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Studies Related to the Concept of Pest - Crop System Design:
1) Adult Parasitoid Activity and Its Relation
to Biocontrol and 2) Forest Harvesting
and the Spruce Budworm

presented by

Jan Peter Nyrop

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# STUDIES RELATED TO THE CONCEPT OF PEST - CROP SYSTEM DESIGN:

1) ADULT PARASITOID ACTIVITY AND ITS RELATION

TO BIOCONTROL AND 2) FOREST HARVESTING

AND THE SPRUCE BUDWORM

bу

Jan Peter Nyrop

### A THESIS

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#### **ABSTRACT**

STUDIES RELATED TO THE CONCEPT OF PEST-CROP SYSTEM DESIGN:

1) ADULT PARASITOID ACTIVITY AND ITS RELATION TO BIOCONTROL AND

2) FOREST HARVESTING AND THE SPRUCE BUDWORM

By

#### Jan Peter Nyrop

Damage to crops from pest organisms can be controlled two ways. First, the structure of the crop production system can be accepted as is and the pest organisms controlled in some fashion. Second, the structure of the crop production system can be manipulated so that pest damage is minimized. In this thesis the latter concept is explored in two studies.

In the first project, the temporal and spatial dynamics of an adult parasitoid (Glypta fumiferanae Vierick) were assessed and related to the theory of parasitoid-host dynamics. This was done because adult parasitoids and factors influencing them may be objects of control for system management.

Field data on adult parasitoid activity was used to construct a model relating activity to weather. Historical data were then examined to determine if weather induced changes in parasitoid activity were reflected in changes in parasitism. I concluded cool, wet weather inhibited parasitoid host seraching and attack.

Observations of G. <u>fumiferanae</u> attacking hosts suggested the parasitoid does not forage optimally. An experiment was conducted to determine whether light and/or temperature influenced parasitoid foraging behavior. The experiment indicated this was the case.

The empirical studies were related to parasitoid-host theory which was then revised. This revision consisted of redefining the attributes of a successful biocontrol agent to be one which has a high searching efficiency but need not aggregate in areas of high host density.

In the second project, the use of forest harvesting was explored as a way to change the structure of the spruce budworm/forest system and thereby improve system control. A theoretical basis for the strategy was developed, an economic analysis was made and the strategy was field tested. Results from the field test did not substantiate the theory on the effect of partial harvesting on budworm dynamics. As a result, the test was inconclusive.

to Jessica Erin

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#### INTRODUCTION

An inherent aspect of most crop production systems is the necessity to take into account the influence of crop damaging organisms. Two different, though not mutually exclusive, approaches can be adopted. First, the structure of the crop production system can be accepted as is and pests can be controlled in some fashion. The structure of a system refers to the physical and institutional properties of the system and policies which, once implemented, usually cannot be changed. The structure of crop production systems involves a number of diverse factors such as crop variety, crop spacing and mixing, the genetic makeup of plants and pests, crop markets, and society's expectations of crop consumption. With the first approach, chemical toxicants are most often used to either drive a pest species as close to extinction as possible or preserve the integrity of the host plant.

Increasing social and economic costs of chemical pesticides, reports of their deleterious effects, and the significant rate at which they fail to prevent crop damage make it apparent that the application of pesticides can no longer be an exclusive pest control policy. With the advent of integrated pest management, inputs of pesticides have been reduced in some crop production systems. For the most part, however, this has been accomplished by improving the decision making process of when and how much of a pesticide to apply as opposed to reducing reliance on pesticides as a control tool.

The second approach, which might also be considered the second phase of pest management, is to consider pests and factors which influence pest dynamics as an integral part of the crop production system and design a system structure so that the effects of pest organisms are minimized (Haynes et al. 1980). This will be

a formidable task. As in the design of an electronic amplifier, it will necessitate the wedding of theory (often couched in terms of mathematics) and empirical observations. However, unlike the amplifier, pest crop systems are "badly defined systems" (Beck 1981, Young 1978). With such systems, a priori theory usually cannot predict the nature of the system and design cannot come about from theory alone. Furthermore, planned experiments with these systems are often difficult and may at times be impossible to implement. Even in situ data of the "normal operation" of the system may be difficult to acquire.

Although the obstacles to this approach seem insurmountable, it must be pursued vigorously if pest management is to become what the name implies. In this thesis, I present the results of two projects which were motivated by the concept of design. In both, an effort is made to make use of theory and empirical data in an effort to propagate the concept of system design and to develop principles appropriate to each area of study. In the first project, the temporal and spatial dynamics of an adult parasitoid of the spruce budworm were investigated. This was done to develop methods for sampling this important life stage of parasitoids and to develop concepts appropriate to design-mediated biological control. In the second project, the use of forest harvesting was explored as a way to change the structure of the spruce budworm/spruce fir forest system and thereby improve system control. No pretense existed of actually being able to meet this entire objective. However, the project was pursued in hopes of establishing a basis for further study in this area.

Adult Parasitoid Activity and
Its Relation to Biocontrol

#### Introduction

Actions taken to achieve or improve control of crop damaging insects using parasitoids fall into three endeavors: importation of exotic parasitoids, augmentation of established exotic or endemic parasitoids, or management of existing parasitic insects. Management entails any action taken to improve the effectiveness of a parasitoid as a control agent. When applicable, management should be the first action taken. Furthermore, when management can be implemented, it will likely be more cost effective than importation or augmentation.

Knowledge of factors which influence the life system of a parasitic insect and information on how these factors may be manipulated are prerequisites for successful parasitoid management. One of the most important aspects of the life system of parasitic insects is host searching since the maximum number of progeny that can be produced by a generation of parasitoids is determined by their host searching efficiency. Therefore, the adult female parasitoid and factors which influence the number and distribution of hosts attacked by the female may be important objects of control in any management effort.

Numerous theoretical investigations have examined the relationships between the host distribution, parasitoid host searching behavior and resultant dynamics of parasitoid-host systems. Based on these studies, the characteristics of successful biological control agents have been advanced (Beddington et al. 1978). In all of these studies the only factors influencing parasitoid host searching are the density and distribution of hosts. However, factors other than the host influence parasitoid searching behavior. Few field studies have been undertaken to elucidate these factors. This is in part because such studies should

focus on the temporal and spatial dynamics of the adult parasitoid. These organisms are small, highly mobile and most often few in number and as such are difficult to sample. Even fewer studies have sought to juxtapose empirical observations of the adult parasitoid and the theory of parasitoid-host systems. Investigations of this type are necessary though if the chasm between empirical observations and parasitoid-host theory is to be bridged and, concurrently, our ability to manage parasitoids broadened.

In this paper we present the results of an investigation of the spatial and temporal dynamics of adults of the parasitoid Glypta fumiferanae Vierick. G. fumiferanae is a common, specific, univoltine parasitoid that attacks first and second instar spruce budworm (Choristoneura fumiferana (Clemens)). It overwinters in the budworm larvae and emerges from either fifth or sixth instar hosts. It then spins a pupal case in the tree and emerges as an adult at approximately the time when budworm eggs are eclosing. Details of the biology of G. fumiferanae are provided by Brown 1946, Wilkes et al. 1948, Dowden et al. 1948, and Miller 1960.

The paper consists of three sections. First, information on the temporal and spatial dynamics of adult <u>G. fumiferanae</u> is presented. Data was obtained through field studies of the activity patterns of the parasitoid and through a laboratory investigation of host searching behavior in a gradient of abiotic environments. Second, the relationship between the temporal dynamics of the adult parasitoid and its searching efficiency is examined using historical data of the interaction between <u>G. fumiferanae</u> and its host. Finally, the general implications of our findings are assayed. This is done by relating our observations to established parasitoid-host theory and to extensions of this theory which we have developed.

#### Methods and Results

<u>Field Studies</u>: Data was collected on the temporal and spatial activity patterns of adult <u>G</u>. <u>fumiferanae</u> during 1980 and 1981 in northern Michigan. One forest stand was sampled in 1980, and two stands were sampled in 1981 (fig. 1). The study area used in 1980 was not used in 1981 because it was cut for timber.

Based on previous sampling of adult parasitoids (Julliet 1963, Price 1971, Reardon et al. 1977, Ticehurst and Reardon 1977, Simmons unpubl. data), malaise traps were selected as the principal tool for measuring the activity patterns of adult G. fumiferanae. Malaise traps are ideally suited for this task because they are passive traps and snare actively flying insects. The traps were designed and constructed for placement in tree crowns and on the forest floor; this way trap catch could be easily inspected. Details of trap design and construction are given in figure 2.

During early July 1980, 14 of these traps were placed in the mid to upper crowns of 14 balsam fir (Abies balsamea (L.) (Miller)), and 14 were placed on the forest floor in stand 1 (fig. 1). Traps were placed on the forest floor because it was hypothesized that G. fumiferanae might use this habitat when foraging for food. The forest floor provides a habitat for many plants which produce nectar and pollen. These plant products have been cited as food for adult parasitoids by many authors (Chermokova 1960; Leuis 1960, 1961a, 1961b, 1963, 1967; Shahjahan 1974; Simmons et al. 1975; Syme 1966, 1975; Thorpe and Caulde 1938).

Stand 1 consisted of <u>ca.</u> 60% mature balsam fir. Trees in which traps were placed were selected on the basis of ease of access for trap placement and good crown condition. Traps were positioned in the trees so that one open side of the

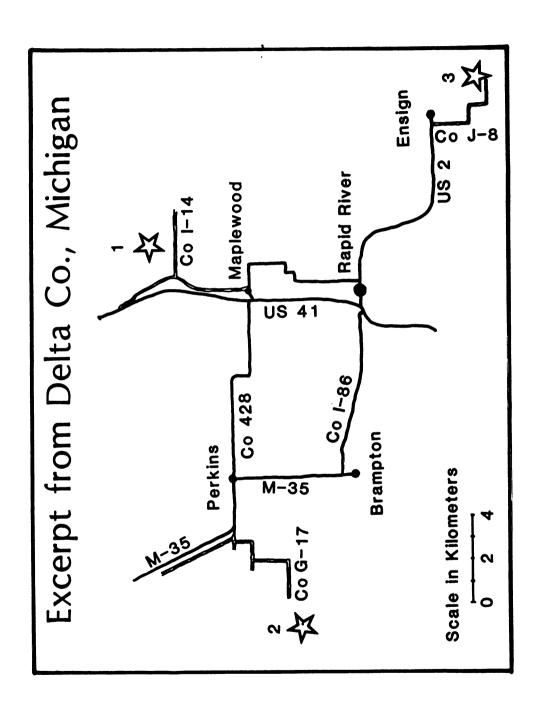


Figure 1. Location of study plots in Michigan's Upper Peninsula.

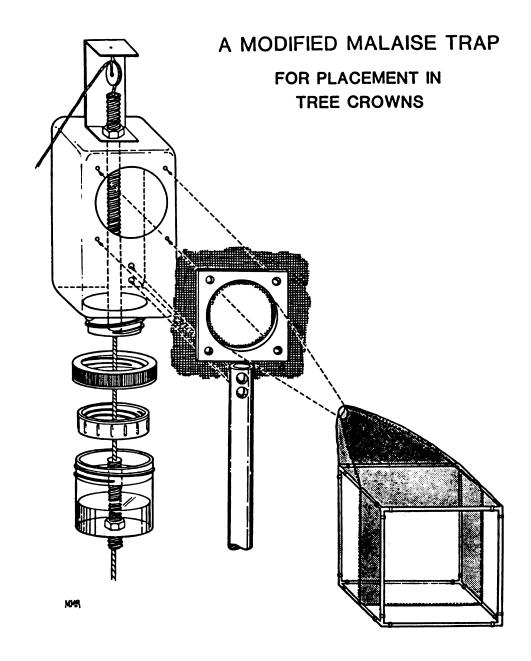


Figure 2. Details of a malaise trap designed for placement in tree crowns. Trap volume is 1 m<sup>3</sup>. The frame is constructed of PVC pipe and the screening of Saran 20 mesh screening. The intermediate and final collection jars and the face plate are plastic. The final collection jar is affixed to a nylon cord which passes through a threaded pipe which is in turn attached to the intermediate collection jar. Trap catch is inspected by lowering the final collection jar.

trap was tangent to the tree crown. G. fumiferanae pupal density per m<sup>2</sup> balsam fir foliage was 7.61 (s=10.02, n=48). Traps were emptied each morning between 10 July and 10 August. In total, 39 females were trapped in the trees (fig. 3), and none were caught on the forest floor.

We felt that the low trap catch in 1980 may have been due to the orientation of the traps in the tree crowns. Therefore, in 1981 the traps were oriented so that the open sides were perpendicular to the crown. Thirteen traps were located in balsam fir and 13 on the forest floor in stands 2 and 3 (fig. 1). Stand 2 was 100% mature balsam fir and stand 3 ca. 50% mature balsam fir. The density of G. fumiferanae pupae per m² balsam fir in stands 2 and 3 was 10.45 (s=11.28, n=40) and 9.87 (s=9.03, n=40). Traps were emptied each morning between 2 July and 15 August. During the peak flight of females, traps in stand 2 were emptied bi-hourly from 800 to 2200 hours for 3 days. In total, 125 males and 294 females were trapped in trees in stand 2, and 72 males and 182 females were trapped in trees in stand 3 (fig. 4). Clearly, the orientation of the traps in the trees during 1981 was superior and indeed necessary for capturing significant numbers of G. fumiferanae. No parasitoids were trapped on the ground.

The flight activity as measured by trap catch of both males and females in stands 2 and 3 during 1981 are remarkably similar (fig. 4). Hence, we hypothesized that weather was a major determinant of flight activity. This hypothesis was supported by the bi-hourly trap catch data for females in stand 2 (fig. 5). This data indicated that parasitoid flight activity was strongly depressed by rain. On 17 July rain began ca. 1500 hours resulting in a decline in female activity as measured by trap catch. Activity also followed the daily temperature profile.

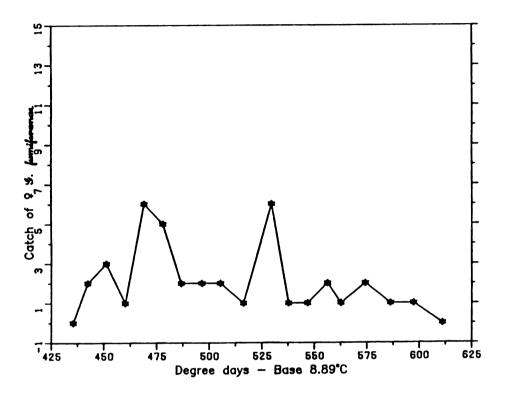
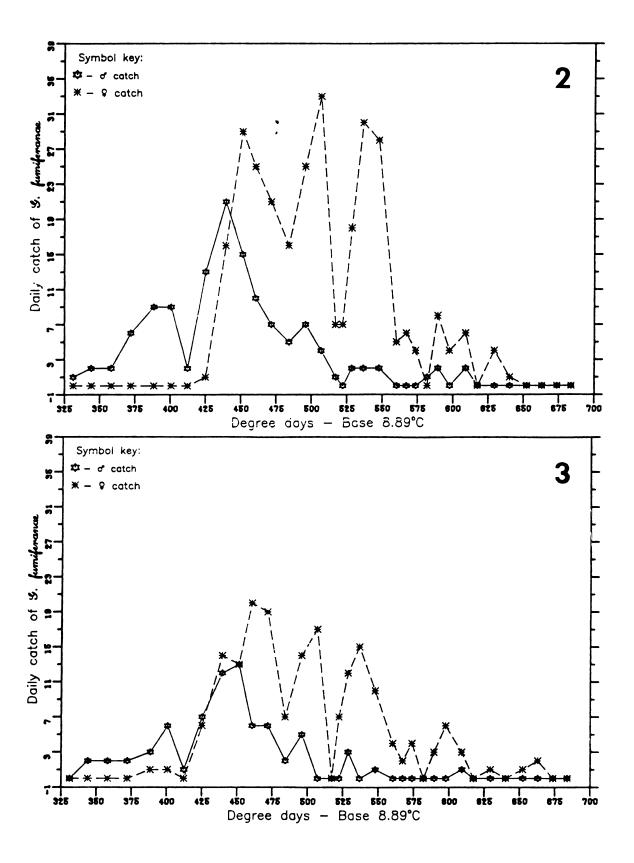


Figure 3. Catch of female <u>Glypta fumiferanae</u> from 14 malaise traps located in balsam fir trees in stand 1 during 1980.

Figure 4. Catch of male and female adult <u>Glypta fumiferanae</u> from 13 malaise traps located in balsam fir trees in stands 2 and 3 during 1981.



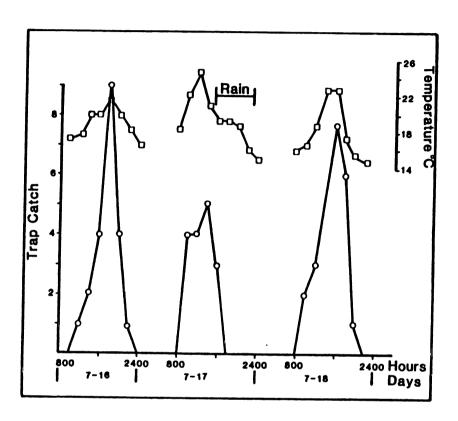


Figure 5. Bi-hourly trap catch of female <u>Glypta</u> <u>fumiferanae</u> from 13 malaise traps located in balsam fir trees in stand 2 and the temperal distribution of temperature and rainfall.

In order to examine the influence of rainfall and other possible weather factors more extensively, a model was constructed so that the relationship between weather and the time series of trap catches could be examined. A modelling approach was adopted because conventional statistical methods could not be used in analyzing this relationship. This is because trap catch in the absence of any other influencing factors would be proportional to the density of adult parasitoids. This density was unknown. However, with the assumptions outlined below, an index of this density could be derived from the trap catch data.

In the model, predicted trap catch (TC) of <u>G</u>. <u>fumiferanae</u> was expressed as a function of adult density (Ad) and adult activity (Aa). In addition, the activity of the adults was related to a set of weather variables (W). These relationships are concisely written as:

$$TC = f(Ad, Aa)$$
 (1)

$$Aa = f(W)$$

The time-specific adult density, Ad(t), is not known. However, it is a function of the emergence, dispersal and death processes operating until time t. With the assumptions that dispersal into and out of an area balance and that either a constant or no external mortality operates on the adult population, Ad(t) is a function of the emergence rate and physiological time interval between emergence and death. With these assumptions, our modelling methodology can be outlined as follows: 1) specify a function for the change in adult density through physiological time, 2) specify a function relating trap catch to adult density and weather, 3) parameterize a model based on these functions through the use of a nonlinear optimization algorithm, and 4) evaluate model fit by

comparing predicted trap catch to the data with which the model was parameterized and to an independent data set. Physiological time was approximated with degree days (DD) (base 8.89°C), and the model was parameterized with the data collected from stand 2.

The general structure of the model is given in figure 6. Ad(t) is the integral of the difference between the emergence and adult mortality rates. Each of these rates was modelled as a distributed delay. These delays can be used to model aggregate processes which are made up of entities in which each entity has an output related to its input via a pure time delay. In this case the emergence of a parasitoid is related to some initial time point and its death is related to its emergence via two physiological time delays (PTD).

For a population of parasitoids, a PTD is a random variable. It can be shown that the distributed delay is based on the probability density function (pdf) of a PTD (appendix 1). If f(t) is the pdf of the physiological time delay between emergence and death, and p(t) and m(t) are the emergence and mortality rates, then:

$$m(t) = \int_{0}^{t} p(v)f(t-v)dv$$
 where  $t = physiological time$ .

The adult density is given by:

$$Ad(t) = \int_0^t p(v) - m(v) dv$$

The parameters for the emergence rate were estimated from data on the cumulative emergence of <u>G. fumiferanae</u> collected in stand 2. Those for mortality were estimated jointly with other model parameters from the trap catch data. The gamma density function with an integer shape parameter (k) was used for f(t). The shape parameter was determined by initially selecting a value based on the shape of the distribution of p(t) and Ad(t) and then adjusting it

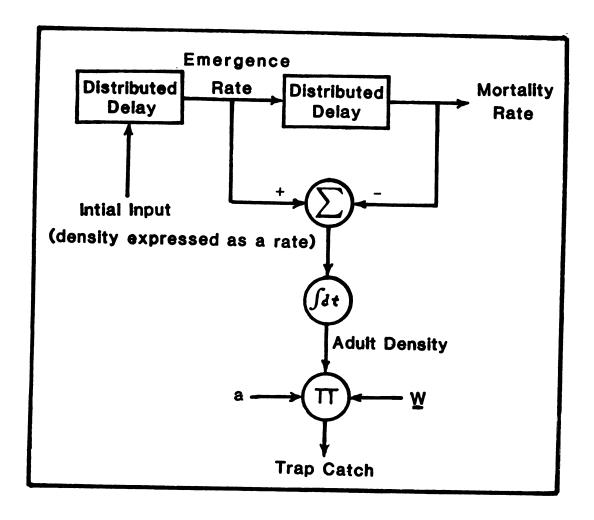


Figure 6. General structure of the model used to analyze the temporal distribution of malaise trap catches of Glypta fumiferanae.

W is a vector of weather variables and a is a proportionality constant.

prior to determining the other model parameters until a best fit was obtained. k was estimated independent of the optimization because it must be an integer and the optimization algorithm used requires continuous decision variables. The mean of PTD (PTD) and k are sufficient statistics for f(PTD).

Variables in the set  $\underline{W}$  included rainfall, relative humidity, temperature, and barometric pressure. Hourly measures of these variables were averaged over 800 to 2200 hours, which was the period  $\underline{G}$ . <u>fumiferanae</u> was active. The values of  $\underline{W}$  and TC used in the model are constant for a given day while Ad changes continuously according to the degree days accumulated on that date. Therefore, the value of Ad midway through a day was used in the function TC = f(Ad, Aa).

To estimate model parameters, a linear form of (1) was adopted. Predicted trap catch is therefore given by:

$$\hat{\mathrm{TC}}_n = (\mathbf{a} + \underline{\mathbf{b}}\underline{\mathbf{W}}) \; (\int_0^{t_n} \mathbf{p}(\mathbf{v}) - \int_0^{t_n} \mathbf{\tilde{\mathbf{I}}}(\mathbf{t}_n - \mathbf{z}) \mathbf{p}(\mathbf{z}) \mathrm{d}\mathbf{z} \mathrm{d}\mathbf{v})$$

where: n is a daily index, t<sub>n</sub> are degree days accumulated on day n, and underlined variables denote vectors. The parameters a, <u>b</u> and <u>PTD</u> of the mortality rate were estimated using the complex nonlinear optimization algorithm due to Box (Kuester and Mize 1973). The objective function for the optimization is given by:

Minimize 
$$F = \sum_{n=1}^{m} (T\hat{C}_n - TC_n)^2$$

With this objective function, the value assigned to the parameters of a particular model minimizes a sum of squares about the observed trap catch.

Owing to the importance of the female parasitoid, the model was parameterized for female trap catch only. To initiate parameter estimation, the initial input (i.e., total number of pupal female parasitoids) was arbitrarily selected. This was done for two reasons. First, the absolute number of pupal G.

fumiferanae in the trapping area was not known. Second, we were interested in the qualitative aspects of the model, and knowing this value was therefore unnecessary. As a result, model parameters except <u>PTD</u> are scaled by the initial input.

As stated previously, the emergence rate p(t) was determined from emergence data. The rate of adult female emergence was separated from the male emergence rate by assuming that the initial trap catch of females indicated the onset of female emergence and that the shape of the female emergence curve was similar to the joint male and female emergence curve. The first assumption is based on the fact that G. fumiferanae was not collected in any habitat other than the forest canopy. Hence, it is unlikely that females emerged at the same time as males but initially used a habitat other than the trees containing budworm in order to find food or alternative hosts. Parameters for the distributed delay used to describe female emergence were selected so that predicted emergence resembled the combined male and female curve but was appropriately delayed in time (fig. 7).

Results of the parameter estimation are given in table 1. Only models which had some ability to predict trap catch are presented. Since the objective function for the optimization process can be thought of as a residual sum of squares, an R<sup>2</sup> was computed for each model. A test of significance cannot be made, though, due to correlation among variables within the model and because the distribution of the various sum of squares is unknown. The best prediction of trap catch was achieved with precipitation and temperature as independent variables. Wet and cool weather was associated with declines in trap catch. Reasonably large changes in the residual sum of squares associated with the

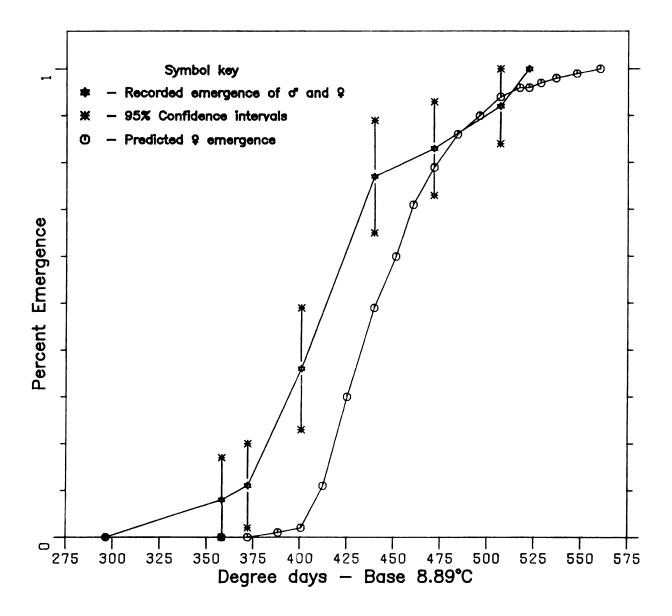


Figure 7. Observed combined emergence of adult male and female <u>Glypta</u> fumiferanae in stand 2 and predicted emergence of female <u>G. fumiferanae</u> as a function of degree days base 8.89 °C.

Table 1. Models used to predict the temporal distribution of malaise trap catch of female Glypta fumiferanae. Variables used are:

= predicted trap catch

= average temperature between 800 and 2200 hours on day n RH<sub>n</sub> = average relative humidity between 800 and 2200 hours on day n

= rainfall in inches between 800 and 2200 hours on day n = a measure of adult female density on day n

= expected number of degree days for emergence from pupae from an initial time point to

PTD = expected adult temale life span in degree days

K, K = shape parameters of gamma density function used to describe PTD

In all models the emergence rate is given by a distributed delay with  $\overline{\text{PTD}}$  = 55.5, k<sub>p</sub> = 4 and t<sub>o</sub> = 388 degree days base 8.89°C. The value of k<sub>m</sub> was 8.

