



THERIS



This is to certify that the

thesis entitled

Infectious Agents of Bovine Abortion-Laboratory Diagnosis

presented by

Valeria Moojen

has been accepted towards fulfillment of the requirements for

Master of Sciengegree in _____Microbiology

Major professor

Date August 06th, 1979.

O-7639



OVERDUE FINES ARE 25¢ PER DAY PER ITEM

Return to book drop to remove this checkout from your record.

INFECTIOUS AGENTS OF BOVINE ABORTION -

.

LABORATORY DIAGNOSIS

By

Valeria Moojen

A THESIS

Submitted to Michigan State University in partial fulfillment of the requirements for the degree of

MASTER OF SCIENCE

Department of Microbiology and Public Health

ABSTRACT

INFECTIOUS AGENTS OF BOVINE ABORTION -LABORATORY DIAGNOSIS

By

Valeria Moojen

Specimens from 98 aborted bovine fetuses were collected at the Animal Health Diagnostic Laboratory of Michigan State University, between June 1977 and September 1978. The fetuses were mostly from Holstein-Friesian herds from many different areas within the State of Michigan. The primary cause of abortion was recognized in 43 (43.8%) of the aborted fetuses examined. Noninfectious factors were associated with 6.1% of the abortions and infectious agents were incriminated in 37.7%. Bacteria accounted for 22.4%, viruses for 14.3%, and fungi for 1%. Antibodies to leptospira were found in 6 fetuses, and 4 of the 50 fetal fluids examined had antibodies to parainfluenza-3. Electron microscopy was the most effective method in demonstrating viral agents in fetal tissues. The presence of herpesviruses was detected in 8.2% of the fetuses, correlating in all instances with positive fluorescent antibody (FA) results for infectious bovine rhinotracheitis (IBR) virus.

Herpesviruses were demonstrated in 3 additional fetuses that were negative for IBR by FA. Nutritional deficiencies were noted in 3 fetuses and congenital anomalies were found in 2 fetuses.

ACKNOWLEDGEMENTS

I wish to express my sincere appreciation to all who aided me in the completion of this study. I would especially like to thank Dr. Gordon R. Carter, chairman of my guidance committee, for his advice and support throughout the study, and Mr. A. Wayne Roberts for his unstinting technical guidance and constant encouragement.

This research involved the assistance of many individuals and thanks is expressed to all of them. Dr. K. K. Keahey, Director of the Animal Health Diagnostic Laboratory, and his associates provided the specimens used in this research. Mrs. Marian Bennett and Dr. S. B. Singh assisted in the leptospiral and chlamydial serology; Mr. Paul Watkins assisted with the fluorescent antibody tests; and Mrs. Dorothy Boettger helped with the preparation of glassware and supplies. Dr. A. L. Trapp read a draft of this thesis and offered valuable suggestions.

I would like to thank other members of the committee, Dr. S. D. Sleight and Dr. L. F. Velicer, Dr. A. Haberman and Dr. S. B. Singh, for their efforts on my behalf. I would especially like to acknowledge the excellent editorial assistance of Dr. Sleight.

ii

This study was made possible by support of the PEAS Project (Brazil/MSU) and by a grant from the Michigan Agricultural Experiment Station. The College of Veterinary Medicine, Federal University of Rio Grande do Sul, Porto Alegre, Brazil, generously provided me the opportunity for this advanced study.

TABLE OF CONTENTS

	Page
INTRODUCTION	1
REVIEW OF THE LITERATURE	3
Factors Associated with Bovine Abortion	3
Genetic Factors	3
Environmental Factors	4
Infectious Agents	5
Viral agents	6
Bacterial agents	9
Mycotic agents	13
Protozoan agents	14
Laboratory Diagnosis of Infectious Agents	
of Bovine Abortion	14
Serology	17
Virus Concentration.	
Electron Microscopy.	
Viral Interference	21
	21
LITERATURE CITED	22
ARTICLE: INFECTIOUS AGENTS OF BOVINE ABORTION -	
LABORATORY DIAGNOSIS	34
Summary \ldots \ldots \ldots \ldots \ldots \ldots	35
Summary	35
Material and Methods	37
Results	
Discussion	52
Literature Cited	59

LIST OF TABLES

Table		Page
1	Infectious agents isolated or demonstrated in studies of 98 bovine abortions in Michigan, 1977-1978	44
2	Results of microbiological, pathological and serological examination of 98 bovine aborted fetuses, Michigan, 1977-1978	4 5
3	Results of bacteriological examination of 98 aborted bovine fetuses in Michigan, 1977-1978	46
4	Results of the virological examination of aborted bovine fetuses	48
5	Histopathological lesions associated with infectious agents of aborted bovine fetuses .	50

INTRODUCTION

Bovine abortion is recognized as one of the major economic losses in dairy and beef cattle production, and infectious agents are considered to play a major role. However, a definitive diagnosis is obtained in only 25% of the total cases submitted to most laboratories. This low percentage is likely due to three factors:

- 1. Most abortions are not due to infectious agents.
- Inappropriate specimens are examined (either incorrect tissues or tissues in poor condition).
- 3. Inappropriate tests are utilized.

The purpose of this study was to increase the effectiveness of the diagnostic laboratory in dealing with infectious agents of bovine abortion. This was accomplished by supplementing conventional tests (routine bacteriologic tests including darkfield examination of stomach contents, routine virologic procedures including fluorescent antibody examination for infectious bovine rhinotracheitis (IBR) and bovine virus diarrhea (BVD), and pathologic examination) with serologic examination of fetal fluids and electron microscopy. Serologic examination of fetal fluids takes advantage of the relative early immune competence of the fetus, and the fact that

immunoglobulins do not normally cross the placental membranes in the cow. Electron microscopy permits the detection of infectious agents not normally associated with abortion, and allows the demonstration of known infectious agents no longer viable as a result of autolytic changes of the fetus.

REVIEW OF THE LITERATURE

Factors Associated with Bovine Abortion

Factors associated with the cause of bovine abortions have been categorized mainly as genetic, environmental, and infectious, according to Hubbert et al. (1973) and Miller (1977).

Genetic Factors

Gilmore (1949) reviewed the inherited causes of reproductive failure. He discussed and characterized the lethal genes responsible for death such as those responsible for mummification and subsequent abortion, and "the Ljutikow's lethal" that results either in abortion, stillbirth or death shortly after birth.

Knudsen (1956) postulated that bulls with normal appearing semen may have abnormally arranged genes which might contribute to unbalanced genetic constitution of the embryo resulting in its death. Bishop (1964) also discussed the male influence on embryonic death. Attention was directed to the often undetected recessive lethal factors which could contribute to a large economic loss based on their wide dissemination as a result of artificial insemination.

Mylrea (1963) reported an outbreak of abortion in a Shorthorn herd in which a genetic cause was suspected. The abortions occurred in late pregnancy, and no significant organisms were detected on bacteriological examination. Also, no evidence of brucellosis, leptospirosis, vibriosis or trichomoniasis was found.

Stevens and King (1968) reported cases of midterm abortion in a Holstein-Friesian herd. By analyzing pedigrees and breeding histories and eliminating suspect animals from the mating program, they found evidence to support an autosomal recessive lethal gene involved in the abortion problem.

Wijeratne and Stewart (1971) studied the data from abortion in cattle over an 8-year period, with special reference to genetic factors. They found that rates of abortion increased with the number of inseminations, and that there was also a slight increase among twin fetuses. A significant difference was observed among the breeds Friesian, Guernsey, Jersey, Ayrshire, Dairy Shorthorn, Hereford, Aberdeen-Angus, and Charolais.

Environmental Factors

Crane (1965) provided evidence of an interrelationship between IBR related abortion and nutrition. Deficiencies such as phosphorus, vitamin A, protein, and some of the trace minerals were found in California at certain times of the year. Research with 800 cows in which supplemented feed and controlled vaccination for IBR,

BVD, leptospirosis, and brucellosis were provided, showed an increase in the weaned calf crop. Neighboring ranches with analogous vaccination programs, but without supplemented feed, did not have a reduction in their loss rate.

Andrews (1972), studying cattle from different environmental areas, found nutrition, management and breed to be important contributors to reproductive failures. For example, poorer body condition was found in nonconceiving cows compared to those that were pregnant, and it was suggested that supplementary feeding to correct the protein concentration would increase reproduction. Age of the stock was also considered to influence reproductive performance.

A study by Hignett (1959) showed that cows with high levels of phosphorus in relation to calcium in the diet had their resistance to infectious agents decreased. Woelffer (1953) reported that pregnancies were maintained in cows by the administration of progesterone. Schiefer and Moffatt (1974) described a case of bovine abortion associated with renal oxalosis in the fetus. They emphasized the need for further studies to determine the effects of oxalate on the fetus.

Infectious Agents

Viral, bacterial, mycotic and protozoan agents are well documented as causes of abortion. General articles have been published by Gilman (1939), Lawson (1963), Ashfar (1965), Dennis (1969), Woelffer (1972), Hubbert

et al. (1973), and Miller (1977). The studies by Dennis (1969) and Woelffer (1972) were also directed at the diagnosis of infectious agents.

Viral agents. Among the viruses causing bovine abortion, IBR virus is one of the more frequent. This herpesvirus, first known to cause a respiratory disease (Miller, 1955), was isolated from an aborted fetus by McKercher and Wadda in 1964. Reports by Crane et al. and Lukas et al. in the same year also incriminated IBR virus as a cause of abortion. Experimentally induced abortion in cattle with IBR virus was reported in 1964 by Chow et al., and Mitchell (1974) described epizootic abortions after inoculation with live IBR vaccine. Reports have indicated that IBR continues to be a major cause of bovine abortion (Kirkbride et al., 1973; Miller and Quinn, 1975; Kradel, 1978). IBR abortion was experimentally produced and the lesions in the fetus were described (Owen et al., 1964; Sattar et al., 1967; Kendrick and Straub, 1976). Kennedy and Richards (1964) studied the changes produced in the aborted fetuses by IBR virus infection acquired naturally, experimentally and by vaccination and concluded that they were essentially the same. Lesions consisted of foci of necrosis found consistently in the liver and irregularly in other organs. Owen et al. (1968) studied the pathogenesis of IBR virus infection in the fetus and correlated fetal and placental lesions with viral isolations.

Herpesviruses other than IBR have also been isolated from aborted fetuses. Crandell et al. (1976) reported the isolation of a virus designated BH-1247 with characteristics fitting the herpes group of viruses. This new isolate differed from IBR, DN-599 and MOVAR 33/63 viruses, and its significance as an abortifacient agent was not determined. Reed et al. (1979) reported a concurrent infection of an aborted bovine fetus with BVD virus and a herpesvirus. This latter isolate was serologically indistinguishable from DN-599 and MOVAR 33/66. Experimental studies by Kendrick et al. (1976) have suggested that this virus does not cause abortion. Malignant catarrhal fever (MCF) virus has been reported to infect wildebeest in utero (Plowright et al., 1960), but in cattle transplacental infection could not be demonstrated (Plowright, 1968). Even though abortion is reported in some cases of MCF, it is attributed to a lowered resistance of the animal due to the disease and not to the virus per se (Berkman and Barner, 1958). Cytomegalovirus was isolated from an aborted bovine fetus by Schiefer (1974); its role as a cause of abortion in cattle has not been established.

Bovine virus diarrhea was first described by Olafson et al. (1946), and abortion was one of the clinical signs observed. Cytopathic strains (Scott et al., 1972) as well as noncytopathic strains (Gillespie et al., 1967) capable of producing abortion have been described. The pathological changes produced by BVD virus in the bovine

fetus were characterized by Casaro et al. (1971), Braun et al. (1973) and Brown et al. (1974).

Sattar et al. (1965) first isolated bovine parainfluenza (PI-3) virus from an aborted bovine fetus. Its possible role in abortion was investigated experimentally by Sattar et al. (1967) and Swift and Kennedy (1972). Serological studies by Dunne et al. (1973) and Ruth et al. (1974) on aborted fetuses suggested the possible role of this paramyxovirus in bovine abortions.

