.00 0.00 0.00 .22 2.27 5.21 9.27 18.31 10.33 7.08 2.31 .25	0.00 0.00 0.00 .10 .11 1.39 4.51 10.69 10.80 9.61 3.69 1.20 .18

CROP HEIGHT			LARVAL DENSITY				
DD	0(Y)	E(Y)	O(Y)	E(Y)	F(Y)		
FIELD: 6	12.						
380 399 487 570 640 756 815 879 931	8.00 7.00 13.00 17.00 18.00 20.00 20.00 27.00 28.00 32.00	8.01 12.93 17.73 21.20 25.56 25.75 26.08 28.98 30.36	0.00 .20 .20 1.00 1.60 .30 0.00 0.00 0.00	0.00 2.18 4.35 5.20 4.98 2.37 .83 .20 0.00	.20 1.19 2.68 3.40 2.64 1.18 .41 .10 0.00		
FIELD: 10	024.						
262 282 330 354 374 387 487 570 640 756 815 879 931 1025	3.00 4.00 6.00 6.00 12.00 12.00 13.00 17.00 22.00 28.00 28.00 29.00 36.00	3.03 3.54 4.78 5.41 8.71 16.29 17.90 19.39 24.32 26.11 30.00 31.30 34.14	0.00 0.00 0.00 0.00 0.00 .40 1.30 3.50 9.60 4.80 1.70 .30 .10	0.00 0.00 0.00 0.00 0.00 4.03 9.19 12.76 17.13 9.82 3.84 1.20	0.00 0.00 0.00 0.00 .40 2.67 6.35 11.18 10.97 5.76 2.07 .65		
FIELD: 1135.							
330 354 380 399 499 591	6.00 6.00 6.00 8.00 3.00 12.00	6.04 7.71 8.11 14.12 14.13	0.00 .30 .90 .60 .50	0.00 .79 1.50 9.10 19.91	.30 .84 1.05 4.80 10.20		

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