MODEL	PTDm	Residual So	um s R <sup>2</sup>
$1 \text{ TC}_{n} = .605 \text{ Ad}_{n}$	81	1300	.51
$2 \text{ TC}_{n} = (.012 + .852 \text{ T}_{n}) \text{ Ad}_{n}$	84	1117	. 58
$3 \text{ TC}_{n} = (1.24892 \text{ RH}_{n}) \text{ Ad}_{n}$	76	1046	.61
$4 \text{ TC}_{n} = (154.6 - 1.89 \text{ R}_{n}) \text{ Ad}_{n}$	86	708	.74
$5 \text{ TC}_{n} = (.662714 \text{ RH}_{n} + .599 \text{ T}_{n}) \text{ Ad}_{n}$	87	982	.63
$6 \text{ TC}_{n} = (.029 - 1.232 \text{ R}_{n} + .9553 \text{ T}_{n}) \text{ Ad}_{n}$	87	546	.80

stepwise inclusion of each variable in this model ensured that the model was not over-parameterized. The predicted trap catch and predicted temporal distribution of adult females using this model and actual trap catch for stand 2 are shown in figure 8. The predicted temporal distribution of adults corresponds to the index of adult density.

As stated above, it was not possible to evaluate the statistical significance of the relationship between wet and cool weather and diminished trap catch. Nonetheless, one is led to the conclusion that the relationship is not spurious for two reasons. First, the probability of obtaining the relationship by chance is very small. Based on the model-generated population index, adult female G. fumiferanae were most abundant between 450 DD and 560 DD. At the same time, there were 3 days when trap catch was greatly reduced (517 DD, 522 DD, 560 DD) and 1 day when trap catch was somewhat reduced (484 DD).

The trap catch at 528 DD was not considered a reduced level even though in stand 2 the number caught on this day (18) was approximately the same as that caught at 484 DD (16). The reasons for excluding this date are twofold. First, in stand 3 the number caught on this date is close to the average number trapped during what we assume to be favorable weather conditions. Second, the trap catch on this date followed a period of extremely depressed parasitoid activity and the resumption of higher levels of activity may not occur immediately after the restoration of favorable environmental conditions.

During the four days of reduced trap catch and only during these days was precipitation recorded. In addition, for the three days with greatly reduced trap catch the temperature was lower than the average for the 12-day period. For the purpose at hand, these weather conditions will be classified as wet and cool.

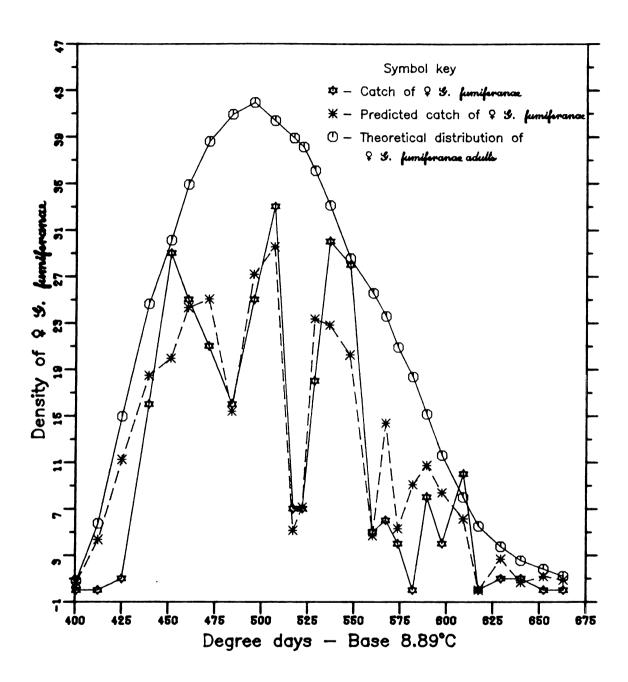


Figure 8. Predicted and actual malaise trap catch of adult female

Glypta fumiferanae in stand 2 and the theoretical, relative
temporal distribution of adult females. Malaise traps
were located singly in 13 balsam fir trees.

A measure of the likelihood that no relationship exists between trap catch and wet, cool weather can be computed by determining the probability that these two sets of events occurred on the same days by chance.

The number of ways in which 4 depressed trap catches can be distributed among 12 days without respect to order is given by the binomial coefficient  $\binom{12}{4}$ . Suppose only the 3 days with greatly reduced trap catch are classified as low catch days and the other as a normal catch day. There then occurred 3 wet and cool days during which trap catch was reduced and 1 wet and cool day during which no reduction in trap catch occurred. The probability of this happening by chance is given by  $\binom{4}{3}$   $\binom{8}{1}/\binom{12}{4} = .065$ . The probability of either this event or the more extreme event that all 4 days are classified as reduced is realized is then given by  $.065 + \binom{4}{4}$   $\binom{8}{0}/\binom{12}{4} = .067$ . In contrast, the most likely single outcome would be for 1 reduced trap catch day to occur during the 4 wet and cool days and the rest to occur during the other 8 days (p=.453). Clearly, the probability that the partitioning of wet and cool days with reduced trap catch occurred by chance is very low. It is therefore likely that a relationship does exist.

The second reason for believing that the relationship between weather and trap catch is not spurious is that the model closely predicts the actual trap catch in stand 3 (fig. 9). Recall that the data from this stand was not used to parameterize the model. In this case, the initial input of pupal parasitoids was set equal to the input from stand 2 scaled by the ratio of estimated pupal densities in stands 3 and 2. The close fit of the model is not surprising considering the similarity in the trap catch from stands 2 and 3. What the relationship between the model and data from stand 3 does is reinforce the important fact that the trap catch from structurally different and spatially separated forest stands was strongly influenced by the same weather patterns.

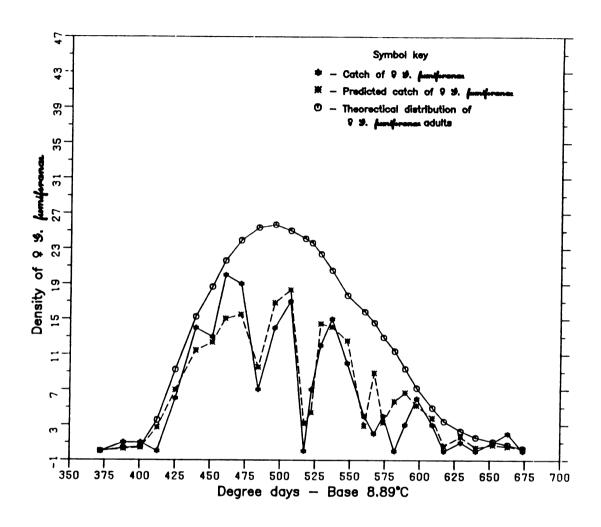


Figure 9. Predicted and actual malaise trap catch of adult female Glypta fumiferanae in stand 3 and the theoretical, relative temporal distribution of adult females. Malaise traps were located singly in 13 balsam fir trees.

We have established the fact that rainfall and cool temperatures reduce the malaise trap catch of G. fumiferanae. An important question is whether the reduced trap catch is indicative of a reduction or cessation of host searching. Based on observations of G. fumiferanae's host searching behavior made in 1981, the answer is yes. These observations will be discussed more fully later; however, two aspects are relevant here. First, very few G. fumiferanae were observed attacking hosts on cool, wet days. Second, the parasitoid frequently flew from one part of the tree to another when searching for hosts. This was especially evident when the parasitoids failed to discover hosts in a particular location. Since malaise traps are passive traps, trap catch will increase with activity of the parasitoid; in this case, activity is closely tied to host searching.

The estimated mean and variance of the lifespan of adult <u>G</u>. <u>fumiferanae</u> is given by the mean and variance of the gamma density function used to describe the physiological time delay between pupation and death of the adult parasitoid. The mean is  $\underline{PTD} = 87$  DD and the variance is  $(\underline{PTD})^2/k = 946$  DD. The mean corresponds to <u>ca</u>. 10.5 days in 1981. A similar lifespan was evident in 1980. This conclusion is based on the fact that the degree day interval during which female parasitoids were trapped in 1980 is approximately the same as that in 1981.

In the laboratory, we found that adult female  $\underline{G}$ . fumiferanae died within 3 days if they were not fed. This implies that in the natural environment, adult  $\underline{G}$ . fumiferanae do indeed feed. They do not, however, use the vegetation on the forest floor as a food source. This assertion is based on two observations. First, although many parasitoids were caught in malaise traps on the forest floor, no  $\underline{G}$ . fumiferanae were snared. Second, extensive sweep-netting of the ground

vegetation failed to reveal the presence of <u>G. fumiferanae</u>. While it is evident that the adult parasitoids do feed, they apparently do so in the forest canopy. Futhermore, forest composition appears to have no influence on the parasitoids' ability to secure food. This is founded on the equivalence of the parasitoid lifespans in stands 2 and 3, and the fact that stand 2 was exclusively balsam fir and stand 3 a mixture of balsam fir and other coniferous and deciduous trees.

<u>Laboratory Study</u>: In the laboratory, we sought to elucidate whether a temperature and light intensity gradient influenced <u>G</u>. <u>fumiferanae</u>'s host searching behavior. The study was catalyzed by the theory of optimal foraging and information which indicated that <u>G</u>. <u>fumiferanae</u> searched for hosts in what would appear to be a non-optimal fashion.

It is realistic to assume that parasitoids adopt host searching behaviors which maximize the number of hosts parasitized at the end of their lifetime. Natural selection will favor those strategies which result in a reproductively efficient distribution of the parasitoids' offspring among available hosts. Most hosts of parasitoids are distributed in a non-random fashion and often in patches of variable density. Theoretical and laboratory investigations of parasitoid host searching (Royama 1970, Cook and Hubbard 1977, Hubbard and Cook 1978, Waage 1979, Nachman 1981) and general foraging strategies (Charnov 1976; Parker and Stuart 1976; Pyke, Pullman and Charnov 1977) have led to the principle of optimal patch use. Briefly, this means that all patches of the resource are reduced to some common harvesting rate. Thus, if a parasitoid's rate of encounter with healthy hosts is to be maximized over the period of time during which hosts are available, then the optimal solution requires that when presented with host patches of different density, the parasitoid will reduce all the areas it uses to the same rate of encounter between itself and healthy hosts.

The distribution of 159 attacks by G. fumiferanae on budworm observed in the field is given in table 2. Observations were made by individuals positioned in tree crowns who recorded the activity of individual female G. fumiferanae. The length of time that a particular parasitoid was observed varied from 30 seconds to over 45 minutes. More attacks were observed in the outer portion of the tree crown than on the interior. This distribution may be biased since parasitoids in different parts of the trees may be more easily detected. However, because the observer was positioned within the tree crown, this bias should have deflated the number of attacks observed in the exterior part of the crown.

According to an optimal foraging strategy, an approximately equal distribution of attacks between the exterior and interior portions of the crown would be expected if the hosts were equally distributed in these two areas. This is not the case, though. Lewis (1960) found that only 25% of the hibernating budworm population were found on the terminal 15-inch twigs of whole branches sampled from balsam fir. He also found a trend of increasing parasitism by G. fumiferanae on the exterior of the crown. Obviously, G. fumiferanae responds to factors other than the host distribution when searching for hosts. The following experiment was conducted to determine if physical factors associated with the interior and exterior of the tree crown influenced the parasitoids' host searching behavior.

A chamber was constructed in which: (1) a temperature and light gradient could be generated and (2) parasitoids would be free to range over this environmental spectrum for hosts. The chamber was a rectangular tube of dimensions  $80 \times 10 \times 10$  cm and was manufactured from plexiglass 1.2 cm thick. On the outside walls of the chamber, 5 series of two 25 watt, 725 ohm heat dissipating resistors were mounted and wired in parallel. A controller, which

Table 2. Distribution of attacks by  $\underline{Glypta}$   $\underline{fumiferanae}$  on spruce budworm larvae in balsam fir trees (n = 16). The exterior of the tree denotes the first 18 inches of the canopy.

Location of Attack								
EXTERIOR					INTERIOR			
Bud Tip	Lichen	Bark Scale or Branch Fork	Flower Bract or Needle Base	Bud Tip	Lichen	Bark Scale or Branch Fork	Flower Bract or Needle Base	
	_		ATTA	CKS				
39	13	14	27	3	21	35	10	
Total		93			(	69		
		57%			43%			

produced pulses of electricity whose duration were regulated by potentiometers, was connected to each set of resistors. Pulses of different durations were distributed among the five sets of resistors, resulting in differential warming of the chamber. A circuit diagram for the controller is provided in appendix 2.

The chamber was placed in a constantly cooled (10°C) and darkened room and illuminated from above by two 30 watt flourescent bulbs. The experiments were conducted in the cooled room in order to obtain the desired temperatures within the chamber and so that the chamber would not have to be recalibrated due to changes in room temperature. The light intensity in selected portions of the chamber was regulated by placing a piece of Saran\* 20-mesh screening over these sections.

G. fumiferanae pupae were collected and males and females reared at 22°C and a 14 hour light/8 hour dark photoperiod. To provide hosts for G. fumiferanae, budworm pupae were collected and reared, and adult males and females were placed in small paper bags on which the females oviposited. Eggs were collected and placed in a petri dish which was then covered with gauze and Parafilm. The dishes were placed in a paper sleeve in which an opening was cut to allow light to pass through the gauze and Parafilm. When the eggs hatched, the photopositive larvae crawled onto the gauze and spun hibernaculae.

Parasitoids used in the experiment were between 3 and 6 days old. This age range was imposed to avoid confounding results with age-dependent behavior. In addition, all experiments were conducted between 1100 and 1400 hours to eliminate the possibility of diel periodicity as a source of variation. Ideally, the influence of temperature and light on the behavior of female <u>G. fumiferanae</u> should have been studied independently and then jointly. However, the short time in which adult G. fumiferanae were available precluded such an approach.

A temperature and light gradient was arranged so that there was a fully illuminated warm environment, a shaded cool environment, and a spectrum between these limits (table 3). Four patches of gauze, each with five budworm hibernaculae, were placed in two warm and light regions and two cool and shaded No hibernaculae were placed in the center cell into which female parasitoids were introduced. After the parasitoids moved at least one cell away from the entry point, their behavior was recorded for 1 to 1-1/2 hours. This consisted of recording the time intervals each parasitoid spent in each environment searching for hosts, attacking hosts, and resting. The number of ovipositions in each cell was also recorded. Searching behavior consisted of G. fumiferanae probing and examining the gauze and surrounding areas with its antennae in an effort to locate hosts. Attack behavior consisted of probing with the ovipositor and oviposition. It was possible to determine when oviposition occurred, as the females, upon inserting the ovipositor in the host, remained completely motionless for ca. 15 seconds. Resting behavior consisted of immobility, preening, and non-directed movement.

The average total time spent, average time spent attacking hosts, and average time spent not attacking hosts in each of the different environmental conditions is given in table 3. Attack time consisted of both attack and searching behavior, when searching behavior occurred between attacks separated by a short time interval. Attack time for the two warm-light and cool-shaded environments were summed. This was done because few attacks occurred in the cells at the extreme end of the chamber and this result cannot be attributed to environmental conditions. It is just as likely that most attacks occurred in the cells closest to the entry because this is where hosts were first encountered.

Table 3. Total time and time spent in attack and non-attack behavior by Glypta fumiferanae in different environmental regimes. Time is in minutes and tenths of minutes. The variable W is the number of cases (C<sub>i</sub>) in which time spent in cell 4 is greater than that in cell 2 when parasitoids were observed in either of these cells. ♦ is the probability that w ≤ W given that W/∑C<sub>i</sub> = 0.50. Attack times in cells 1 and 2 and in 4 and 5 are summed.

		Environ	mental	Conditi	lons		
Cell	1	2	3	4	5		
Temperature <sup>O</sup> C	31	27	22	19	15		
Lighting		F U	LL/	P A R	TIAL		
Hosts present	yes	yes	no	yes	yes	$x_i$	ф
Total time n = 23							
Mean	8.29	26.76	11.49	7.24	5.16		
Std. Dev.	13.78	21.00	12.90	17.00	13.43		
W				4		23	.001
Non-attack time n = 21							
Mean	7.81	13.37	11.49	1.68	2.37		
Std. Dev.	13.93	12.86	12.90	3.16	7.15		
W				4		21	.004
Attack time n = 14							
Mean	28.05 12.77						
Std. Dev.	23.05 19.53		. 53				
W				5		14	.212

Analysis of the data is confined to observations from cells 2 and 4, except for attack time in which the data is summed for cells 1 and 2 and for cells 4 and 5. Reasons for this restriction are as follows: The center cell (3) was not included because hosts were not placed in this region. The cells at the ends of the chamber (1 and 5) were not included since the conditional probabilities for a parasite to enter these cells, regardless of environmental influences, are not the same as those for cells 2 and 4. This is a result of the linear arrangement of the environments.

Since independence and normality assumptions could not be met, the following method of analysis was used. Let T<sub>i</sub> be the time a parasitoid spends in cell i. Let v = 1 if  $T_2 > T_4$  and v = 0 if  $T_4 < T_2$ . Define W as  $\Sigma v$ . Then W is distributed as a binomial random variable. If the time spent in cells 2 and 4 were the same, then the parameter p of this binomial distribution would equal 0.5. The probabilities of obtaining the observed or more extreme values of W with the hypothesis that p equals 0.5 are given in table 3. These probabilities are very low for total time and non-attack time; hence, the null hypothesis is rejected. The value of a more extreme value of W for attack time has approximately a 1 in 5 chance of occurring, given p = 0.5. Hence, the null hypothesis cannot be outright rejected; however, there is an indication that differences exist. A more definitive analysis of the allocation of attacks can be made by considering the distribution of ovipositions. In cells 1 and 2, 258 ovipositions were recorded; in cells 4 and 5, 154 were recorded. If temperature and light produced no effect on the oviposition distribution, the proportion of attacks in cells 1 and 2 should be 0.5. The actual proportion was significantly different from this value (normal approximation,  $\alpha < .001$ ).

The data suggests that the distribution of attacks by <u>G. fumiferanae</u> observed in the tree are a result of the light and/or temperature differential which exists between the interior and exterior of the tree crown. If temperature does play a role, it is due to increased radiant heat since only a very small ambient temperature differential would exist between the interior and exterior of the crown.

<u>Historical Data</u>: Based on the field studies of <u>G</u>. <u>fumiferanae</u>'s activity patterns, we concluded that wet and cool weather would inhibit the parasitoid's searching efficiency. In this section, this relationship is explored further by examining historical data on the interaction between the parasitoid and its host.

Extensive data on this interaction was collected from 1950 to 1958 during the Green River Project in northwestern New Brunswick. Details of the study can be found in Morris (1963). This data has previously been analyzed by Miller (1960) who also thought that rainfall might influence the attack efficiency of G. fumiferanae. He did not, however, examine this in detail. Furthermore, Miller employed a specific model to describe the attack dynamics of the parasitoid. In contrast, we have analyzed the Green River data without the encumbrance of a particular parasitoid attack model by exploring the relationship between the ratio of hosts to parasitoids (H/P), rainfall, and percentage of hosts parasitized. In this case, this is a better approach since no a priori assumptions about parasitoid searching behavior are necessary. The influence of temperature on attack efficiency was not explored because the data lacked the resolution necessary to evaluate this factor.

Original data on spruce budworm densities and on the rate of budworm parasitism by G. fumiferanae in the years 1950 to 1958 at the Green River field

station was obtained from the Maritime's Forest Research Center, Fredericton, New Brunswick. The data from 3 plots (G2, G4, G5) were selected for use in the analysis based on the plots having a data series spanning 5 or more years, having budworm densities relatively constant, and having no appreciable tree mortality. The latter two criteria were established to minimize the possible influence of changes in the host's density and distribution and of changes in the forest environment on the parasitoid's searching behavior.

Parasitism rates had been determined by dissecting samples of third and fourth instar budworm. These rates were employed as an index of the parasitism of second instar budworm, which is the host stage G. fumiferanae attacks. The use of this index, however, does introduce a potentially large bias into the estimates of parasitism. Between the second and third instars, budworm undergo dispersal during which a large number of insects die. The propensity for dispersal is lower for parasitized larvae than for unparasitized ones (Lewis 1960). As a result, depending on the differences between the dispersal rates of parasitized and unparasitized larvae, assessment of parasitism in the spring will be inflated over the true parasitism rate. This would not be a problem if this inflation did not vary from year to year. Unfortunately, such an assumption cannot be met.

Two factors influence the proportion of third instars lost during dispersal: stand composition and weather (Kemp et al. 1980). Stand composition did not change in the plots from which the data was collected. Weather, on the other hand, was a dynamic variable. Results from a simulation model indicate that the proportion of third instars lost during dispersal might vary by as much as a factor of 2 between years as a result of weather conditions during the dispersal period (Kemp et al. 1980). The net result is that the apparent parasitism rate may

change dramatically from one year to another while the actual rate changes little. There is no way to determine this bias; however, one must be cognizant of the potential error.

An index of the density per 10 m<sup>2</sup> foliage of female <u>G</u>. <u>fumiferanae</u> was computed by first assuming that the survival of the parasitoid from the larval stage located in the budworm to the adult stage was time invariant and that the sex ratio of <u>G</u>. <u>fumiferanae</u> was also time invariant. The index was then determined by multiplying the density of third and fourth instar budworm by one-half the parasitism rate.

The index of host density (second instars) was based on egg counts less egg mortality and a constant dispersal loss of first instars. An assumption of a constant first instar dispersal loss is supported by references contained in and the simulation results obtained by Kemp et al. (1980).

To determine rainfall during the flight period of the females, the appproximate calendar dates over which this activity occurred were determined by assuming that the occurrence of the female parasitoids flight activity with respect to degree days would be the same in northwestern New Brunswick as it was in Michigan. The time period of activity in Michigan was determined from the malaise trap catches of 1980 and 1981. Minimum and maximum degree day values for which females were caught for these two years were 388.44 and 662.88. However, the magnitude of activity was not uniform over this interval. Hence, total precipitation for the interval 428 DD - 584 DD was computed since it was during this period that peak activity occurred. The dates and precipitation for this interval during the years 1950 to 1957 in the Green River area are given in table 4.

Table 4. Julian dates for the degree day interval 428 - 584 (base 8.89°C) and precipitation during this period in the years 1950 to 1957 at the Green River field station, New Brunswick.

Year	Julian Dates	Precipitation in inches
1950	215 - 244	2.87
1951	216 - 240	3.07
1952	203 - 221	2.57
1953	204 - 232	2.00
1954	214 - 241	5.87
1955	203 - 225	2.79
1956	221 - 248	3.00
1957	217 - 245	0.70

The data from each plot is presented in figure 10. Note that the percentage of hosts attacked in year i is plotted at year i + 1. This was done to preserve the temporal sequence in which the data was collected; the parasitoid searched for and attacked hosts in late summer while the assessment of this attack was completed in early spring the following years. Of particular interest is the year 1954. During this year the host density declined, the parasitoid density increased, and parasitism declined dramatically in all three plots. This decline in the parasitism rate is contradictory to what would be expected. However, during 1954, more precipitation fell than in any other year. Because the decline in parasitism was evident in all three geographically separated plots, it is logical to conclude that the decline was in fact related to inordinate rainfall.

A more complete picture of the relationship between rainfall and parasitism is obtained by using the data from all 3 plots to generate a parasitism surface as a function of the H/P ratio and rainfall (fig. 11). The pronounced influence of rainfall on the parasitism rate is readily apparent at both high and low H/P ratios. There are, however, two unexplainable irregularities in the parasitism surface. With moderate rainfall and high H/P ratios, a distinct valley exists when the parasitism rate should either be nondecreasing or nonincreasing with respect to the independent variables. There is also a lack of response in the parasitism rate to a decline in the H/P ratio when associated with a moderate to high rainfall. It must be remembered, though, that the data used in the analysis is subject to a great deal of potential bias. This bias may be due to the affect of spring dispersal on the measured parasitism rate and also due to year-to-year differences in the temporal sychrony between G. fumiferance and the budworm

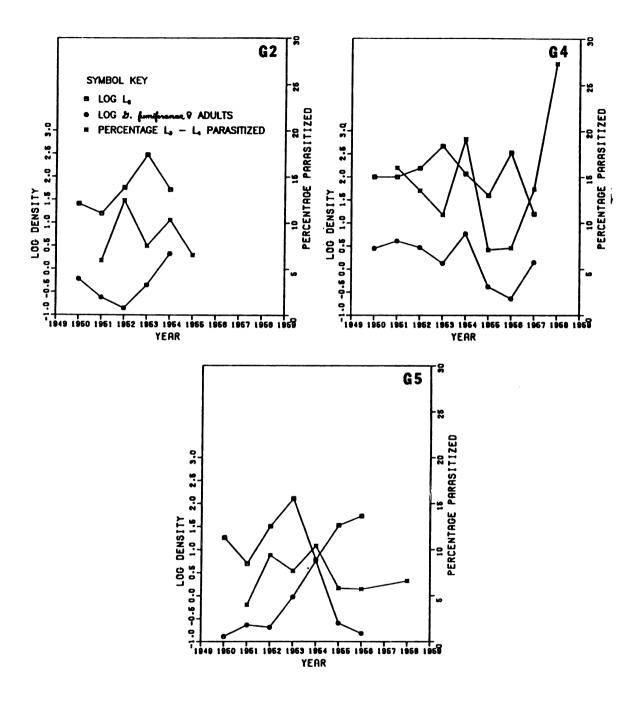


Figure 10. Temporal relationships between the density per  $10 \text{ m}^2$  foliage of second instar  $(L_2)$  spruce budworm, the density of adult female Glypta fumiferanae and the percentage of third and fourth instar  $(L_3-L_4)$  budworm parasitized by G. fumiferanae in three plots at the Green River field station, New Brunswick. (Data obtained form Maritimes Forest Research Center Frederiction, New Brunswick)

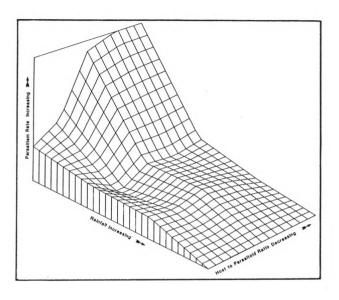


Figure 11. Parasitism of spruce budworm larvae by Glypta fumiferanae as a function of the ratio of hosts to parasitoids and rainfall during the period of adult parasitoid activity.

host. In addition, a large amount of sampling variability must be present, although information on this variability is unavailable. Finally, temperature has not been included as an independent variable even though it has been shown to influence the activity of the female parasitoid. It is somewhat surpising that any discernable relationship between rainfall and parasitism was produced. In fact, in order to be visible, the relationship must be quite strong. When this relationship is combined with the observation that rainfall impedes host searching by the parasitoid, a conclusion can be drawn that excessive rainfall greatly diminishes the attack efficiency of the parasitoid.