Moll and Davis (1957) and Moll and Finlayson (1959) isolated enteric viruses (BEV) from feces of cattle and from animals having respiratory tract infection. Abortion was observed in those herds. Continuing the study, Moll (1964) experimentally exposed guinea pigs to BEV and produced abortion and stillbirths. Later, Dunne et al. (1973) reported the results of serologic examination of aborted bovine fetuses and incriminated BEV as potentially important agents in bovine abortion.

Parvoviruses are known to cause reproductive failures in other animal species (Kirkbride and McAdaragh, 1978). The possibility of this group of viruses causing abortion in cattle was investigated by Storz et al. (1978) by inoculation of the virus into pregnant cows, recovery of the virus from the aborted fetuses, and demonstration of hemagglutination inhibition antibodies in those animals.

Bluetongue (BT) virus was first isolated in the United States from cattle in 1959 (Bowne et al., 1968); Luedke et al. (1970) isolated BT virus from the spleen of

an aborted fetus. The heifer that aborted had clinical signs suggesting BT infection. Experimental studies confirmed the abortogenic nature of the BT virus in cattle (Luedke et al., 1977).

Dierks et al. (1976) isolated subgroup 2 adenoviruses from aborted bovine fetuses, and suggested that they may be involved with intrauterine infection and abortion.

A member of the Simbu group of the Bunyaviridae family of viruses, Akabane virus, is known to cause abortions in cattle. It has been reported in Australia (Blood, 1956), Kenya (Metselaar, 1976), and Japan (Kurogi et al., 1976), but not in the United States. Weiss et al. (1956) studied Wesselsbron virus and discussed its association with abortion in domestic animals. Their findings suggested that Wesselsbron virus infection in cattle could clinically mimic brucellosis. Of 252 clinical cases of brucellosis that were serologically negative, 39.62% had specific antibody to Wesselsbron virus.

The agent of sheep infection, Border disease, produced abortion when inoculated into pregnant heifers (Gibbons et al., 1974).

<u>Bacterial agents</u>. Bovine leptospirosis was clinically described first in 1935 in Russia by Michin and Azinov (1937), followed by reports in Australia (Johnson, 1943), the United States (Jungherr, 1944), and Palestine (Bernkopf et al., 1947; Freund, 1947). Leptospira were

not isolated until 1948 by Baker and Little in the United States. Stoenner et al. (1956) investigated the epizootiology of bovine leptospirosis, and serological studies on leptospiral abortion in cattle were done by Burk and Wiesmann (1963). The pathogenesis of leptospiral abortion was studied experimentally by Murphy and Jensen (1969). The difficulty in isolating the bacterium from the fetus was attributed to autolytic changes occurring in the tissues (Hanson, 1977). There are reports on instances of abortions in which significant levels of antibodies, specific for *Leptospira* species, were found in the fetus (Knott and Dadswell, 1970; Ellis et al., 1978).

Campylobacter fetus (Vibrio fetus) was first isolated from bovine fetuses in 1919 by Smith. Experimental studies on its etiological significance were carried out by the same worker in 1923. Based on serum-agglutination titers and isolation, Plastridge and Williams (1943) reported cases of bovine abortion associated with this bacterium. Many additional reports on isolation of the organism from aborted fetuses have been published (Apice, 1956; Mitchel, 1960; Hubbert et al., 1973). Whitford et al. (1977) reported the isolation of Campylobacter fetus ss. intestinalis and other campylobacters from aborted bovine fetuses.

Brucellosis has a worldwide distribution, but has been eradicated from Norway (1952), Sweden (1957), Finland (1960), and Japan (1974) (15). The United

States is currently conducting a program of control and eradication. Although bovine brucellosis (contagious epizootic abortion) was recognized as early as the 18th century (Jensen and Mackey, 1974), the causative agent was not isolated until 1897 by Bang in Denmark. Reports by Bolton et al. (1969), Kirkbride et al. (1973) and Ewalt and Harrington (1979) emphasized the continued importance of this disease in the United States in spite of efforts at control. The latter investigators stressed the problem of animal condemnation resulting from unofficial adult cattle vaccination.

A Salmonella species was reported by Gibson (1965) and Richardson and Watson (1971) to be associated with abortion in cattle. Maddox (1976) described an "abortion storm" due to Salmonella typhimurium, and Hall and Jones (1977) studied the pathogenesis of experimentally induced abortion in cattle due to S. dublin.

The association of bovine abortion with Corynebacterium pyogenes was reviewed by Hinton (1972). The organism was considered to play a minor role in abortions in the studies of Langneau (1964) and Dennis (1969). Hinton (1974) examined paired-serum samples from cases in which only C. pyogenes was isolated and concluded that it was the primary cause of abortions.

Epstein et al. (1972) reported an abortion associated with concurrent infection of the fetus with *Listeria monocytogenes* and IBR virus. Reports by Siddique et al.

(1976) and Miller (1977) have also incriminated L. monocytogenes as a cause of bovine abortion.

Although epizootic bovine abortion (EBA) has been referred to as a disease syndrome, not necessarily infectious, a chlamydial agent has been demonstrated in aborted fetuses from cows manifesting this particular syndrome (Wada et al., 1976; McKercher et al., 1976). Reed et al. (1971), in investigations of EBA in Colorado and South Dakota, suggested the possibility of other agents being responsible for this syndrome in addition to *Chlamydia*. Experimental studies on chlamydial abortion were done by Storz and McKercher (1962). They isolated EBA agent from 5 aborted bovine fetuses and successfully fulfilled Koch's postulates.

Sporadic abortions have been attributed to the following bacteria: Haemophilus somnus (Chladek, 1975; vanDreumel and Kierstad, 1975); Aeromonas hydrophila (Wohlgemuth et al., 1972); and Pastuerella (Yersinia) pseudotuberculosis (Langford, 1969).

Streptococcus species, Escherichia coli, Staphylococcus species, and Pseudomonas species have also been incriminated as occasional causes of bovine abortion, although their significance as abortifacient agents is doubted by some (Bolton et al., 1973; Dennis, 1969; Woelffer, 1972; Hubbert et al., 1973; Siddique et al., 1976).

Mycoplasma species have been isolated from aborted bovine fetuses (Langford, 1975; Ball et al., 1978), and have been shown to produce abortion experimentally by Stalheim and Proctor (1976).

<u>Mycotic agents</u>. Mycotic infections of the bovine fetus, placenta, and uterus were reported as early as 1920 by Smith, and experimental studies on abortion associated with *Mucor* were done by Gilman and Birch in 1925. Reports of mycotic pneumonia and associated abortion were presented by Cordes et al. (1964) and Harcourt and Thompson (1969).

Kirkbride et al. (1971), studying the diagnosis of mycotic abortion in cattle, called attention to the importance of examination of the placental tissue in diagnosing bovine abortion. In a 3-year period, 1,556 aborted bovine fetuses were examined in which 485 placentas were included. Mycotic placentitis was diagnosed in 3.85% of the cases.

Aspergillus species and Mucor species were reported as causes of abortion by Bolton et al. (1969), Siddique et al. (1976), and Williams et al. (1977).

A CONTRACTOR AND A CONTRACT OF A CONTRACT OF

Isolated cases of mycotic abortion due to other fungi have been reported: *Torulopsis glabrata* (Kirkbride et al., 1972; Knudtson et al., 1976); *Candida tropicalis* (Wohlgemuth and Knudtson, 1973); *Mortirella wolfii* (Harcourt and Thompson, 1969; Wohlgemuth and Knudtson, 1977). <u>Protozoan agents</u>. An unidentified protozoan was reported by Corner et al. (1963) as the cause of abortion in cows. This report prompted Fayer et al. (1976) to study abortion in cattle experimentally inoculated with sporocysts of *Sareocystis fusiformis*. *Trichomonas foetus*, an organism usually associated with infertility, has also been reported as a cause of abortion in cattle (Dennis, 1969; Woelffer, 1972). Abortion is described as a clinical sign of toxoplasmosis in other animal species (Hartley and Marshall, 1957), and a study by Dennis (1969) on the diagnosis of bovine abortion drew attention to the fact that toxoplasmosis may play a role in bovine abortion. Abortions due to this protozoan were reported in studies of bovine abortion (Schiefer and Moffatt, 1974; Miller and Quinn, 1975).

Laboratory Diagnosis of Infectious Agents of Bovine Abortion

The specimens most often received at the laboratory for the determination of the cause of bovine abortion are fetal tissues such as lung, liver, spleen, kidney, and stomach. Placenta is seldom submitted. The routine procedures applied to these specimens are bacterial and viral isolation attempts, darkfield microscopy (examination for *Leptospira*, *Campylobacter* and *Trichomonas*), fluorescent antibody tests for IBR and BVD, and gross and histologic examinations (Kirkbride et al., 1973; Schultz, 1973; Schiefer and Moffatt, 1974).

Reports have generally shown a figure in the range of 23% to 40% for the successful diagnosis of the cause of bovine abortion. Mitchel (1960) examined 227 cases of bovine abortion in the Ottawa (Canada) area and reported a diagnostic success of 33%, of which 8.8% was attributed to *B. abortus*, 22.5% to *L. pomona*, and 1.72% to *V. fetus*, *S. pyogenes*, and *C. pyogenes*. Sattar et al. (1965) examined 28 bovine fetuses for viruses, by virus isolation, and found IBR virus in 6 (21.4%) and PI-3 virus in 1 (3.5%).

In 1968, in a work published by Faulkner, it is stated that only 20 to 30% of cases of bovine abortion submitted to diagnostic laboratories yielded a positive diagnosis. Bolton et al. (1969) examined 1,412 aborted bovine fetuses in Vermont and reported a diagnostic success of 26.48%, of which 1.56% was due to mycotic infection and 24.92% to bacterial causes. Hubbert et al. (1973) reported that 23% of bovine abortions examined in an 11-year period from 5 Northeastern states had an infectious cause: 3.49% were viruses, 5.11% fungi, and 14.64% bacteria. The most frequently diagnosed infections were aspergillosis, IBR, streptococcosis, leptospirosis, vibriosis, and infection due to *Corynebacterium* species.

Kirkbride et al. (1971) and Kirkbride et al. (1973), examining fetal specimens and fetal placental tissue from 2,544 aborted bovine fetuses during a 4-year period in the Northern Plains States, successfully determined the etiologic agent in 35.3%. IBR virus was the most commonly

found agent (16%). Mycotic abortion was diagnosed in 3.5%, and vibriosis in 3.0%. Other bacterial agents, dystocias, and anomalies accounted for 7.4% of the abortions or stillbirths.

According to data obtained from the New Zealand Animal Disease Data Bank records (Kirkbride et al., 1977) for 1974, 34.8% of 15,741 bovine abortions examined had an etiologic diagnosis which included 9% of abortions due to brucellosis.

In a study carried out in Canada, of 1,509 abortion cases examined, 17.83% were due to viruses, 10.66% to bacteria, 3.98% to fungi, 0.07% to protozoan agents, and 7.82% to noninfectious causes (Schiefer and Moffatt, 1974).

Miller and Quinn (1975) compared data from aborted and nonaborted bovine fetuses in Ontario, using pathological, microbiological, and immunological criteria. They considered a positive finding in at least one of the three categories as indicative of a positive etiological diagnosis. This analysis resulted in a positive diagnostic rate of 48%.

Kradel (1978), in a study of the abortion problem in bovine herds, stated that etiological agents are determined in less than 25% of the cases submitted to most diagnostic laboratories, but that this figure could likely be increased to 34-50% with an improvement in diagnostic methods.

Serology

Serological studies currently being done on bovine fetuses are contributing to an increase in the number of positive diagnoses of abortions. The placental transfer of maternal immunoglobulins (Ig) to the fetus does not normally occur in the bovine (Brambell, 1958), and early studies by Fennestad and Borg-Petersen (1958) provided evidence of the bovine fetus' capability to respond immunologically. Later, in 1962, the same workers emphasized that this capability to respond immunologically probably depended upon the animal species, period of gestation when infected, and the antigen. In this study, bovine fetuses were infected with *Leptospira saxkoebing* and specific antibody was detected as early as the 132nd day of fetal life.