## Discussion

Factors other than the host which influence the number and distribution of hosts attacked by a parasitoid may be classified as being either controllable or uncontrollable. Controllable factors are those which can be ameliorated or accentuated by management actions. In this study we have identified two ways in which uncontrollable factors affect G. fumiferanae. First, the attack efficiency of the parasitoid is greatly diminished by excessive rainfall and/or cool temperatures. This is because host searching by the parasitoid occurs at a reduced rate during these weather conditions. Second, the spatial pattern of host searching by G. fumiferanae is influenced by light intensity and/or temperature. Similar types of phenomena have been observed for other parasitoids.

Temperature, sunlight, rainfall, and dew have been reported as influencing the ability of parasitoids to locate and parasitize hosts. Klomp (1959) found that the attack efficiency of the tachinid parasitoid, <u>Carelia obesa</u>, was correlated with the amount of sunshine during the period of egg deposition. Burnett (1951,

1954, 1958) demonstrated that as the temperature declined, the number of sawfly pupae attacked by a chalcid parasitoid decreased. This decrease was attributable to a decline in the capacity of the parasitoid for oviposition and also to a reduction of the number of hosts contacted by the insect. In field experiments, he noted that parasitism was inhibited by rain and heavy dew. Messenger (1968) found that temperature extremes affected the aphid parasitoid, <a href="Praon exsoletum">Praon exsoletum</a>, by inducing a shift in the proportion of ovipositional activities relative to other activities engaged in by the females, and by reducing the proportion of successful attacks. In laboratory studies of <a href="Bracon mellitor">Bracon mellitor</a>, Barfield <a href="Eact al">et al</a>. (1977) found that the parasitoid exhibited a narrow temperature range in which total fecundity was maximized.

The spatial interplay between parasitoid host searching and abiotic factors has also been observed before. Weseloh (1976) reviews a number of studies which indicate that responses to humidity, light, and temperature are at least partially responsible for the spatial distribution of parasitoids in forest habitats. Munster-Swendsen (1980) has suggested that the aggregation of the parasitoid Apanteles tedellae with respect to its host may be explained by an evolution of a similar preference for physical conditions as opposed to a direct response to host densities.

An important factor influencing adult parasitoids and, one that may be controllable, is the availability of nutrients and moisture. These resources are essential for the survival of many adult parasitoids (Townes 1958, Hassan 1967). We found no evidence that a particular habitat was a better source of nutrients or moisture for <u>G</u>. <u>fumiferanae</u>. Examples exist, however, which demonstrate the importance of habitat in providing food sources for adult parasitoids (e.g., Syme 1975, Calderon 1977).

The importance of abiotic and biotic factors other than the host in determining the number and distribution of hosts attacked is clear. As a result, it is desirable to know, at least in a qualitative manner, how these factors influence the dynamics of parasitoid-host systems and the capacity of a parasitoid to act as a biological control agent. One way of elucidating these influences is to incorporate these factors, in a general way, in the mathematical models used to describe and study these systems. This is done below.

The dynamics of coupled parasitoid-host systems with discrete generations have been extensively analyzed with the following model:

$$H_{t+1} = H_t F(H_t) f(H_t, P_t)$$
 (2)

$$P_{t+1} = H_t(1-f(H_t, P_t))$$

Because the basic structure of this model corresponds to many real-world systems, it will serve as a basis for the discussion. In (2),  $H_t$  and  $P_t$  are host and parasitoid densities,  $F(H_t)$  is the net rate of increase of the host in the absence of  $P_t$ , and  $f(H_t, P_t)$  is a parasitoid attack function. This function can be interpreted as the probability of  $H_t$  hosts escaping discovery and subsequent parasitization by  $P_t$  parasitoids.

Model (2) is nonlinear. As a result, analytical analysis is generally limited to determination of the equilibrium states and stability properties of these states. Equilibrium states are values of state variables which are either constant or are periodic through time. Periodic equilibrium states are called limit cycles. In (2), the state variables are the host and parasitoid densities. An equilibrium point or cycle is stable if, when a state variable is perturbed from its equilibrium, system dynamics drive the variable back to or asymptotically close to this state. An equilibria is locally stable if the magnitude of the perturbation

is restricted to some neighborhood about the equilibrium. It is globally stable if such a restriction is not necessary.

Limiting the analysis of (2) to determination of equilibria and stability of these equilibria is not overly restrictive because these properties convey, within the context of (2), the salient features of a successful biological control agent. These features are that a parasitoid can depress and/or help maintain a host population at a density well below the host's carrying capacity. This does not imply that the parasitoid must singularly be able to do this, since  $F(H_t)$  may contain complex density dependent relations. What is necessary is that all mortality factors acting in combination are able to do so, and that in the absence of the parasitoid, regulation would not be possible or would not be as strong. The aim here is to examine the ties between factors which influence adult parasitoids in their search for hosts and the equilibrium states and stability properties of (2). This will be done by first relating the empirical observations to forms of (2) for which equilibria and stability have been laid bare. An analysis is then made of a variation of (2) which provides a different insight into the importance of factors influencing adult parasitoid dynamics.

A sufficient condition for stability in simple one parasitoid-one host models is that spatial or temporal incoincidence (or both) between the host and parasite occur<sup>1</sup> (Hassel 1978, Nachman and Munster-Swendsen 1978). A stable system is

Stability can also be generated if: (1) F(H<sub>t</sub>) is a nonlinear function such that the host is limited by the carrying capacity of its habitat (Beddington et al. 1975) or (2) the host and parasitoid populations are comprised of cells and dispersal of hosts and parasitoids occurs among these cells (Crowley 1981 and references therein). However, (1) results in host equilibrium densities which are unrealistically high (Beddington et al. 1978), and (2) might be considered as a component of spatial and/or temporal incoincidence.

generated when either of these events is realized at appropriate intensities. Stability transpires because the parasitoids do not over-exploit the host resource. Excessive incoincidence leads to runaway host populations while too little incoincidence leads to diverging oscillations in the host and parasitoid densities. Here, attention is restricted to spatial incoincidence.

Two factors lead to spatial incoincidence: (1) contagiously distributed hosts, and (2) parasitoids which forage in response to this patchy distribution and possibly also in response to other biotic and abiotic factors. The essential aspects of spatial incoincidence can be captured in (2) by incorporating non-random host searching by the parasitoid. This has been done a number of ways (Hassel and May 1974, Murdoch and Oaten 1975, Munster-Swendsen 1982). A detail independent and analytically tractable method first proposed by Griffiths and Holling (1969) and fully developed by May (1978) is used here.

May proved that the zero term of the negative binomial distribution

$$(1 - aP/k)^{-k}$$
 (3)

could be used to model f(H,P) when the hosts are patchily distributed and the parasitoids search randomly (i.e., do not discriminate previously attacked hosts) within a particular host patch. This is a descriptive model unlike other models which have been derived from the geometrical attributes of parasitism (Royama 1971). Nonetheless, it is adequate for our purposes because we are only interested in being able to describe the outcome of the parasitoids' attack.

In (3), the parameter (a) is an index of the searching efficiency of a parasitoid within a patch and k is an index of its aggregation with respect to the host distribution. Note that an unrealistic linear relationship between the number of hosts available and number attacked is assumed. In general, the

number of hosts attacked by a constant density of parasitoids must reach an asymptotic limit as host density increases (Holling 1959, Royama 1970). For the present, this additional complexity is omitted, although it will be discussed later.

May has shown that a sufficient condition for stability of an equilibrium of (2) employing (3) and allowing  $F(H_t)$  to be a constant is k < 1. This means that a sufficient degree of aggregation must be realized for stability. Searching efficiency (a) has no influence on the stability of the equilibrium in the one parasitoid-one host model. It does, however, scale the equilibrium values.

Factors such as temperature, rainfall, and adult food sources which influence the searching efficiency of a parasitoid can be incorporated in (2) through the parameter (a). For instance, if adverse weather causes a reduction or cessation of host searching activity, this can be incorporated by writing (a) as (a't) where a' is an instantaneous index of searching efficiency and t is the amount of time favorable weather occurs. Likewise, those factors which influence the spatial distribution of parasitoids can be partially described in terms of k. They can only be partially described since k can only incorporate the influence of factors up to the point where the spatial distribution of parasitoid attack is random and because a single statistical parameter can not capture all aspects of a complex spatial pattern. Nonetheless, this simple model indicates that it is essential that parasitoid aggregation with respect to the host distribution not be interrupted by extrinsic factors if the parasitoid is to be an effective biological control agent. The model also indicates that factors which influence the searching efficiency of the parasitoid cannot alter system stability but simply influence the equilbrium densities. Similar conclusions can be made for all single parasitoid-single host models which incorporate non-random parasitoid search.

An important question is whether these models are representative of any real-world situation. We feel that they may describe cases of introduced pests and exotic parasitoids. Additional complexity is needed, though, to describe other, more common parasitoid-host systems. As demonstrated below, these added components dramatically alter the effect of factors that influence adult parasitoids on the stability properties of a particular system.

Populations of many insects exist for long periods of time at low densities and periodically erupt to high densities. This is most noticeable in temporally continous and undisturbed ecosystems but may also occur in agro-ecosystems. The underlying mechanism of such phenomena is density dependent mortality which at first becomes more effective with increasing densities but whose effectiveness then diminishes beyond some threshold density. This model, often called a double equilibria population process, was proposed as an explanation of population dynamics by Ricker (1954), Morris (1963), and Takahashi (1964), and has been extensively reviewed by Holling (1973), May (1976), Royama (1977), and Berryman (1978). The salient features of the models are two equilibrium points or cycles, one at low densities which is locally stable, the other at higher densities which is unstable. A population will reside in the low density state until moved beyond the unstable point or until the unstable point is itself moved below the existing population density. Because these types of population processes are likely to be quite common, we have analyzed a form of (2) which incorporates such processes along with a parasitoid.

Consider a situation where a host population is limited by a group of factors which, when acting in combination, produce a double equilibrium population process. To add substance, let this group consist of predators whose density

is constant and which display a sigmoidal functional response and a parasitoid. The predation can be incorporated in (2) by writing  $F(H_{+})$  as:

$$\delta \exp \left[ \left( -\gamma P_b H_t \right) / \left( \alpha^2 + H_t^2 \right) \right]$$
 (4)

The parameter ( $\delta$ ) in (4) is the net rate of increase of H in the absence of parasitism and predation by  $P_b$  predators. The exponential function describes the probability of H prey escaping predation by these predators. It is based on Real's (1977) extension of models of enzyme kinetics for describing functional responses. The parameter ( $\gamma$ ) is the maximum number of prey captured per predator, and  $\alpha$  is the density of prey at which the predator's consumption level is  $1/2 \gamma$ .

The model obtained by incorporating (4) and (3) in (2) is:

$$H_{t+1} = H_t \delta \exp \left[-\gamma P_b H_t / (\alpha^2 + H_t^2)\right] (1 + aP/k)^{-k}$$
 (5)

$$P_{t+1} = H_t [1 - (1 + aP/k)^{-k}]$$

Predation is assumed to occur after parasitism and is complementary.

In order to analyze (5), numerical procedures had to be adopted. These procedures consisted of: (1) evaluating equilibrium densities for a particular set of parameters via Newton's method for determining the roots of a function, (2) ascertaining the stability of these equilibria, and (3) determining a region of stability about each equilibrium point by iteratively perturbing equilibria and simulating system dynamics. Because numerical methods were necessary, it was impossible to completely elucidate model behavior. However, the results do indicate its qualitative properties.

Suppose that the predators by themselves are incapable of regulating the prey population. That is, the net recruitment curve in the absence of parasitism is greater than one for all prey densities (fig. 12). By incorporating a parasitoid

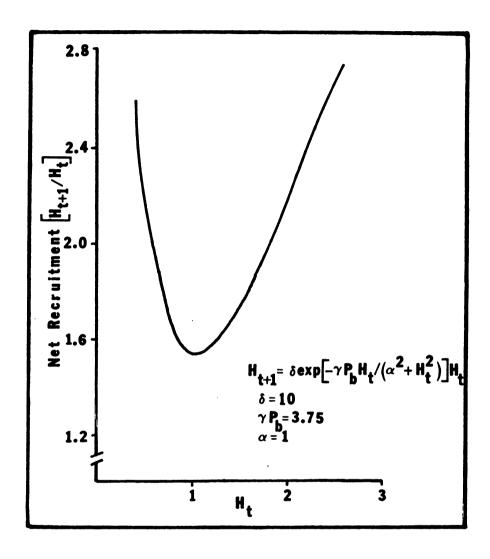


Figure 12. Net recruitment for an insect population preyed upon by a constant density of predators which exhibit a sigmoidal functional response. H is the insect density,  $P_b$  is the predator density,  $\delta$  is the rate of increase of H,  $\gamma$  is the maximum number of H consumed by a predator and  $\alpha$  is the density of H for which predator consumption is  $\gamma/2$ .

with an appropriate searching efficiency, a stable, low-density equilibrium can be generated. The requirement placed on the searching efficiency is that the host equilibrium falls in a particular region of the cusp of the recruitment curve.

The equilibrium densities of the parasitoid and host and eigenvalues of model (5) linearized about the equilibrium points for  $\delta = 10$ ,  $\gamma P_b = 3.75$ ,  $\alpha = 1.0$ , and various values of parasitoid searching efficiency and aggregation are presented in table 5. The methodology for obtaining these values is outlined in appendix 3. The eigenvalues indicate which equilibria are locally stable, since a necessary and sufficient condition for local stability of an equilibrium is that the real part of the eigenvalue has modulus less than 1.

The most noticeable aspect of this model is that aggregation is no longer necessary for local stability. Even with random parasitoid search  $(k = \infty)$ , stable equilibria exist. Moreover, one measure of system resilience, the speed with which perturbed populations return to the vicinity of an equilibrium, is higher with less aggregated parasitoid search. System resilience is the ability of a system to absorb changes in state variables or parameters without abrupt changes in system trajectory. Here, system resilience relates to the host population remaining in an endemic phase. A system will be more resilient with a faster return time since it will be less likely that disturbing effects in year i and then again in year i + 2 will induce instability. The speed with which perturbed populations return to the vicinity of an equilibrium is expressed in terms of the largest eigenvalue  $(\lambda)$  of the linearized model by  $1/(1-|\lambda|)$  for  $|\lambda|$  < 1 (Beddington, Free, and Lawton 1976). With strong parasitoid aggregation (k = 0.75), an equilibrium was stable or no equilibrium existed  $(\alpha = 1.0)$ . Nonexistence of an equilibrium was first indicated by Newton's method; however, because this

Table 5. Equilibrium points and eigenvalues of the following parasitoid-

host model linearized about these points.

H =  $\delta$ H exp( $-\gamma$ P H /( $\alpha$ +H<sup>2</sup>))(1+aP /k) -k

Pt+1 = H (1-(1+aP /k) -k)

H is the host density, P is the parasitoid density, P, is the density of background predators,  $\delta$  is the rate of increase of H,  $\gamma$  is the maximum number of H consumed per predator,  $\alpha$  is the density of H for which predator consumption is  $\gamma/2$ , (a) is a measure of the parasitoid's searching efficiency with efficiency increasing with larger (a) and K is a measure of the parasitoid's aggregation with respect to host density with low k representing strong aggregation. Values for parameters not given below are:  $\delta = 10.0, \gamma P_1 = 3.75, \alpha = 1.0.$ 

	Equilibr	Eigenvalues		
<u>a</u>	H	P		
	<u>k =</u>		<b></b>	
3.0	.661	.290	.420 ± .627i	
2.5	.734	.295	.509 🕇 .6081	
2.0	.852	.310	.509 ± .608i .650 ± .580i	
1.5	1.120	.399	.907 ± .490i	
1.0	*	*		
	<u>k =</u>	2.0	1	
3.0	.558	.283	.371 ± .743i	
2.5	.627	.288	.444 ± .705i	
2.0	.730	.295	.444 ± .705i .565 ± .666i .780 ± .622i	
1.5	.918	.323	.780 ± .622i	
1.0	1.539	.685	1.1664791	
	<u>k =</u>	6.0	1	
3.0	.513	.277	.365 ± .817i .429 ± .763i	
2.5	.582	.284	.429 🚡 .763i	
2.0	.683	.291	.539 🛨 .7081	
1.5	.859	.311	.539 ± .708i .742 ± .657i	
1.0	1.325	.521	1.117 ± .580i	
	k =	∞ **		
3.0	.489	.274	.363 + .3861	
2.5	.558	.283	.425 ± .362i .529 ± .353i	
2.0	.659	.290	.529 🛨 .3531	
1.5	.831	.307	.726 ± .370i 1.096 ± .356i	
1.0	1.256	.475	1.096 ± .3561	

<sup>\*</sup> No equilibrium point found 
\*\* When  $k=\infty$  (1 +  $aP_t/k$ ) 
\*\* exp(- $aP_t$ ) and parasitoid search is

procedure may not converge to a root of a function, extensive simulations were also employed. The simulations failed to uncover either an equilibrium or a limit cycle. It was also observed that as aggregation increased, the searching efficiency where equilibria ceased to exist became larger (i.e., for k = 0.5, no equilibrium could be found for a = 1.5).

The eigenvalue analysis of the linearized model provides an initial indication of the stability properties of the system. It is, however, of limited usefulness because it only pertains to the immediate neighborhood about an equilibrium point. Real-world populations are not restricted to "small" changes, and local stability analysis may be quite misleading because parameters may produce local stability; however, the region about the stable point where the model remains stable may be extremely small. In order to determine the global stability properties of (5), simulation was used to outline the boundaries of the host and parasitoid densities for which the system remained stable.

The influence of decreasing parasitoid aggregation on the stability region (i.e., the set of all host and parasitoid densities for which stability is maintained) of the model is portrayed in figure 13. With k = 0.75 and k = 2.0, the stability region is not closed. This is a result of truncating the analysis at a particular parasitoid density. The region would eventually close with further increases in parasitoid numbers. It is clear that while parasitoid aggregation is not essential, it does improve the system's resilience to perturbation in host or parasitoid numbers. The effect of changes in searching efficiency on the stability region is illustrated for two levels of parasitoid aggregation in figure 14. As with the more simple models, the equilibrium host density increased, though only slightly, with a decline in searching efficiency. More importantly, the stability region became much smaller.

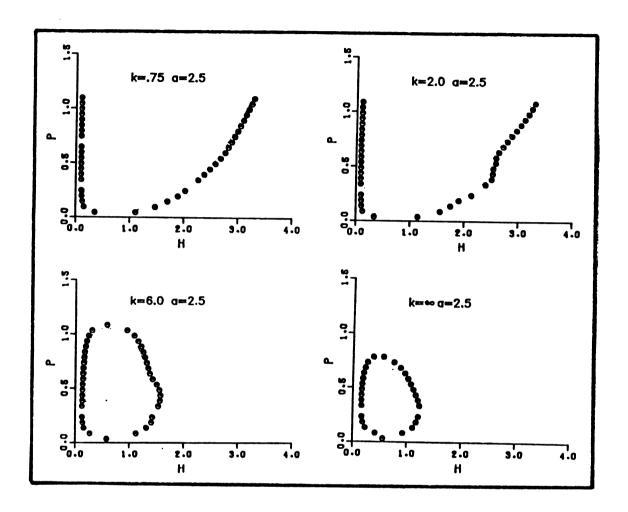
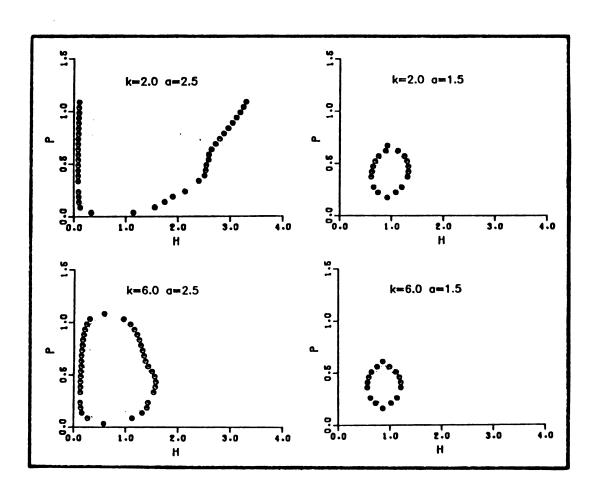


Figure 13. Influence of parasitoid aggregation with respect to host density on the stability region of the following parasitoid-host model:

 $H_{t+1} = H_t \delta \exp(-\gamma P_b H_t / (\alpha^2 + H_t^2)) (1 + a P_t / k)^{-k}$   $P_{t+1} = H_t (1 - (1 + a P_t / k)^{-k})$ 

The stability region is indicated by the shaded areas. k is a measure of parasitoid aggregation with low k reflecting strong aggregation and (a) is a measure of parasitoid searching efficiency. H is the host density, P is the parasitoid density,  $P_b$  is the density of background predators,  $\delta$  is the rate of increase of H,  $\gamma$  is the maximum number of H consumed per  $P_b$  and  $\alpha$  is the density of H for which predator consumption is  $\gamma/2$ . Here,  $\gamma P_b = 3.75$ ,  $\alpha = 1.0$  and  $\delta = 10.0$ .



The implications of the empirical observation are far different when viewed in the context of this model as opposed to the simpler one parasitoid-one host models. In this case, in addition to parasitoid aggregation, the attack efficiency of the parasitoid is of paramount importance in establishing a stable and resilient system. This is not a new revelation, since biological control scientists have long emphasized the importance of host searching (Huffaker et al. 1976). Factors which influence the searching efficiency of a parasitoid or group of parasitoids now do not simply determine an equilibrium level but also help determine system stability. As a result, determination of controllable and uncontrollable factors which influence parasitoid-host searching are important aspects of parasitoid management. Aggregation is still a valuable property of a parasitoid from the biological control point of view. However, parasitoids that do not aggregate can still be important natural enemies, and improved pest control can be obtained from management actions taken to improve their searching efficiency.

This is important because empirical data suggests that more times than not, parasitoids fail to significantly aggregate in areas of high host density (Morrison and Strong 1980). These observations may be explained three ways. First, as indicated by our study of G. fumiferanae and studies of other parasitoids, the spatial distribution of these insects is influenced by factors other than host numbers. Second, as discussed by Royama (1970), parasitoids may respond to areas of higher host densities but only in a limited capacity due to a less advanced nervous system. When aggregation is measured in terms of parasitism, slight aggregation will result in constant or declining parasitism when expressed as a function of host density. Finally, these observations may reflect

an inappropriate scale for measuring aggregation since it is well known that the distribution of a population of samples is a function of the size of the sample unit. For instance, G. fumiferanae does not aggregate with respect to the host within a tree but may do so between trees. In passing, we note that this was investigated in our study; however, no relationship was found between host numbers and trap catch in different trees. Further studies of parasitoid behavior, of spatial variation in the distribution of adult parasitoids, and of the spatial variation in parasitism intensity are needed to fully answer the important questions of parasitoid aggregation and thereby determine if aggregation can be manipulated through habitat management.

The results obtained with (5) are not restricted to the particular case examined, since the results will hold for any set of mortality factors which produce a net recruitment curve similar to that portrayed in figure 12. Furthermore, these mortality factors may in themselves produce a stable equilibrium, in which case the parasitoid serves to enlarge the stability region about an equilibrium. The inclusion of a limited capacity for parasitism by each parasitoid so that the number of hosts parasitized does not increase indefinitely as the number of hosts increases will not alter the general results obtained. The affect of inclusion of this limitation will be a contraction of the stability region, the size of which will be dependent on the density at which limitation occurs.

## Conclusions

In this paper, we have presented data which indicate that: (1) the searching efficiency of <u>G</u>. <u>fumiferanae</u> is reduced in wet and cool weather, (2) habitat does not influence the parasitoids' ability to obtain nutrients and moisture, and (3) the spatial distribution of the adult female parasitoid and of

the ovipositions made by these insects within a tree crown is influenced by temperature and light. These results and those obtained by a number of other authors point to the fact that abiotic factors and biotic factors other than the host play an important role in determining the number and distribution of hosts attacked by a parasitoid. This, in turn, may have a pronounced affect on the ability of a parasitoid to act as a biological control agent. In some cases, the factors which influence the intensity and distribution of the parasitoids' searching activity will be amenable to manipulation and should then serve as an object of control for parasitoid management. However, identification of these factors requires detailed studies of the temporal and spatial dynamics of the adult parasitoids.

Forest Harvesting and the Spruce Budworm

#### Introduction

The eastern spruce budworm is native to North America and is distributed throughout the northern boreal forest from Alberta to Maine and the Maritimes. The preferred hosts are balsam fir (Abies balsamea (L.) Mill.), white spruce (Picea glauca (Muench) Voss), red spruce (P. rubens Sarg.), and to a lesser extent, black spruce (P. mariana (Mill.) B.S.P.) (Greenbank 1963). Defoliation of balsam fir and spruce by spruce budworm occurs at periodic intervals. Depending upon defoliation intensity, the death of individual trees or whole stands can occur. Spruce and fir are valued for paper production and lumber. Budworm outbreaks often kill more trees than can be used rapidly. In addition, a future shortage of fiber can result from extensive budworm-induced tree mortality.

Outbreaks of the budworm occur at 20 to 40 year intervals. The outbreaks themselves may extend only 8-10 years, but the effects are longer lasting. In the past, especially in the East, large-scale spraying programs minimized the budworm damage. However, concerns have been voiced about the effectiveness and environmental impact of the spray programs. In the Lake States, recent supplies of spruce and fir have exceeded demand. As a result, few aerial sprayings of forests were conducted. Instead, greater emphasis was placed on cutting to salvage damaged material or to presalvage stands.

Management techniques are needed that take demand for fiber into account, take commercial management objectives into account, and provide resistance to spruce budworm. Without such management techniques, the future will hold the same scenario that has developed during recent budworm outbreaks; a large acreage of killed timber with no hope of marketing all of the product.

There are several techniques that can be used. These techniques can be grouped into three general approaches: (1) those which are directed at the forest, (2) those which are directed at the insect, and (3) those which combine (1) and (2).

# GROUP I.

The first group of approaches, those directed at the forest, largely involve the way the forest is harvested or the timing of harvest. The following illustrate what is meant:

- 1) No Action This is the deliberate avoidance of any action by man. This approach is appropriate where the cost of utilizing a particular stand is greater than the value of the product it will yield. Stands categorized as beyond salvage fall into this category.
- 2) Salvage This is the marking, selling, and harvesting of stands which are in the process of dying but have not deteriorated too far to be uneconomical to cut. The prime limitations of this tactic are lack of manpower to carry out the action and lack of spruce-fir markets where loggers can sell the pulpwood. During a spruce budworm outbreak, an oversupply of material results in both of the above.
- 3) Presalvage This is the marking, selling, and harvesting of stands which are healthy but vulnerable to spruce budworm (i.e., these stands, because of age and composition, would suffer damage if subjected to budworm outbreak). Presalvage is difficult to carry out efficiently and effectively during an outbreak because both manpower availability and market availability are limiting.
- 4) Changing Stand Composition This involves the outright removal of some spruce and fir from stands to encourage a species mix. Although an elegant concept, realization of such conditions over a large area is unrealistic.

5) 40-year Rotation — This is the deliberate utilization and marketing of stands such that no individual stand ever exceeds 40 years of age. This is based on the supposition that spruce budworm creates the greatest damage in older stands (usually older than 50 years of age). In concept, this approach is perhaps the most appealing of all techniques available today. However, it is too idealistic. Many spruce-fir stands already exceed 40 years of age. In the next rotation, this is a target that forest managers may be able to aim toward.

# GROUP II.

The second group of techniques are activities directed at the spruce budworm itself and, for the most part, involve chemical and biological pesticides. These techniques are often inappropriate. The prime reason is that techniques directed at the insect are often uneconomical for stands being managed for timber production.

#### GROUP III.

The third group of techniques are those which are combinations of methods directed at the forest and at the insect. By this we mean that through forest directed activities we affect factors which influence spruce budworm numbers. This group of techniques relies on small block cutting which adds a "checker-board" pattern to existing healthy stands (those which we expect to hold for 5 to 20 years and then market). These techniques are explicitly designed to add resistance to residual stands through cutting by enhancing predator and parasite populations and increasing mortality that occurs during the larval dispersal period.

A project was begun in 1979 to examine how the problem of budworm, mature forests, and the demand for wood fiber could be addressed by partial

harvesting of existing, living stands. The following is a report of work done during this project. It is comprised of three parts: (1) a theoretical basis for partial cutting, (2) the economic aspects of the strategy, and (3) a measurement of the response of the biological system to partial cuts.

### A Theoretical Basis for a Strategy of Partial Cuts

Fluctuations in spruce budworm numbers have been the focus of scientific study for several decades. It has been suggested that population changes can be described by a model whose general form has been used for many different biological populations ranging from bacteria to big game (e.g.; Noy-Meir 1975, Gulland 1975, Clark and Mangell 1979, Petermen et al. 1979, Weckham and Botsford 1980, Fleming 1980, Campbell and Sloan 1977, Clark et al. 1979, McLeod 1979, Southwood and Cummins 1976, Ludwig et al. 1978). The components of such a model include an intrinsic logistic growth rate coupled with mortality factors that increase in intensity as the population density increases (density-dependent). The growth rate and density-dependent mortality factors are also often coupled to habitat conditions.

With such models, it is possible to analyze population changes over a range of densities from very low (endemic) to very high (epidemic). When this is done, analysis often reveals conditions where the rate of increase is balanced by an equal mortality rate. Such conditions are termed equilibrium states. If a population tends to remain at an equilibrium, it is considered stable. In the case of the spruce budworm, two stable equilibrium states may be evident: one at very low densities and one at very high densities.

The rate of change of the budworm population as a function of budworm density is portrayed in figure 15. In this figure, the solid line depicts the intrinsic biological growth rate of the budworm population. The dashed lines are losses due to predation and parasitism (M). Where the dashed line lies above the solid line, the net growth rate of the budworm population is negative, and where the dashed line lies below the solid line the net growth rate is positive. Points of intersection of the dashed and solid lines indicate possible equilibrium points. For an intermediate value of M, there are stable low and high density equilibrium points (A and C) and an unstable intermediate equilibrium point (D). Budworm numbers will remain low provided their density remains below T. If they exceed T, the population density will move to C.

The forest plays an important role in determining fluctuations in budworm numbers. This role is manifested through the influence of forest composition and forest structure on the intrinsic growth rate of the budworm population and on the level of budworm predation and parasitism.

Forest composition and structure influence the intrinsic growth rate of the budworm population through egg mortality from parasitism and through the dispersal loss of early instars. Typically, the intrinsic growth rate of an insect population is determined by the number of eggs laid by females. However, with budworm, most significant density-dependent mortality occurs after the budworm are established in feeding sites in the spring. Hence, the number of surviving third instars represents the intrinsic growth rate for a particular year.

Parasitism of budworm eggs has been found to increase with an increasing proportion of non-host trees in a stand (Kemp and Simmons 1979a). Unfortunate-

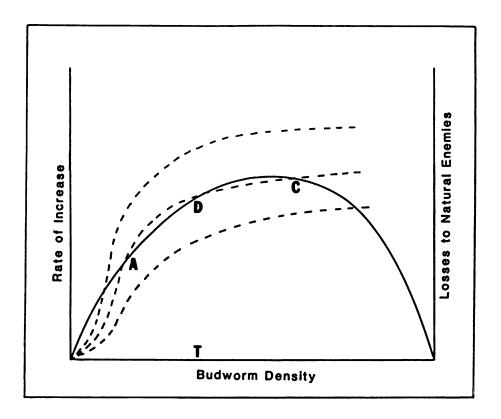


Figure 15. The relationship between the rate of increase of spruce budworm populations (solid line) and different levels of predation and parasitism (dashed lines)

ly, little is known of egg parasitism at low budworm densities. At high densities, it is an insignificant mortality factor.

Dispersal loss of budworm larvae occurs in the fall and spring and results in a 60% to 85% reduction in the population (Miller 1958, 1975). Mortality factors during the dispersal period are: (1) air dispersal to non-host material, (2) predation, (3) failure to spin hibernaculae, and (4) diapause-free development (Miller 1958). Of these, the most important is air dispersal to non-host material. Kemp and Simmons (1979b) found that a higher proportion of non-host trees in the understory, reduced crown closure, and the presence of aspen in the overstory resulted in higher mortality during the dispersal period. Similar results were obtained in a simulation study of spruce budworm dispersal (Kemp et al. 1980). A recent report on the closely related jack pine budworm (Choristoneura pinus) (Freeman) indicated that dispersal losses increased with decreased stand stocking (Batzer and Jennings 1980).

The picture painted is that, in stands with non-host trees and open areas, the intrinsic rate of increase of the budworm is reduced. As a forest stand matures and crown closure increases, the growth rate of the budworm population increases. Depending on the magnitude of the increase in the growth rate of the budworm population, the change in early larval survival may be sufficient to catalyze a budworm outbreak. However, forest dynamics also have a significant effect on natural control agents. The most important of these natural enemies are those which attack the large larvae and pupae.

Stand composition and structure can influence the mortality of large larvae and pupae through bird predation. Views on the importance of bird predation in maintaining budworm populations at low numbers are varied. We feel that it is

unlikely that bird predation alone is responsible for maintaining budworm at low densities. However, when complemented with other mortality factors, bird predation is important.

The bird predator guild can be classified into two groups: ground feeders and crown feeders. Gage et al. (1970) have shown that ground feeders feed less frequently on budworm than crown feeders, while their mean consumption per individual is much higher. If we assume that differences in feeding frequency and comsumption rates tend to cancel each other, then the important aspect of the bird predator guild is not the species mix, but rather the density of all bird predators. Data presented by Titterington et al. (1979) and Gage and Miller (1978) suggest that the densities of the predator guild reach a maximum in stands characterized by a mix of hardwoods, softwoods, and stand openings. As a stand matures, crown closure increases, tree species diversity declines, and the number of avian predators decreases.

In addition to changes in bird predator numbers, forest dynamics may also influence the searching efficiency of bird predators. Birds search foliage in order to find budworm. As the forest matures, the foliage area to be searched increases and the efficiency of birds in locating and capturing budworm likely declines.

A large numbers of parasitoids attack the late instars and pupae of spruce budworm. The role of these parasitoids as mortality agents has been largely discounted since they depend on alternate hosts and their parasitism rates are generally low. However, no study has attempted to determine their influence on budworm dynamics at low densities. Stand characteristics influence the parasitism of budworm by these parasitoids. Simmons et al. (1975) found that pupal

parasitism of budworm increased as stand diversity increased and crown closure decreased. The reasons for this are twofold: (1) increased stand diversity provides more habitat for alternate hosts required by the pupal parasitoids, and (2) stand openings provide sources of food and water for the adult parasitoids.

The net effect of forest-induced changes on avian predators and insect parasitoids is a reduction in their ability to limit budworm numbers as the forest becomes more uniform in composition and as open area declines.

The contentions raised here are supported elsewhere. In particular, it has been found that budworm-induced tree mortality is related to stand composition and often declines with increases in the density of non-host trees and along borders of stand openings (MacLean 1980).

It is clear that the dynamics of the spruce-fir forest and the budworm are closely related. In general, as a forest matures, the food resources of the budworm increase and the impact of various mortality factors acting on the budworm are diminished. Forest management might therefore make an effort to reduce the impact of budworm by either maintaining or transforming the forest to a successional state where these mortality factors can better limit budworm numbers. Such a state may be achieved through the use of partial cuts.

#### Mathematical Analysis

A theoretical basis for partial cutting to reduce budworm impact has been presented. In essence, a system (the forest, the budworm, and a harvesting strategy) has been envisioned through which a set of objectives (minimizing fiber loss, maintaining a steady supply of fiber, or maximizing economic returns) might be met. The next step is to analyze this system through the use of a

mathematical model. In so doing, we hope to increase understanding of real world phenomena and better develop management or design policies.

If, instead of seeking better ways to manage spruce fir stands, we were asked to design a controller for an industrial chemical process, the task would be relatively straightforward. Mathematical models could be employed to design the controller and analyze its operation. However, mathematical models of biological processes are usually too abstract to capture the real world. For this reason, they normally cannot be used alone as a design tool. Instead, they should be used to explore a range of possible outcomes, and then one or some of the logical management approaches can be field tested. This is the philosophy adopted here.

As stated earlier, the swift change that can occur in budworm numbers depends in part on slow changes in forest composition and forest structure. In order to capture the complete dynamics of the spruce budworm and spruce fir forest, these fast and slow changes must be coupled. However, we are not interested in the dynamics of the budworm from the beginning of one outbreak to the next one, but rather in the transition from the endemic to the epidemic states. This is because our objective is to prevent, delay, or lessen the severity of a budworm outbreak in the residual forest stand following a partial cut.

The rate of change in budworm numbers for a particular forest condition is given by the intrinsic rate of increase of the budworm population (feeding third instars) minus the rate at which budworm are lost to mortality factors. This rate can be described in terms of a mathematical model:

$$dB/dt = rBs - (B^2/(\alpha^2 + B^2))\delta$$
 (1)

In (1), B is the budworm density per  $m^2$  foliage, r is the rate of increase of the budworm, S is the number of  $m^2$  units of foliage per hectare,  $\alpha$  is the density per  $m^2$  foliage of budworm at which one half the maximum efficiency of budworm natural enemies is reached, and  $\delta$  is the maximum number of budworm per hectare lost to natural enemies. A differential equation is an approximation of what is actually a discrete time process. However, adoption of a continous representation facilitates analysis and will not invalidate any general conclusions reached.

Ludwig, Jones, and Holling (1978) used a form of (1) and concluded that  $\delta$  is principally attributable to bird predation. To begin, we will adopt their conclusions and use their estimates of parameters of (1) which place the budworm population in a transition between the endemic and outbreak states. Equation (1) can then be written as:

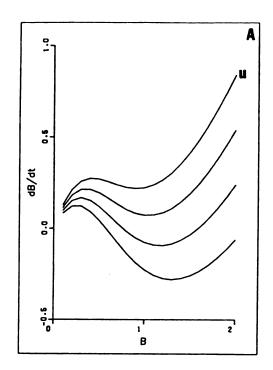
$$dB/dt = 1.52(B)37050 - (B^2(1.1^2 + B^2))106210$$

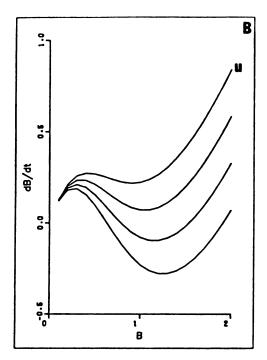
The relationship between the rate of increase of the budworm population and the budworm density for these values is presented as curve **u** in figure 16. It can be seen that the rate of change of the budworm is greater than zero for all budworm densities, and budworm numbers will therefore quickly increase. The effect of reducing the rate of increase by 10%, 20%, and 30% is portrayed in figure 16. With a 20% and 30% reduction of the intrinsic rate of increase, an outbreak would not develop unless budworm numbers exceeded 1.5 and 2 larvae per m<sup>2</sup>, respectively. The reduction in this rate may be brought about as a result of increased dispersal loss in areas subjected to a partial harvesting strategy.

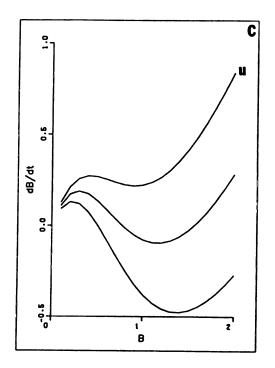
Figure 16. The influence of different levels of dispersal loss and predation on the rate of change of spruce budworm numbers. This rate is described by;

$$dB/dt = rBS - (B^2/(\alpha^2+B^2))\delta$$

B is the number of budworm per  $m^2$  foliage, r (1.52) is the growth rate of B, S (37050) is the number of  $m^2$  units of foliage per hectare,  $\alpha$  (1.1) is the density of budworm at which predation is 1/2 the maximum and  $\delta$  (106210) is the maximum predation of budworm per hectare. The topmost curve in A,B and C correspond to these values. In A, dispersal loss is increased 10, 20 and 30%; in B predation is increased 10, 19 and 26%; and in C both dispersal loss and predation are increased 10 and 10% and 20 and 19% respectively.







In addition to a reduction in the intrinsic rate of increase, a partial cutting strategy will also result in an increase in losses to natural enemies. If these losses are wholly attributable to bird predation, this comes about as a result of an increase in bird densities and a decrease in foliage area.

The effect of increasing losses to natural enemies by 10%, 19%, and 26% is illustrated in figure 16. Once again, an outbreak will not develop unless the critical thresholds of budworm densities are exceeded.

By combining the influence of partial harvesting on the intrinsic rate of increase and on the loss to natural enemies, dramatic changes in the rate of increase of the budworm population can be achieved. This is illustrated in figure 16 for a 10% reduction in the rate of increase and 10% increase in losses to natural enemies, and a 20% reduction in the rate of increase and a 19% increase in losses to natural enemies, respectively. In the latter case, the threshold density for an outbreak is approximately 2.5 budworm per m<sup>2</sup>.

Unfortunately, the analysis presented cannot be considered conclusive. While the general forms of the relationships presented are likely correct, questions exist with respect to model components and the actual parameters used. First, it is unlikely that avian predators are the only density-dependent mortality factors. The influence of parasitic insects and infectious diseases has been ignored. Second, the maximum consumption rate of the avian predators is a gross estimate and may deviate by ± 30%. Finally, it is not possible to predict the actual effect of a partial cut on the intrinsic rate of increase or on losses to natural enemies. The important point, though, is that modest changes in the intrinsic rate of increase of the budworm and in the number of budworm lost to

natural enemies can have significant effects on the rate of increase of the budworm population.

An obvious confounding factor is the immigration of budworm into stands which have been partially cut. This immigration need not be extraordinary to push budworm densities beyond the threshold of 2 to 5 larvae per m<sup>2</sup> foliage for which budworm numbers cannot be regulated by natural enemies. Miller et al. (1978) estimated that moths immigrating into a test plot from surrounding areas with moderate to high budworm densities deposited ca. 10 egg masses per m<sup>2</sup> foliage. Assuming 20 eggs per mass and 80% cumulative mortality of eggs and small larvae, the density of large larvae will exceed the mortality caused by (functional response) natural enemies. For this reason, a partial harvest may not be able to prevent high budworm densities in treated stands.

Within the context of the model, once the rate of increase of the budworm population exceeds natural enemy induced mortality, the population quickly increases to the carrying capacity of the habitat and remains there until the food supply is exhausted. However, there are instances in which the budworm population has increased well beyond an endemic density but did not destroy the habitat. This model, therefore, cannot be considered representative of all real world situations. It is conceivable that, by decreasing the intrinsic rate of increase of the budworm and increasing losses to natural enemies, an outbreak may not be prevented but may be diminished in intensity. As a result it is important to examine the dynamic interaction between the budworm and forest when budworm numbers are greater than endemic levels.

Ludwig et al. (1978) suggested using two variables to describe the forest: the number of units of an area of foliage (S), and an artificial variable to

describe the physiological state of the trees (E). In this case, S is the number of m<sup>2</sup> units of foliage per hectare. The rate of change of these variables is described by the following model:

$$dS/dt = r_S S(1 - (SK_S)(K_F/E))$$
 (2)

$$dE/dt = r_E E(1 - (E/K_E)) - P(B/S)$$
 (3)

In (2), the increase in S is limited by a maximum  $(K_S)$  which we have set equal to 49,400 per hectare. A similar situation exists for E; however, the peak physiological condition  $(K_E)$  is equal to 1. Where the physiological condition of the tree is poor, the value of  $S/K_S$  is multiplied by  $K_E/E$  so that S does not increase. In (3), B/S is budworm larvae per  $m^2$  foliage and P is the rate of consumption of foliage per larvae. It is assumed that a decline in the physiological condition of the tree is directly proportional to the loss in foliage. S does not decline directly as a result of budworm feeding, but rather through a decline in the physiological well-being of the tree. Ludwig, et al. estimated P as  $1.5 \times 10^{-3}$ ,  $r_S$  as .10 and  $r_E$  as 1.0. We will use these values initially.

For a given budworm density, (2) and (3) constitute a dynamic system which has no analytical solution. In order to gain insight into the properties of this system, graphical methods are used. In figure 17, the values of S and E for which the rates of change of each is zero are plotted.

The points where these lines intersect are equilibrium points. For any initial value of S and E and a given value of B, the rate of change of the complete system (dS/dE) can be evaluated. By so doing, the dynamics of S and E can be determined. Approximate system trajectories for various values of S and E are given in figure 17. If the equilibrium points exist, then the forest will remain alive, providing the physiological condition of the trees is good.

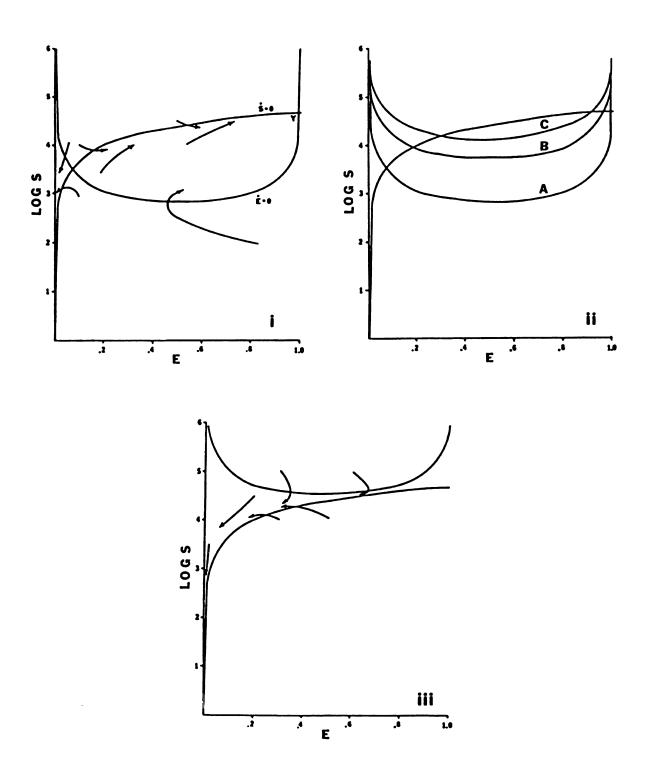
Figure 17. The influence of spruce budworm defoliation on the survival of host trees. The dynamics of the host trees is described by two equations ;

$$dS/dt = r_SS(1-(S/K_S)(K_E/E))$$

$$dE/dt = r_E E(1-(E/K_E)) - P(B/S)$$

Where S is the number of m<sup>2</sup> units of foliage per hectare,  $r_S(.1)$  is the growth rate of S,  $K_S$  (49,400) is the maximum S per hectare, E is the physiological condition of the tree,  $K_E$  (1.0) is the maximum E,  $r_E$  (1.0) is the growth rate of E, P (.0017) is the proportion of a m<sup>2</sup> unit of foliage consumed by 1 budworm and B is the number of budworm per hectare. In (i) B =  $1 \times 10^5$ . The arrows indicate general system dynamics for various initial conditions. In (ii) B =  $1 \times 10^5$  for A,  $8 \times 10^5$  for B, and  $2 \times 10^6$  for C. In (iii) B =  $5 \times 10^6$  and extensive tree mortality occurs.

Ludwig, D., D.D. Jones, and C.S. Holling. 1978. Qualitative analysis of insect outbreak systems: The spruce budworm and forest. J. Anim. Ecol. 47:315-332.



The forest (S) will succumb to budworm under two conditions. First, the equilibrium points may vanish. This will occur if budworm densities exceed some critical level. Second, the physiological state of the trees may decline below a critical value as a result of budworm feeding.

The second condition can also be evaluated graphically. In figure 18, equation (3) is portrayed for different values of B/S and  $r_E$ . As shown, the tree remains vigorous even for relatively high densities of budworm and values of  $r_E$  less than that given by Ludwig et al. In the real world, it is likely that sustained moderate defoliation will result in a decline in  $r_E$ , and the physiological condition of the tree may then decline. In addition, parameter values used are gross estimates and will vary from site to site. Accurate predictions of the budworm density required to cause extensive tree mortality cannot be made. However, this is not the purpose of the analysis. The important point is that extensive tree mortality will not occur with moderate budworm densities for a number of years. Hence, if a policy of partial harvesting can lessen the intensity of an outbreak in the residual stand, mortality of the remaining trees can at least be delayed and may be reduced.

To conclude this section we have shown that a reduction in the rate of increase of budworm populations coupled with an increased loss of budworm to natural enemies dramatically alters the dynamics of the insect. Immigration of moths from surrounding stands may, however, cancel these effects. The forest can tolerate moderate defoliation by the budworm for many years. It is conceivable that, as a result of a partial cut, the intensity of an outbreak is diminished. Without more information, though, a mathematical analysis can go no further. A real world evaluation of a partial harvest policy is desirable and

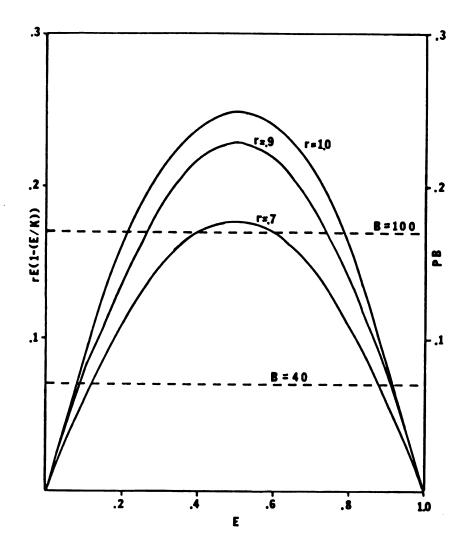


Figure  $^{18}$ . Influence of spruce budworm on the physiological condition of host trees. The physiological condition is described by  $^{1}$ 

$$dE/dt = rE(1-(E/K)) - PB$$

where E is the physiological condition, r is the rate of increase of E, K (1.0) is the maximum E, P (.0017) is the rate of consumption of a  $m^2$  unit of foliage per larvae and B is the budworm density per  $m^2$  foliage. The dashed lines are PB and the solid lines are rE(1-(E/K)).

Ludwig, D., D.D. Jones, and C.S. Holling. 1978. Qualitative analysis of insect outbreak systems: The spruce budworm and forest. J. Anim. Ecol. 47:315-332.

necessary but also difficult to conduct. Nevertheless, an effort has been made to perform such an analysis.

## Implementation and Assessment of a Partial Cut Strategy

A policy of partial harvesting was evaluated on three sites in the Hiawatha National Forest in Delta County in Michigan's Upper Peninsula. The specific sites are all within 30 miles of the city of Escanaba. The spruce-fir type is scattered throughout the region, with individual stands ranging from about 30 acres up to about 100 acres in size.

Three forest stands (fig. 1) were selected for implementing the partial harvest strategy based on the following criteria: (1) species composition: spruce fir type with some hardwood component, either aspen or northern hardwoods; (2) area size: a minimum of 40 acres with a control plot of no less than 10 acres, preferably adjacent to the treatment plot; (3) budworm density: low budworm numbers at the time of harvesting; and (4) harvestability: sufficient volume of timber so that partial cutting may be commercially feasible, if road-building costs and distance to mills is not prohibitive. In fall 1979, stands 1 and 3 (fig. 1) were identified. In fall 1980, stand 2 (fig. 1) was added, although budworm densities were already at moderate population levels.

In stand 1, cuts laid out as patches were used to partially harvest the stand. The layout of the cuts and the control plot is shown in figure 19. Harvesting was conducted in the winter of 1979/1980. In stand 2, cuts were laid out in strips and were made during the winter of 1981. Harvested strips are 40 m wide and the uncut strips are 60 m wide. The layout of the strips and the control plot is shown in figure 19. In stand 3, cuts were laid out in strips 20 m wide with an uncut

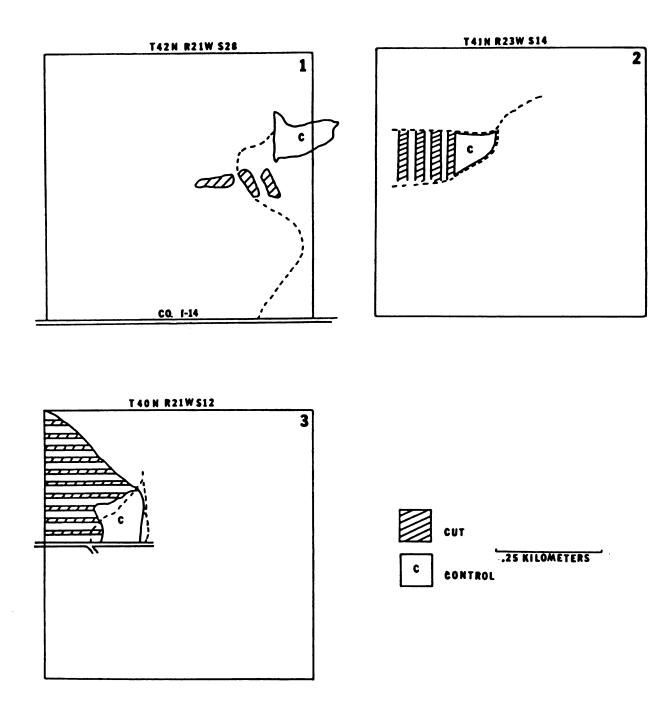


Figure 19. Layout of cut and control areas in stands 1, 2 and 3.

strip 40 m wide separating each cut (fig. 19). Harvesting of this area took place in the winter of 1979/1980.