Valuable studies have been carried out on the bovine immune system. Buttler (1969) studied the different bovine immunoglobulins, and a symposium was conducted in 1970 on the bovine immune system (Buttler et al., 1971). Schultz et al. (1973) provided important information concerning the ontogeny of the immune system in the bovine fetus. In their work, 106 fetuses of various ages were studied. The development of the bovine immune response and the human immune response was compared. Although fetuses were not inoculated with infectious agents, the results suggested the need of antigenic stimulation for the immune system to show morphological and functional activities. A review by Schultz published in the same

year (1973) stated that periods in which IgM and IgG producing cells were recognized in the spleen were found to be 56 and 145 days, respectively. The appearance of immunologic competence of specific antigens occurred chronologically, and was approximately 90 to 120 days for BVD, IBR, and PI-3.

In view of the aforementioned considerations, the presence of significant levels of Ig in fetal serum and body fluids is indicative of intrauterine antigenic stimulation (Tyzard, 1977). The applicability of the results of these studies may be of value in the laboratory diagnosis of bovine abortion. For instance, Kahrs et al. (1971) concluded that serologic tests could be a valuable source . of information in the diagnosis of bovine abortion and congenital defects. They worked with BVD and IBR viruses, and 11 aborted fetuses. Sawyer et al. (1973) reported the use of the radialimmunodiffusion (RID) test for detection of congenital infections in the bovine fetus, and Dunne et al. (1973), based on results of serologic examination of fetal fluids, presented evidence that viruses other than IBR and BVD were associated with bovine abortion. The serologic tests were done specifically for IBR, BVD, BEV, and PI-3 viruses. The number of positive diagnoses was increased by 58% compared to standard diagnostic procedures. Ruth et al. (1974) also showed serologic tests of fetal fluids to be useful in the diagnosis of bovine abortion. The presence of IgG was determined by RID, and it was often associated with

fetuses aborted due to infectious agents. The fetal fluids were specifically tested for antibodies to IBR, BT, BVD, PI-3, parvoviruses, and *Leptospira*. Kirkbride et al. (1977) compared levels of Ig (IgG and IgM) in aborted and abattoir fetuses, and correlated the presence of Ig to lesions in these fetuses. They found a significant difference in the level in aborted fetuses as compared to fetuses from the abattoir. According to the results obtained, the measurement of fetal Ig by RID may be of value as a first step in the diagnosis of infectious abortion.

Virus Concentration

Various techniques have been described and applied in an attempt to concentrate different viruses. Among the methods utilized was the alcohol precipitation of proteins from solutions containing influenza virus (Cox et al., 1947). He and co-workers found methanol to be superior to ethanol, and concluded that if precipitation was conducted under controlled conditions, such as temperature, methanol concentration and pH, the technique was applicable to a wide range of viruses.

Later, in 1949, Pollard et al., following essentially the procedure of Cox et al. (1947), studied the effects of methanol on 5 viruses from 4 different groups. In this study, the optimal pH for elution was given, and it was observed that the nitrogen content of the virus suspension decreased after precipitation. Goodheart et

al. (1974), working with oncogenic herpesviruses concentrated by methanol precipitation, called attention to the usefulness of this technique in virus concentration trials. Verrilli (1978) successfully used the methanol precipitation procedure in his electron microscopy study of representative viruses of veterinary significance.

Electron Microscopy

Morphology is an important feature of viruses that can be determined only by electron microscopy (EM) and which obviously is useful in viral identification.

Spradbrow and Francis (1969), using virus infected cell culture lysates, emphasized the possibility of EM playing an important role in diagnosis of viral infections, and Gibbs and Johnson (1970), working with udder and teat lesions, stated that the examination of clinical specimens by EM provides a satisfactory method for a rapid and accurate differential diagnosis of cowpox, pseudocowpox, and bovine herpes mammilitis.

Marsolais et al. (1971) reported the use of EM for the rapid diagnosis of avian coronavirus infection, and McFerran et al. (1971) discussed the application of negative contrast EM to routine virus diagnosis. They worked with different animal viruses obtained directly from clinical material or isolated in eggs or cell cultures.

England et al. (1976) reported the greater efficiency of EM for demonstration of viruses associated with neonatal calf diarrhea as compared to viral isolation and

the fluorescent antibody techniques, but Draayer and Kirkbride (1977) doubted the need for the routine examination of fetal tissue by EM. Verrilli (1978), working on negative contrast EM of viruses of veterinary significance, stressed the usefulness of its application in clinical virology.

Viral Interference

The usefulness of viral interference to detect noncytopathogenic viruses was emphasized by Gillespie et al. (1967) in their report on the isolation of noncytopathogenic BVD virus from 2 aborted fetuses. Dellers (1977), in his evaluation of the diagnostic capability of veterinary virology, drew attention to the importance of using the interference test to detect noncytopathogenic viruses.

Nuttall et al. (1977) used the interference test in the examination of the bovine fetal serum employed in cell cultures for possible virus contaminants. Results indicated that the test was less sensitive than the quantitative plaque technique, and probably would not detect low levels of interference. LITERATURE CITED

.

LITERATURE CITED

- Afshar, A. 1965. Virus diseases associated with bovine abortion and infertility. Vet. Bull. 35(12):735-752.
- Andrews, L. G. 1972. The major non-infectious causes of reproductive wastage in beef cattle in the Northern territory. Aust. Vet. Jour. 48:41-46.
- Apice, M. d'. 1956. Ocorrencia do aborto bovino no Estado de Sao Paulo devido ao Vibrio fetus. O Biologico. Sao Paulo. 22(1):15-18.
- 4. Baker, J. A. and Little, R. B. 1948. Leptospirosis in cattle. Jour. Exptl. Med. 88:295-307.
- Ball, H. J., Neill, S. D., Ellis, W. A., O'Brien, J. J. and Ferguson, H. W. 1978. The isolation of mycoplasma from bovine foetuses and their dams. Br. Vet. J. 134(6):584-589.
- 6. Bang, B. 1897. The etiology of epizootic abortion. Jour. Comp. Path. and Therap. 10:125-149.
- Berkman, R. N. and Barner, R. D. 1958. Bovine malignant catarrhal fever. 1. Its occurrence in Michigan. J.A.V.M.A.:243-248. Mar. 15.
- Bernkopf, H., Olitzki, L. and Stuczynski, L. A. 1947. Studies on bovine and human leptospirosis. J. Infect. Dis. 80:53-63.
- 9. Bishop, M. W. H. 1964. Paternal contribution to embryonic death. J. Reprod. Fertil. 7:383-396.
- Bolton, W. D., Durrell, W. B., Wadsworth, J. R. and Murray, R. W. 1969. A survey of abortions in Vermont dairy cattle. J.A.V.M.A. <u>155</u>(3):500-503.
- Bowne, J. G., Luedke, A. J., Jochim, M. M. and Metcalf, H. E. 1968. Bluetongue disease in cattle. J.A.V.M.A. 153(6):662-668.

- 12. Brambell, F. W. R. 1958. The passive immunity of the young mammal. Biol. Rev. 33:488-531.
- Braun, R. K., Osburn, B. I. and Kendrick, J. W. 1973. Immunologic response of bovine fetus to bovine viral diarrhea virus. Am. J. Vet. Res. 34(9):1127-1132.
- Brown, T. T., deLaunta, A., Bistner, S. I., Scott, F. W. and McEntee, K. 1974. Pathogenetic studies of infection of the bovine fetus with bovine viral diarrhea virus. Vet. Path. <u>11</u>: 486-505.
- 15. Brucellosis Summary. Animal and Plant Health Inspection Service Cooperative State-Federal Brucellosis Eradication Program. Notes. 1977. May. In J.A.V.M.A. 171(5):430. 1977.
- Burk, F. and Weismann, E. 1963. Serological diagnosis of leptospiral abortion in cattle. Wien. Tierarztl. Mschr. 50:748-761.
- Buttler, J. E. 1969. Bovine immunoglobulins: A review. J. Dairy Sci. 52(12):1895-1909.
- Buttler, J. E., Winter, A. J. and Wagner, G. G. 1971. Symposium: bovine immune system. J. Dairy Sci. 54(9):1309-1314.
- Casaro, A. P. E., Kendrick, J. W. and Kennedy, P. C. 1971. Response of the bovine fetus to bovine viral diarrhea-mucosal disease virus. Am. J. Vet. Res. 32(10):1543-1562.
- 20. Chladek, D. W. 1975. Bovine abortion associated with Haemophilus somnus. Am. J. Vet. Res. <u>36</u>: 1041.
- 21. Chow, T. L., Molello, J. A. and Owen, N.V. 1964. Abortion experimentally induced in cattle by infectious bovine rhinotracheitis virus. J.A.V.M.A. 144:1005-1007.
- 22. Cordes, D. O., Dodd, D. C. and O'Hara, P. J. 1964. Bovine mycotic abortion. N. Z. Vet. J. 12:95-100.
- 23. Corner. A. H., Mitchell, D., Meads, E. B. and Taylor, P. A. 1963. Dalmeny disease. An infection of cattle presumed to be caused by an unidentified protozoan. Can. Vet. J. <u>4</u>: 252-264.

- 24. Cox, H. R., van der Scheer, J., Aiston, S. and Bohnel, E. 1947. The purification and concentration of influenza-virus by means of alcohol precipitation. J. Immunol. 56:149-166.
- 25. Crandell, R. A., Sells, D. M. and Gallina, A. M. 1976. The isolation and characterization of a new bovine herpesvirus associated with abortion. Theriogenology 6(1):1-19.
- 26. Crane, C. S. 1965. Infectious bovine rhinotracheitis abortion and its relationship to nutrition in California beef cattle. J.A.V.M.A. 147(12):1308-1309.
- 27. Crane, C. S., Lukas, G. N. and Watkins, W. W. 1964. Infectious bovine rhinotracheitis abortion in California beef cattle. J.A.V.M.A. 144(1):13-18.
- Dellers, R. W. 1977. Diagnostic virology in veterinary medicine: An evaluation, pp. 173-186. <u>In</u> 20th Ann. Proc. Amer. Assoc. Vet. Lab. Diag.
- Dennis, S. M. 1969. Laboratory diagnosis of infectious bovine abortion. J.A.V.M.A. <u>155</u>(12): 1913-1922.
- 30. Dierks, R. E., Malcom, H., Smith, M. H. and Gollehon, D. 1976. Isolation and characterization of adenoviruses from aborted fetuses and calves with weak calf syndrome, pp. 339-404. In 19th Ann. Proc. Amer. Assoc. Vet. Lab. Diag.
- 31. Draayer, H. A. and Kirkbride, C. A. 1977. An electron microscopic investigation of bovine abortion, pp. 47-54. <u>In</u> 20th Ann. Proc. Amer. Assoc. Vet. Lab. Diag.
- 32. van Dreumel, A. A. and Kierstead, M. 1975. Abortion associated with Hemophilus somnus infection in a bovine fetus. Can. Vet. Jour. <u>16</u>(12):367-370.
- 33. Dunne, H. W., Ajinkya, S. M., Bubash, G. R. and Griel, L. C. 1973. Parainfluenza-3 and bovine enteroviruses as possible important causative factors in bovine abortion. Am. J. Vet. Res. 34(9):1121-1126.
- 34. Ellis, W. A., Logan, E. F., O'Brien, J. J., Neill, S. D., Ferguson, H.W. and Hanna, J. 1978. Antibodies to leptospira in the sera of aborted bovine fetuses. Vet. Rec. 103:237-239.