The structure and composition of each cut and control area was described by measuring stand parameters in five fixed-radius plots in the residual stand of the treatment areas and in the control areas. In the strip cuts, plots were established in the center of each residual strip. In the patch cut, plots were located 20 m from the edge of the cut. In all control areas, plots were located 60 m or more away from a cut area.

Plot measurements were made according to procedures developed by Mog and Witter (1979). This data is presented in tables 6 through 8. The live tree species composition and tree condition parameters for budworm hosts in the cut and control areas for stands 1 and 2 are very similar. In stand 3, the total percentage of budworm hosts is approximately the same; however, the control area has a larger percentage of white spruce.

Evaluation of a policy for managing or designing a system must address two things: (1) Is the policy feasible and (2) are the objectives as originally set forth met by applying the policy? The feasibility of partial harvesting is determined by profitability and compatibility with other forest management objectives. Whether partial cutting can help meet these objectives is determined by how this policy affects the budworm population.

### Feasibility of a Policy of Partial Harvests

Any feasibility analysis of a partial cut strategy is based on determining when and how much of a forest stand to harvest. The timing of timber harvest involves two considerations: (1) stand growth, and (2) financial maturity of a

Table 6. Comparison of species composition of live trees and tree condition parameters for spruce budworm host trees by treatment in stand 1. Species composition is based on data collected from 5.04 hectare plots and tree parameters are based on data from 5.02 hectare plots.

	Cut				Control				
	<u>Trees</u> hectare	z	Basal <sub>2</sub> area m hectare	z	<u>Trees</u> hectare	z	Basal <sub>2</sub> area m <sup>2</sup> hectare	z	
Balsam fir	554	58	12.6	40	563	53	13.7	39	
White spruce ,	20	2	1.2	4	25	2	1.7	5	
Trembling aspen	198	21	12.1	39	237	22	12.3	35	
Paper birch	44	4	1.0	3	124	12	2.9	8	
N. white cedar	114	12	3.1	10	89	8	3.7	10	
Sugar, red maple	5	1	0.2	1	20	2	0.4	1	
Other hardwoods	15	2	0.8	3	10	1	0.2	ī	
Hemlock	0	0	0	0	5	0	0.4	ī	
All species	950	100	31.0	100	1073	100	35.3	100	
Parameter	x		s <del>_</del>	n	×		s <del>_</del>	n	
Balsam fir									
DBH (cm)	17.5		0.69	53	15.9		0.42	62	
Tree height (m)	14.8		0.41	53	14.5		0.34	62	
Crown position <sup>2</sup>	2.7		0.15	53	2.8		0.10	62	
Live crown length (m)	7.2		0.37	53	8.0		0.36	62	
Dead crown length (m)	1.0		0.15	53	1.7		0.15	62	
Proportion trees dead	0.11		0.04	53	0.17		0.05	62	
site index	60.0			29	60.0			27	
White spruce									
DBH (cm)	32.6		3.46	3	26.4		4.25	4	
Tree height (m)	18.0		1.56	3	17.8		1.85	4	
Crown position <sup>2</sup>	1.0		0.0	3	2.2		0.5	4	
Live crown length (m)	10.5		1.79	3	8.8		0.95	4	
Dead crown length (m)	3.7		0.0	3	4.0		0.0	4	
site index	52.0			12	55.0			13	

<sup>1</sup> Includes balsam poplar

<sup>&</sup>lt;sup>2</sup>Crown position ranking: 1-dominant, 2-co-dominant, 3-intermediate, 4-suppressed

<sup>&</sup>lt;sup>3</sup>Based on USDA Forest Service Lake States Exp. Sta. Tech. Notes 465 and 474 by S. R. Gevorkiantz

Table 7. Comparison of species composition of live trees and tree condition parameters for spruce budworm host trees by treatment in stand 2. Species composition is based on data collected from 5 .04 hectare plots and tree parameters are based on data from 5 .02 hectare plots.

		C	ut		Control			
Species	<u>Trees</u> hectare	z	Basal <sub>2</sub> area m hectare	2	<u>Trees</u> hectare	z	Basal 2 area m hectare	z
Balsam fir	763	71	18.12	66	788	85	16.29	79
White spruce	0	0	0	ő	6	1	0.07	/9
Aspen	313	29	9.18	34	131	14	4.36	70
Total	1076	100	27.30	100	925	100	20.72	20 100
Parameter	x		s <del>_</del>	n	×		<del>s_</del>	n
Balsam fir								
DBH (cm)	16.85		0.51	54	15.70		0.41	53
Tree height (m)	14.66		0.30	54	14.09		0.37	53
Crown position 1	2.40		0.12	54	2.72		0.16	53
Live crown length (m)	7.78		0.33	54	6.03		0.28	53
Dead crown length (m)	.50		0.16	54	0.35		0.14	53
Proportion, trees dead	0.16		0.05	54	0.19		0.05	53
site index*	62			12	65		0.03	12

<sup>1</sup>Crown position rankings: 1-dominant, 2-co-dominant, 3-intermediate, 4-suppressed

<sup>&</sup>lt;sup>2</sup>Based on USDA Forest Service Lake States Exp. Sta. Tech. Notes 465 and 474 by S. R. Gevorkiantz

Table 8. Comparison of species composition of live trees and tree condition parameters for spruce budworm host trees by treatment in stand 3. Species composition is based on data collected from 5 .04 hectare plots and tree parameters are based on data from 5 .02 hectare plots.

	Cut				Control				
	<u>Trees</u> hectare	z	Basal <sub>2</sub> area m hectare	z	<u>Trees</u> hectare	x	Basal <sub>2</sub> area m hectare	z	
Balsam fir	469	55	12.3	39	183	29	5.4	16	
White spruce ,	35	4	1.3	4	247	27	7.4	21	
Trembling aspen	193	23	13.5	43	114	12	11.0	31	
Paper birch	84	10	2.3	7	247	27	6.3	18	
N. white cedar	64	7	2.0	6	124	13	4.1	12	
Sugar, red maple	5	1	0.3	1	10	1	0.7	2	
Other hardwoods	0	0	0	0	0	ō	0	ō	
Hemlock	0	0	0	0	Ö	Ö	Ö	ŏ	
All species	850	100	31.7	100	925	100	34.9	100	
Parameter	x		s <del>_</del>	n	<del>-</del> x		s <del>_</del>	n	
Balsam fir									
DBH (cm)	17.6		0.50	62	20.6		1.31	17	
Tree height (m)	13.5		0.42	62	16.2		0.92	17	
Crown position <sup>2</sup>	2.9		0.13	62	2.5		0.32	17	
Live crown length (m)	8.1		0.36	62	7.6		0.78	17	
Dead crown length (m)	0.5		0.06	62	1.0		0.15	17	
Proportion trees dead	0.04		0.02	62	0.1		0.08	17	
site index	64.0		****	27	59.0		0.00	18	
White spruce								10	
DBH (cm)	20.5		2.25	4	19.3		1.0	27	
Tree height (m)	10.7		1.45	4	15.2		0.64	27	
Crown position2	3.0		0.60	4	2.5		0.23	27	
Live crown length (m)	6.0		3.70	4	6.2		0.44	27	
Dead crown length (m)	0.0		0.0	4	0.6		0.06	27	
site index	52.0		-	11	50.0		3	33	

<sup>1</sup> Includes balsam poplar

<sup>&</sup>lt;sup>2</sup>Crown position ranking: 1-dominant, 2-co-dominant, 3-intermediate 4-suppressed

<sup>&</sup>lt;sup>3</sup>Based on USDA Forest Service Lake States Exp. Sta. Tech. Notes 465 and 474 by S. R. Gevorkiantz

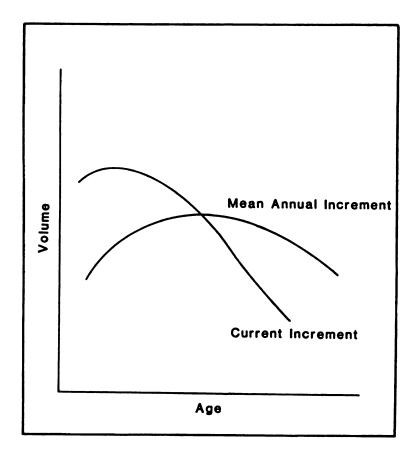


Figure 20. Relationship between the mean annual increment and the current increment in timber volume growth. The maximum mean annual increment occurs when this variable equals the current increment. This can also be shown with a first order condition since volume = F(age) and the mean annual increment = F(age)/age so d(F(age)/age)/dt = (ageF'(age)-F(age))/age<sup>2</sup> = 0

F'(age) = F(age)/age which is equivalent to the current increment equal to the mean annual increment.

stand. When determining a time to harvest, most foresters opt for maximizing the mean annual growth increment (fig. 20) (Clawson 1977). However, growth rates change slowly over a span of years. Any harvest timing which seeks to take advantage of a growth relationship can be varied without significant consequences. Further, this policy ignores forest management costs, markets for fiber, and prices for stumpage. If the objective of forest management is to maximize the volume of wood produced by a parcel of land, then this is a proper strategy. Most often, economic considerations should also be included in a harvesting decision.

The financial maturity of a stand involves costs and returns of growing timber. Three separate costs can be identified: (1) inputs of capital, labor and materials throughout the rotation of a stand; (2) interest on the returns that might be realized from the immediate harvest of a stand; and (3) rent on the bare land. Given these costs and knowledge of tree growth and future returns from this growth, the age of financial maturity can be computed. This is the point in time when net returns are maximized. The age of financial maturity will almost always be sooner than when the maximum mean annual growth increment is reached.

In practice, things are not as simple. A number of factors confound computation of financial maturity. First, the markets for the wood and the effect of the harvest volume on the market price have not been considered. Changes in either of these factors may dramatically influence the point in time when financial maturity is reached. In an extreme case, markets could cease to exist and render the concept of financial maturity meaningless. Second, the timing and intensity of harvest will partially determine the stand type and

number of trees available for the next harvest. This consideration must also be part of the harvest decision. Third, age/growth relationships are not precise. Thus, financial maturity is not a point, but an interval in time. Finally, a harvest decision may be motivated by factors other than an optimal economic return. For instance, a steady flow of fiber, minimizing loss of trees to damaging organisms, or conversion of a site to different tree species are other considerations which may dictate harvest of a forest stand.

In addition to the general considerations discussed, harvest decisions are strongly dependent upon timber ownership. There are three broad categories of timber owners: (1) small, private landowners; (2) timber firms which grow trees and use this fiber and fiber from other sources to produce a product (a vertically integrated forest firm); and (3) state or federal agencies.

Small, private forest owners are well advised to use financial maturity in deciding when to harvest. In addition, though, the owner must accept the market as given and adjust operations to it. If prices for fiber fluctuate, a strictly economic analysis of an optimal date for a timber sale is useless. Faced with the prospects of such variability, a small forest owner may seek to minimize risk by cutting prior to financial maturity.

For a vertically integrated forest firm, the cost of wood taken from the forest is a small, though not insignificant, part of total costs for processing and marketing the finished wood products. Clawson (1977) has estimated that these costs are on the order of 15% to 20% of total expenditures. As a result of operating costs, it is important that the firm continue operating at or near full capacity. This is especially true of pulp and paper plants.

Under these circumstances, low wood costs are desirable but not as important as a constant supply of wood. The vertically integrated forest firm usually chooses harvest dates within very wide limits for processing and marketing efficiency. The date of financial maturity is an important consideration, but it is not dominant in decisions about the date of harvest. These firms will cut more or less of their own timber or buy more or less from other sources for market and processing reasons and not for efficiency of growth in the forest stand.

When the Forest Service or other public agencies offer wood for sale, the market for fiber can be influenced because of the large land holdings of these agencies. In addition, the Forest Service, and perhaps other public agencies, have traditionally paid great attention to local economic stability. Judgements of timber harvest to meet these conditions may override economic and biological analysis. Finally, the Forest Service is directed through the National Forest Management Act of 1976 to grow and market timber within specific guidelines. All of these factors influence the decision as to when and how much timber to harvest.

In summary, a number of factors, other than the characteristics of stand growth and financial maturity, influence decisions to harvest timber. These considerations vary depending upon forest ownership.

For each category of timber owner, the feasibility of a partial cut strategy is strongly influenced by forest management objectives. For the small, private landowner, the principle management objective will likely be to secure the maximum economic return with the least risk. For this objective, a partial cut strategy is likely not feasible because the residual stand will probably not

increase in value faster than returns are accrued from the immediate full harvest of the stand. This is more fully explained in appendix 5. Hence, the best policy for a small, private landowner is, if possible, to completely presalvage a stand. If maximization of economic returns with least risk is not a management objective then a policy analysis for the small, private landowner is similar to that for the vertically integrated forest firm or the public agency.

For a vertically integrated forest firm or a public agency, we will assume that it is: (1) desirable to spread the harvest, marketing or processing of spruce-fir fiber over a longer time interval than would result from complete presalvage; or (2) harvest the stand to promote regeneration of multi-aged spruce-fir which is less vulnerable to budworm. Feasibility is then a question of whether a partial cut can be profitably implemented.

To determine profitability, questionnaires (appendix 6) were sent to the loggers responsible for harvesting each stand. Only one questionnaire was returned. The information from this questionnaire is given in table 9. It can be seen that the strip cut operation in stand 3 was profitable. In all likelihood, the operations in stands 1 and 2 were also profitable, since it was quite easy to secure a contractor for these harvests. Hence, a forest owner could implement a partial cut in a profitable manner. For the same reasons that a small, private landowner would likely not maximize returns from a partial harvest, neither will a vertically integrated firm or a public agency. However, as explained earlier, these forest owners do not make harvest decisions based solely on maximizing economic returns (ie; harvest at financial maturity). Forest management objectives and fiber processing and marketing considerations are also heavily

Table 9. Costs and revenues incurred by logger in harvesting stand 3.

Balsam fir	pulpwood	35	8.90	) :	11.50	
3/ Species	Product	Quantity (cunits)	Price (\$/cun:		ecies	
cost for jo	b = (annual cost / working days p	er yr.) (f days wor	ked on job)	where:		or chain a 10% purcha .job = 28
2/Determination	of capital cost of	equipment:	$\frac{(1+i)^n}{1+i)^n-1}$		interest rate life of equu	pment in y
1/Ivan	chain saws purchase	-4 107 <b>0</b>				
NET REVENUE						3,180.6
TOTAL REVENUE					14,028.00	14,028.0
Product: Pulpwood Sawtimber Products-Cedar	Mill: Head Paper Co. Escanaba Lumber Habitant Fence		orda:	Value/cd: 33.58 35.00 35.00	9,268.00 1,120.00 3,640.00	
REVENUES  Delivered stumps	<b>L</b>					
TOTAL COSTS						10,845.5
Stumpage 3/					1,392.08	1,392.0
Transportation co	et subtotel				3,926.40	3,926.4
Transportation o	mills (hired out a f crew to and from		y)		- 0 - 3,708.00 218.40	
Transportation						
Equipment cost s	ubtotal				1,519.05	1,519.0
Capital equipme Fuel Maintenance Insurance	nt cost-				661.15 295.40 91.50 471.00	
3 Husqvarna cha Operating costs:				900.00 39,600.00	443.34	
Equipment Original cost1/: I-4500 Ford iro	n mule			38,700.00		
Labor cost subtot	<b>a</b> 1				4,008.00	4,008.0
	: 45 sticks/cord = \$ ce	9.00/cord)			300.00	
(\$.20/stick x Employee insuran					3,708.00	

3/ Species	Product	Quantity (cunits)	Price (\$/cunit)	Species Total (S)
Balsam fir	pulpwood	35	8.90	311.50
White spruce	pulpwood	19	16.14	306.66
Cedar	products	80 .	5.03	402.40
Paper birch	pulpwood	58	3.94	228.52
Aspen	pulpwood	120	1.15	138.00
Mixed hardwoods	pulpvood	5	1.00	5.00
Sale tot	al	317 cunits		\$1.392.08

weighed. Whether or not these objectives can be reached with this policy depend upon the dynamics of the budworm and the forest in the residual stands.

## Biological Response of the System

The objective here was to assess the biological response of the system to a partial cut. To do this, budworm survival and balsam fir defoliation was evaluated in cut and control areas of each stand. Budworm survival was selected as an indicator of all the processes operating on the budworm population.

In late summer 1980, egg mass densities were estimated in the cut and control areas of stands 1 and 3. Egg mass densities were not estimated in stand 2 because the harvest of this stand had not been completed. Three branches were clipped with a pole pruner from the mid to uppercrown of 16 host trees in the cut and control areas of each stand. The number of white spruce or balsam fir sampled was based on percentage composition. From each branch, the current year's viable egg masses and parasitized egg masses were removed and recorded. If an egg mass was incompletely parasitized, those with less than half of the eggs attacked were classified as viable. Each branch was checked a second time by a different worker for overlooked egg masses. The foliage surface area of each branch was measured with a grid. Egg mass densities were converted to egg densities by assuming there were 20 eggs per viable egg mass.

In spring and summer 1981 the densities of small and large larvae, pupae, and adults were estimated in the cut and control areas in each stand. The dates for each sample and the budworm life stages sampled are given in table 10. Four branches were clipped with a pole pruner from either 5 or 10 randomly selected balsam fir trees. The foliage area was estimated similarly to the egg mass

Table 10. Dates and degree days (base 5.56 C) for five sampling periods during which spruce budworm densities were determined in three stands. Peak 3rd instar occurs at c. 167 DD, peak 6th instar at c. 416 DD and initial emergence of male moths at c. 472 DD.

	Stand 1			2		3	
Sample		Cut	Control	Cut	Control	Cut	Control
3rd Instar	Date Degree Days Base 5.56 C	6-09 218	6-08 208	6-04 168	6-03 160	6-05 183	6-06 192
4th Instar	Date Degree Days Base 5.56 C	6-12 250	6-12 250	6-17 309	6–17 309	6-11 239	6 <b>-11</b> 239
5th Instar	Date Degree Days Base 5.56 C	6–23 364	6-23 364	6-19 329	6–19 329	6-18 321	6–18 321
Pupae S <sub>1</sub>	Date Degree Days Base 5.56 C	7-03 478	7-03 478	7-02 464	7-02 464	7-01 451	7-01 451
Pupae S <sub>2</sub>	Date Degree Days Base 5.56 C	7-08 560	7 <b>-</b> 08 560	7-07 543	7 <b>-</b> 08 560	7-07 543	<b>7-</b> 07 543

Miller, C. A., D. C. Eidt, and G. A. McDougall. 1971. Predicting spruce budworm development. Dept. Environ. Can. For. Serv., Bi-mon. Res. Notes. 27:33-34.

sample. Samples of 3rd, 4th, and 5th instars were examined in the field and densities expressed as larvae per m<sup>2</sup> foliage. Larvae and pupae found during the first and second pupal samples (Pupae S<sub>1</sub>, Pupae S<sub>1</sub>) were collected and reared. From these rearings, the mortality from parasitism and unknown causes was Adult densities were determined from rearings and from pupal determined. cases from which adults had emerged in the field. Defoliation of old and new foliage was estimated during the two pupal samples by classifying the percentage of foliage missing per branch into 5 groups: 0-20%, 21-40%, 41-60%, 61-80%, and 81-100%. Differences in budworm densities in the cut and uncut areas in each stand were statistically tested after necessary transformations with ANOVA. Differences in percentage mortality were tested by comparing the parameter p of a binomial distribution from the cut and control areas through a Defoliation estimates were tested for differences normal approximation. between the cut and control areas with a Mann-Whitney U test.

<u>Dispersal loss</u>—Estimates of budworm population densities for the different life stages sampled in the cut and control areas of stands 1, 2, and 3 are given in tables 11, 12, and 13. No significant differences in egg mass densities occurred in the cut and control areas from stands 1 and 3. Based on the close proximity and similar stand composition of the cut and control areas in stand 2, we can infer that equality of egg mass numbers likely occurred here also.

The density of feeding 3rd instars was significantly less in the control areas as compared to the cut areas in all three stands. In stands 1 and 3, this apparently indicated a greater mortality of early instars in the control areas. If egg mass densities were equivalent in the cut and control areas of stand 2, the same can be said for this plot. Since dispersal loss is apparently the major

Table 11. Density per m<sup>2</sup> balsam fir foliage of various life stages of the spruce budworm in 1981 and percentage mortality due to parasitism and unknown causes in stand 1. ANOVA (F) or a normal approximation of a binomial distribution (Z) were used to test for treatment differences. C indicates the statistic was not computed due to a small sample size.

Treatment	c	Control			Cut			
Sample Stage	x	s <sub>x</sub>	n	x	s <sub>∓</sub>	n	Foor Z*	P(F≥F <sub>0</sub> ) or P(z≤Z or z≥Z)*
Egg								
hatched	63.01	6.20	48	62.09	5.90	48	.01	.915
parasitized	17.13	2.98	48	21.94	3.25	48	1.19	.278
Larvae								
3rd instar	124.31	7.80	40	218.20	14.81	40	23.56	.001
4th instar	142.37	16.64	20	191.26	25.39	20	2.60	.115
Sth instar	95.72	10.13	20	111.62	16.15	20	.10	.754
4th + 5th instar	119.05	6.53	40	151.44	10.22	40	1.41	.239
Pupae S,								
total	51.71	7.04	40	66.52	6.27	40	2.47	.120
larvae	14.75	2.77	40	14.96	2.43	40	.003	.955
% parasitized	. 536	.059	71	.512	.054	86	0.207*	.417*
% unknown	••••		•			•		
mortality	.338	.056	71	.233	.046	86	1.24 *	.108*
pupae	36.96	5.17	40	51.56	5.21	40	3.96	.050
% parasitized	.167	.025	216	.225	.025	284	1.45 *	.075*
2 unknown		***	-10			204		
mortality	.106	.021	216	.056	.001	284	2.00 *	.023*
adults	31.68	4.40	40	40.68	4.68	40	1.96	.166
Pupae S <sub>2</sub>	31.00	*****	70			40	2.70	1200
total	45.37	5.09	20	54.35	6.58	20	1.17	. 287
larvae	1.84	.87	20	7.52	2.05	20	5.36	.026
% parasitized	1.0	0.0	-4	.450	.111	20	C	C
7 unknown	1.0	•••	•			20	•	•
mortality	0.0	0.0	4	.500	.112	20	С	С
pupae	43.53	5.44	20	46.82	5.65	20	.176	.677
% parasitized	.112	.033	89	.185	.033	135	1.35 *	.09 *
2 unknown		.033	07	,		133	1.33 "	.07
mortality	.157	.039	89	.059	.020	135	2.28 *	.01 *
adults	32.45	4.49	20	36.23	4.80	20	.332	.5681

Table 12. Density per m<sup>2</sup> balsam fir foliage of various life stages of the spruce budworm in 1981 and percentage mortality due to parasitism and unknown causes in stand 2. ANOVA (F) or a normal approximation of a binomial distribution (Z) were used to test for treatment differences. C indicates the statistic was not computed due to a small sample size.

Treatment	C	Control			Cut			
Sample Stage	x	S <del>_</del>	n	x	S	n	F or Z*	$P(F \ge F)$ or $P(z \le Z) \ne 0$
Egg								. (222 01 222)
hatched								
parasitized		NOT	COL	LECTE				
Larvae								
3rd instar	174.19	18.48	40	276.32	22.09	40	12.57	.001
4th instar	132.09	13.09	20	201.86	19.12	20	9.96	.003
5th instar	135.05	18.37	20	137.52	11.79	20	3.60	.552
4th + 5th instar	133.57	7.04	40	169.69	7.73	40	5.74	.019
Pupae S <sub>1</sub>								
total *	74.50	7.31	40	66.31	6.07	40	.74	.392
larvae	12.34	1.58	40	10.98	1.96	40	.22	.592
% parasitized	.449	.060	69	. 576	.061	66	1.460*	.072*
Z unknown								
mortality	.174	.046	69	.197	.049	66	.288*	.384*
pupae	62.17	7.18	40	55.33	5.15	40	.60	.442
% parasitized	.137	.020	299	.146	.020	308	.290*	.386*
<b>Z unknown</b>								
mortality	.197	.023	299	. 208	.023	308	.289*	.385*
adults	44.81	4.67	40	37.04	3.86	40	1.64	.204
Pupae S <sub>2</sub>								
total	85.28	12.14	20	62.12	6.27	20	1.49	.230
larvae	1.83	.89	20	1.34	.77	20	.18	.675
% parasitized	.50	.25	4	.50	.25	4	c	C
% unknown						•	•	•
mortality	.50	.25	4	. 50	. 25	4	С	С
pupae	83.44	6.43	20	60.78	6.43	20	1.28	.266
% parasitized	. 201	.028	209	.164	.029	165	1.489*	.068*
Z unknown								
mortality	.072	.018	209	.079	.021	165	.241*	.405*

Table 13. Density per  $m^2$  balsam fir foliage of various life stages of the spruce budworm in 1981 and percentage mortality due to parasitism and unknown causes in stand 3. ANOVA ( $F_0$ ) or a normal approximation of a binomial distribution (Z) were used to test for treatment differences. C indicates the statistic was not computed due to a small sample size.

Treatment	C	ontrol			Cut			
Sample Stage	Ř	s <sub>x</sub>	n	x	s <sub>x</sub>	n	For Z*	P(F≥F ) or P(z≤Z or z≥Z)*
<u>Egg</u>								
hatched	43.80	4.43	48	36.80	3.96	48	1.39	.241
parasitized	12.22	2.36	48	11.84	1.67	48	.228	.635
Larvae								
3rd instar	95.86	7.82	40	198.83	15.83	40	38.90	.001
4th instar	96.67	7.98	20	127.85	15.56	20	1.67	.204
5th instar	77.53	9.88	20	142.43	18.48	20	10.59	.002
4th + 5th instar	87.11	6.46	40	135.14	11.98	40	10.88	.002
Pupae S,								
total	40.89	4.76	40	79.87	6.83	40	20.59	.001
larvae	14.24	1.95	40	32.81	3.92	40	9.07	.004
<pre>% parasitized</pre>	.467	.052	92	.635	.035	189	2.69 *	.004*
2 unknown								
mortality	.152	.037	92	.212	.030	189	1.204*	.115*
pupae	26.65	5.88	40	47.06	5.38	40	10.55	.002
% parasitized	.114	.025	158	.142	.022	253	.829*	.203*
Z unknown								
mortality	.165	.030	158	.198	.025	253	.850*	.198*
adults	23.80	2.93	40	35.23	3.78	40	4.98	.028
Pupae S <sub>2</sub>								
total	22.96	4.57	20	37.76	6.15	20	3.34	.075
larvae	.50	.34	20	8.18	1.93	20	15.35	.001
% parasitized	1.0	0	2	. 905	.064	21	С	С
% unknown								-
mortality	0.0	0	2	.095	.064	21	С	С
pupae	22.46	4.57	20	29.57	5.89	20	.76	.390
% parasitized	.162	.036	105	.158	.042	76	.073*	.472*
% unknown							· <del>-</del>	=
mortality	.086	.027	105	.092	.033	76	.093*	.464*
adults	16.98	3.42	20	21.19	4.52	20	.165	.687

mortality factor during this period of the budworm's life cycle, the results suggest that dispersal loss was greater in the control areas. This contradicts the theoretical basis we developed for the partial cut strategy. As indicated in that section, dispersal loss of budworm has been found to increase with increases in the number of non-host trees and increases in stand openings. It is possible that stand factors which influence dispersal loss function on a finer spatial pattern than that provided by the strip and patch cuts. However, a comparison of stand parameters in the cut and control areas of each stand (tables 6, 7, and 8) provides no indications of contrasts between these areas which might explain the large differences in dispersal loss. The sampling methodology has also been reviewed in an effort to uncover a systematic bias which may have led to an underestimation of egg mass numbers in the cut areas or an overestimation in the control areas. We have found none.