- 35. England, J. J., Frye, C. S. and Enright, E. A. 1976. Negative contrast electron microscopic diagnosis of viruses of neonatal calf diarrhea. Cornell Vet. <u>66</u>:172-182.
- 36. Epstein, B., Turnes, C. G. and Etchverrigaray, M. E. 1972. Aislamiento de feto bovino abortado de virus de la rinotraqueitis bovina infecciosa y *Listeria monocytogenes*. Revista de Medicina Veterinaria 53:99-102.
- 37. Ewalt, D. R. and Harrington, R. 1979. Isolation of Brucella abortus and Brucella abortus, strain 19, from cattle. J.A.V.M.A. 174(2):172-173.
- Faulkner, L.C. (ed.). 1968. Abortion Diseases of Livestock. Charles C. Thomas Publ., Illinois, U.S.A.
- 39. Fayer, R., Johnson, A. J. and Lunde, M. 1976. Abortion and other signs of disease in cows experimentally infected with Sarcocystis fusiformis from dogs. J. Infec. Dis. 134(6):624-628.
- 40. Fennestad, K. L. and Borg-Petersen, C. 1958. Fetal leptospirosis and abortion in cattle. J. Infec. Dis. 102:227-236.
- 41. Fennestad, K. L. and Borg-Petersen, C. 1962. Antibody and plasma cells in bovine fetuses infected with Leptospira saxkoebing. J. Infec. Dis. 110:63-69.
- 42. Freund, S. 1947. Leptospirosis in cattle in Palestine. J. Comp. Path. and Therap. 57:62-66.
- Gibbons, D. F., Winkler, C. E., Shaw, I. G., Terlecki, S., Richardson, C., and Done, J. T. 1974. Pathogenicity of the Border disease agent for the bovine foetus. Br. Vet. J. <u>130</u>(4):357-361.
- 44. Gibbs, E. P. J. and Johnson, R. H. 1970. Differential diagnosis of virus infections of the bovine teat skin by electron microscopy. J. Comp. Path. <u>80</u>:455-463.
- 45. Gibson, E. A. 1965. Salmonella infection in cattle. Journal of Dairy Research <u>32</u>:97-134.
- Gillespie, J. H., Bartholomew, P. T., Thomson, R. G. and McEntee, K. 1967. The isolation of noncytopathic virus diarrhea virus from two aborted bovine fetuses. Cornell Vet. <u>57</u>:564-571.

- 47. Gilman, H. L. 1939. Some causes of abortion in cattle free from Bang's disease. Cornell Vet. 29:153-165.
- Gilman, H. L. and Birch, R. R. 1925. A mould associated with abortion in cattle. Cornell Vet. <u>15</u>:81-89.
- 49. Gilmore, L. O. 1949. The inheritance of functional causes of reproductive inefficiency: A review. J. Dairy Sci. 32:71-91.
- 50. Goodheart, C. R., Armstrong, G. R., Ablashl, D. V., Pearson, G. and Orr, T. W. 1974. Concentration of oncogenic herpesviruses by methyl alcohol precipitation. Appl. Microbiol. 27:988-990.
- 51. Hall, G. A. and Jones, P. W. 1977. A study of the pathogenesis of experimental *Salmonella dublin* abortion in cattle. J. Comp. Path. 87:53-65.
- 52. Hanson, L. E. 1977. Immunology of bacterial diseases, with special reference to leptospirosis. J.A.V.M.A. 170(9):991-994.
- 53. Harcourt, R. A. and Thompson, F. G. A. 1969. Mycotic abortion and mycotic pneumonia in a cow. Vet. Rec.: 199-200. Aug.
- 54. Hartley, W. J. and Marshall, S. C. 1957. Toxoplasmosis as a cause of ovine perinatal mortality. New Zealand Vet. J. 5:119-124.
- 55. Hignett, S. L. 1959. Some nutritional and other inter-acting factors which may influence the fertility of cattle. Vet. Rec. 71:247.
- 56. Hinton, M. 1972. Bovine abortion associated with Corynebacterium pyogenes. Vet. Bull. <u>42(12)</u>: 753-756.
- 57. Hinton, M. 1974. Corynebacterium pyogenes and bovine abortion. J. Hyg. Camb. 72:365-368.
- 58. Hubbert, W. T., Booth, G. D., Bolton, W. D., Dunne, H. W., McEntee, K., Smith, R. E. and Tourtellotte, M. E. 1973. Bovine abortions in five Northeastern States, 1960-1970: Evaluation of diagnostic laboratory data. Cornell Vet. 63:291-316.
- 59. Jensen, R. and Mackey, D. R. 1974. Diseases of Feedlot Cattle, 2nd Ed. Lea and Febiger, Philadelphia, pp. 95-101.

- 60. Johnson, D. W. 1943. Epidemiology of Weil's disease. Brit. Med. J. 2:659.
- 61. Jungherr, E. 1944. Bovine leptospirosis. J.A.V.M.A. 105:276.
- 62. Kahrs, R. F., Scott, F. W. and Hillman, R. B. 1971. An appraisal of fetal serology for the diagnosis of bovine abortion and congenital defects, pp. 588-594. In 75th Proc. Ann. Meet. U. S. Anim. Health Assoc.
- 63. Kendrick, J. W., Osburn, B. I. and Kronlund, N. 1976. Pathogenicity studies on a bovine herpesvirus. Theriogenology 6:447-462.
- 64. Kendrick, J. K. and Straub, O. C. 1967. Infectious bovine rhinotracheitis-infectious pustular vulvovaginitis virus infection in pregnant cows. Am. J. Vet. Res. 28(126):1269-1282.
- 65. Kennedy, P. C. and Richards, W. P. C. 1964. The pathology of abortion caused by the virus of infectious bovine rhinotracheitis. Path. Vet. 1:7-17.
- 66. Kirkbride, C. A., Bicknell, E. J., Knudtson, W. U., and Reed, D. E. 1972. Bovine abortion associated with *Torulopsis glabrata*. J.A.V.M.A. <u>161</u>(4): 390-391.
- 67. Kirkbride, C. A., Bicknell, E. J., Reed, D. E., Robl, M. G., Knudtson, W. U. and Wohlgemuth, K. 1973. A diagnostic survey of bovine abortion and stillbirth in the Northern Plains States. J.A.V.M.A. 162:556-560.
- 68. Kirkbride, C. A., Knudtson, W. U., Bicknell, E. J. and Robl, M. G. 1971. Diagnosis of mycotic abortion in cattle, pp. 580-587. <u>In Proc. 75th</u> Ann. Meet. U.S. Anim. Health Assoc.
- 69. Kirkbride, C. A., Martinovich, D. and Woodhouse, D. A. 1977. Immunoglobulins and lesions in aborted bovine foetuses. New Zealand Vet. J. 25:180-187.
- 70. Kirkbride, C. A. and McAdaragh, J. P. 1978. Infectious agents associated with fetal and early neonatal death and abortion in swine. J.A.V.M.A. 172(4):480-483.
- 71. Knott, S. G. and Dadswell, L. P. 1970. An outbreak of bovine abortions associated with leptospirosis. Aust. Vet. Jour. 46:385-386.

- 72. Knudsen, O. 1956. Some aspects of chromosomes in andrological diagnosis. Proc. IIIrd Int. Congr. Anim. Reprod. Cambridge Section 1:42.
- 73. Knudtson, W. U., Ruth, G. R., Kirkbride, C. A. and Tinant, M. 1976. Pneumonia associated with Torulopsis glabrata in an aborted bovine fetus. Sabouraudia 14:43-45.
- 74. Kradel, D. C. 1978. IV. Abortion storms projection for the future. Cornell Vet. 68 suppl. 7:195-199.
- 75. Kurogi, H., Inaba, Y., Takahashi, E., Sato, K., Omori, T., Miura, Y., Goto, Y., Fujiwara, Y., Hatano, Y., Kodama, K., Fukuyama, S., Sasaki, N. and Matumoto, M. 1976. Epizootic congenital arthrogryposis-hydranencephaly syndrome in cattle: Isolation of Akabane virus from affected fetuses. Arch. Virol. 51(1-2):67-74.
- 76. Lagneau, F. 1964. Considerations d'actualite sur les avortements brucelliques et nonbrucelliques chez la vache. Recl. Med. Vet. 140:1049-1055.
- 77. Langford, E. V. 1969. Pasteurella pseudotuberculosis associated with abortion and pneumonia in the bovine. Can. Vet. Jour. 10(8):208-211.
- 78. Langford, E. V. 1975. Mycoplasma recovered from bovine male and female genitalia and aborted foeti, pp. 221-227. <u>In</u> 18th Ann. Proc. Amer. Assoc. Vet. Lab. Diag.
- 79. Lawson, J. R. 1963. Bovine abortion associated with infectious agents other than *Brucella*. Bull. Off. Int. Epizoot. <u>60</u>:295-311.
- 80. Luedke, A. J., Jochim, M. M., Bowne, J. G. and Jones, R. H. 1970. Observations on latent bluetongue virus infection in cattle. J.A.V.M.A. 156(12):1871-1879.
- 81. Luedke, A. J., Jochim, M. M. and Jones, R. H. 1977. Bluetongue in cattle: Effects of Culicoides variipennis-transmitted bluetongue virus on pregnant heifers and their calves. Am. J. Vet. Res. 38(11):1687-1695.
- 82. Lukas, G. N., Weidenbach, S. J., Palmer, K. G., Dickie, C. W., Duncan, R. F. and Barrera, L. 1964. A bovine fetal virus isolate neutralized by IBR immune serum as a cause of abortion in cattle, pp. 108-128. <u>In Proc. 67th Ann. Meet.</u> U.S. Livestock Sanit. Assoc.

- 83. Maddox, J. G. 1976. Bovine abortion. Vet. Rec.: 517. Dec.
- 84. Marsolais, G., Berthiaume, L., DiFranco, E. and Marois, P. 1971. Rapid diagnosis by electron microscopy of avian coronavirus infection. Can. J. Comp. Med. <u>35</u>:285-288. Oct.
- 85. McFerran, J. B., Clarke, J. K. and Curran, W. L. 1971. The application of negative contrast electron microscopy to routine veterinary virus diagnosis. Res. Vet. Sci. 12:253-257.
- 86. McKercher, D. G., Theis, J. H., Wada, E. M., Loomis, E. C., Bolton, V. and Ito, H. 1976. Recent studies in epizootic bovine abortion. Theriogenology 6(23):251-261.
- 87. McKercher, D. G. and Wada, E. M. 1964. The virus of infectious bovine rhinotracheitis as a cause of abortion in cattle. J.A.V.M.A. <u>144</u>(2):136-142.
- 88. Metselaar, D. and Robin, Y. 1976. Akabane virus isolated in Kenya. Vet. Rec. 99:86. Jul.
- 89. Michin, N. A. and Azinov, S. A. 1935. Spirochaetal jaundice of cattle in North Caucasus. Sovyet Vet. 10:23. In Vet. Bull. 7:419, 1937.
- 90. Miller, N. J. 1955. Infectious necrotic rhinotracheitis of cattle. J.A.V.M.A. 126:463-467.
- 91. Miller, R. B. 1977. A summary of some of the pathogenetic mechanisms involved in bovine abortion. Can. Vet. Jour. 18(4):87-95.
- 92. Miller, R. B. and Quinn, P. J. 1975. Observations on abortions in cattle: A comparison of pathological, microbiological and immunological findings in aborted foetuses and foetuses collected at abattoirs. Can. J. Comp. Med. 39:270-290.
- 93. Mitchell, D. 1960. Bovine abortion an analysis of 227 cases. Can. Vet. J. 1:337-343.
- 94. Mitchell, D. 1974. An outbreak of abortion in a dairy herd following inoculation with an intramuscular infectious bovine rhinotracheitis virus vaccine. Can. Vet. J. 15(5):148-151.
- 95. Moll, T. 1964. Abortion and stillbirth of guinea pigs resulting from experimental exposure to bovine enteric virus. Am. J. Vet. Res. <u>25(109)</u>: 1757-1762.

- 96. Moll, T. and Davis, A. D. 1959. Isolation and characterization of cytopathogenic enteroviruses from cattle with respiratory disease. Am. J. Vet. Res.: 27-32. Jan.
- 97. Moll, T. and Finlayson, A. V. 1957. Isolation of cytopathogenic viral agent from feces of cattle. Science 126:401-402.
- 98. Murphy, J. C. and Jensen, R. 1969. Experimental pathogenesis of leptospiral abortion in cattle. Am. J. Vet. Res. <u>30(5):703-713.</u>
- 99. Mylrea, P. J. 1963. A suspected genetic cause of abortion in cattle. Aust. Vet. Jour. 39:35-36.
- 100. Nuttall, P. A., Luther, P. D. and Stott, E. J. 1977. Viral contamination of bovine foetal serum and cell cultures. Nature 266:835-837. Apr.
- 101. Olafson, P., MacCallum, A. D. and Fox, F. H. 1946. An apparently new transmissible disease of cattle. Cornell Vet. 36(1):205-213.
- 102. Owen, N. V., Chow, T. L. and Molello, J. A. 1964. Bovine fetal lesions experimentally produced by infectious bovine rhinotracheitis virus. Am. J. Vet. Res. 25(109):1618-1625.
- 103. Owen, N. V., Chow, T. L. and Molello, J. A. 1968. Infectious bovine rhinotracheitis: Correlation of fetal and placental lesions with viral isolations. Am. J. Vet. Res. 29(10):1959-1965.
- 104. Plastridge, W. N. and Williams, L. F. 1943. Observations on Vibrio fetus infection in cattle. J.A.V.M.A. 102:89-95.
- 105. Plowright, W. 1968. Malignant catarrhal fever. J.A.V.M.A. 152(6):795-806.
- 106. Plowright, W., Ferris, R. D. and Scott, G. R. 1960. Blue wildebeest and the aetiological agent of bovine malignant catarrhal fever. Nature <u>188</u> (4757):1167-1169.
- 107. Pollard, M., Connolly, J., and Fromm, S. 1949. The precipitating effect of methanol on viruses. Proc. Soc. Exp. Biol. Med. 71:290-293.
- 108. Reed, D. E., Langpap, T. J. and Bergeland, M. E. 1979. Bovine abortion associated with mixed MOVAR 33/63 type herpesvirus and bovine viral diarrhea virus infection. Cornell Vet. <u>6</u>9:54-66.

- 109. Reed, D. E., Pierson, R. E., Kirkbride, C. A., and McAdaragh, J. P. 1971. Investigations of epizootic bovine abortion in Colorado and South Dakota, pp. 574-579. <u>In Proc. 75th Ann. Meet.</u> U.S. Anim. Health Assoc.
- 110. Richardson, A. and Watson, W. A. 1971. A contribution to the epidemiology of *Salmonella dublin* infection in cattle. Br. Vet. J. 127:173-183.
- 111. Ruth, G. R., Kirkbride, C. A. and Langpap, T. J. 1974. Fetal serology as an aid to diagnosis of bovine abortion, pp. 9-18. <u>In</u> 17th Proc. Ann. Meet. Amer. Assoc. Vet. Lab. Diag.
- 112. Sattar, S. A., Bohl, E. H. and Senturk, M. 1965. Viral causes of bovine abortion in Ohio. J.A.V.M.A. 147(11):1207-1210.
- 113. Sattar, S. A., Bohl, E. H. and Trapp, A. L. 1967. Abortion in cattle caused by experimental infection with infectious bovine rhinotracheitis virus. Cornell Vet. 57:438-454.
- 114. Sattar, S. A., Bohl, E. H., Trapp, A. L. and Hamdy, A. H. 1967. In utero infection of bovine fetuses with myxovirus parainfluenza-3. Am. J. Vet. Res. 28(122):45-49.
- 115. Sawyer, M., Osburn, B. I., Knight, H. D. and Kendrick, J. W. 1973. A quantitative serologic assay for diagnosing congenital infections of cattle. Am. J. Vet. Res. 34(10):1281-1284.
- 116. Schiefer, B. 1974. Bovine abortion associated with cytomegalovirus infection. Zbl. Vet. Med. B. <u>21</u>:145-151.
- 117. Schiefer, B. and Moffatt, R. E. 1974. Bovine abortion associated with renal oxalosis in the fetus. Can. Vet. J. <u>15(3):57-65</u>.
- 118. Schultz, R. D. 1973. Developmental aspects of the fetal bovine immune response: A review. Cornell Vet. 63:507-535.
- 119. Schultz, R. D., Dunne, H. W. and Heist, C. E. 1973. Ontogeny of the bovine immune response. Infec. Immun. 7(6):981-991.
- 120. Scott, F. W., Kahrs, R. F. and Parsonson, I. M. 1972. A cytopathogenic strain of bovine viral diarrhea-mucosal disease virus isolated from a bovine fetus. Cornell Vet. <u>6</u>2:74-84.

- 121. Siddique, I. H., Blackwell, J. G. and McKenzie, B. E. 1976. Organisms associated with abortion and reproductive problems in cattle. Modern Vet. Practice:809-811. Oct.
- 122. Smith, T. 1919. The etiological relation of spirilla (Vibrio foetus) to bovine abortion. J. Exptl. Med. 30:313-323.
- 123. Smith, T. 1920. Mycosis of the bovine fetal membranes due to a mould of the genus Mucor. J. Exptl. Med. 31:115-122.
- 124. Smith, T. 1923. Further studies on the etiological significance of Vibrio foetus. J. Exptl. Med. 37:341-346.
- 125. Spradbrow, P. B. and Francis, J. 1969. Electron microscopy as an aid to the rapid identification of animal viruses. Vet. Rec.: 244-246. Mar.
- 126. Stalheim, O. H. V. and Proctor, S. J. 1976. Experimentally induced bovine abortion with Mycoplasma agalactiae subsp. bovis. Am. J. Vet. Res. <u>37</u>: 879-883.
- 127. Stevens, R. W. C. and King, G. J. 1968. Genetic evidence for a lethal mutation in Holstein-Friesian cattle. Journal of Heredity 59:366-368.
- 128. Stoenner, H. G., Crews, F. W., Crouse, A. E., Taschner, L. E., Johnson, C. E. and Wohleb, J. Jr. 1956. The epizootiology of bovine leptospirosis in Washington. J.A.V.M.A. 129:251-259.
- 129. Storz, J. and McKercher, D. G. 1962. Etiological studies on epizootic bovine abortion. Zentralbl. Veterinaermed. 9:411-427 and 520-541.
- 130. Storz, J., Young, S., Carroll, E. J., Bates, R. C., Bowen, R. A. and Keney, D. A. 1978. Parvovirus infection of the bovine fetus: Distribution of infection, antibody response, and age-related susceptibility. Am. J. Vet. Res. 39(7):1099-1102.
- 131. Swift, B. L. and Kennedy, P. C. 1972. Experimentally induced infection of *in utero* bovine fetuses with bovine parainfluenza-3 virus. Am. J. Vet. Res. <u>33</u>(1):57-63.
- Tyzard, I. R. 1977. An Introduction to Veterinary Immunology. W. B. Saunders Company, Pennsylvania, pp. 155-168.

- 133. Verrilli, M. R. 1978. Negative Contrast Electron Microscopy of Veterinary Viruses. M.S. Thesis, Dept. Microbiology and Public Health, Michigan State University, USA.
- 134. Wada, E. M., McKercher, D. G., Castrucci, G. and Theis, J. H. 1976. Preliminary characterization and pathogenicity studies of a virus isolated from ticks (Ornithodoros coriaceus) and from tick-exposed cattle. Am. J. Vet. Res. 37: 1201-1206.
- 135. Weiss, K. E., Haig, D. A. and Alexander, R. A. 1956. Wesselsbron virus - A virus not previously described, associated with abortion in domestic animals. Onderstepoort Jour. Vet. Res. <u>27</u>(2): 183-195.
- 136. Whitford, H. W., Brown, L. N. and Bryner, J. H. 1977. The isolation of Campylobacter fetus ss intestinalis and other vibrios from aborted bovine fetuses, pp. 55-68. <u>In</u> 20th Ann. Proc. Amer. Assoc. Vet. Lab. Diag.
- 137. Wijeratne, W. V. S. and Stewart, D. L. 1971. Population study of abortion in cattle with special reference to genetic factors. Anim. Prod. <u>13</u>: 229-235.
- 138. Williams, B. M., Shreeve, B. J., Hebert, C. N. and Swire, P. W. 1977. Bovine mycotic abortion: Some epidemiological aspects. Vet. Rec. <u>100</u>: 382-385.
- 139. Woelffer, E. A. 1953. Use of progesterone to control habitual abortion in cattle. J.A.V.M.A.: 505-507. Dec.
- 140. Woelffer, E. A. 1972. Diagnosis of bovine abortion. J.A.V.M.A. <u>161</u>(11):1284-1287.
- 141. Wohlgemuth, K. and Knudtson, W. 1973. Bovine abortion associated with Candida tropicalis. J.A.V.M.A. <u>162</u>(6):460-461.
- 142. Wohlgemuth, K. and Knudtson, W. 1977. Abortion associated with Mortierella wolfii in cattle. J.A.V.M.A. <u>171</u>(5):437-439.
- 143. Wohlgemuth, K., Pierce, R. L. and Kirkbride, C. A. 1972. Bovine abortion associated with Aeromonas hydrophila. J.A.V.M.A. 160(7):1001-1002.

ARTICLE

INFECTIOUS AGENTS OF BOVINE ABORTION -LABORATORY DIAGNOSIS

By

V. Moojen, A. W. Roberts and G. R. Carter

SUMMARY

Specimens from 98 aborted fetuses were collected at the Animal Health Diagnostic Laboratory of Michigan State University between June 1977 and September 1978. The fetuses examined were mostly from Holstein-Friesian herds coming from many different areas within the State of Michigan. The primary cause of abortion was recognized in 43 (43.8%) of the 98 aborted fetuses examined. Noninfectious factors were associated with 6.1% of the abortions and infectious agents were incriminated in 37.7%. Bacteria accounted for 22.4%, viruses for 14.3%, and fungi for 15%. Fluids from 50 fetuses were tested and antibodies to leptospira were found in 6 fetuses, and 4 had antibodies to PI-3. Electron microscopy was the most effective method in demonstrating viral agents in fetal tissues. The presence of herpesviruses was detected in 8.2% of the fetuses, correlating in all instances with positive fluorescent antibody (FA) results for infectious bovine rhinotracheitis (IBR) virus. Herpesviruses were demonstrated in 3 additional fetuses that were negative for IBR by FA. Nutritional deficiencies were noted in 3 fetuses and congenital anomalies were found in 2 fetuses.

INTRODUCTION

Bovine abortion is recognized as one of the major economic losses in dairy and beef cattle production, and infectious agents are considered to play a major role. However, a definitive diagnosis is obtained in only 25%

of the total cases submitted to most laboratories (5,21, 36,37,41). This low percentage is likely due to three factors:

1. Most abortions are not due to infectious agents.

- Inappropriate specimens are examined (either incorrect tissue or tissues in poor condition).
- 3. Inappropriate tests are utilized.

The purpose of this study was to increase the effectiveness of the diagnostic laboratory in dealing with infectious agents of bovine abortion. This was accomplished by supplementing conventional tests (routine bacteriologic tests including darkfield examination of stomach contents, routine virologic procedures including fluorescent antibody examination for IBR and BVD, and pathologic examinations) with serologic examination of fetal fluids and electron microscopy. Serologic examination of fetal fluids takes advantage of the relative early immune competence of the fetus, and the fact that immunoglobulins do not normally cross the placental membranes in the cow (6). Examination of fetal fluid previously showed an increase in the number of positive diagnoses of abortion (13,42). Electron microscopy has played an important role in diagnosis of viral infections (16,35,51,54) and should permit the detection of infectious agents not normally associated with abortion as well as allowing the demonstration of known infectious agents no longer viable as a result of autolytic changes of the fetus.

MATERIAL AND METHODS

COLLECTION OF SPECIMENS

Specimens from 98 aborted bovine fetuses were collected at the Animal Health Diagnostic Laboratory of Michigan State University between June 1977 and September 1978. The gestational age of the fetuses ranged from 2 to 8 months.

Necropsy was performed on the fetuses and gross pathologic changes were recorded. Portions of lung, liver, kidney and abomasal contents were collected aseptically for microbiological examination and serosanguineous fluid was collected for serologic tests. Samples were processed immediately or frozen at -20 C for subsequent testing. Portions of selected tissues were fixed in 10% buffered formalin for histopathological examination.

BACTERIOLOGIC EXAMINATION

Lung, liver, kidney and abomasal contents were cultured for bacteria using standard laboratory procedures, and darkfield examination was performed on abomasal contents for the detection of *Leptospira* species and *Campylobacter* species (28).