The empirical evidence, therefore, seems to refute the theory which suggests that partial cuts will lower the intrinsic rate of increase of the budworm. In fact, it appears as though this rate is higher in the cut areas. However, it must be remembered that only a snapshot of a long-term process has been taken. If differences in dispersal loss are a random phenomena there is a l-in-8 chance that observed differences occurred by chance alone. Taking into account the data presented here and that discussed previously, little is actually known of budworm dispersal. Before abandoning the idea that partial cuts can increase dispersal loss, it would be wise to collect long-term data on actual dispersal losses.

<u>Predation</u> and <u>parasitism</u>—The survival of budworm from the 3rd instar to the adult stage was less in all three cut areas than in the check areas. In fact, in

stand 2, the combined density of large larvae and pupae (Pupae  $S_1$ ) is less in the cut area than in the control plot. Survival curves constructed from data in tables 11, 12, and 13 are presented in figures 21, 22, and 23. In these figures, the survival rate between successive sampling points is represented by the slope of the line joining the density estimates at these points. Survival curves in the cut areas in each stand are similar and those for the control areas in each stand are also similar. This suggests that the higher mortality rate in the cut areas may be attributable to increases in natural enemies following harvest. However, no systematic and statistically significant difference in parasitism of large larvae or pupae was found in the cut and control areas. On the other hand, it has been demonstrated that bird densities increase following a partial harvest (Titterington et al. 1979). A breeding bird census (appendix 7) in the three stands substantiates this.

Another explanation is that budworm survival is a function of density. In the cut areas, the density of 3rd instars is much greater than in the control plots. The survival of budworm between the 3rd and 4th instars is much less in the cut areas than in the controls. This suggests that survival during this period is a function of density. If true, then, at least at moderate densities, mortality during the 3rd and 4th instars compensates for mortality during the dispersal period.

<u>Defoliation</u>—Defoliation of old and new balsam fir foliage in the cut and control areas of each stand is given in table 14. Defoliation of the new foliage was significantly less in the cut areas of stand 2 and 3, and no difference was found in stand 1. Defoliation of old foliage was significantly less in stands 1 and 2, while no difference was found in stand 3. This data seems contraditory to the

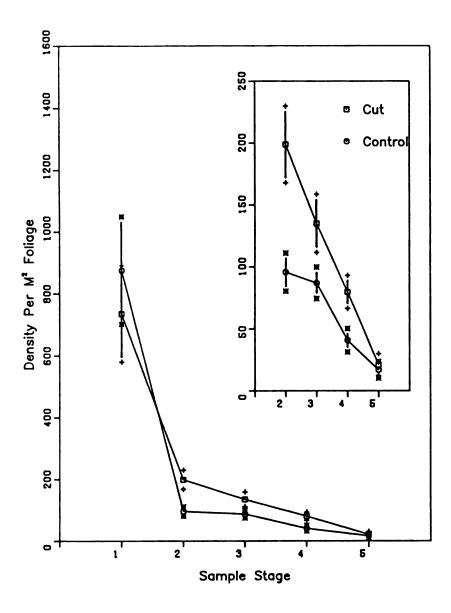


Figure 21. The density per m<sup>2</sup> foliage of various life stages of the spruce budworm in the cut and control areas of stand 3. The life stages are; 1-egg, 2-3rd instar, 3-4th and 5th instar, 4-6th instar and pupa, and 5-adult.

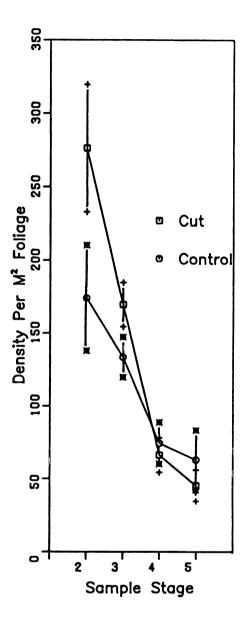


Figure 22. The density per m<sup>2</sup> foliage of various life stages of the spruce budworm in the cut and control areas of stand 2.

The life stages are; 2-3rd instar, 3-4th and 5th instar, 4-6th instar and pupae, and 5-adult.

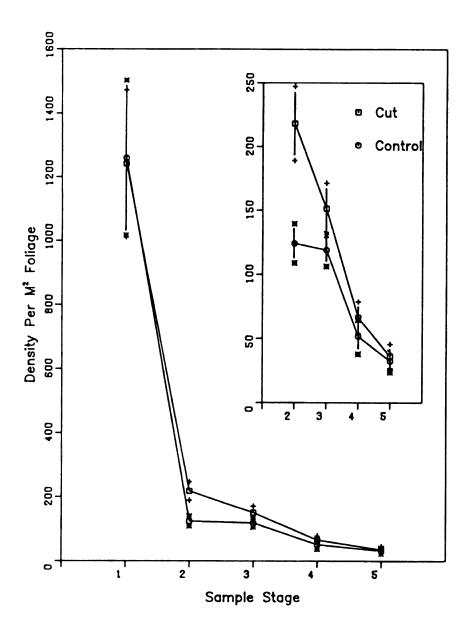


Figure 23. The density per m<sup>2</sup> foliage of various life stages of the spruce budworm in the cut and control areas of stand 1. The life stages are; 1-egg, 2-3rd instar, 3-4th and 5th instar, 4-6th instar and pupae, and 5-adult.

Table 14. Defoliation of balsam fir by spruce budworm in three stands in which part of the stand was partially harvested and the other part served as a control. Defoliation was ranked on the following scale; 0: 0-20% foliage missing, 1: 21-40% foliage missing, 2: 41-60% foliage missing, 3: 61-80% foliage missing, 4: 81-100% foliage missing. Treatment effects were tested with the Mann-Whitney U test. The sample size in each case is 60.

		Def	oliation of new foliage.	Defoliation of old foliage.		
Stand	Treatment	x	Probibility of a Type I error	x	Probibility of a Type I error	
1	cut control	1.23 1.42	.6753	1.45 1.98	.0018	
2	cut control	1.38 1.95	.0074	1.12 1.82	.0001	
3	cut control	1.57 2.70	.0001	1.37 1.53	.5277	

<sup>&</sup>lt;sup>1</sup>Defoliation occurred in 1981

<sup>&</sup>lt;sup>2</sup>Defoliation occurred in or prior to 1981

population estimates in tables 11, 12, and 13 since most defoliation by budworm result from feeding by 5th and 6th instars. This discrepancy may result from two things. First, there may have been a systematic bias in the estimation of defoliation. Second, the cut may have influenced the trees in the residual stand in some manner to produce the observed result.

In summary, no substantiating proof of the hypothesized system response can be offered. The effect of partial cuts on the dispersal loss of budworm seems to be completely opposite from what we hypothesized. In the three stands in which a partial harvest was tested, the survival of budworm from the 3rd instar to the adult life stage was less in the cut areas. It is, however, unclear as to what caused this reduction in survival. Defoliation of old and new balsam fir foliage was generally less in the cut areas than in the control plots; however, the cause of this difference is unknown.

## Conclusion

The partial harvest of spruce/fir stands has been proposed as a strategy to help reduce the impact of spruce budworm. A theoretical basis for this strategy has been developed, the feasibility of the strategy has been assessed, and a superficial check on the response of the system to the strategy has been made. At this point, sufficient information is not available to prescribe the strategy. However, when the study was initiated, there was no pretense of accomplishing such an objective.

For many years, information has been collected on the dynamics of the forest and budworm and, more recently, conceptual models have been advanced to explain these dynamics. We felt it was important to take these empirical

observations and theory and begin to mold them into a holistic strategy for dealing with the budworm problem. This report has demonstrated one way this might be done. It also serves as a basis for future work.

The major shortcoming of the proposed strategy is the unanswered question of what affect a partial harvest has on budworm and forest dynamics in the residual stands. In order for the strategy to be successful, the vulnerability of the residual stand to the budworm must be reduced. The data we collected to address this question offers no clear-cut answer. In order to adequately answer this question it will be necessary to collect information over a number of years. This is because the response of the system to a partial harvest will not be immediate and a long term series of data is indispensible for analyzing complex biological systems. Although we tried to implement the partial harvests in stands with low budworm numbers, budworm populations were well beyond the endemic level once the harvests were completed. In future work, it will be important to ensure that the strategy is implemented at low budworm densities.

Even with these shortcomings, we feel a strategy has been identified which might help forest managers deal with the spruce budworm. It certainly merits further investigation. Furthermore, the philosphy presented, that of manipulating the structure of a cropping system, should have broad applicability to other pest management problems.

### Conclusions:

The results of two studies which were motivated by the concept of designing crop production systems in order to reduce the impact of crop-damaging pests have been presented. The first study explored the dynamics of an adult parasitoid and used these findings to catalyze extensions to parasitoid host theory. Three important results were generated: (1) A methodology for assessing the dynamics of adult parasitoids was developed; (2) It was demonstrated that parasitoids respond to factors in addition to the host when searching for these organisms; and (3) the attributes of a successful biological control agent were revised, the most important attribute being host searching. The theoretical results provide a foundation for further work directed toward parasitoid management. The sampling method provides a tool for acquiring information necessary for managing these biocontrol agents. In addition, this methodology might also prove useful in on-line pest management (Tummala and Haynes 1977) if it evolves to explicitly include biological control agents.

On-line pest management seeks to use models driven by monitored environmental variables to improve knowledge of the state of crop-pest systems. Biological control agents are important components of these states. Berryman (1982) has synthesized the concept of intolerant biological control agents. An intolerant biological control agent is one which is made ineffective as a result of changes in the biotic or abiotic environment. Intolerant biological control agents may give rise to thresholds which separate endemic and epidemic pest populations. Factors which contribute to intolerance are often part of the monitored environment or part of the system structure. The sampling method presented here may be used to develop models of natural enemies driven by monitored variables and can be incorporated in on-line pest management systems.

Use of the sampling method in other systems may require further development of the model used to describe parasitoid activity. In particular, daily activity patterns should be included. This is discussed with reference to another data set in appendix 7.

The second study sought to change system structure through the use of forest harvesting in order to reduce spruce budworm damage. The feasibility of this system design rested on increasing the mortality of spruce budworm in residual forest stands. Previous studies suggested that this was a likely outcome; this study, however, did not substantiate these findings.

Both studies point to a considerable gap in empirical data and theory. This gap must be bridged if design-mediated pest management is to be successful. Undoubtedly, theory cannot do for pest management what it has done for physics, chemistry, and the engineering sciences. It can, however, serve as a template upon which empirical studies can be shaped. Studies which seek to develop crop system designs will require a long-term time commitment. This is perhaps the most serious shortcoming of the spruce budworm/forest harvesting study. Currently, there is considerable difficulty in securing funding for such long-term work. Finally, many design studies will necessarily have to be conducted on endemic populations. Before this can be done, sampling and analysis methods for such studies will have to be developed.



# Appendix 1.

In this appendix we briefly show that if the transition of an insect from one stage to another occurs via a random time delay with no mortality within and between stages, then an aggregate approximation of a population of these transitions can be made using the probability density function of the delay. Further details can be found in Pugh (1963) and Manetsch and Park (1977).

Let  $K_i^{j}(t)$  be an indicator variable which is equal to one if insect i enters stage j at time t and is zero otherwise. Define u(t) as a unit step function such that:

$$u(t) = \begin{cases} 1 \text{ for } t > 0 \\ 0 \text{ for } t \leq 0 \end{cases}$$

Also , let  $\tau_i$  be the delay in physiological time between stages j and j + 1 for insect i. We note that this variable is a random variable with probability density function f ( $\tau$ ).

For a population of n insects:

$$K_{1}^{j}(t) = K_{1}^{j-1}(t-\tau_{1})u(t-\tau_{1})$$

$$K_{n}^{j}(t) = K_{n}^{j-1}(t-\tau_{n})u(t-\tau_{n})$$
(1)

Taking the Laplace transform of both sides of (1) produces:

$$K_{i}^{j}(s) = K_{i}^{j-1}(s)e^{-\tau_{i}s}$$
 (2)

By summing both sides of (2) over i, assuming  $K_i^{j-1}$  and  $\tau_i$  are independent and taking expectations we have:

$$E[N^{j}(s)] = E[N^{j-1}(u)] E[e^{-\tau s}]$$
 (3)

where

$$N^{j}(s) = \sum_{i=1}^{n} K_{i}^{j}(s)$$

Because  $\tau$  is a random variable  $E[e^{-\tau s}]$  can be expressed as:

$$E[e^{-ts}] = \int_{a}^{\infty} f(\tau)e^{-\tau s}d\tau = F(s)$$

where F(s) is the one-sided Laplace transform.

Substituting this result in (3) yields:

$$E[N^{j}(s)] = E[N^{j-1}(s)]F(s)$$
 (4)

Multiplication in the s domain is equivalent to convolution in the time domain. Hence, assuming F(s) is time invariant (4) becomes:

$$E[n^{j}(t)] - \int_{0}^{t} E(n^{j-1}(z))f(t-z)dz.$$

In other words, the expected aggregate rate of insects entering stage j is a convolution of the rate they enter stage j-1 and the probability density function of the time interval between these stages.

If mortality does occur within or between stages this can be easily incorporated (see Manetsch and Park 1977).

# Appendix 2

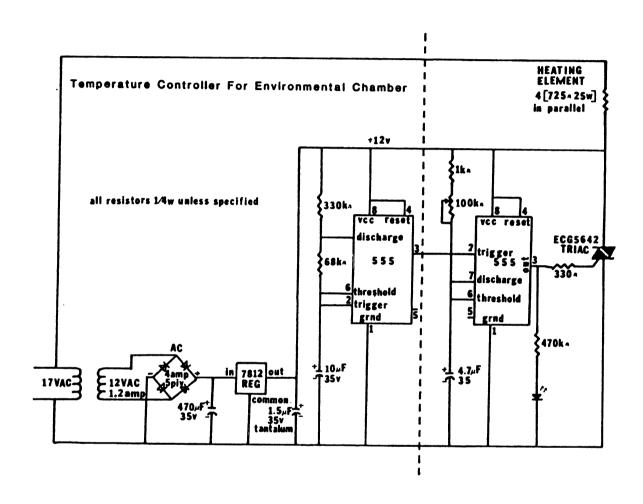


Figure 24. Circuit diagram for temperature controller of an environmental chamber. The portion to the right of the dashed line is replicated for each group of heating elements. The controller was designed by James Pieronek, Michigan State University.

## Appendix 3.

In this appendix the method employed in the analysis of the parasitoid-host model is outlined. A coupled, discrete generation parasitoid-host model was given in the main text as:

(1) 
$$H_{t+1} = F(H_t)H_tf(P_t,H_t)$$
$$P_{t+1} = H_t(1-f(P_t,H_t))$$

At equilibrium (H,P) the following conditions hold:

$$f(P, H) = F^{-1}$$

$$P = H(1-F^{-1})$$

The local stability of (1) can be determined by examining the dynamics of a pertebation model defined as

$$x_{t+1} = F(\ddot{H} + x_t)(\ddot{H} + x_t)f(\ddot{P} + y_t, \ddot{H} + x_t) - F(\ddot{H})\ddot{H}f(\ddot{P}, \ddot{H})$$

$$y_{t+1} = (\ddot{H} + x_t)(1 - f(\ddot{P} + y_t, \ddot{H} + x_t)) - \ddot{H}(1 - f(\ddot{P}, \ddot{H}))$$

and linearized about H and P.

For stability, x(t) and  $y(t) \longrightarrow 0$  as  $t \longrightarrow \infty$ .

The linearized model obtained through a Taylor expansion and after substitution of equilibrium conditions is given as

$$\begin{bmatrix} x_{t+1} \\ y_{t+1} \end{bmatrix} = \begin{bmatrix} 1 + F(H)HF(H)^{-1} + F(H)H\frac{\partial f}{\partial H} & HF(H)\frac{\partial f}{\partial P} \\ 1 - F(H)^{-1} - H\frac{\partial f}{\partial H} & -H\frac{\partial f}{\partial P} \end{bmatrix} \begin{bmatrix} x_t \\ y_t \end{bmatrix}$$
(A)

A necessary and sufficient condition for stability of the linearized model is that the dominant eigenvalue of matrix A has modulus less than 1.

For model (5) in the text an equilibrium value for  $\overset{\bigstar}{H}$  can be found by solving the function

$$H\Psi(1+\frac{aH(\Psi-1)}{\Psi K})^{-k}-H=0$$

where  $\Psi = \lambda \exp[-\gamma H/(\alpha^2 + H^2)]$ . This was done using Newton's method for determining the roots of an equation. An equilibrium value for P can then be easily found using the equilibrium conditions. Matrix A for model (5) is given as

$$A = \begin{bmatrix} 1 - \frac{(\alpha^2 - H^2)\gamma H}{(\alpha^2 + H^2)^2} & H(1 + \frac{ap}{k})^{-1} a \\ \\ 1 - (1 + \frac{ap}{k})^{-k} & H(1 + \frac{ap}{k})^{-(k+1)} a \end{bmatrix} \begin{vmatrix} * \\ H \\ * \\ P \end{vmatrix}$$

The stability region can be found by solving (2) through simulation for various values of  $\mathbf{x}_t$  and  $\mathbf{y}_t$ .