VIROLOGIC EXAMINATION

PREPARATION OF SPECIMENS

A 10 to 20% tissue suspension of pooled lung, liver and kidney was made by grinding the tissues with a mortar and pestle, and sterile sand, using sterile distilled

water as a diluent. Two milliliters of the tissue suspension was immediately added to an equal volume of 2X concentrated Eagle's minimal essential medium with Earle's base supplemented with 0.5% lactalbumin hydrolysate, sodium pyruvate and non-essential amino acids (EMEM). Also contained in the medium was a high antibiotic concentration (1,000 mg/ml streptomycin, 1,000 units/ml penicillin G, and 500 mg polymyxin B sulfate). The remainder of the sample was used for electron microscopy. Samples were examined immediately or frozen at -20 C.

VIRUS ISOLATION

The tissue suspension was centrifuged at 2,000 rpm for 10 minutes, in a clinical centrifuge, and the supernatant fluid was used as inoculum. Amounts of 0.1 and 0.3 ml were inoculated into each of 2 tubes containing early passages of bovine fetal kidney (BFK) cells or bovine fetal peritoneal (BFP) cells. The maintenance medium was EMEM with 2% bovine fetal serum. The inoculated tubes were incubated at 37 C and observed for cytopathic effect (CPE) daily for 7 days and then frozen. Second and third blind passages were made by following the same procedure as above. In the third passage, coverslips were included in the tubes for hematoxylin and eosin staining (41). Third passage samples (at 8 days post-inoculation) were also inoculated with 0.2 ml of the NADL strain of BVD virus containing 100 TCID₅₀ to test for the presence of noncytopathogenic BVD virus. The cultures were examined daily for CPE for 7 days (18).

ELECTRON MICROSCOPY

Virus Concentration

Samples from the 10 to 20% tissue suspensions in distilled water were used. After these lysates were thawed and frozen twice, dry KCL was added to achieve a final concentration of 1 M (54). This suspension was mixed for 5 to 10 minutes at room temperature, and then centrifuged at 10,000 g for 10 minutes at 0 C. The supernatant fluid was collected and precipitated with methanol, essentially as described by Goodheart et al. (1974).

Briefly, one-half volume of chilled methanol (-65 C) was added slowly to one volume of magnetically stirred tissue suspension in an ice bath (2 C). The temperature of the mixture was not allowed to rise above 3 C. The tissue suspension-methanol mixture was allowed to stir at 2 C in an ice bath for 3 hours, followed by centrifugation at 10,000 g for 20 minutes at 0 C. The pellets obtained were collected in small volumes of phosphate buffered saline (PBS) (0.2 M, pH 7.4) to achieve an approximately final volume concentration of 20 to 30 fold. Elution was allowed to proceed at room temperature for 10 minutes, followed by centrifugation at 10,000 g in an Eppendorf microcentrifuge at 2 C for 30 minutes. The supernatant fluid, if clear, was collected, placed into small glass vials, and stored at -70 C for EM examination. The non-clear supernatants were recentrifuged for an additional 10 minutes.

Electron Microscopic Examination

The methanol precipitated samples as well as unprecipitated samples were negatively stained as described by Verrilli (1978). Two grids were used for each sample: one grid for the methanol and another for the direct preparation. A Philips 300 electron microscope operated at 80KV was used for the examination; grids were scanned initially at instrumental magnification of 20,000X and 60,000X to determine the presence of virus. Fifteen minutes were allowed for each preparation, as suggested by McFerran et al. (1971). A second grid was prepared if the first preparation was unsatisfactory.

FLUORESCENT ANTIBODY TECHNIQUE

The fluorescent antibody (FA) tissue section technique was done essentially as described by Reed et al. (1971) and recommended by Carbrey et al. (1971) for diagnosis of IBR and BVD. Briefly, frozen sections 8 μ thick were made from lung, liver and kidney. The sections were placed on slides, fixed in acetone for 10 to 20 minutes at room temperature, and overlaid with fluorescent conjugates to IBR and BVD viruses. The tissue sections were allowed to stain for 30 minutes in a humid chamber at 37 C followed by 10 minutes wash in 0.85% saline. Slides were then mounted with a coverslip using a drop of equal parts of saline and glycerol.

The slides were observed under a Zeiss fluorescence microscope with a halogen lamp source using a Schott OG5 barrier filter. The presence of IBR or BVD virus was indicated by the presence of bright focal areas of fluorescence or by the characteristic staining of individual cells.

SEROLOGIC TESTS

Serologic tests were done on serosanguineous fluid or pooled fluids extracted from lung, liver and kidney of aborted fetuses. Fluid samples were obtained from 50 fetuses.

Microtiter serum neutralization (SN) tests were performed according to Carbrey et al. (1971) to determine antibodies specific for IBR, BVD (NADL), and BEV types 2, 4, 5, and 6. The screening dilutions were 1:8 and 1:16 (final).

Hemagglutination inhibition (HI) tests were used to detect antibodies to PI-3 and bovine parvovirus (HADEN), at the screening dilutions of 1:8 and 1:16. The technique employed was based on that described by Carbrey et al. (1971), except a microtiter system was used. A complement fixation (CF) test was performed for antibodies to *Chlamydia* according to the technique used for the viruses of the encephalitides as described by Senne and Pearson

(1978). A standard serum agglutination procedure similar to that employed for brucellosis diagnosis was used to detect antibodies to *Campylobacter fetus* (2), and antibodies to the following serogroups of *Leptospira* were determined by the microscopic agglutination test (11): *pomona*, *hardjo*, *grippotyphosa*, *icterohaemorrhagiae* and *canicola*. The standard plate agglutination test was employed to examine for antibodies to *Brucella abortus* (2). After an initial screening test, positive samples were retested at higher dilutions. Negative and positive controls were periodically included in all serologic tests.

HISTOLOGIC EXAMINATION

Specimens were fixed in 10% buffered formalin and sectioned at 6 μ , stained with hematoxylin and eosin, and examined under the microscope. Histologic examinations were not carried out on fetuses that were too decomposed.

RESULTS

The fetuses examined were mostly from Holstein-Friesian herds coming from many different areas within the State of Michigan. Histories indicated that 15.3% of these herds were vaccinated for IBR, 11.2% for BVD, 10.2% for leptospirosis, 6.1% for brucellosis, 5.1% for PI-3, and 3.1% for vibriosis. The majority of abortions occurred between 6 and 7 months of gestation.

Infectious causes of abortion were recognized in 37 (37.7%) of 98 aborted fetuses examined (Table 1). In addition, noninfectious causes were associated with 6 (6.1%) aborted bovine fetuses. Frequently the fetuses had no gross changes with the exception of autolysis. Infectious conditions in the aborted bovine fetuses were detected by the tests indicated in Tables 2 and 4.

BACTERIOLOGY

The bacteriologic results are summarized in Table 2. Darkfield examination of the fetal abomasal contents demonstrated the presence of vibrio and leptospiral organisms in 5 and 3 fetuses, respectively, and C. *pyogenes* was isolated in pure culture or as a predominant organism from 6 fetuses. Bacteria isolated but not generally known to induce abortion are listed in Table 3. Abortion was attributed to these agents only when they were isolated in pure culture or as the predominant organism from abomasal contents and/or other fetal tissues, when inflammatory lesions were observed in the fetal tissues, and in the absence of other known causes of abortion.

Antibodies to leptospira were found in 6 fetuses, of which 4 reacted to *L. hardjo* and 4 to *L. pomona* and, among them, 2 fetuses reacted to both serotypes, with titers ranging from 10 (+) to 40 (++). In one of the reactors, enterovirus-like particles were found in the

actors	Number	Percentage
NFECTIOUS:	<u> </u>	
Herpesvirus IBR virus IBR virus + L. monocytogenes IBR virus + K. pneumoniae Others	3 1 1 3	(8.2)
Togavirus BVD virus	3	(3.1)
Picornavirus Enterovirus	2	(2.0)
Parvovirus	1	(1.0)
Paramyxovirus PI-3 virus	4	(4.1) ^a
C. pyogenes C. pyogenes C. pyogenes + Parvovirus	5 1	(6.1)
Campylobacter species Campylobacter species Campylobacter species + Entero-	4	(5.1)
virus	1	
Leptospira species	9	(7.1) ^b
E. coli	1	(1.0)
Aspergillus species	1	(1.0)
Miscellaneous	3	(3.1)
TOTAL	43	(37.7)

^aPercentage not reflected in the total because of overlap with other significant findings.

^bPercentage calculated on the basis of 7 fetuses; the other 2 were not included because of overlap with other significant findings.

Table 1. Infectious agents isolated or demonstrated in studies of 98 bovine abortions in Michigan,

1977-1978

Table 2. Results of microbiological, pathological and bovine aborted fetuses, Michigan, 1977-1978	obiologica fetuses, M	l, patho] ichigan,	logical and 1977-1978	serolo	.cal ^a exam:	examination of 98
Bacteria & Fungi		DFb	Total DF ^b ISOL ^c	AGG ^d	PREC ^e	PREC ^e PATHOL ^f
Campylobacter species	5	ъ	hDh	I	NA	+
Leptospira species	3	3	ND	Q	NA	+
C. pyogenes	6	NA ^g	6	NA	NA	+
$E. \ coli$	1	NA	1	NA	NA	+
Aspergillus species	1	NA	1	NA	ı	+
Miscellaneous	4	NA	4	NA	NA	+
^a Only 50 fetal sera	were	tested.				
^b Darkfield examinati	tion					
^c Isolation of the microorganism	nicroorgan	ism				
d _{Agglutination test}						
^e Precipitation test						
^f Pathology: lesions	ıs on Table	е Л				
^g Not applicable						
h _{Not} done						

Isolates	Number of Specimens
E. coli	27
Streptococcus species	17
Acinetobacter species	11
Bacillus species	9
Aeromonas species	8
Enterobacter species	8
Proteus species	8
Moraxella species	7
C. pyogenes	6
Klebsiella species	5
Pseudomonas species	3
Staphylococcus species	2
Pasteurella species	1
L. monocytogenes	1
Haemophilus species	1

Table 3.Results of bacteriological examination of 98aborted bovine fetuses in Michigan, 1977-1978

tissues by EM and, in another, lesions of arthrogryposis and cleft palate were observed.

VIROLOGY

The results of virological examinations are summarized in Table 4. Electron microscopy (EM) was the most effective method utilized, demonstrating viruses in 15 of the 85 fetuses. Eight were herpesviruses, 2 were toga-like viruses, 3 were entero-like viruses, and 2 were parvolike viruses. Of the 8 herpesviruses, 5 were presumed to be IBR on the basis of positive FA tests on original tissues. The FA test did not detect any IBR virus that was not demonstrated by EM. One of two fetuses positive for toga-like virus by EM was also positive for BVD by FA, and FA revealed BVD virus in 1 fetus that was negative for toga-like virus by EM. Methanol precipitated material was generally more suitable for EM examination, being somewhat cleaner, but variability in concentration was occasionally noticed.

Virus isolation attempts were least effective. Of 97 fetuses examined, only 1 yielded a positive isolation, viz., a noncytopathic BVD virus. Although the FA test on original tissue was negative for BVD, EM examination revealed the presence of a toga-like virus.

Serological results were negative for all viruses tested except for PI-3 virus, to which there were 4 reactors. These 4 fetuses corresponded to 4 in which other causative agents of abortion were demonstrated.

Virus	Total	ISOL ^a	ЕМ ^b	FA ^C	SN ^d	HIe	PATHOL ^f
Entero	3	-	3	NA	-	NA	+
BVD	3	1	2	2	-	NA	+
IBR	5	-	5	5	-	NA	+
Other Herpes	3	-	3	NA	-	NA	+
Parvo	2	-	2	NA	NA	-	-
PI-3	4	-	-	NA	NA	4	-
No. of fetuses examined		97	85	98	50	50	76

Table 4.	Results	of the	virological	examination	οf
	aborted	bovine	fetuses		

^aIsolation

^bElectron microscopy

^CFluorescent antibody test

^dSerum neutralization test

e_{Hemagglutination-inhibition test}

f_{Pathology:} lesions on Table 5

PATHOLOGY

Some of the lesions found in the different organs examined are tabulated in Table 5 with the corresponding microbiological findings. Four of eight fetuses in which herpesvirus was demonstrated had areas of necrosis in the liver. One of these four fetuses also had similar lesions in the lung, while another had such lesions in the spleen. The remaining 4 fetuses had no significant lesions (NSL) or were too decomposed to be examined.

Hydrocephalus and severe malacia of the cerebrum in areas adjacent to the lateral ventricles were found in the fetus whose tissue preparations yielded a positive isolation of a noncytopathic strain of BVD virus. Two other cases of BVD showed NSL.

Pathologic changes found in 2 fetuses in which EM examination of the tissue preparations revealed presence of enterovirus-like particles consisted of slight inflammatory reactions in the liver, kidney and lung with focal hemorrhages in 1 fetus and the presence of thrombi in the blood vessels of the brain, lung, myocardium, spleen and kidney in the other. Significant lesions were not found in the examination of the fetus whose tissues contained only particles resembling parvovirus.

From the 6 *Corynebacterium*-associated abortions, 1 fetus had small areas of necrosis in the liver, 3 had neutrophilic infiltration in the lung, and 1 of these also had bacterial colonies in the blood vessels and areas

tious	agents o	i aborted	bovine fet	uses
		IBR + L. mono-	-	<i>C</i> .
Histopathology	Herpes	cytogenes	moniae	pyogenes
Areas of necros Lung Liver Spleen Kidney	is: + +	+	+ +	+ + + +
	C. pyogenes	C. pyoge + Parvo	enes E. coli	
Neutrophilic in: Lung Liver Blood vessel:	+ +	n: +	+	+ +
		Campylo	bacter sp.	
Lymphocytic inf: Liver	iltration	:	+	
		C. pyog	renes	
Bacterial colon: Blood vessels		+		
	Enterovi	rus Mi	xed cultur	e of bacteria
Thrombi: Brain Myocardium Lung Spleen Kidney	+ + + +			+
	Mixed cu	lture of h	acteria A	spergillus sp.
Necrotic dermat: Generalized Eyelid	itis:	+		+

Table 5. Histopathological lesions associated with infectious agents of aborted bovine fetuses of necrosis in the spleen and kidney. Collections of lymphocytes in the liver were found in 1 of the 5 fetuses aborted due to Campylobacter species; the other 4 fetuses had NSL. In a case in which L. monocytogenes was isolated from fetal tissue and the FA for IBR was positive, the histopathologic findings consisted of foci of necrosis in the liver. Similar findings in the liver and spleen were found in a fetus which had a positive FA reaction for IBR virus and presence of Klebsiella pneumoniae. Tissues from 3 fetuses yielded the following mixed culture of bacteria and fungi: a) E. coli and P. mirabilis; b) E. coli, Bacillus sp. and Rhizopus sp.; c) Klebsiella sp. and alpha hemolytic Streptococcus sp. The lesions were bile duct hyperplasia and dermatitis of the eyelid, neutrophilic infiltration of the blood vessels, and foci of inflammatory cells in the liver and thrombi in the brain, respectively.

Necrotic dermatitis with presence of fungal hyphae was found in the tissues from the fetus in which Aspergillus sp. was isolated.

Abortions associated with noninfectious causes were identified by pathological findings in 6 fetuses. Histopathologic examination of 2 fetuses demonstrated increased cellularity of the skeletal muscles, and fragmentation and loss of striation of individual muscle fibers. One of these also had signs of interstitial pneumonia. The placenta contained areas of neutrophilic accumulation and 1 to 2 areas with degenerative changes. These conditions

were identified as nutritional muscular dystrophy (NMD). Iodine deficiency goiter was diagnosed on 1 fetus on the basis of hyperplasia of the thyroid follicles which had a columnar epithelium and scant colloid. Histopathologic examination of another aborted fetus revealed oxalate-like crystals in the kidneys, and inflammatory changes in the Developmental anomalies were present in 2 of the lung. fetuses. One had an interventricular septal defect and severe chronic passive congestion of the liver, degenerative changes of the myocardium consisting of atrophy and hyalinization of muscle fibers, and severe and extensive encephalomalacia. No other cause of abortion was found. The anomalies found in the other fetus consisted grossly of cleft palate and arthrogryposis; microscopic examination demonstrated increased connective tissue in the liver. This particular fetus was serologically positive for L. pomona (1:40).

DISCUSSION

In the present study, noninfectious factors were associated with 6.1% of the abortions and infectious agents were incriminated in 37.7%. Of these 37.7%, bacteria accounted for 22.4%, viruses 14.3%, and fungi 1%. The number and types of infectious agents obtained varied considerably when compared with earlier studies (5,36,37,43) and was likely due to variability of specimens, different herd health programs, and epizootiological factors as well as the availability of diagnostic methods.

The number of mycotic abortions diagnosed (1%) agrees with the results of earlier studies (5,13), but is considerably lower when compared to other reports (21,30,36). This low percentage was anticipated as only a small number of placental tissues (4%) were submitted. It has been demonstrated (30, 57, 58) that abortions due to fungi are usually a result of placentitis and that the fetus is often not infected or does not show specific lesions. In view of the fact that placental tissue is usually not submitted, it was thought that the demonstration of antibody to fungal antigen might be useful in reaching a diagnosis of mycotic abortion. However, of 50 fluids examined, none showed evidence of antibody to Aspergillus To determine whether or not our results indicated sp. the ineffectiveness of fetal serology for the diagnosis of fungal abortions will require further study using larger sample size, and the inclusion of additional fungal antigens.

Similarly disappointing results were obtained in the serologic examination of fetal fluids for the detection of antibodies to various viruses (IBR, BVD, PI-3, BEV, Parvo), and to *Chlamydia*, *Campylobacter* and *Brucella*. There were no antibodies detected to any of these with the exception of PI-3 virus, to which there were 4 reactors. These results differed markedly from earlier reports. In 1971, Kahrs et al. reported serological evidence to BVD and IBR in 18.8% and 25%, respectively, of aborted fetuses examined. Dunne et al., in 1973,

examined 100 aborted fetuses and demonstrated that 10% had antibodies to IBR, 3% to BVD, 56% to PI-3 and 41% to BEV. Much lower frequencies were found in work by Ruth et al. in 1974. In analyzing the data of these earlier reports, considerable variation was found in what was considered a positive antibody response (e.g., serum dilutions from 1:2 to greater than 1:40). In our study, all fetal fluids were screened at dilutions of 1:8 and 1:16. This was necessary as most of the fluids were in poor condition and could not be tested at lower dilutions. Thus, the poor condition of the samples may have contributed to our lower success rate.

More rewarding results were obtained when fetal fluids were examined for antibodies to *Leptospira*. Six reactors were found distributed within the serotypes *L. pomona* and *L. hardjo*. None of these reactions correlated with results of darkfield examination. Whether or not this was related to the acuteness of the infection and consequent lack of antibody or to membranous materials that were confused for *Leptospira* is unknown. These results compared favorably with those obtained in a recent report (15).

Electron microscopy was the most effective method in demonstrating viral agents in fetal tissues. The presence of herpesviruses was detected in 8.2% of the fetuses, correlating in all instances with positive FA results for IBR and demonstrating herpesviruses in 3 additional fetuses that were negative for IBR by FA.

Considering the 91.5% reported accuracy of the FA test in the diagnosis of IBR abortion (29), these 3 herpesviruses may represent other bovine herpesviruses (12,40) whose role in abortion has not been elucidated. The inconsistency of methanol precipitation as a concentration technique was thought to be a result of "virus trapping" due to the large amount of tissue proteins that were precipitated. The use of ultrasonic vibration and absorption procedures is presently being attempted to rectify this problem.

Contrary to results obtained by others (29,43), viral isolation attempts were practically useless as a diagnostic aid. Only one isolate (a noncytopathogenic BVD virus) was obtained, and this virus was demonstrated directly in tissue emulsions by EM. The failure to isolate other viruses was attributed to lack of preservation and autolytic changes of fetal tissues examined. Failures in isolation were not due to the lack of susceptibility of the cell cultures employed, as these cells were previously shown to support the growth of viruses such as IBR, BVD, PI-3 and BEV. Experimentally induced viral abortions (7,10,26,38,44,45,52,53) revealed that recovery of virus from the fetus was inversely related to the length of time between infection and collection of specimens, and that fetuses with marked autolysis yielded few isolates.

Autolysis was less a factor in histopathological examination. Consistent with observations of others

(26,27,44), focal areas of necrosis were consistently found in the liver and irregularly in lung and spleen of fetuses that were shown to be infected with herpesviruses. There were no distinguishable differences in the fetuses in which IBR was confirmed and others in which herpesvirus was demonstrated but were negative for IBR virus by FA.

Histopathological changes associated with other viruses were nil or inconsistent. A correlation between BEV and degeneration of arterial walls has been suggested (13), and in the present study 1 of the 3 fetuses in which enterovirus-like particles were demonstrated had lesions consisting of thrombi in small vessels of various tissues. No lesions were observed in the other 2 fetuses or in the fetus with parvovirus-like particles. Hydrocephalus was found in a fetus whose tissues contained togavirus-like particles as seen by EM. A noncytopathic BVD virus was also isolated. Although studies have established BVD virus as a teratogenic agent (8,10,25,55), and hydrocephalus has been associated with this virus in sheep (56), hydrocephalus produced by BVD virus has not been demonstrated experimentally in cattle.

Alveolar infiltration of neutrophils and mononuclear cells, and the presence of bacteria, were observed in the fetus infected with *C. pyogenes*. Hepatic necrosis was evident in the fetus whose tissues yielded *L. monocytogenes*. These lesions are consistent with those described by other investigators (1,17,49,50). However, the fetus infected with *L. monocytogenes* was also infected

with IBR virus, which may have been responsible for the lesions observed. No specific lesions were observed in abortion associated with other bacteria.

Nutritional factors were associated with 3 abortions. Congenital nutritional muscular dystrophy has been reported in association with abortion (23,36), and 2 fetuses which we examined had lesions consistent with this myopathy. Iodine deficiency goiter was identified in an additional fetus, but its association with abortion is unknown.

Although renal oxalosis has not been established as a cause of abortion, it has been observed in aborted bovine fetuses (4,46) and in 1 fetus examined in this study.

The significance of arthrogryposis and cleft palate in 1 fetus was difficult to assess. These congenital anomalies have been associated with genetic (22,32), nutritional (14,34,48), and infectious factors including Akabane virus (31) and BVD virus (20,33). This particular fetus had serological evidence of leptospiral infection and was from a herd with a history of an abortion epidemic. An interventricular septal defect was found in 1 fetus examined. This genetic anomaly has been reported in young calves, but no causal relation to abortion was suggested (3).

In conclusion, the capability of the diagnostic laboratory in dealing with bovine abortion was enhanced by the implementation of additional aids. Our results clearly indicate the value of electron microscopy in

detecting viruses, and it is anticipated that more effective concentration and purification methods will increase the effectiveness of this tool. Such procedures should be relatively inexpensive, simple, and not unduly timeconsuming. Ultracentrifugation does not appear to be the answer, as a considerable amount of extraneous proteins are pelleted, viruses are occasionally distorted, and excessive use of the centrifuge leads to frequent servicing.

Although our results were not as impressive as those of previous studies (13,42), serologic examinations of fetal fluids appears to be a useful adjunct to routine procedures. The value of serologic tests using fetal fluids will probably be more fully realized as additional antigens are screened.

Finally, the effectiveness of the diagnostic laboratory would be increased if placental membranes and fetuses in good condition were obtained. Thus, an effort should be made to communicate this requirement to practicing veterinarians through extension media. To facilitate more satisfactory specimens, diagnostic laboratories should probably provide suitable shipping containers.