# Appendix 4.

Computer program listings.

```
*JOBCARD*, JC2000, RG2.
                                                                         100
ATTACH, B, BNPGCOMPLEX.
                                                                         110
FIN
                                                                         120
LOAD. B.
                                                                          130
LGO.
                                                                         140
*EOS
                                                                          150
C THIS PROGRAM USES AN OPTIMAZATION ALGORITHM TO ASSIGN VALUES
                                                                         160
C TO PARAMETERS OF A MODEL DESCRIBED IN SUBROUTINE FUNC. THE
                                                                         170
C OPTIMIZATION ALGORITHM IS BOX'S COMPLEX ALGORITHM DESCRIBED IN;
                                                                         180
                                                                         190
          KEUSTER, J.L., AND J.H. MIZE. 1973. OPTIMIZATION
                                                                         200
          TECHNIQUES WITH FORTRAN. MCGRAW-HILL. NEW YORK. 500PP.
                                                                         210
                                                                         220
C REQUIRED DATA IS CONTAINED IN THE SUBROUTINES AND AT THE END OF
                                                                         230
C THE PROGRAM AS FOLLOWS
                                                                         240
                                                                         250
C CARD COLUMNS CONTENTS
                                                                         260
        1 - 5
                 N (NUMBER OF VARIABLES)
                                                                         270
                 M (NUMBER OF CONSTRAINTS)
        6 - 10
C
                                                                         280
        11 - 15 K (NUMBER OF POINTS IN COMPLEX)
C 1
                                                                         290
C 1
        16 - 20 ITMAX (MAXIMUM ITERATIONS)
                                                                         300
        21 - 25 IC (NUMBER OF IMPLICIT VARIABLES)
С
  1
                                                                         310
        26 - 30 IPRINT (PRINT CONTROL)
                                                                         320
        1 - 10
                 ALPHA (REFLECTION FACTOR)
C 2
                                                                         330
        11 - 20
                 BETA (CONVERGENCE PARAMETER)
                                                                         340
        21 - 25
                 GAMMA (CONVERGENCE PARAMETER)
                                                                         350
        1 - 8
C 3
                 INITIAL X(1)
                                                                         360
        9 - 16
                                                                         370
C 3
                  INITIAL X(2)
        17 - 24 INITIAL X(3)
C 3
                                                                         380
        25 - 32 INITIAL X(4)
C 3
                                                                         390
        1 - 8
                 G(1) (LOWER CONSTRAINT)
C 4
                                                                         400
C 4
                                                                         410
                 H(1) (UPPER CONSTRAINT)
                                                                         420
C 5
                                                                         430
C SUBROUTINE FUNC
                                                                         440
                                                                         450
C PURPOSE
                                                                         460
      MODEL OF MALAISE TRAP CATCH OF GLYPTA FUMIFERANAE WHICH
                                                                         470
C
      IS BASED ON THE TEMPORAL CHANGES IN THE NUMBER OF ADULTS
C
                                                                         480
      AND WEATHER FACTORS WHICH INFLUENCE ADULT ACTIVITY. THE
                                                                         490
      SUBROUTINE IS USED BY THE OPTIMIZATION ALGORITHM TO FIT
C
                                                                         500
      PARAMETERS TO THE MODEL;
                                                                         510
           CATCH=(PROPOR+AW)+FLYAD
                                                                         520
C
       WHERE:
                                                                         530
           PROPOR IS A PROPORTIONALAITY CONSTANT
                                                                         540
           A IS A VECTOR OF PARAMETERS TO BE DETERMINED
C
                                                                         550
           W IS A VECTOR OF WEATHER FACTORS
                                                                         560
           FLYAD IS THE DENSITY OF ADULT G. FUMIFERANAE
C
                                                                         570
      THE INITIAL INPUT (PUPIN) IS ARBITRARY AS THE VARIABLES IN THE
                                                                         580
      MODEL ARE SCALED ACCORDING TO THIS VALUE
                                                                         590
                                                                         600
C DESCRIPTION OF SUBROUTINE PARAMETERS
                                                                         610
      N - NUMBER OF EXPLICIT INDEPENDENT VARIABLES
                                                                         620
      M - NUMBER OF CONSTRAINTS
                                                                         630
      K - NUMBER OF POINTS IN COMPLEX
C
                                                                         640
      X - INDEPENDENT VARIABLES (PARAMETERS TO BE FITTED TO MODEL)
                                                                         650
      F - OBJECTIVE FUNCTION
                                                                         660
      I - POINT INDEX
                                                                         670
                                                                         680
C DESCRIPTION OF VARIABLES USED IN SUBROUTINE
                                                                         690
C
      RP. RA1 - INTERMEDIATED RATES IN DISTRIBUTED DELAYS
                                                                         700
С
      NDD48 - DEGREE DAYS BASE 48F PER DAY
                                                                         710
      PRECIP - PRECIPTATION IN INCHES BETWEEN 800 AND 2200 HRS
                                                                         720
```

```
RH - AVERAGE RELATIVE HUMIDITY BETWEEN 800 AND 2200 HRS
                                                                         730
С
      TEMP - AVERAGE TEMPERATURE (F) BETWEEN 800 AND 2200 HRS
                                                                         740
C
С
      TRUCAT - ACTUAL CATCH OF G. FUMIFERANAE IN MALAISE TRAPS
                                                                         750
                                                                          760
C
                                                                          770
С
      THE OPTIMIZATION ALGORITHM ASSIGNS VALUES TO X TO MINIMIZE THE
                                                                          780
С
      SUM OF THE SQUARED DIFFERENCES BETWEEN THE TRUE AND PREDICTED
                                                                         790
C
      TRAP CATCH. IT USES SUBROUTINE DELAY2 WHICH IS A DISTRIBUTED
                                                                         800
C
                                                                          810
      DELAY AND IS DESCRIBED IN
C
                                                                          820
С
           MANETSCH.T.J., AND G.L. PARK. 1974. SYSTEM ANALYSIS AND
                                                                          830
С
С
           SIMULATION WITH APPLICATION TO ECONOMIC AND SOCIAL SYSTEMS
                                                                         840
Ċ
           PART II. MICH. STATE UNIV., E.LANSING. 239PP.
                                                                          850
                                                                         860
С
                                                                          870
C
                                                                         880
      SUBROUTINE FUNC(N,M,K,X,F,I)
      DIMENSION X(20,20), F(20), RP(10), RA1(10), NDD48(27)
                                                                         890
     +,PRECIP(27),TRUCAT(27),TEMP(27),RH(27)
                                                                          900
                                                                         910
C ASSIGN VALUES TO VARIABLES
      DATA RP /0.,0.,0.,0.,0.,0.,0.,0.,0.,0.
                                                                         920
      DATA RA1 /0.,0.,0.,0.,0.,0.,0.,0.,0.,0./
                                                                         930
      DATA NDD48 /24,22,23,22,14,15,16,18,14,20,12,5,11.
                                                                         940
                                                                         950
     +14, 19, 12, 8, 11, 11, 15, 19, 24, 22, 21, 20, 17, 19/
                                                                         960
      DATA PRECIP /0.,0.,0.,0.,0.,01,.26,0.,0.,.4,.3,0.,
     +0.,0.,.35,0.,.32,.11,0.,0.,0.,2.34,0.,.36,.01,0./
                                                                          970
      DATA RH /34.8,59.8,74.,46.8,60.5,52.6,73.4,89.7,80.,65.5,
                                                                         980
     +93.1,66.6,49.5,56.6,64.5,79.1,59.2,44.5,49.2,53.1,57.1,
                                                                         990
                                                                          1000
     +61.6,85.6,78.8,84.5,80.2,54.9/
      DATA TEMP /.809,.765,.757,.755,.663,.698,.661,.698,.656,
                                                                          1010
                                                                          1020
     +.735,.623,.552,.646,.690,.712,.612,.608,.646,.629,.711,
                                                                          1030
     +.726,.773,.708,.718,.709,.675,.737/
      DATA TRUCAT /0.,0.,1.,16.,29.,25.,21.,16.,25.,33.,7.,
                                                                          1040
                                                                          1050
     +7.,18.,30.,28.,5.,6.,4.,1.,8.,4.,5.,4.,1.,1.,0.,0./
C DELAY OF PUPAL STAGE
                                                                          1060
                                                                          1070
      DELPUP = 100
C K'S OF ERLANG DENSITY FUNCTIONS USED IN DISTRIBUTED DELAYS
                                                                          1080
                                                                          1090
      KPIIP=4
                                                                          1100
      KAD=8
C INITIALIZE STATE VARIABLES
                                                                          1110
                                                                          1120
      ADULT=RMORT=TADULT=TMORT=O.
                                                                          1130
      DT=.125
                                                                          1140
C INITIALIZE INPUT INTO SYSTEM
                                                                          1150
      PUPIN=50/DT
C ASSIGN VARIABLES IN MODEL TO DECISION VARIABLES OF OPTIMIZATION
                                                                          1160
                                                                          1170
C ALGORITHM
      DELAD=X(I,1)
                                                                          1180
                                                                          1190
      PROPOR=X(I,2)
                                                                          1200
      ALPHA=X(I,3)
                                                                          1210
      BETA=X(I,4)
                                                                          1220
C SET OBJECTIVE FUNCTION TO ZERO
                                                                          1230
      U=O.
                                                                          1240
C DETERMINE SIMULATION ITERATIONS
                                                                          1250
C SIMULATION IS RUN FOR N DAYS AND X DD48 FOR EACH DAY
                                                                          1260
      T=0.0
                                                                          1270
      DO 10 I1=1,27
      IEND=NDD48(I1)/DT+.00001
                                                                          1280
                                                                          1290
      NN1=IEND/2
C NN1 IS THE DEGREE DAYS MIDWAY THROUGH A DAY. ADULT DENSITY AT
                                                                          1300
C THIS TIME IS USED TO RELATE TRAP CATCH TO OTHER VARIABLES
                                                                          1310
      DO 20 12=1, IEND
                                                                          1320
                                                                          1330
      T=T+DT
```

```
C CALL DELAYS WHICH MODEL THE PUPAL AND ADULT LIFE STAGE
                                                                           1340
      CALL DELAY2(ADULT, RMORT, RA1, DELAD, DT, KAD)
                                                                           1350
      CALL DELAY2(PUPIN, ADULT, RP, DELPUP, DT, KPUP)
                                                                           1360
C COMPUTE STORAGE IN DELAYS WHICH IS THE NUMBER OF PUPAE OR
                                                                           1370
C ADULTS AT ANY POINT IN TIME (PUP, FLYAD)
                                                                           1380
                                                                           1390
      VAR1=VAR2=O.
                                                                           1400
      DO 15 J=1.KPUP
                                                                           1410
      VAR1=VAR1+RP(J)
15
                                                                           1420
      DO 16 J=1,KAD
                                                                           1430
      VAR2=VAR2+RA1(J)
16
                                                                           1440
      PUP=DELPUP/KPUP+VAR1
                                                                           1450
      FLYAD=DELAD/KAD+VAR2
                                                                           1460
      IF (I2.EQ.NN1) FLYAD2=FLYAD
      PUPIN=O.
                                                                           1470
                                                                           1480
      CONTINUE
20
C COMPUTE MULTIPLIER TO RELATE TRAP CATCH, ADULT DENSITY AND
                                                                           1490
                                                                           1500
C EXTRINSIC FACTORS
      RMULT=PROPOR+(ALPHA+PRECIP(I1))+(BETA+TEMP(I1))
                                                                           1510
      IF (RMULT.LT.O) RMULT=O.
                                                                           1520
                                                                           1530
      CATCH=RMULT+FLYAD2
C COMPUTE OBJECTIVE FUNCTION
                                                                           1540
                                                                           1550
      U=U+(CATCH-TRUCAT(I1))++2
                                                                           1560
      F(I)=-U
                                                                           1570
      CONTINUE
10
                                                                           1580
      END
                                                                           1590
C
                                                                           1600
С
С
                                                                           1610
      SUBROUTINE DELAY2 (RINR, ROUTR, CROUTR, DEL, DT, K)
                                                                           1620
                                                                           1630
      DIMENSION CROUTR(1)
      DEL1=DEL/(FLOAT(K)+DT)
                                                                           1640
                                                                           1650
      RIN=RINR
                                                                           1660
      DO 1 I=1,K
      ABC=CROUTR(I)
                                                                           1670
      CROUTR(I)=ABC+(RIN-ABC)/DEL1
                                                                           1680
                                                                           1690
1
      RIN=ABC
                                                                           1700
      ROUTR=CROUTR(K)
      RETURN
                                                                           1710
                                                                           1720
      END
                                                                           1730
C
                                                                           1740
С
                                                                           1750
С
C SUBROUTINE CONST
                                                                           1760
                                                                           1770
C PURPOSE
                                                                           1780
      SPECIFIES EXPLICIT AND IMPLICIT CONSTRAINT LIMITS. IT
                                                                           1790
С
C
      MUST BE INCLUDED EVEN IF NO IMPLICIT CONSTRAINTS ARE USED.
                                                                           1800
      PARAMETERS ARE DEFINED ABOVE.
                                                                           1810
С
                                                                           1820
С
      SUBROUTINE CONST(N,M,K,X,G,H,I)
                                                                           1830
                                                                           1840
      DIMENSION X(20,20),G(20).H(20)
      RETURN
                                                                           1850
                                                                           1860
      END
*EOS
                                                                           1870
                                                                           1880
                 100
                         0
                              0
               8
                                                                           1890
       1.3
                 50.0
                         5
180.0
                                                                           1900
        0.75
                         0.0
                 0.0
                                                                           1910
110.0
        0.0
                 -2.0
                         0.0
                                                                           1920
250.0
                 2.0
                         2.0
        2.0
                                                                           1930
*EOP
```

```
PROGRAM BOX(INPUT, OUTPUT)
      DIMENSION X(20,20),R(20,20),F(20),G(20),H(20),XC(20)
      INTEGER GAMMA
      KOUNT = O
                                                                  5
      READ 1.N.M.K.ITMAX, IC, IPRINT
                                                                  6
      FORMAT (815)
                                                                  7
      READ 2, ALPHA, BETA, GAMMA
      FORMAT(2E10.4,15)
2
                                                                  9
      DELTA=0.0001
                                                                  10
      READ 4. (X(1,J).J=1,N)
                                                                  11
      READ 4. (G(I), I=1.M)
      READ 4. (H(I), I=1,M)
                                                                  12
                                                                   13
      FORMAT(9F8.4)
                                                                   14
      DO 100 II=2,K
                                                                   15
      DO 3 JJ=1,N
                                                                   16
      R(II,JJ)=RANF(1.0)
                                                                   17
100
      CONTINUE
                                                                  18
      PRINT 10
      FORMAT(1H1.//, 18X, 24HCOMPLEX PROCEDURE OF BOX)
                                                                  19
10
                                                                  20
      PRINT 18
18
      FORMAT(//, 2X, 10HPARAMETERS )
                                                                  21
      PRINT 11,N,M,K,ITMAX,IC,ALPHA,BETA,GAMMA,DELTA
                                                                  22
  O11 FORMAT (//.2X.4HN = .I2.3X.4HM = .I2.3X.4HK = .I2.2X.8HI23
                                                                  23.5
     +TMAX =
     114,2X,5HIC = ,12,//.2X,8HALPHA = ,F5.2,5X,7HBETA = ,F10.24
     +5.3X.
                                                                  24.5
     28HGAMMA = ,12,3X,8HDELTA = ,F6.5)
                                                                  25
                                                                  26
      PRINT 900
900
      FORMAT(//,2X,16HINITIAL X VALUES)
                                                                  27
                                                                  28
      DO 200 J = 1,N
                                                                  29
      PRINT 16, J, X(1,J)
      CONTINUE
   50 CALL CONSX (N.M.K.ITMAX,ALPHA.BETA,GAMMA,DELTA,X,R.F.IT.31
                                                                  32
     11EV2,NO,G,H,XC,IPRINT)
                                                                  33
      IF (IT-ITMAX) 20,20,30
                                                                  34
   20 PRINT 14. F(IEV2)
  O14 FORMAT (///.2X.30HFINAL VALUE OF THE FUNCTION = .E20.8) 35
      PRINT 15
                                                                  36
                                                                  37
  O15 FORMAT (//,2X,14HFINAL X VALUES)
                                                                  38
      DO 300 J=1.N
  PRINT 16, J, X(IEV2,J)
016 FORMAT (/,2X,2HX(.I2,4H) = ,E20.8)
                                                                  39
                                                                  40
  300 CONTINUE
                                                                  41
                                                                  42
      PRINT 9, IT
      FORMAT (//* THE NUMBER OF ITERATIONS IS
                                                   *. I5)
                                                                  43
                                                                  44
      GO TO 999
                                                                  45
  30 PRINT 17, ITMAX
  017 FORMAT (///.2X.38HTHE NUMBER OF ITERATIONS HAS EXCEEDED 46
                                                                  46.5
     +, 14, 10X,
     118HPROGRAM TERMINATED)
                                                                  47
                                                                  48
  999 STOP
                                                                  49
      END
      SUBROUTINE CONSX (N.M.K.ITMAX, ALPHA, BETA, GAMMA, DELTA, X, R50
                                                                  50.5
                                                                  51
     1IT, IEV2, NO. G, H, XC, IPRINT)
      DIMENSION X(20,20),R(20,20),F(20),G(20),H(20),XC(20)
                                                                  52
                                                                  53
      INTEGER GAMMA
                                                                  54
      IT = 1
                                                                  55
      KODE = O
      IF (M-N) 20,20,10
                                                                  56
                                                                  57
   10 KODE = 1
   20 CONTINUE
                                                                  58
                                                                  59
      DO 40 II=2.K
```

```
DO 30 J=1.N
                                                                   60
   30 \times (II,J) = 0.0
                                                                   61
   40 CONTINUE
                                                                   62
                                                                   63
      DO 65 II=2,K
      DO 50 J=1.N
                                                                   64
                                                                   65
      I = II
                                                                   66
      CALL CONST (N,M,K,X,G,H,I)
      X(II,J) = G(J) + R(II,J) \cdot (H(J) - G(J))
                                                                   67
                                                                   68
   50 CONTINUE
      K1 = II
                                                                   69
      CALL CHECK (N.M.K.X.G.H.I.KODE.XC.DELTA.K1)
                                                                   70
                                                                   71
      IF (II-2) 51, 51, 55
                                                                   72
   51 IF (IPRINT) 52,65,52
                                                                   73
   52 PRINT 18
  O18 FORMAT (//.2X.30HCOORDINATES OF INITIAL COMPLEX)
                                                                   74
                                                                   75
      IO = 1
  PRINT 19. (IO. J. X(IO.J). J=1.N)
019 FORMAT (/.3(2X.2HX(.I2.1H..I2.4H) = .1PE13.6))
                                                                   76
                                                                   77
                                                                   78
   55 IF (IPRINT) 56,65,56
                                                                   79
   56 PRINT 19, (II, J, X(II,J), J=1.N)
                                                                   80
   65 CONTINUE
                                                                   81
      K1 = K
                                                                   82
      DO 70 I=1,K
      CALL FUNC(N,M,K,X,F,I)
                                                                   83
                                                                   84
70
        CONTINUE
                                                                   85
       KOUNT = 1
                                                                   86
       IA=O
                                                                   87
      IF(IPRINT) 72.80.72
                                                                   88
72
      PRINT 21
      FORMAT(/.2X.22HVALUES OF THE FUNCTION )
                                                                   89
21
                                                                   90
      PRINT 22 , (J,F(J), J=1,K)
22
      FORMAT(/,3(2X,2HF(,12,4H) = ,1PE13.6))
                                                                   91
                                                                   92
80
      IEV1=1
                                                                   93
      DO 100 ICM=2,K
       IF(F(IEV1)-F(ICM)) 100,100,90
                                                                   94
                                                                   95
90
      IEV1=ICM
                                                                   96
      CONTINUE
100
                                                                   97
      IEV2=1
      DO 120 ICM=2,K
                                                                   98
      IF(F(IEV2)-F(ICM)) 110,110,120
                                                                   99
  110
        IEV2=ICM
                                                                   100
                                                                   101
120
      CONTINUE
      IF(F(IEV2)-(F(IEV1)+BETA)) 140,130,130
                                                                   102
                                                                   103
       KOUNT = 1
130
                                                                   104
      GO TO 150
        KOUNT = KOUNT + 1
                                                                   105
140
                                                                   106
      IF (KOUNT-GAMMA) 150, 240, 240
       CALL CENTR(N,M,K,IEV1,I,XC,X,K1)
                                                                   107
150
                                                                   108
      DO 160 JJ=1,N
       X(IEV1,JJ)=(1.0+ALPHA)+(XC(JJ))-ALPHA+(X(IEV1,JJ))
                                                                   109
160
                                                                   110
      I=IEV1
      CALL CHECK(N,M,K,X,G,H,I,KODE,XC,DELTA,K1)
                                                                   111
      CALL FUNC(N,M,K,X,F,I)
                                                                   112
                                                                   113
       IEV2=1
170
                                                                   114
      DO 190 ICM=2,K
      IF(F(IEV2)-F(ICM)) 190.190.180
                                                                   115
                                                                   116
180
      IEV2=ICM
                                                                   117
190
      CONTINUE
      IF(IEV2-IEV1) 220,200,220
                                                                   118
                                                                   119
200
      DO 210 JJ=1.N
                                                                   120
      X(IEV1,JJ)=(X(IEV1,JJ)+XC(JJ))/2.0
```

```
121
       CONTINUE
210
                                                                  122
      I=IEV1
      CALL CHECK(N.M.K.X.G.H.I.KODE.XC.DELTA.K1)
                                                                  123
                                                                  124
      CALL FUNC(N.M.K.X.F.I)
                                                                  125
       GO TO 170
                                                                  126
220
       CONTINUE
                                                                  127
      IF (IPRINT) 230,228,230
      PRINT 23.IT
                                                                  128
230
      FORMAT(//.2X.17HITERATION NUMBER .15)
                                                                  129
23
                                                                  130
      PRINT 24
      FORMAT(/.2X.30HCOORDINATES OF CORRECTED POINT)
                                                                  131
24
      PRINT 19. (IEV1, JC, X(IEV1, JC), JC=1,N)
                                                                  132
                                                                  133
      PRINT 21
                                                                  134
      PRINT 22, (I,F(I), I=1,K)
                                                                  135
      PRINT 25
      FORMAT(/,2X,27HCOORDINATES OF THE CENTROID)
                                                                  136
25
      PRINT 26. (JC,XC(JC),JC=1.N)
                                                                  137
                                                                  138
      FORMAT(/,3(2X,2HX(,12.6H,C) = ,1PE14.6.4X))
26
      IT=IT+1
                                                                  139
228
                                                                  140
      IF(IT-ITMAX) 80,80,240
                                                                  141
240
      RETURN
      END
                                                                  142
                                                                  143
      SUBROUTINE CHECK(N.M.K.X.G.H.I.KODE.XC.DELTA.K1)
      DIMENSION X(20,20),G(20),H(20), XC(20)
                                                                  144
                                                                  145
10
       KT=0
                                                                  146
      CALL CONST(N,M,K,X,G,H,I)
                                                                  147
      DO 50 J=1.N
                                                                  148
      IF(X(I,J)-G(J)) 20,20,30
       X(I,J) = G(J) + DELTA
                                                                  149
20
                                                                  150
      GO TO 50
      IF(H(J)-X(I,J)) 40.40.50
                                                                  151
30
                                                                  152
      X(I,J)=H(J)-DELTA
40
                                                                  153
50
      CONTINUE
      IF(KODE) 110, 110,60
                                                                  154
                                                                  155
60
      NN=N+1
                                                                  156
      DO 100 J=NN.M
                                                                  157
      CALL CONST(N.M.K.X.G.H.I)
                                                                  158
      IF(X(I,J)-G(J)) 80,70.70
      IF(H(J)-X(I,J))80,100,100
                                                                  159
70
                                                                  160
80
      IEV1=I
                                                                  161
      KT = 1
      CALL CENTR (N.M.K.IEV1, I.XC, X, K1)
                                                                  162
      DO 90 JJ=1,N
                                                                  163
      X(I,JJ) = (X(I,JJ) + XC(JJ))/2.0
                                                                  164
                                                                  165
90
      CONTINUE
      CONTINUE
                                                                  166
100
                                                                  167
      IF (KT) 110, 110, 10
                                                                  168
      RETURN
110
                                                                  169
      END
                                                                  170
      SUBROUTINE CENTR (N.M.K.IEV1, I.XC.X.K1)
                                                                  171
      DIMENSION X(20,20),XC(20)
                                                                  172
      DO 20 J=1.N
                                                                  173
      XC(J) = 0.0
                                                                  174
      DO 10 IL=1.K1
                                                                  175
   10 XC(J) = XC(J) + X(IL.J)
                                                                  176
      RK=K1
                                                                  177
   20 \text{ XC(J)} = (\text{XC(J)-X(IEV1,J)})/(\text{RK-1.0})
                                                                  178
      RETURN
                                                                  179
      END
```

```
PROGRAM STABLE (INPUT, OUTPUT)
                                                                 100
C THIS PROGRAM COMPUTES THE REGION OF STABILITY FOR STABLE
                                                                 110
C EQUILIBRIA OF THE FOLLOWING MODEL:
                                                                 120
      H(T+1)=H(T)EXP(-BH(T)/C**2+H(T)**2)(1+AP/K)**-K
                                                                 130
C
                                                                 140
      P(T+1)=H(T)(1-(1+AP/K)**-K)
C DATA REQUIREMENTS ARE EQUILIBRIA VALUES AND VALUES FOR
                                                                 150
                                                                 160
C PARAMETERS AND ARE INPUT IN DATA STATEMENTS.
C THE STABILITY REGION IS DETERMINED ITTERATIVELY BY
                                                                 170
C PERTERBING HE BY SOME VALUE DELTA AND CHECKING THROUGH
                                                                 180
C SIMULATION CONVERGENCE OF THE MODEL BACK TO HE AND PE.
                                                                 190
C PRECISION IS DETERMINED BY THE TEST VALUE OF DO LOOP 1.
                                                                 200
                                                                 210
C K=O FLAGS K=INFINITY
      DIMENSION DRK(4), DA(4), DHE(4,4), DPE(4,4)
                                                                 220
                                                                 230
      DATA DRK/.75,2.0,6.0,0./
      DATA DA/1.5,2.0,2.5,3.0/
                                                                 240
      DATA (DHE(1,M),M=1,4)/1.12039,.85213,.73440,.66097/.
                                                                 250
                                                                 260
     +(DHE(2,M),M=1,4)/.91802,.73042,.62695,.55780/,
                                                                 270
     +(DHE(3,M),M=1,4)/.85923,.68300,.58156,.51270/
     +, (DHE(4,M),M=1,4)/.83137,.65886,.55793,.48903/
                                                                 280
      DATA (DPE(1,M),M=1,4)/.39855,.30951,.29494,.28999/.
                                                                 290
                                                                 300
     +(DPE(2,M),M=1,4)/.32348,.29462,.28796,.28280/,
     +(DPE(3,M),M=1,4)/.31079,.29133,.28484..27710/
                                                                 310
     +, (DPE(4,M), M=1,4)/.30607,.28987,.28281,.27422/
                                                                 320
                                                                 330
      Y = 10.0
      B=3.75
                                                                 340
                                                                 350
      C=1.0
                                                                 360
      DO 300 L=1.4
                                                                 370
      DO 301 M=1,4
      RK=DRK(L)
                                                                 380
                                                                 390
      A=DA(M)
                                                                 400
      HS=DHE(L,M)
      PS=DPE(L.M)
                                                                 410
      PRINT 100, B,C,Y,A,RK,HS,PS
                                                                 420
      FORMAT ("OB=",F4.2,1X,"C=",F3.1,1X,"Y=",F4.1,1X,"A=",F3.430
100
                                                                 435
     +1.1X.
     +"RK=",F3.1,1X,"HE=",F7.5,1X,"PE=",F7.5)
                                                                 440
      PRINT 101
                                                                 450
      FORMAT ("OMAX HOST AND PARASITE DENSITIES FOR WHICH MODE460
101
     +L IS STABL
                                                                 465
     +E")
                                                                 470
                                                                 480
      SIGMA=.05
      DO 1 I=1,20
                                                                 490
                                                                 500
      DO 2 I2=1,2
      SIGN=1.
                                                                 510
                                                                 520
      IF(12.EQ.2)SIGN=-1.
      P1=PS+(SIGMA+I+SIGN)
                                                                 530
      IF(P1.LT.O.) GO TO 21
                                                                 540
                                                                 550
      P=P1
                                                                 560
      CHECK=5+HS
                                                                 570
      DO 3 J=1,2
      SIGN=1.
                                                                 580
      IF(J.EQ.2)SIGN=-1.
                                                                 590
                                                                 600
      DELTA=.05
                                                                 610
      H1=HS
                                                                 620
      CONTINUE
      DELTA=DELTA+2
                                                                 630
      H3=HS+(DELTA+SIGN)
                                                                 640
                                                                 650
      IF(H3.LT.O.) H3=.001
                                                                 660
      H=H3
                                                                 670
      CALL DYNAM (P.H. INDEX. Y.B.C.RK. A. CHECK)
      P=P1
                                                                 680
                                                                 690
      IF(INDEX.EQ. 1) GO TO 4
                                                                 700
      CONTINUE
```

```
710
      IF(H3.LT.H1) VAR1=H1-H3
      IF(H1.LT.H3) VAR1=H3-H1
                                                                 720
                                                                 730
      IF(VAR1.LT..005) GO TO 6
      H2=(H1+H3)/2
                                                                 740
                                                                 750
      H=H2
      CALL DYNAM (P,H,INDEX,Y,B,C,RK,A,CHECK)
                                                                 760
                                                                 770
      P=P1
                                                                 780
      IF(INDEX.EQ.1) H1=H2
      IF(INDEX.EQ.O) H3=H2
                                                                 790
                                                                 800
      GO TO 5
      CONTINUE
                                                                 810
6
      PRINT 102, P1, H1
                                                                 820
      FORMAT (" PARASITE DENSITY = ",F7.5, "HOST DENSITY = ",F7.5830
102
                                                                 840
3
      CONTINUE
      GO TO 25
                                                                 850
                                                                 860
21
      CONTINUE
      PRINT 22
                                                                 870
      FORMAT (" PARASITE DENSITY LESS THAN ZERO")
                                                                 880
22
                                                                 890
25
      CONTINUE
                                                                 900
      CONTINUE
2
                                                                 910
      CONTINUE
301
      CONTINUE
                                                                 920
                                                                 930
      CONTINUE
300
                                                                 940
      END
                                                                 950
С
                                                                 960
С
Č
                                                                 970
      SUBROUTINE DYNAM (P.H.INDEX, Y.B.C.RK, A.CHECK)
                                                                 980
      INDEX=1
                                                                 990
                                                                 1000
      DO 1 I=1,100
                                                                 1010
      IF(RK.EQ.O.) GO TO 10
      V=(1/(1+A+P/RK))++RK
                                                                 1020
                                                                 1030
      GO TO 11
      CONTINUE
                                                                 1040
10
                                                                 1050
      V=EXP(-A+P)
      CONTINUE
                                                                 1060
11
                                                                 1070
      P=H+(1-V)
      H=H+Y+EXP(-B+H/((C++2)+(H++2)))+V
                                                                 1080
                                                                 1090
      IF(H.GT.CHECK) INDEX=0
                                                                 1100
      CONTINUE
                                                                 1110
      RETURN
                                                                 1120
      END
```

```
PROGRAM EIGENV (INPUT, DUTPUT)
C THIS PROGRAM COMPUTES THE EIGENVALUES OF THE FOLLOWING PARASI10
C ITE
C HOST MODEL LINEARIZED ABOUT THE EQUILIBRIUM POINT
                                                                120
                                                                130
      H(T+1)=H(T)YEXP(-BH(T)/C++2+H(T)++2)((1+AP/K)++-K)
C
                                                                 140
C
      P(T+1)=H(T)(1-(1+AP/K)++-K)
C DATA REQUIREMENTS ARE VALUES FOR ALL PARAMETERS AND EQUILIBR150
C TUM
C DENSITY OF THE HOST POPULATION (K=O FLAGS INFINITY, H=O FLAG160
C S NO
                                                                 170
C EQUILIBRIUM)
      DIMENSION DC(3), DRK(4), DA(5), DH(3,4,5)
                                                                 180
      DATA (DH(1,1,1),1=1,5)/.66097,.7344,.85213,1.12039,0./. 190
     +(DH(1,2,I),I=1,5)/.5578,.62695,.73042,.91902,1.53906/.
                                                                200
     +(DH(1,3,I),I=1,5)/.5127,.58156,.683,.85923,1.32473/,
                                                                210
     +(DH(1,4,I),I=1,5)/.48903,.55793,.65886,.83137,1.25647/
                                                                220
     +,(DH(2,1,I),I=1,5)/.76831,.86262,1.02186,1.47683,0./,
                                                                230
     +(DH(2,2,I),I=1,5)/.61619,.69588,.81580,1.03479,1.74045/,231
     +(DH(2,3,I),I=1,5)/.55489,.63154,.74510,.94161,1.44326/, 232
     +(DH(2,4,1),I=1,5)/.52304,.59908,.71039,.89939,1.34973/, 233
     +(DH(3,1,I),I=1,5)/.55807,.61367,.69855,.86409.0./,
                                                                234
     +(DH(3,2,I),I=1,5)/.49852,.55404,.64030,.79407,1.29533/,
                                                                235
     +(DH(3,3,I),I=1,5)/.46634..52631..61441..76741,1.18364/.
                                                                236
     +(DH(3,4,I),I=1,5)/.45033,.51133,.60064,.75400,1.14571/
                                                                237
                                                                270
      DATA DA/3.,2.5,2.,1.5,1./
      DATA DC/1.,1.1,.9/
                                                                280
                                                                290
      DATA DRK/.75,2.,6.,0./
                                                                300
      Y = 10.
                                                                310
      B = 3.75
                                                                320
      DO 1 L=1.3
                                                                330
      DO 2 M=1.4
                                                                340
      DO 3 N=1.5
                                                                350
      FLAG=0.
                                                                360
      C=DC(L)
      RK=DRK(M)
                                                                370
                                                                380
      A=DA(N)
                                                                390
      H=DH(L,M,N)
      PRINT 10, C, RK, A, H
                                                                400
      FORMAT ("OC= ",F3.1,1X,"K= ",F4.2,1X,"A= ",F3.1,1X,"HOST410
10
     + EOUIL= ".F7.3)
                                                                420
                                                                430
      IF(H.EQ.O.) GO TO 100
                                                                440
C COMPUTE PARASITE EQUIL. DENSITY
                                                                450
      F=Y*EXP(-B*H/((C**2)+(H**2)))
      P=((F-1)/F)+H
                                                                460
                                                                470
C COMPUTE FOUR TERMS OF MATRIX FOR LINEARIZ
                                                MODEL
      SIGN=-1
                                                                480
                                                                490
      X=C++2
                                                                500
      V=H++2
      T11=(1-(((X-V)+B+H)/((X+V)++2)))+SIGN
                                                                510
                                                                520
C IF K=INFINITY USE POISSON ZERO TERM
      IF(RK.EQ.O.) GO TO 110
                                                                530
      W=(A+P/RK)+1
                                                                540
                                                                550
      T12=H+A/W
                                                                560
      T21=(1-((1/W)**RK))*SIGN
                                                                570
      T22=(((1/W)++(RK+1))+H+A)+SIGN
                                                                571
      GO TO 120
                                                                580
110
      CONTINUE
      W=-A+P
                                                                590
      T12=W+H+SIGN
                                                                600
      T21=(1-EXP(W))+SIGN
                                                                610
                                                                620
      T22=H+EXP(W)+A+SIGN
                                                                621
     CONTINUE
120
C TERMS OF QUADRATIC EQUATION
                                                                630
```

```
QT1=((T11+T22)/2.) *SIGN
                                                                640
      QI1=T11+T22
                                                                650
      QI2=T12+T21
                                                                660
      QI3=QI1-QI2
                                                                670
      QI4=((QT1+2)++2)-(4+QI3)
                                                                680
      IF(Q14.LT.O.) FLAG=1
                                                                690
                                                                700
      IF(QI4.LT.O.) QI4=QI4+SIGN
      QT2=(SQRT(QI4))/2
                                                                710
      PRINT 20,QT1,QT2
                                                                720
20
      FORMAT("OEIGENVALUES= ",F7.5,1X,"PLUS AND MINUS",1X,F7.5730
                                                                735
      IF(FLAG.EQ.O.) GO TO 3
                                                                740
      PRINT 30
FORMAT (" PLUS AND MINUS TERM IS COMPLEX")
                                                                750
30
                                                                760
      GO TO 3
                                                                770
100
      CONTINUE
                                                                780
      PRINT 40
                                                                790
40
      FORMAT (" NO EQUIL. FOUND, NO EIGENVALUES COMPUTED")
                                                                800
      CONTINUE
3
                                                                810
2
      CONTINUE
                                                                820
      CONTINUE
                                                                830
      END
                                                                840
```

## Appendix 5.

In this appendix we show why a partial cutting strategy is probably not compatible with maximizing economic returns with least risk. The return  $(R_t)$  from the sale of a tract of timber at time t is determined by price  $(P_t)$  and volume  $(V_t)$ :

$$R_t = P_t \cdot V_t \tag{1}$$

We will assume that the only costs associated with growing timber are foregone returns from immediate harvest of the trees. (The conclusions reached will be the same even if this is not true). The value (Val<sub>t</sub>) of a stand at some future point in time (t) is then given by:

$$Val_t = R_t - R_{t_0} e^{i(t-t_0)}$$
 (2)

In (2),  $R_{t_0}$  are the current returns and i is the interest rate obtained on investments. The exponential function is used to approximate the future value of the current stand.