LITERATURE CITED

- 1. Addo, P. B. and Dennis, S. M. 1979. Experimental production of *Corynebacterium pyogenes* abortion in sheep. Cornell Vet. 69:20-32.
- Alton, G. G. and Jones, L. M. 1967. Laboratory techniques in brucellosis. World Health Organization, Monograph ser. 55.
- Belling, T. H. 1962. Genetic effect of cardiac ventricular septal defect in Hereford cattle. Vet. Med. <u>57</u>:965.
- Bergeland, M. E., Kirkbride, C. A., Ruth, G. R., Chladek, D. W. and Reed, D. E. 1974. Lesions and microorganisms in aborted bovine fetuses with bovine viral diarrhea antibodies, pp. 321-325. In 17th Proc. Ann. Meet. Vet. Lab. Diag.
- Bolton, W. D., Durrell, W. B., Wadsworth, J. R. and Murray, R. W. 1969. A survey of abortions in Vermont dairy cattle. J.A.V.M.A. <u>155</u>(3):500-503.
- 6. Brambell, F. W. R. 1958. The passive immunity of the young mammal. Biol. Rev. 33:488-531.
- Braun, R. K., Osburn, B. I. and Kendrick, J. W. 1973. Immunologic response of bovine fetus to bovine viral diarrhea virus. Am. J. Vet. Res. 34(9):1127-1132.
- Brown, T. T., DeLaunta, A., Bistner, S. I., Scott, F. W. and McEntee, K. 1974. Pathogenic studies of infection of the bovine fetus with bovine viral diarrhea virus. Vet. Path. 11:486-505.
- 9. Carbrey, E. A. and Stewart, W. C. 1971. Applications of recommended fluorescent antibody techniques for viral diseases in veterinary diagnostic laboratories, pp. 564-573. <u>In Proc. 75th Ann.</u> Meet. U.S. Anim. Health Assoc.
- Casaro, A. P. E., Kendrick, J. W. and Kennedy, P. C. 1971. Response of the bovine fetus to bovine viral diarrhea-mucosal disease virus. Am. J. Vet. Res. 32(10):1543-1562.
- 11. Cole, J. R. 1973. Spirochetes, pp. 157-169. In G. R. Carter (ed.), Diagnostic Procedures in Veterinary Microbiology. Charles C. Thomas Publ., Illinois, USA.

- 12. Crandell, R. A., Sells, D. M. and Gallina, A. M. 1976. The isolation and characterization of a new bovine herpesvirus associated with abortion. Theriogenology <u>6</u>(1):1-19.
- 13. Dunne, H. W., Ajinkya, S. M., Bubash, G. R. and Griel, L. C. 1973. Parainfluenza-3 and bovine enteroviruses as possible important causative factors in bovine abortion. Am. J. Vet. Res. 34(9):1121-1126.
- 14. Dyer, A., Cassaft, J. and Rao, R. R. 1964. Manganese deficiency in the etiology of deformed calves. Bioscience 14:31-32.
- 15. Ellis, W. A., Logan, E. F., O'Brien, J. J., Neill, S. D., Ferguson, H. W. and Hanna, J. 1978. Antibodies to leptospira in the sera of aborted bovine fetuses. Vet. Rec. 103:237-239.
- England, J. J., Frye, C. S. and Enright, E. A. 1976. Negative contrast electron microscopic diagnosis of viruses of neonatal calf diarrhea. Cornell Vet. <u>66</u>:172-182.
- 17. Epstein, B., Turnes, C. G. and Etchverrigarary,
 M. E. 1972. Aislamiento de feto bovino abortado de virus de la rinotraqueitis bovina infecciosa y Listeria monocytogenes. Revista de Medicina Veterinaria 53:99-102.
- 18. Gillespie, J. H., Bartholomew, P. T., Thomson, R. G. and McEntee, K. 1976. The isolation of noncytopathic virus diarrhea virus from two aborted bovine fetuses. Cornell Vet. 57:564-571.
- Goodheart, C. R., Armstrong, G. R., Ablashl, D. V., Pearson, G. and Orr, T. W. 1974. Concentration of oncogenic herpesviruses by methyl alcohol precipitation. Appl. Microbiol. 27:988-990.
- 20. Gratzek, J. 1968. Discussion, pp. 769. In J. H. Gillespie. Comments on bovine viral diarrheamucosal disease. J.A.V.M.A. 152(6):768-770.
- Hubbert, W. T., Booth, G. D., Bolton, W. D., Dunne, H. W., McEntee, K., Smith, R. E. and Tourtellotte, M. E. 1973. Bovine abortions in five Northeastern States, 1960-1970: Evaluation of diagnostic laboratory data. Cornell Vet. 63:291-316.
- 22. Hutt, F. B. 1934. A hereditary lethal muscle contracture in cattle. J. Hered. 25:41-46.

- 23. Jenkins, K. J. and Hidiroglou, M. 1972. A review of selenium/vitamin E responsive problems in livestock: A case for selenium as a feed additive in Canada. Can. J. Anim. Sci. 52(4):591-620.
- 24. Kahrs, R. F., Scott, F. W. and Hillman, R. B. 1971. An appraisal of fetal serology for the diagnosis of bovine abortion and congenital defects, pp. 588-594. <u>In</u> 75th Proc. Ann. Meet. U.S. Anim. Health Assoc.
- 25. Kendrick, J. W. 1971. Bovine viral diarrheamucosal disease virus infection in pregnant cows. Am. J. Vet. Res. 32(4):533-544.
- 26. Kendrick, J. K. and Straub, O. C. 1967. Infectious bovine rhinotracheitis-infectious pustular vulvovaginitis virus infection in pregnant cows. Am. J. Vet. Res. 28(126):1269-1282.
- 27. Kennedy, P. C. and Richards, W. P. C. 1964. The pathology of abortion caused by the virus of infectious bovine rhinotracheitis. Path. Vet. 1:7-17.
- 28. Kirkbride, C. A. 1975. Laboratory diagnosis of bovine abortion (C. A. Kirkbride, ed.), pp. 331 (1-62). In Proc. 17th Ann. Meet. Amer. Assoc. Vet. Lab. Diag., Virginia.
- 29. Kirkbride, C. A., Bicknell, E. J., Reed, D. E., Robl, M. G., Knudtson, W. U. and Wohlgemuth, K. 1973. A diagnostic survey of bovine abortion and stillbirth in the Northern Plains States. J.A.V.M.A. 162:556-560.
- 30. Kirkbride, C. A., Knudtson, W. U., Bicknell, E. J. and Robl, M. G. 1971. Diagnosis of mycotic abortion in cattle, pp. 580-587. <u>In</u> Proc. 75th Ann. Meet. U.S. Anim. Health Assoc.
- 31. Kurogi, H., Inaba, Y., Takahashi, E., Sato, K., Omori, T., Miura, Y., Goto, Y., Fujiwara, Y., Hatano, Y., Kodama, K., Fukuyama, S., Sasaki, N. and Matumoto, M. 1976. Epizootic congenital arthrogryposis-hydranencephaly syndrome in cattle: Isolation of Akabane virus from affected fetuses. Arch. Virol. 51(1-2):67-74.
- 32. Lauvergne, J. J. and Blin, P. C. 1968. Hereditary determinism of the cleft palate associated with ankylosis of limbs in Charolais cattle. 12th Internat. Conf. Genetics:1.

- 33. Leipold, H. W., Cates, W. F., Radostits, O. M. and Howell, W. E. 1970. Arthrogryposis and associated defects in newborn calves. Am. J. Vet. Res. 31(8):1367-1374.
- 34. Leipold, H. W., Husby, F., Brundage, A. L. and Shupe, J. L. 1977. Congenital defects of calves on Kodiak Island. J.A.V.M.A. 170(12):1408-1410.
- 35. McFerran, J. B., Clarke, J. K. and Curran, W. L. 1971. The application of negative contrast electron microscopy to routine veterinary virus diagnosis. Res. Vet. Sci. 12:253-257.
- 36. Miller, R. B. and Quinn, P. J. 1975. Observations on abortions in cattle: A comparison of pathological, microbiological and immunological findings in aborted foetuses and foetuses collected at abattoirs. Can. J. Comp. Med. 39:270-290.
- 37. Mitchell, D. 1960. Bovine abortion An analysis of 227 cases. Can. Vet. J. 1:337-343.
- 38. Owen, M. V., Chow, T. L. and Molello, J. A. 1964. Bovine fetal lesions experimentally produced by infectious bovine rhinotracheitis virus. Am. J. Vet. Res. 25(109):1618-1625.
- 39. Reed, D. E., Bicknell, E. J., Larson, C. A., Knudtson, W. U. and Kirkbride, C. A. 1971. Infectious bovine rhinotracheitis virus-induced abortion: Rapid diagnosis by fluorescent antibody technique. Am. J. Vet. Res. 32(9):1423-1426.
- 40. Reed, D. E., Langpap, T. J. and Bergeland, M. E.
 1979. Bovine abortion associated with mixed MOVAR 33/63 type herpesvirus and bovine viral diarrhea virus infection. Cornell Vet. 69:54-66.
- 41. Rovozzo, G. C. and Burke, C. N. 1973. A Manual of Basic Virological Techniques. Prentice-Hall, Inc., Englewood Cliffs, NJ.
- Ruth, G. R., Kirkbride, C. A. and Langpap, T. J. 1974. Fetal serology as an aid to diagnosis of bovine abortion, pp. 9-18. In Proc. 17th Ann. Meet. Amer. Assoc. Vet. Lab. Diag.
- 43. Sattar, S. A., Bohl, E. H. and Senturk, M. 1965. Viral causes of bovine abortion in Ohio. J.A.V.M.A. 147(11):1207-1210.

- 44. Sattar, S. A., Bohl, E. H. and Trapp, A. L. 1967. Abortion in cattle caused by experimental infection with infectious bovine rhinotracheitis virus. Cornell Vet. 57:438-454.
- 45. Sattar, S. A., Bohl, E. H., Trapp, A. L. and Hamdy, A. H. 1967. In utero infection of bovine fetuses with myxovirus parainfluenza-3. Am. J. Vet. Res. 28(122):45-49.
- Schiefer, B., Pantekoek, J. F. C. A. and Moffatt, R. E. 1974. The pathology of bovine abortion due to Corynebacterium pyogenes. Can. Vet. J. 15(11):322-326.
- 47. Senne, D. A. and Pearson, J. E. 1978. Microtitration complement fixation test for identification of mouse brain isolates of VEE, EEE, and WEE viruses, pp. 39-46. In Serologic Microtitration Techniques. U.S. Dept. Agriculture, National Veterinary Services Laboratories, Ames, Iowa.
- Shupe, J. L., Binns, W., James, L. F. and Keeler, R. F. 1967. Lupine, a cause of crooked calf disease. J.A.V.M.A. 151(2):198-203.
- Smith, H. A., Jones, T. C. and Hunt, R. D. 1972. *Veterinary Pathology*, 4th Ed. Lea and Febiger, USA.
- 50. Smith, R. E., Reynolds, I. M., Clark, G. W. and Milbury, J. A. 1971. Fetoplacental effects of Corynebacterium pyogenes in sheep. Cornell Vet. 61:573-590.
- 51. Spradbrow, P. B. and Francis, J. 1969. Electron microscopy as an aid to the rapid identification of animal viruses. Vet. Rec.:244-246. Mar.
- 52. Storz, J., Young, S., Carroll, E. J., Bates, R. C., Bowen, R. A. and Keney, D. A. 1978. Parvovirus infection of the bovine fetus: Distribution of infection, antibody response, and age-related susceptibility. Am. J. Vet. Res. 39(7):1099-1102.
- 53. Swift, B. L. and Kennedy, P. C. 1972. Experimentally induced infection of *in utero* bovine fetuses with bovine parainfluenza-3 virus. Am. J. Vet. Res. 33(1):57-63.
- 54. Verrilli, M. R. 1978. Negative Contrast Electron Microscopy of Veterinary Viruses. M.S. Thesis, Dept. Microbiology and Public Health, Michigan State University, USA.

- 55. Ward, G. M. 1971. Bovine viral diarrhea-mucosal disease implicated in a calf with cerebellar hypoplasia and ocular disease. A case report. Cornell Vet. <u>61</u>:224-228.
- 56. Ward, G. M. 1971. Experimental infection of pregnant sheep with bovine viral diarrhea-mucosal disease virus. Cornell Vet. 61:179-191.
- 57. Williams, B. M., Shreeve, B. J., Hebert, C. N., and Swire, P. W. 1977. Bovine mycotic abortion: Some epidemiological aspects. Vet. Rec. <u>100</u>: 382-385.
- 58. Wohlgemuth, K. and Knudtson, W. 1973. Bovine abortion associated with Candida tropicalis. J.A.V.M.A. 162(6):460-461.