Suppose some proportion ( $\delta$ ) of the stand is harvested prior to a budworm outbreak where  $0 \le \delta \le 1$ . Then (2) can be written as:

$$Val_{t} = (1 - \delta)R_{t} + (2\delta - 1)R_{t_{0}}e^{i(t-t_{0})}$$
(3)

The rate at which the value of the stand increases is given by:

$$\frac{dVal_t}{dt} = (1 - \delta) dR_t/dt + (2\delta - 1)iR_{to}e^{i(t-t_0)}$$
(4)

This is a complicated function; however, its general nature can be determined by evaluating three harvesting policies.

If the complete stand is harvested at time  $t_0$ , then  $\delta = 1$ , and (4) becomes:

$$dVal_t/dt = iR_{t_O}e^{i(t-t_O)}$$
 (5)

The rate of increase of the stand is equal to the rate of return on an investment. If it is possible to harvest the stand, this policy entails no risk.

If none of the stand is harvested prior to the outbreak, (4) becomes:

$$dVal_t/dt = dR_t/dt - iR_{t_0}e^{i(t-t_0)}$$
(6)

In order that the rate of increase in value be positive, price and volume must increase in combination at a rate faster than returns are accrued from an immediate harvest. This is an unlikely situation since the volume of spruce fir stands greater than 50 years in age generally increases at a rate less than or equal to 5% of the present volume (Bowman 1944). This increase may decline or even be less than zero during a budworm outbreak. It is even more improbable that this policy will be superior to the first one or:

$$dR_t/dt > 2iR_{t_0}e^{i(t-t_0)}$$
 (7)

If half of the stand is harvested prior to an outbreak, (4) becomes:

$$dVal_{t}/dt = .5dR_{t}/dt$$
 (8)

For this policy to be superior to the first one, equation (7) must be satisfied. Once again, this is unlikely. Therefore, complete presalvage is the best strategy for maximizing returns and minimizing risks.

Appe	ndix	6.								
Form	sent	to	loggers	to	determine	costs	and	returns	of	harvests
Loca	tion:									
Sale	number	::								
Logge	er:									
thos	Please e estim	e fil	l in the you cons	blani ider	cs as accurat to be very i	cough est	ossib timate	le. Write s.	an	(X) by
A. 3	Wages									
	1. Num	nber	of people	етр.	loyed in the	harvest	of th	is sale ar	ea _	
	2. Num	ber	of days s	pent	working in	this area	·			
	3. Hou	ırly	wage Tate	and	total 'man-	nours' w	orked	at each ra	te	
	<u> </u>	7age	rate	tota	l hours	wage	rate	total h	ours	,
	1) _					4)				
	2) _					5)				
	3) _					5)				•
	4. Ins	surar	nce cost (	Work	man's Comp.)					
	5. Oth									
<b>3.</b> 1	Equipme	nt								
•	l. Wha	t eq	uipment w	as us	sed to harve:	t this	sale a	rea?		
	<b>a</b> )									
	ь)				<del></del>					
	c)									
1					for each equ			bove:		
					-	-				
	2. Inv	es to	ent in eq	uipm	ent (origina)	cost)				

	Fuel cost					
	a)		d)			
	b)					
	c)		-/	<del></del>	<del></del>	
4.	Maintenance co					
•	a)		4)			
	b)					
	c)					
٥.	Insurance cost					
	a)			<del></del>		
	b)		e)	<del></del>		
6.	• · · · · · · · · · · · · · · · · · · ·					
	a)					
	b)		e)			
_						
7.	Scheduled opera	ting hours:		<del></del>		
	nsportation Road-building	cost (if any)				
2.	Other costs in	this category	<b>':</b>			
2.	Other costs in	this category	<b>':</b>			
		this category	·:			
Rev	enues				d14	
	enues			gs, pulpwoo	d, or chi <sub>l</sub>	os were sold
Rev	enues			gs, pulpwoo	d, or chip	os were sold
Rev	enues			gs, pulpwoo	d, or chi	os were sold
Rev	enues			gs, pulpwoo	d, or chip	os were sold
Rev	enues			gs, pulpwoo	d, or chi	os were sold
Rev	enues Name and locat		that sawlo	gs, pulpwoo	d, or chip	os were sold
Rev	enues Name and locat	ion of mill(s)	that sawlo	gs, pulpwoo	d, or chi	os were sold
Rev	enues Name and locat	ion of mill(s)	that sawlo	gs, pulpwoo	d, or chip	os were sold
Rev	Name and locat	ion of mill(s)	that sawlo	gs, pulpweo	d, or chi	os were sold
Rev	Total volume  product pulpwood	ion of mill(s)	that sawlo	gs, pulpwoo	d, or chi	os were sold

# Appendix 7.

Bird census data for stands 1, 2 and 3.

### KEY TO BIRD CODES ----

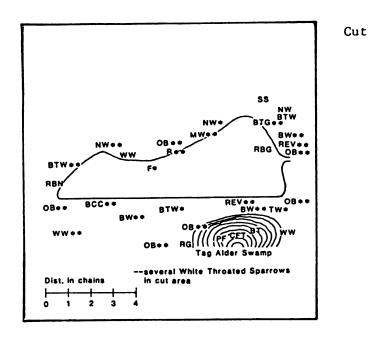
AF - Alder Flycatcher OB - Ovenbird BTW - Black and White Warbler - Purple Finch PF BB - Blackburnian Warbler (or BW) PS - Pine Siskin - Brown Creeper PW - Palm Warbler BCC - Black Capped Chicadee (or CD) PWP - Pileated Woodpecker BHC - Brown Headed Cowbird BT - Bittern - Robin RBG - Rose Breasted Grosbeak BTG - Black Throated Green Warbler RBN - Red Breasted Nuthatch (or NH) BW - Blackburnian Warber (or BB) REV - Red Eyed Vireo RG - Ruffed Grouse RS - Red Start CMW - Cape May Warbler CS - Chipping Sparrow SCW - Chestnut sided Warbler SS - Song Sparrow CYT - Common Yellow Throat (or YT) ST - Scarlet Tanager DW - Downy Woodpecker EP - Eastern Pheobe SV - Solitary Vireo TW - Tennessee Warbler EWP - Eastern Wood Peewee v - Veery WC - Woodcock
WT - Wood Thrush - Flicker GCK - Golden Crowned Kinglet WTS - White Throated Sparrow HT - Hermit Thrush HWP - Hairy Woodpecker WW - Winter Wren IB - Indigo Bunting MGW - Magnolia Warbler YBC - Yellow Billed Cuckoo YBSS - Yellow Bellied Sapsucker MW - Mourning Warbler NH - Nuthatch (or RBN) YRW - Yellow Rumped Warbler
YT - Common Yellow Throat (or CYT) - Nashville Warbler

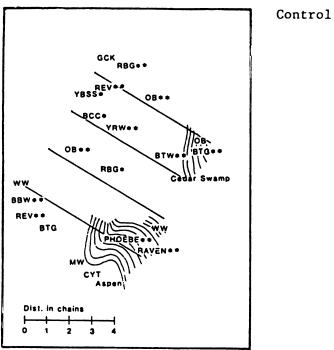
# Legend for Bird Sighting Maps

aa - One sightingaa\* - Two sightingsaa\*\*- Territory confirmed

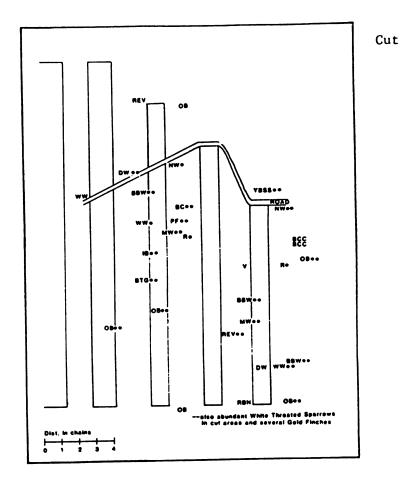
Methods: The bird census was conducted by mapping singing males. This was done by transecting each plot on 6-1, 6-3 and 6-4, 1981.

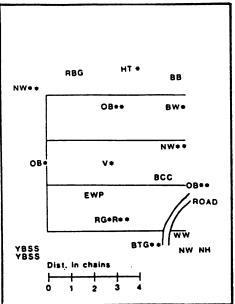
Stand 1.





Stand 2.

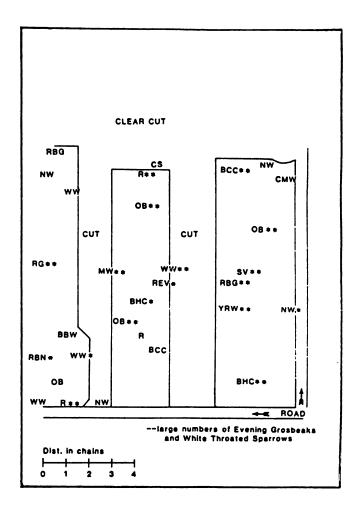


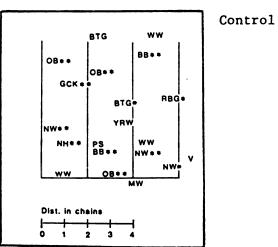


Control

Cut

Stand 3.





## Appendix 8.

The malaise traps and method presented for analyzing data collected by these traps can be used with other parasitoids. It may be necessary, though, to base the model on the daily activity of these animals. This was unnecessary with G. fumiferanae attacking spruce budworm because most adult activity occurred during an identifiable part of the day. As a result, average temperature and total rainfall during this time period were good measures of adult activity.

Data on G. fumiferanae attacking jack pine budworm (Elliot, unpubl.) indicate that, in this habitat, G. fumiferanae is active throughout the day (fig. 25). When this data and appropriate weather information (table 15) were inserted in the model, a poor fit was obtained. One reason model parameters could not be fit to this data was that rain often fell for only part of the day while the rest of the day was conducive to parasitoid activity (i.e., 1068 DD and 1091 DD, table 15). This did not happen in the spruce fir habitat. In order for the model to work, weather (or other influencing variables) and trap catch must be assessed at regular intervals throughout each day. The part of the model which predicts adult parasitoid density will still be appropriate, but activity must be modeled as a convolution of density, intrinsic activity patterns (if any), and factors (i.e., weather) which influence activity.

The inability of the model to describe the data collected on <u>G</u>. <u>fumiferanae</u> in jack pine does not contradict the idea that parasitoid activity patterns are strongly influenced by weather. This data was collected from two locations and is plotted as a function of degree days (base 8.9°C) in figure 26. The pattern of trap catch is similar in both plots. This similarity can be evaluated statistically.

Let  $x_1(i)$  equal 1 if the trap catch on day i is greater than the catch on day i-1, and 0 if it is less in one of the two locations. The variable  $x_2(i)$  is defined similarly for the other location. No pattern is expected between  $x_1$  and  $x_2$  when adult parasitoid density is relatively constant if weather does not influence trap catch. If  $x_1(i)$  and  $x_2(i)$  are equal then let  $x_{12}(i)$  be 1, and 0 otherwise. The variable  $x_t$ , defined as the summation of  $x_{12}(i)$  over i, is a binomial random variable. A test of a relationship between  $x_1$  and  $x_2$  is whether p of this distribution equals 0.5. Ignoring trap catch prior to c. 70% emergence of the adult female parasitoids (< 1037 DD) and the tail end of trap catch (> 1477 DD)  $x_t = 17$  and the sample size (N) = 23. Using a one-sided test (we expect p > 0.5) the probability of this occurring is .017 (p( $x_t \ge 17$ );  $x_t = (B, N = 23, p = .5)$ ). The null hypothesis (p = 0.5) is rejected. Hence, the pattern of trap catch in the two locations is similar, which suggests that weather influenced G. fumiferanae's adult activity. This is reinforced by the observation that the dramatic decline in parasitoid activity at 1263 DD was accompanied by prolonged cool, wet weather.

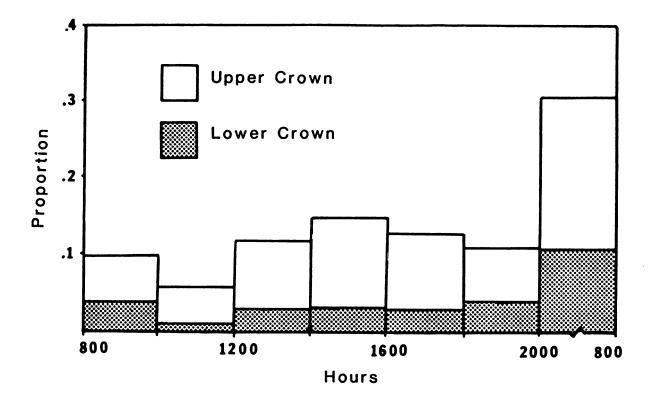


Figure 25. Proportion of total malaise trap catch of female Glypta fumiferanae for 7 time intervals within a day. The sample period was 6 days (7-19 to 7-24). One trap was located in 48 jack pine trees. Twenty four traps were in the upper crown and 24 in the lower crown. Data collected by N. Elliott in Grand Traverse Co., MI.

Table 15. Malaise trap catch of female Glypta fumiferanae from two plots and weather data in 1982. One trap was located in 24 jack pine in each plot. Twelve traps were in the upper crown and 12 in the lower crown. Data collected by N. Elliott in Grand Traverse Co., MI.

Julian Date	Degree Days Base 8.9° C.	Malaise Trap l	Catch 2	Temperature <sup>1</sup>	Precipitation <sup>2</sup>
191	924	0	1	25.78	.97
192	938	0	0	16.85	.58
193	955	2	4	20.78	0
194	972	4	12	25.63	0
195	991	10	11	23.74	.03
196	1012	22	28	27.21	0
197	1037	35	15	29.67	0
198	1068	14	30	27.15	7.85
199	1091	45	28	23.71	.76
200	1106	28	19	20.44	0
201	1122	33	25	21.71	0
202	1137	54	38	24.56	0
203	1160	40	31	22.89	0
204	1181	64	30	24.37	0
205	1200	92	54	26.74	0
206	1225	41	43	26.11	.03
207	1249	35	24	22.29	.03
208	1263	4	3	18.07	1.04
209	1276	9	9	21.82	0
210	1289	25	4	23.41	0
211	1309	39	25	21.33	0
212	1329	27	17	22.71	0
213	1350	20	20	23.26	0
214	1369	14	8	20.37	0
215	1392	28	22	24.26	0
216	1416	10	14	20.74	0
217	1435	13	11	22.11	0
218	1455	11	6	23.52	0
219	1477	19	9	26.29	0
220	1498	6	2	24.48	0
221	1510	3	2	15.48	.08
222	1517	5	1	15.93	0
223	1528	0	1	17.89	0
224	1537	1	1	19.48	0
225	1551	0	0	21.67	0

 $<sup>^{1}</sup>$ average  $^{o}$ C between 800 and 2200 hours

<sup>2</sup> total precipation in cm. between 800 and 2200 hours

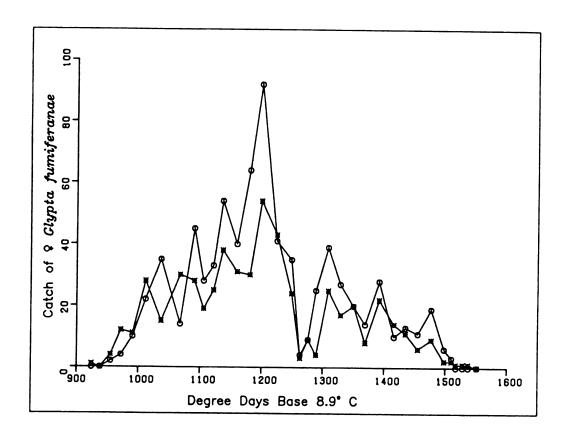


Figure 26. Malaise trap catch of female <u>Glypta fumiferanae</u> from two plots in relation to degree days base 8.9°C. in 1982. One trap was located in each of 24 jack pine in each plot. Twelve traps were in the upper crown and 12 in the lower crown. Data collected by N. Elliott in Grand Traverse Co., MI.

## Appendix 9.

Spruce budworm population estimation data from forest harvesting study

9.1 Egg mass densities in stands 1 and 3.

Description by columns; 1-stand, 2-treatment; (1) cut, (2) control, 3-blank, 4 and 5-tree, 6-blank, 7-branch, 8-blank, 9-species; (1) balsam fir, (2) white spruce, 10-blank, 11 through 14-branch surface area (cm<sup>2</sup>), 15-blank, 16 and 17-hatched egg masses, 18-blank, 19 and 20-parsitized egg masses. Repeat 3 times.

9.2 Early and mid instar densities in stands 1, 2 and 3.

Description by column: 1-stand, 2-treatment; (1) cut, (2) control, 3-sample number, 4-blank, 5 through 7-date, 8-blank, 9 and 10-tree, 11-branch, 12-blank, 13 and 14-budworm, 15 blank, 16 through 19-branch surface area (100 cm<sup>2</sup>). Repeat 2 times.

9.3 Late instar, pupal and adult densities in stands 1, 2, 3.

Description by columns: 1-stand, 2-treatments; (1) cut, (2) control, 3-sample number, 4 and 5-tree, 6-blank, 7-branch, 8-blank, 9 and 10-branch surface are (100 cm<sup>2</sup>), 11-blank, 12-defoliation of all foliage; (0) 0-20%, (1) 21-40%, (2)41-60%, (3) 51-80%, (4)81-100%, 13-defoliation of new foliate (same ranking), 14-blank, 15 and 16-adults, 17-blank, 18-parasitized pupae, 19-blank, 20-pupal mortality (unknown cause), 21-blank, 22 and 23-Meteorus trachynotus pupae, 24-blank, 25-G. fumiferanae pupae, 26-blank, 27-Apanteles fumiferanae pupae, 28 and 29-blank, 30 and 31-total budworm at time of sample, 32-blank, 33 and 34-larvae at time of sample, 35-blank, 36 and 37-parasitized larvae, 38-blank, 39-larval mortality (unknown cause). Repeat.

12 05 3 2 2000 16 02

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11 01 1 1 0900 05 01 12 06 1 2 2050 05 01 31 10 2 2 2050 05 02 32 14 3 1 0900 02 02 11 01 2 1 0600 03 00 12 06 2 2 1900 05 00 31 10 3 2 1200 04 08 32 15 1 1 0900 02 00
11 01 3 1 0750 03 02 12 06 3 2 2100 13 09 31 11 1 2 1500 04 03 32 15 2 1 0800 00 00
           1500 06 01 12 07 1 1 1500 05 04 31 11 2 2 1250 03 03 32 15 3 1 0800 00 01 1300 06 02 12 07 2 1 1400 06 00 31 11 3 2 1500 06 00 32 16 1 2 1800 06 03
11 02 2 1 1300 06 02 12 07 2 1 1400 06 00 31 11 3 2 1500 06 00 32 16 1 2 1800 06 03 11 02 3 1 0650 06 00 12 07 3 1 1200 01 00 31 12 1 2 1800 07 04 32 16 2 2 1100 03 01
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                                                              1200 04 00
11 04 2 1 1100 01 02 12 09 2 1 0800 05 01 31 13 3 2
11 04 3 1 1700 12 02 12 09 3 1 1000 14 04 31 14 1 2
                                                              1500 02 00
                                                              1700 08 00
      1 1 1600 05 00 12 10 1 1 0600 03 00 31 14 2 2
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11 05 2 1 0500 02 00 12 10 2 1 0850 05 00 31 14 3 2
                                                              0800 05 01
11 05 3 1 0800 05 00 12 10 3 1 0850 02 01 31 15 1 2
                                                              1750 03 01
           2400 02 03
                        12 11 1
                                  1 0950 06 01 31 15 2
                                                            2
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11 06 2 1 2100 04 00 12 11 2 1 0650 09 01 31 15 3 2
                                                              1900 00 00
11 06 3 1 1200 04 00 12 11 3 1 0350 05 01 31 16 1 2
                                                              2400 10 04
                         12 12 1 1
                                     1300 03 02 31 16 2 2
11 07 1 1 1000 04 02
                                                              1100 07 00
11 07 2 1 2100 08 02 12 12 2 1 2300 22 11 31 16 3 2 1600 14 01
11 07 3 1 1500 00 01 12 12 3 1 1000 04 02 32 01 1 1
                                                              1400 01 00
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11 08 1 1
                        12 13 2 1 0800 08 02 32 01 3 1 0950 00 00
            1400 05 02
                                    1300 12 09 32 02 1 1 0800 01 01
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           0900 10 02 12 14 1 1 1000 04 01 32 02 2 1 1200 03 02
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                        12 14 2 1 0850 03 01
                                                  32 02 3 1 0850 08 01
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           1200 06 01 12 15 1 1 1200 04 05 32 03 2 1 2100 15 05
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11 10 2 1 1500 09 01 12 15 2 1 0800 02 00 32 03 3 1 1400 05 02
            1400 12 03 12 15 3 1 0900 05 01
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11 11 2 1 0800 04 01 12 16 2 1 0900 03 00 32 04 3 1 1900 06 08
            1400 06 04 12 16 3 1 0900 10 00
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            1050 08 01 31 01 1 2 2100 15 01 32 05 2 1 1900 10 01
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112 612 032 09 06.0	121 609 091 26 15.0	
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211	603	062	38	06.	0					17.0						09.0
	603							-		12.0			617			10.0
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_	603									07.0			619	-		12.0
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	617			17.		221	604	034	23	14.0	)	223	619	041	19	08.0
	617			16.		221	604	041	12	11.5	5	223	619	042	10	15.0
	617						604			06.0		223	619	043	02	08.0
	617			06.	_	221	604	043	17	15.0	)	223	619	044	12	07.0
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						221	604	051	13	11.0	)	223	619	052	11	15.0
						221	604	052	09	04.0	?					16.0
						221	604	053	18	14.5		223	619	054	22	17.0
						221	604	054	03	07.5	•					
						221	604	061	20	12.0	<b>,</b>					
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						221	604	064	99	17.0	)					
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						'										

9.2

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17 03 02 0

04 01 01 0

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00 00 00 0

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03 02 02 0

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06 00 00 0

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05 01 00 0

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07 03 01 1

10 07 02 0

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04 01 01 0

06 01 01 0

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18 00 00 0

16 00 00 0

13 00 00 0

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### APPENDIX 10

## Record of Deposition of Voucher Specimens\*

The specimens listed on the following sheet(s) have been deposited in the named museum(s) as samples of those species or other taxa which were used in this research. Voucher recognition labels bearing the Voucher No. have been attached or included in fluid-preserved specimens.

No. have been attached or included	in fluid-preserved specimens.					
Voucher No.:1982-4	· · · · · · · · · · · · · · · · · · ·					
Title of thesis or dissertation (or	other research projects):					
Studies Related to the Concept of Pest - Crop System Design: 1) Adult Parasitoid Activity and Its Relation to Biocontrol and 2) Forest Harvesting and the Spruce Budworm						
Museum(s) where deposited and abbre	eviations for table on following sheets:					
Entomology Museum, Michig	gan State University (MSU)					
Other Museums:						
	•					
	Investigator's Name (s) (typed)					
	Jan Peter Nyrop					
	Date11-13-82					
*Reference: Yoshimoto, C. M. 1978. Voucher Specimens for Entomology in North America. Bull. Entomol. Soc. Amer. 24:141-42.						

Deposit as follows:

Original: Include as Appendix 10in ribbon copy of thesis or

dissertation.

Copies: Included as Appendix 10in copies of thesis or dissertation.

Museum(s) files.

Research project files.

This form is available from and the Voucher No. is assigned by the Curator, Michigan State University Entomology Museum.

## APPENDIX 10.1

# Voucher Specimen Data

Page 1 of 1 Pages

	Museum where depos- ited	Entomology Department Michigan State University	, n
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	Label data for specimens collected or used and deposited	Q Delta Co., MI T21N R23W Sec14 caught in malaise traps located in balsam fir trees 7-5-81 Jan P. Nyrop G Delta Co., MI T21N R23W Sec14 caugh in malaise traps located in balsam fir trees 7-13-81 Jan P. Nyrop	sary)  Voucher No. 1982-4  Received the above listed specimens for deposit in the Michigan State University  Farymology Muserm  Curator  Date
	Species or other taxon	Glypta fumiferanae (Vierick)	(Use additional sheets if necessary) Investigator's Name(s) (typed) Jan Peter Nyrop  Date 10-18-82

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